

Effect of Processing Unripe Plantain (*Musa paradisiaca*) Extracts on Some Biochemical Parameters in Alloxan-Induced Wistar Rats

ABSTRACT

Unripe plantain is used for management of diabetes mellitus in Nigeria, the possible effect of its methods of processing on some biochemical parameters has not been investigated. The objective of this study is to determine the effect of methods of processing unripe plantain on blood glucose, Total Cholesterol, Triglyceride, HDL, LDL, serum Alanine aminotransferase (ALT), Aspartate aminotransferase (AST), Alanine Phosphatase (ALP), total protein, albumin, creatinine, uric acid and urea levels in alloxan-induced diabetic rat models. Twenty male albino Wistar rats were used and were divided into 5 groups of 4 rats each. Group 1 (normal control) received standard rat feeds; group 2 (diabetic control) received standard rat feeds; groups 3,4 and 5 received boiled, roasted and dried unripe plantain pellets respectively. In the result, a significant ($P<0.05$) reduced blood glucose levels were seen for those experimental animals given dried, boiled and roasted extracts respectively. Triacylglycerol levels were significantly decreased ($p<0.05$) for those diabetic rats administered dried, boiled and roasted extracts respectively. The Total cholesterol levels were significantly decreased ($p<0.05$) for those administered dried, roasted and boiled extracts respectively. The LDL-cholesterol levels were also significantly decreased ($p<0.05$) for those treated with dried, roasted and boiled extracts respectively. The HDL-cholesterol were significantly increased for the diabetic administered dried extract, roasted extract and a significant decrease for those administered boiled extract. There were significant increase ($p<0.05$) in the levels of serum ALT, AST and ALP in the diabetic control group compared to the normal control are observed. On administration of unripe plantain, the values of these enzymes significantly decreased ($p<0.05$). The boiled extract showed a greater decrease in the level of ALT and ALP whereas the dried extract showed a greater decrease in the level of AST. There were also increase in the level of urea and uric acid in the diabetic control compared to the normal control group is observed. However the dried extract showed a greater decrease in the levels of both urea and uric acid. The creatinine level is seen to increase in the diabetic control group when compared to the normal control group and showed greater increase on administration of unripe plantain, for the group fed with roasted extract having the highest level of creatinine. **Keywords:** Diabetes Mellitus, Unripe Plantain, Alloxan

INTRODUCTION

One of the most challenging diseases of the 21st century is Diabetes. Diabetes mellitus is a metabolic disease characterized by hyperglycemia, resulting from partial or total destruction of pancreatic beta cells (Shepherd, 2005). It affects the essential biochemical pathways of the body (carbohydrate, protein and lipid metabolism) and its prevalence is rising globally, which includes the rural Nigeria

39 populations (Ime *et al.*, 2011; Karau *et al.*, 2012). Due to the inability of the modern therapy to
40 control all the pathophysiological aspects of the disorder, as well as the enormous cost it poses on
41 the economy of the developing nations of the World, alternative strategies are urgently needed
42 (WHO, 2002).

43 The global concern for the diversification of the uses of plant foods to improve normal and
44 therapeutic nutrition for diabetes control has shifted scientists' interest to enhancing the potential
45 sources of beneficial constituents in plant foods. Plant foods have generated increasing research
46 interest because of their anti-diabetic potentials. The diets/medicinal plants that are commonly used
47 in the management of diabetes in Nigeria include: acha (*Digitaria exiles*), breadfruit (*Treculia*
48 *Africana*) and beans (*Phaseolus vulgaris*) (WHO, 2003). However, diabetic patients have often
49 complained of the monotony of staying on a particular diet (personal communication) and this has
50 therefore increased the research into other plants.

51 Plantain (*Musa paradisiaca*) belongs to the *musical* family and it is cultivated in many tropical and
52 sub-tropical countries of the world. Plantain is a source of starchy staple for millions of people in
53 Nigeria. It contains low quantities of minerals and sugars; this can be seen in an unripe plantain.
54 Scientifically, unripe plantain has being documented as a hypoglycemic plant (Nelson *et al.*, 2006). In
55 folklore medicine, unripe plantain is used in the management of diabetes, renal and liver dysfunction
56 (Iweala *et al.*, 2001). Although, unripe plantain is used to manage diabetes mellitus in Nigeria, the
57 possible effect of its methods of processing on some biochemical parameters, including renal and
58 hepatic dysfunction has not been investigated. This study was aimed at examining the effect of
59 methods of processing unripe plantain on some biochemical parameters in alloxan-induced diabetic
60 rats.

