

2 **Antibacterial Potentiality and Brine Shrimp Lethality Bioassay of**
3 **the Methanol Extract of *Trema orientalis* Leaves**

4 **Abstract**

5 **Aim:** The aim of this study was to evaluate the antibacterial and cytotoxic activities of
6 methanol extract of *Trema orientalis* leaves.

7

8 **Materials and Methods:** Antibacterial activity of *Trema orientalis* leaves was tested against
9 two Gram-positive and seven Gram-negative bacteria by disc diffusion assay. The liquid
10 microdilution assay was used for the determination of the minimum inhibitory concentration
11 (MIC). The brine shrimp lethality bioassay analyzed the cytotoxic activity of methanol
12 extract of *Trema orientalis* leaves.

13

14 **Results:** The methanol extract exhibited potent antibacterial activity with the zone of
15 inhibition ranging from 9 ± 0.81 to 14 ± 0.81 mm against both the tested Gram-positive and all
16 tested Gram-negative bacteria except *Pseudomonas denitrificans* and *Xanthomonas*
17 *campestris*. Comparatively, higher antibacterial activity was found against Gram-negative
18 bacteria in case of *Shigella dysenteriae* and *Salmonella typhi* which showed 14 ± 0.81 mm and
19 13 ± 0.81 mm zones of inhibition respectively. *Salmonella typhi* showed resistance against
20 reference antibiotics (Tetracycline, Erythromycin, Gentamicin and Ciprofloxacin) but
21 methanol extract of leaves exhibited potent antibacterial activity against *Salmonella typhi*.
22 The MIC values for tested Gram-positive bacteria was 10 mg/mL while for Gram-negative
23 bacteria were ranged from 1.25 to 20 mg/mL. The methanol extract of *Trema orientalis*

24 leaves showed very low cytotoxicity (LC50, 170.215 $\mu\text{g/mL}$) in comparison with the standard
25 vincristine sulphate having LC50 value 2.477 $\mu\text{g/mL}$.

26

27 **Conclusion:** The results suggest that the methanol extract of *Trema orientalis* leaves has
28 potent antibacterial activity with minimum cytotoxicity and could lead to the development of
29 novel broad-spectrum antibacterial agent.

30

31 **Keywords:** *Trema orientalis*, Antibacterial activity, Cytotoxicity, Disc diffusion, Minimum
32 inhibitory concentration and Brine shrimp lethality bioassay.

33

34 **1. Introduction**

35 Pathogenic microorganisms, such as bacteria, viruses, parasites and fungi cause infectious
36 diseases, which are considered a significant threat to human health because of the
37 unavailability of vaccines, limited chemotherapy and emergence of resistant bacteria against
38 antibiotics [1, 2]. One half of all death in tropical countries is caused due to infectious
39 diseases and responsible for the second leading cause of death worldwide [3]. Most of the
40 current antibiotics have considerable limitations concerning antimicrobial spectrum and side
41 effects on the host including allergic reactions, immune-suppression and hypersensitivity [4,
42 5]. Moreover, their indiscriminate and inappropriate overuse has led to increasing clinical
43 resistance of previously sensitive microorganisms and to the occurrence of different
44 infections [1, 6]. New and re-emerging infectious diseases are rising very rapidly. Due to
45 these problems, attention is now being given to biologically active compounds isolated from
46 plant species because they offer a new source of antimicrobial drugs and are widely perceived
47 as natural and safe, that is, not toxic [5, 7]. Moreover, plant-based medicines contain the
48 diverse chemical structure and novel mechanism of action that work in the way of orchestral

49 ensembles which can target many elements of the complex cell signalling pathways [8]. It is
50 well known that the bioactive plant extracts are promising sources of a majority of drugs. For
51 example, plant-based antibiotics such as Quinine (Cinchona) and berberine (Berberis) are
52 highly effective against *Staphylococcus aureus* and *Escherichia coli* [5]. Therefore, there is
53 an urgent need to search for new and more potent anti-bacterial and bioactive agents that can
54 fight against an infectious pathogen.