61 **MATERIALS AND METHODS**

62 **Preparation of Plant Materials**

63 Matured freshly cut unripe green plantain (*Musa paradisiacal*) where purchased from a local market
64 in Jos Plateau state, Nigeria. The bunch of *Musa paradisiacal* was rinsed with water to remove latex
65 and dirt. These were divided into three portions and prepared differently. To the first portion, it was
66 boiled in boiling water for 30 minutes, allowed to cool before being peeled and cut into pieces, air-
67 dried at room temperature for 6-7 days. The dried pieces were pulverized using a milling machine to
68 obtain a fine powder. The second portion was roasted using a charcoal oven until it is browned. It
69 was peeled, cut into piece pulverized using a milling machine to obtained a fine powder. The last
70 portion was air-dried at room temperature for 6-7 days. The dried plant food was milled into
71 powder.

72 **Preparation of extracts**

73 Each of the processed plantain powder was used for the extraction process. 100g of each powdered
74 plant material was weighed using a weighing balance and transferred into different beakers. Each
75 was dissolved with 1L of distilled water and allowed to stay for twenty-four hour for maximum
76 extraction of the active ingredient(s). The dissolved plant was filtered and the filtrate was kept in an
77 oven at 60°C. This was done to ensure that it undergo evaporation until it becomes dried. The dried
78 extracts were separately transferred into airtight containers and stored in the refrigerator.

79 **Experimental Designs**

80 The experimental animals used were twenty (20) healthy Wistar rats (weighing 185-200kg) obtained
81 from the animal house of University of Jos, Nigeria. They were allowed to acclimatize for 2 weeks,
82 after which they were maintained under a constant 12 hours light and dark cycle and at room
83 temperature.

84

85 **Induction of Experimental Animals**

86 The animals were induced with freshly prepared saline solution of Alloxan and injected into the
87 animals intraperitoneally. After 48 hours of induction of Alloxan, the animals were tested to confirm
88 if they were diabetic.

89

90 **Grouping and Administration of Extract**

91 The plant (plantain extract) from the differently processed method were administered orally to the
92 animals with 150ml/kg body weight measurement. This was given to the experimental animals
93 (albino rats) that were divided into 5 groups of 4 each as follows:

94 Group 1: normal rats administered standard feed pellets (Normal control)

95 Group 2: diabetic control rats administered standard feed pellets (Diabetic control)

96 Group 3: diabetic rats administered boiled plantain extract

97 Group 4: diabetic rats administered roasted plantain extract

98 Group 5: diabetic rats administered dried plantain extract.

99 The extract was administered for the period of 8 days following an interval of 48 hours fasting
100 period. The administration was stopped at the eighth day, the rats were anesthetized with
101 chloroform and their blood samples collected.

102 Ethical issues were observed in line with the regulations of animal usage and approval was obtained
103 as required in the University of Jos ethical committee guide.

104

105 **METHODS**

106 **Determination of Glucose**

107 Glucose reacts with O'toluidine in a glacial acetic acid with heat of field N-glucosyl amine which is
108 blue green in color. The absorbance is measured at 025nm (Bishop *et al.*, 2000).

109 **Determination of Cholesterol Level**

110 The Total Cholesterol was determined by Liebermann Burchard's method (Edward and Morris,
111 1969).

112 **Determination of Creatinine**

113 Creatinine reacts with picric acid to produce a coloured compound creatinine adenine picrate which
114 was photometrically measured. The intensity of the colour is a function of the creatinine in the
115 blood. The total serum proteins, albumin, uric acid and urea were determined using Biosystems
116 Diagnostic kits as described by (Tietz, 1995; Friedman and Young, 2001).

117 **Determination of Serum Urea**

118 The ammonia reacts with phenol in the presence of hypochlorite to form indophenols which give a
119 blue compound in alkaline solution (Bolleter *et al.*, 1961).

120 **Haematological Analysis**

121 The haematological analysis carried out were Packed Cell Volume, haemoglobin, platelet count,
122 white blood cell count, red blood cell count, using standard procedures (Saeed *et al.*, 2011).

123

124 **STATISTICAL ANALYSIS**

125 Data were subjected to analysis using the Statistical Package for Social Sciences (SPSS), version 15.0.
 126 Results were presented as the means standard deviations of triplicate experiments. One way
 127 analysis of variance (ANOVA) was used for comparison of the methods. Differences between
 128 methods were considered to be significant at $P < 0.05$ using the Duncan Multiple Range Test.

129 **RESULTS**

130 The administration of alloxan at a dosage of 65mg/kg body weight to the rats produced a stable
 131 diabetic condition within few days in most of the experimental rats. Administration of unripe
 132 plantain resulted in 510%, 55% and 113% decrease in blood glucose compared to the diabetic
 133 control in the groups administered boiled, roasted and dried extracts respectively (table 1).

134 **Table 1: Result of Effect of Plantain Extracts on Blood Glucose Level (mMol/L) in Diabetic Rat**
 135 **Models**

Treatment Groups		Boiled Extract	Roasted Extract	Dried Extract
A	Normal Control	5.56±0.01	5.56±0.01	5.56±0.01
B	Diabetic Control	19.84±0.02 ^a	19.84±0.02 ^a	19.84±0.02 ^a
C	Diabetic Treated	3.25±0.02 ^{ab}	12.82±0.02 ^{ab}	9.32±0.02 ^{ab}

136 Values are expressed as mean ± SD, n = 4 for each group.
 137 values are significantly different from healthy control group ($P < 0.05$)
 138 ^b values are substantially different from diabetic control group ($P < 0.05$)
 139

140 **Table 2: Result of Effect of Plantain Extracts on Lipid Profile Concentrations in Diabetic Rat Models**

Treatment Groups		TG	TC	LDL	HDL
A	Normal control	1.37±0.02	3.68±0.01	2.26±0.36	1.54±0.02
B	Diabetic control	2.53±0.18 ^a	5.84±0.01 ^a	3.24±0.01 ^a	0.47±0.01 ^a
C	Diabetic + boiled extract	1.73±0.01 ^{ab}	4.43±0.01 ^{ab}	2.61±0.01 ^{ab}	0.08±0.01 ^{ab}
D	Diabetic + roasted extract	1.89±0.01 ^{ab}	4.82±0.01 ^{ab}	2.93±0.02 ^{ab}	0.74±0.02 ^{ab}
E	Diabetic + dried extract	1.53±0.01 ^{ab}	4.02±0.01 ^{ab}	2.24±0.02 ^{ab}	0.94±0.01 ^{ab}

141 Values are expressed as mean ± SD, n = 4 for each group.
 142 values are significantly different from healthy control group ($P < 0.05$)

143 ^b values are substantially different from diabetic control group (P<0.05)

144

145 **Table 3: Result of Effect of Plantain Extracts on Enzymes of Rat Models**

146

Treatment Groups		ALT(mmol/L)	AST (mmol/L)	ALP (mmol/L)
A	Normal Control	11.31±0.01	16.84±0.02	236.62±0.02
B	Diabetic Control	48.18±2.46 ^a	79.87±0.01 ^a	642.24±0.01 ^a
C	Diabetic + boiled extract	31.46±0.01 ^{ab}	60.16 ±0.09 ^{ab}	86.00±0.70 ^{ab}
D	Diabetic + roasted extract	39.85±0.03 ^{ab}	64.41 ±0.01 ^{ab}	431.00±0.70 ^{ab}
E	Diabetic + dried extract	36.36±0.02 ^{ab}	49.42 ±0.01 ^{ab}	411.00± 0.01 ^{ab}

147 Values are expressed as mean ± SD, n= 4 for each group.

148 values are significantly different from normal control group (P<0.05)

149 ^b values are significantly different from diabetic control group (P<0.05)

150

151 **Table 4: Result of Effect of Plantain Extracts on Creatinine, Urea and Uric Levels in the**

152 **Experimental Rat Models**

Treatment Groups		Creatinine (mmol/L)	Urea (mmol/L)	Uric acid (mmol/L)
A	Normal Control	91.54±0.01	7.02 ± 0.01	226.08±0.01
B	Diabetic Control	173.97±1.01 ^a	22.82 ± 0.01 ^a	564.24±0.02 ^a
C	Diabetic + Boiled Extract	182.54 ±0.02 ^{ab}	10.42±0.01 ^{ab}	396.04± 0.01 ^{ab}
D	Diabetic + Roasted Extract	194.64 ±0.02 ^{ab}	12.82±0.02 ^{ab}	401.64± 0.01 ^{ab}
E	Diabetic + Dried Extract	177.54±0.01 ^{ab}	9.32±0.02 ^{ab}	371.84± 0.01 ^{ab}