55

56 *Trema orientalis* is a medicinal shrub or tree belonging to the family Ulmaceae. Locally, it is
57 known as a charcoal tree or gunpowder tree. It is named for Chikan or Jibon in Bengali,
58 Nalita in English and Gio in Hindi. It is a fast-growing and evergreen tree and distributed all
59 over the world including Bangladesh. The young leaves are eaten like spinach, and in
60 combination with lemon juice, the leaves maceration are used for the treatment of bronchitis,
61 cough, pneumonia and pleurisy. The infusion is prepared from fruits and flowers of *Trema*
62 *orientalis* for administration to children as a therapy for pneumonia, pleurisy and bronchitis
63 [9]. The aerial parts, flowers, bark, and seeds of *Trema orientalis* exhibit various
64 pharmacological activities including antidiarrheal, antidiabetic, antiplasmodial, antimalarial
65 and antioxidant activities [10-14]. These pharmacological effects may be mainly because it
66 contains essential biologically active compounds such as scopoletin, 3, 4-hydroxybenzoic
67 acid, epicatechin, lupeol, methylswertianin, catechin, hexacosanoic acid, tannins, saponins,
68 flavonoids, triterpenoid, phytosterols and xanthones [9]. Although there are many pieces of
69 literature reporting the ethnomedicinal values of *Trema orientalis*, there is little scientific
70 proof for further using this plant commercially or in a more useful form. Therefore, an
71 attempt was made to evaluate the antibacterial and cytotoxic activities of the crude methanol
72 extract of

73 *Trema orientalis* leaves to support the pharmacological effects and phytochemical
74 investigation of the plant.

75

76 **2. Materials and Methods**

77 **2.1. Chemicals and Reagents**

78 All standard antibiotic discs used in this study were purchased from Bio-Rad, USA. Nutrient
79 agar media and Nutrient broth media were obtained from Liofilchem, Italy. Methanol from
80 Merck, Germany and the eggs of Brine shrimp were collected from an aquarium shop,
81 Dhaka, Bangladesh.

82

83 **2.2. Plant Material**

84 *Trema orientalis* leaves were collected during January 2015 from Jessore, Bangladesh and
85 were authenticated by a botanist. A voucher (DACB 31285) has been deposited in
86 Bangladesh National Herbarium, Mirpur, Bangladesh for further reference. The collected
87 plant leaves were washed with running tap water and dried in the shade at room temperature.
88 The air-dried leaves were pulverised into fine powder by commercial blender (Philips, South
89 Korea) and stored in sealed container.

90

91 **2.3. Experimental Methods**

92 **2.3.1. Preparation of the Plant Extract**

93 100 g of powder was taken in a 500 mL conical flask added with 350 mL of methanol. The
94 bottle was kept for seven days with continuous shaking at shaking incubator at room
95 temperature. The plant extract was filtered through Whatman no.1 filter paper (Thermo
96 Fisher Scientific, USA.) and then concentrated by using a rotary evaporator (Stuart, UK) and
97 kept at room temperature to evaporate the remaining solvent. After complete evaporation of

98 water, only plant's crude extracts were obtained. The number of oil extracts was 1.0 g which
99 was stored in a refrigerator at 4°C in the sterile container for further use.

100

101 **2.3.2. Tested Bacterial Preparation**

102 Pure culture of Gram-positive bacteria (*Bacillus subtilis* IFO 3026, *Sarcina lutea* IFO 3232)
103 and Gram-negative bacteria (*Escherichia coli* IFO 3007, *Proteus vulgaris* MTTC 321,
104 *Klebsiella pneumonia* ATTC 10031, *Xanthomonas campestris* IAM 1671, *Pseudomonas*
105 *denitrificans* KACC 32026) were used in this study and obtained from the Microbiology
106 Laboratory of Department of Biotechnology and Genetic Engineering, Islamic University,
107 Kushtia, Bangladesh. Another two Gram-negative bacteria, *Salmonella typhi* and *Shigella*
108 *dysenteriae* were kindly provided by the Microbiology laboratory of Department of
109 Microbiology, Jessore University of Science & Technology. Bacteria were cultured in nutrient
110 agar media and nutrient broth media. For antibacterial assay, minimum inhibitory
111 concentration (MIC) determination and the further stock culture preparation, 100 µL of
112 frozen stock culture was inoculated into 125 mL conical flask containing 25 mL of Nutrient
113 broth media and incubated at 37°C with continuous shaking at 250 rpm for culturing the
114 bacteria until mid-log phase of absorbance at 600 nm reached at 0.4 by using UV
115 spectrophotometer (Oasis scientific Inc., USA) for bacterial broth culture.