153 Values are expressed as mean ± SD, n= 4 for each group.

154 values are significantly different from normal control group (P<0.05)

155 ^b values are significantly different from diabetic control group (P<0.05)

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161 **Table 5: Result of Effect of Plantain Extracts on the Total Protein and Albumin Concentrations in**
 162 **Rat Models**

Treatment Groups		Total protein g/L	Albumin g/L
A	Normal Control	75.60±0.01	35.62±0.02
B	Diabetic Control	68.42±0.02 ^a	28.47±0.02 ^a
C	Diabetic + Boiled Extract	72.08±0.01 ^{ab}	32.34±0.04 ^{ab}
D	Diabetic + Roasted Extract	70.38±0.01 ^{ab}	30.30±0.01 ^{ab}
E	Diabetic + Dried Extract	74.04±0.01 ^{ab}	33.18±0.01 ^{ab}

163 Values are expressed as mean ± SD, n= 4 for each group.

164 values are significantly different from healthy control group (P<0.05)

165 ^b values are substantially different from diabetic control group (P<0.05)

166

167 **Table 6: Result of Effect of Plantain Extracts on some Hematological Parameters in Diabetic Rat**
 168 **Models**

Treatment Groups	PCV	HB	WBC	PLT	RBC
Normal Control	32.00±0.81	7.62±0.01	5260±1.63	242000±0.81	5360±201.33
Diabetic Control	30.00±0.81 ^a	3.24±0.01 ^a	9500.00±0.8 ^a	318000.00±163 ^a	9525.00±49.33 ^{ab}
Diabetic + Boiled Extract	35.00±0.81 ^{ab}	8.32±0.01 ^{ab}	7400.00±0.81 ^{ab}	26400±0.81 ^{ab}	4898.00±0.81 ^{ab}
Diabetic + Roasted Extract	33.00±2.94 ^{ab}	4.51±0.01 ^{ab}	7450.00±0.00 ^{ab}	26600±3.26 ^{ab}	7500±1.41 ^{ab}
Diabetic + Dried Extract	35.00±2.89 ^{ab}	5.31±0.01 ^{ab}	6600±0.00 ^{ab}	255000±0.81 ^{ab}	6600.00±0.81 ^{ab}

169 Values are expressed as mean ± SD, n= 4 for each group.

170 values are significantly different from healthy control group (P<0.05)

171 ^b values are substantially different from diabetic control group (P<0.05)

172

173 The serum Alanine aminotransferase, Aspartate aminotransferase and Alanine phosphatase enzyme
 174 levels of the diabetic control were significantly increased (p<0.05) compared to the non-diabetic
 175 control group. Administration of unripe plantain extracts decreased considerably (p<0.05) the levels

176 of these enzymes, especially in the group fed boiled extract (31.46mmol/L) followed by the group
177 fed dried extract (36.36mmol/L), and then the group fed roasted extract (39.85mmol/L) for ALT. For
178 AST, the level of decrease is more in the group supplied dried extract (49.42mmol/L) as against the
179 boiled (60.16mmol/L) and roasted (63.41mmol/L) plantain extracts respectively. There is a similar
180 decrease in ALP in the trend 86mmol/L for boiled, 411mmol/L for dried and 431mmol/L for roasted
181 plantain extract respectively (table 3).

182 There is a significant increase in creatinine level in the diabetic control group compared to the non-
183 diabetic control group. On administration of unripe plantain extracts, there were significant
184 increases ($p<0.05$) especially in the group administered roasted extract (194.64mmol/L). Also, there
185 is substantial increase in the level of urea in the diabetic control group compared to the non-diabetic
186 control group. However, on the administration of unripe plantain, a significant decrease ($p<0.05$)
187 was observed as follows: 9.32mmol/L, 10.42mmol/L and 12.82mmol/L for groups fed dried, boiled
188 and roasted extracts respectively. A significant increase is observed in the level of uric acid in the
189 diabetic control group (564.24 mmol/L) compared to the non-diabetic group (226.08 mmol/L). After
190 treatment with unripe plantain extracts, uric acid level decreased significantly to 396.04 mmol/L,
191 401.64mmol/L and 371.84mmol/L for the groups treated with boiled, roasted and dried extracts
192 respectively.

193 A significant decrease was observed in the level of total protein in the diabetic control group
194 (68.42g/L) compared with the non-diabetic control group (75.60g/L). After administration of unripe
195 plantain extracts, there was noticeable significant increase ($p<0.05$) especially in the group
196 administered dried extract (74.04g/L) followed by boiled extract (72.08g/L) and then the group
197 conducted roasted extract (70.38g/L). Similarly, there is a significant decrease of Albumin level in the
198 diabetic control (28.47g/L) compared with the non-diabetic control (35.62g/L). Administration of
199 unripe plantain significantly increase ($p<0.05$) this value to 32.34g/L, 30.30g/L and 33.18g/L for
200 groups treated with boiled, roasted and dried plantain extracts respectively.

201

202 **DISCUSSION**

203 Different factors can affect or influence blood glucose level. These include the physical form of the
204 food, degree and type of processing example, cooking method and time, amount of heat and
205 moisture used (Pi-sunyer, 2002), and also the type of starch (amylose versus amylopectin). Findings
206 from this study indicate that the boiled plantain extract has the highest hypoglycemic effect while
207 the roasted extract has the least; thus confirming the ability of unripe plantain to ameliorate
208 hyperglycemia. This further explains that the result of moist heating improves the strength of the
209 food substance to enhance the effect of hyperglycemia than drying without direct heating (drying)
210 and dry heating (roasting). Boiling of the plantain allowed the starch granules to swell, gelatinise and
211 increase the availability of amylase digestion and thereby growing starch availability (Bahado *et al.*,
212 2006).

213 Serum total cholesterol, LDL cholesterol and Triglyceride levels of the diabetic control rats are
214 significantly ($p<0.05$) higher than that of the non-diabetic control rats with the decreased level of
215 HDL-Cholesterol (table 2). This is an indication that diabetes mellitus is associated with elevated
216 levels of Total cholesterol, LDL-Cholesterol and Triglyceride with reduced level of HDL-Cholesterol.

217 The result is shown in the table also indicates that the diabetic animals when fed with unripe
218 plantain have significant ($p < 0.05$) decrease in the levels of cholesterol compared with the controls.
219 This is an indication that plantain, however, reduces cholesterol (Gould *et al.*, 1998). Increase in
220 cholesterol is a risk factor associated with arteriosclerosis and cardiovascular diseases (Cooper *et al.*,
221 2007). The experimental group fed with dried plantain extract had the most pronounced effect in
222 lowering serum cholesterol level while the roasted plantain extract had the least. Therefore, dried
223 plantain extract has the highest ability to enhance the impact of arteriosclerosis and cardiovascular
224 disease than the boiled and the roasted extracts. From the same table, triglyceride and Low-density
225 lipid (LDL) concentrations are significantly decreased ($p < 0.05$) in all the diabetic treated groups, with
226 the group treated with dry extract being the most decreased. High levels of LDL-Cholesterol and
227 triglyceride have been associated with heart disease (Nikkila, 1984). In the medical term, high
228 cholesterol and triglyceride levels are referred to as lipid disorder, which increases the risk of
229 atherosclerosis and also heart disease, stroke and high blood pressure (Cooper *et al.*, 2007). The
230 consumption of unripe plantain has been shown to reduce triglyceride level (Kaimal *et al.*, 2010).

231 Measurement of enzymatic activities of aminotransferases (AST and ALT) and phosphatases is of
232 clinical and toxicological importance as changes in their actions are indicative of tissue damage by
233 toxicants or in disease conditions (Tietz, 1995; Radhika *et al.*, 2012; Adesokan *et al.*, 2009).
234 Aminotransferases such as ALT and AST and ALP are common liver enzymes whose activities are a
235 sensitive indicator of liver cell injury and are helpful in recognising hepatocellular diseases such as
236 diabetes. In Harris *et al.*, (2005) studies, it was shown that individuals with type 2 diabetes mellitus
237 (T2DM) have a higher incidence of liver function abnormalities than individuals who do not suffer
238 from diabetes mellitus. This study indicates an increase in the level of the diagnostic enzymes (AST,
239 ALT and ALP) in the serum of alloxan diabetic rat models which is attributable to the toxicity of
240 alloxan to the tissue that expresses GLUT2 transporters such as hepatocytes and renal tubular cells
241 (Eleazu *et al.*, 2013). The effect of alloxan on the levels of these diagnostic enzymes (AST, ALT and
242 ALP) in the serum of alloxan diabetic rat models has remained unravelled. While some authors
243 reported increased activities of AST, ALT (Rajesh, 2012) and ALP (Umesh *et al.*, 2004) in the liver of
244 alloxan diabetic rat models, some others reported no alteration in the levels of these enzymes in the
245 serum of the diabetic rats. The increase observed in the level of these enzymes in the serum of the
246 diabetic control rat models could be as a result of leakage of these enzymes from the liver cytosol
247 into the bloodstream (Navarro *et al.*, 1993) which indicates the hepatotoxicity of alloxan. However,
248 Treatment of the diabetic animals with unripe plantain was able to decrease significantly ($p < 0.05$)
249 the levels of these enzymes in the serum of these rat models indicating the ability of unripe plantain
250 to repair liver damage. The boiled plantain extract has the highest capacity to decrease the levels of
251 AST and ALP, while the dry extract has the highest ability to reduce the activity of ALT.

252 It is the function of the kidney to remove urea from the blood. In kidney impairment, the urea level
253 builds up in the blood because the kidneys are unable to clear the area from the bloodstream
254 (Okechukwu *et al.*, 2013). In this study, the serum urea levels of the diabetic control group increased
255 significantly ($p < 0.05$) compared with the non-diabetic control. This could be as a result of kidney
256 impairment in diabetic control rats. However, treatment of the diabetic animals with unripe plantain
257 resulted to the significant decrease ($p < 0.05$) of urea in the serum of the diabetic rats (table 4). This
258 shows that plantain may possess protective effects on the kidney. The dried unripe plantain has the
259 most useful ability to enhance the impact of urea on the organs of the diabetic animals while the
260 roasted plantain extract has the least effect.

261 Creatinine, a metabolite of creatine is generated from muscle and excreted by the kidney. However,
262 in kidney impairment, creatinine is poorly cleared and therefore builds up in the blood (Okechukwu
263 *et al.*, 2013). The outcome of this study significantly highlighted that consumption of unripe plantain
264 (be it boiled, roasted or dried) as seen in table 4, can induce an elevation of creatinine level, which
265 by implication, suggests that unripe plantain has a higher propensity to cause renal failure due to
266 increase in creatinine level. The diabetic rats treated with roasted plantain extract have the highest
267 level of creatinine, while those treated with the dry extract has the least.

268 The uric acid level of the diabetic control group is higher when compared to the non-diabetic group.
269 This could be as a result of renal failure, resulting to reduced clearance of uric acid by the kidney,
270 since it is the function of the organ to clear out uric acid from the blood. High levels of uric acid can
271 lead to a kidney stone or cause solid crystals to form within joints. This creates a painful condition
272 called gout. If gout remains untreated, these uric acid crystals can build up in the joint or nearby
273 tissues, forming hardy lumpy deposit called tophi. On administration of unripe plantain, the levels of
274 uric acid significantly decrease ($p < 0.05$). This could be as a result of the renal protective effect of
275 unripe plantain. Precisely, the dried plantain extract has the highest uric acid-reducing ability while
276 the roasted extract has the least.

277 The decrease in the serum protein of the diabetic control rats is an indication of proteinuria which is
278 an important clinical marker of diabetics nephropathy, and this decrease can be attributed to
279 increasing protein catabolism while the increase in the serum protein level of the diabetic rats fed
280 unripe plantain extracts is an indication of the protective action of unripe plantain against
281 nephrotoxicity (Jefferson *et al.*, 1983) and also unripe plantain is an excellent source of protein (Foy
282 and Parratt, 1960). The study shows that the dried plantain extract has the highest ability to enhance
283 the effect of nephropathy in the diabetic rats.

284 The low serum albumin level of the diabetic control rats could be attributed to their low serum
285 protein levels suggesting the impaired renal function for the rats of this group, or it may also
286 recommend an impaired liver function for this group. The elevation of the serum albumin levels of
287 the diabetic rats fed unripe plantain suggests that plantain can be used in the management of
288 diabetic renal dysfunction. However, the dried plantain extract suggests better management of renal
289 dysfunction compared to the boiled and roasted extracts.

290 Packed cell volume (PCV), white blood cells (WBC), Platelet, Red Blood Cell (RBC) and HB are of
291 diagnostic importance. A decrease in PCV generally means red blood cell loss from cell destruction,
292 blood loss or failure of bone marrow production. Consumption of plantain could protect the red
293 blood cells due to its content of blood-forming nutrients such as iron. This explains why the PCV of
294 the diabetic control and non-diabetic control (table 6) is significantly different from the diabetic
295 treated. Distinctly, the boiled and dried extracts have higher PCV values.

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298

299 **CONCLUSION**

300 The knowledge of an active processing method for dietary staples to control and enhance the effect
301 of complications of diabetes is essential in the treatment of diabetes. The findings in this study are
302 useful to health care providers and nutritionist in diabetes education. This is because diet
303 management is crucial to control spikes in blood glucose levels. The research indicates that unripe
304 plantain can be used in the management of complications arising from diabetes mellitus. The boiled
305 plantain extract had the highest hypoglycemic effect of all the processed test extracts and can be
306 used to decrease the impact of hyperglycemia. It can be used to improve the aberrations of the
307 enzymes activities of diabetes mellitus, whereas, the dried plantain extract has the highest TG and
308 LDL-cholesterol lowering effect and can effectively be used to manage the heart-damaging effect of
309 diabetes mellitus. The use of dried plantain in the dietary management of diabetes mellitus could be
310 a breakthrough in search of plants that could prevent the development of diabetic nephropathy.

311 REFERENCES

- 312 1. Shepherd J. 2005. Does statin monotherapy address the multiple lipid abnormalities in type
313 2 diabetes? *Atherosclerosis supplements*; 6:15-19.
- 314 2. Ime, F. A., Atangwho I. J., Regina, I., Ejemot-Nwadiaro, I., Edisua, H. I., Essien U.
315 Hypoglycaemic effect and proximate composition of some selected nigerian
316 traditional diets used in management of diabetes mellitus. *Eur J Food Res*; 2011;
317 1(2): 94-101.
- 318 3. Karau, G.M., Njagi, E.M., Machocho, A.K., Wangai L.N. Phytonutrients: Mineral
319 composition and in vitro antioxidant activity of leaf and stem bark powders of
320 *Pappea capensis* (L.). *Pak J Nutrit*; 2012; 11(2): 123-132.
- 321 4. World Health Organization. 2002. WHO launches the first global strategy on traditional
322 medicine: Press release WHO/38. Geneva.
- 323 5. World Health Organization. 2003. Diet, nutrition and the prevention of chronic diseases.
324 Report of a joint WHO/FAO Expert Consultation. Geneva, Switzerland.
- 325 6. Nelson, S.C.; Ploetz, R.C., Kepler, A.K. 2006. *Musa* species (bananas and plantains) in
326 Elevitch, C.R, *Species Profiles for Pacific Island Agroforestry*, Hōlualoa, Hawai'i:
327 Permanent Agriculture Resources (PAR), [retrieved 10 January 2013].
- 328 7. Iweala E.J, Obichi I.C, Omotosh I.E . Biochemical and histological responses of hepatotoxic
329 rats fed *M. paradisiacal* supplemented diet. *International journal of pharmacology*; 2001;
330 7:471-477.
- 331 8. Bishop, M. L., *et al.*, 2000. *Clinical Chemistry: Principles, Procedures, Correlations* (4thed.).
332 Philadelphia, PA: Lippincott Williams & Wilkins.
- 333 9. Edward Kim and Morris Goldberg, 1969. Serum Cholesterol Assay Using a Stable
334 Liebermann-Burchard Reagent. *Clinical Chemistry*. Vol. IS, No. 12.
- 335 10. Tietz, N.W. 1995. *Clinical guide to laboratory test*, 3rd edition. WB Saunders Company,
336 Philadelphia, PA, pp. 518-519.
- 337 11. Friedman L and Young D. L. 2001. *Effects of disease on clinical laboratory tests*, AACC Press,
338 4th ed. 1:133.
- 339 12. Bolleter W. T, Bushman C. J and Tidwell P. W. Spectrophotometric Determination of
340 Ammonia as Indophenol. *Anal. Chem*, 1961, 33 (4), pp 592-594.
- 341 13. Saeed Nazifi, Ahmad Oryan and Fatemeh Namazi. Hematological and Serum Biochemical
342 Analysis in Experimental Caprine Besnoitiosis. *Korean J Parasitol*, 2011, 49 (2): 133-138.
- 343 14. Pi-sunyer FX. Glycemic index and diseases. *Am.J.Clin.Nutr*, 2002, 76:290S - 298S.

- 344 15. Bahado - Sigh, PS, Wheatley M. H, Ahmad EY, Morrison A, and Asemota HN, Br. J. Nutr, 2006,
345 96: 476 – 481.
- 346 16. Gould,A.L., J.E. Rossouw, N.C. Santanello, J.F. Heyse and C.D. Furberg. Cholesterol reduction
347 yields clinical benefit: impact of statin trials. Circulation, 1998, 97:946-952.
- 348 17. Cooper, A., L. Nherera, N. Calvert and N. O’Flynn and N. Turnbull. (2007). Clinical guidelines
349 and evidence review for lipid modification: cardiovascular risk assessement and the primary
350 and secondary prevention of cardiovascular disease. National Collaborating Centre for
351 Primary Care and Royale College of General Practioners, London.
352 <http://www.nice.org.uk/nicemedia/pdf/CG67fullguideline.pdf>. [7 May 2016]
- 353 18. Nikkila, E.A. Plasma lipid and lipoprotein abnormalities in diabetes. In: diabetes and heart
354 diseases, jarret, R.F., (Ed.) Elsevier science publishers, Amsterdam, the Netherlands, 1984,
355 pp: 134-167.
- 356 19. Kaimal, S., K.S. Sujatha and S. George. Hypolipiddaemic and antioxidant effect of fruits of
357 musa AAA (Chenkadali) in alloxan induced diabetic rats. Ind. J Exp. Boil, 2010, 48: 165-173.
- 358 20. Radhika R., Ragavan B., Sharad P.D., Sudarsanam D. Action of marker enzymes of Rheum
359 emodi in alloxan induced diabetic rats. Asian j Exp Biol Sci, 2012, 3(2), 420-423.
- 360 21. Adesokan, A. A., Oyewole O. I., Turay, B. M. Kidney and liver function parameters in alloxan
361 induced diabetic rats treated with *aloe barbadensis* juice extract. Sierra leone j Biomed Res,
362 2009, 1(1), 33-37.
- 363 22. Harris EH. Elevated liver function tests in Type 2 diabetes. Clin Diabetes 2005; 23:3
- 364 23. Eleazu, C. O., Eleazu K. C., Chukwuma S. C., Udeme N. Review of the Mechanism of cell
365 death resulting from streptozotocin challenge in experimental animals, its practical use and
366 potential risks to humans. J diabetes Metab Disorder, 2013, 12, 60.
- 367 24. Rajesh M. Protective effect of *carthamus tinctorius* on streptozotocin induced diabetic
368 complications in rats and possible morphological changes in the liver and kidney. Int J Sci
369 Innov Discov, 2012, 2(5), 502-510.
- 370 25. Umesh, C., Yadav, S., Moorthy, K., Najma, Z.B. Effects of sodium orthovanadate and
371 *Trigonella foenum-graecum* seeds on renal and hepatic lipogenic enzymes and lipid profiles
372 during alloxan diabetes. J Bio Sci, 2004, 29(1), 81-91.
- 373 26. Navarro, C.M., Montilla, P.M., Martin, A., Jimenez, J., Utrilla, P.M. Free radical scavengers
374 and anti- hepatotoxic activity of *Rosmarinus*. Plant Med, 1993, 59, 312-314.
- 375 27. Okechukwu, P.N., Ndyebura, A.W., Chiang, C.N., Akowuah, G.A. The effect of standardized
376 extracts of cosinnium fanestratum stem bark on liver and kidney function parameters in
377 streptozocin-induced diabetic rats. Journal of acute disease; 2013, 2:201-206.
- 378 28. Jefferson L, Liao W, Peavy D, Miller, Appel M, Taylor J. Diabetes-alterations in liver protein
379 synthesis: changes in the relative abundance of nRNA for albumin and other plasma
380 proteins. J Biol Chem; 1983, 258:1369-1375.
- 381 29. Foy, J.M and J.R. Parratt. A note on the presence of nonadrenaline and 5-hydroxytryptamine
382 in plantain (*musa sapientum var. paradisiaca*). J. pharm. Pharmacol., 1960, 12: 360-364.