116

117 **2.3.3. Disc Preparation**

118 The Whatman No. 1 filter paper discs (6 mm diameter) were transferred to a small vial and
119 autoclaved at 15 lb/inch² pressure for 15 minutes at 121°C. The discs were wholly dried in
120 drying oven at 60°C. 400 mg of crude methanol extract of *Trema orientalis* leaves was
121 dissolved into ten mL of methanol, and each disc was impregnated with ten µL of 40 mg/mL
122 (400 µg/disc) of *Trema orientalis* leaves extract. The discs were utterly air dried in the

123 laminar flow cabinet and used for the antibacterial assay. Blank CDS (negative controls)
124 impregnated with ten μL of methanol.

125

126 **2.3.4. Antibacterial Activity Assay**

127 Antibacterial activity of crude methanol extract was tested by the disc diffusion method [15].

128 The prepared discs were placed on the nutrient-agar-medium plate spread with 100 μL of

129 tested bacterial broth culture, and the plates were incubated at 37°C for 24 h. Standard

130 reference antibiotics Tetracycline (30 $\mu\text{g}/\text{disc}$), Erythromycin (15 $\mu\text{g}/\text{disc}$), Gentamicin (10

131 $\mu\text{g}/\text{disc}$) and Ciprofloxacin (5 $\mu\text{g}/\text{disc}$) were used as positive control to ensure the activity of

132 standard antibiotic against the test organisms. The blank discs were used as negative control.

133 After incubation, the culture plates were examined and the inhibition zones formed around

134 each disc were measured in millimeter scale. Each assay in this experiment was replicated

135 three times.

136

137 **2.3.5. Determination of Minimum Inhibitory Concentration (MIC)**

138 Minimum inhibitory concentration (MIC) of methanol extract of *Trema orientalis* leaves was

139 determined by a two-fold serial dilution method as previously described [16]. The methanol

140 crude extract of *Trema orientalis* leaves was dissolved in nutrient broth medium in an

141 Eppendorf tube (Watson Co. Ltd., Japan) to achieve a concentration of 40 mg/mL. The

142 solution of Eppendorf tube was serially diluted to obtain 20, 10, 5, 2.5 and 1.25 mg/mL of

143 concentrations. The 0.5 mL of bacterial broth culture of each tested bacteria was transferred

144 to each Eppendorf tube. Thus, the total amount of solution in each Eppendorf tube was one

145 mL. The control tubes contain 0.5 mL bacterial broth cultures with 0.5 mL nutrient broth

146 media. The resolution of all Eppendorf tubes was appropriately mixed by vortexing and

147 incubated at 37°C for 24 h with continuous shaking at 250 rpm. After producing 24 h, 100 μL

148 of solution from each Eppendorf tube was spread over the nutrient-agar-media plate. The
149 plates were incubated at 37°C for 16 h for bacterial growth, and the number of a colonies was
150 counted for MIC determination.

151

152 **2.3.6. Brine Shrimp Lethality Bioassay**

153 Brine shrimp lethality bioassay is the most convenient system for preliminary assessment of
154 cytotoxicity of plant extracts. The brine shrimp lethality bioassay of the methanol extract of
155 *Trema orientalis* leaves was evaluated as previously described procedure against *Artemia*
156 *salina* as a test organism to monitor the cytotoxicity of a compound [17]. The eggs of Brine
157 shrimp (*Artemia salina*) were collected from an aquarium shop (Dhaka, Bangladesh) and
158 incubated for 28°C with constant oxygen supply and hatched for two days to provide a large
159 number of larvae called nauplii. The different concentrations of crude extract were prepared
160 by dissolving them in DMSO (not more than 50 µL in 5 mL solution) plus seawater (3.8%
161 NaCl in water) to attain serial dilution from 200-1.562 µg/mL. The standard vincristine
162 sulphate was used as a positive control. The varying concentration of the solution of
163 vincristine sulphate was prepared by serial dilution into DMSO to attain serial dilution from
164 200-1.562 µg/mL. A vial containing 50 µL of DMSO diluted to 5 mL simulated seawater
165 used as a control. Twenty mature shrimps were placed into each of the experimental bottles.
166 After 24 h, the bottles were inspected using a magnifying glass, and the number of surviving
167 nauplii in each jar was counted. From this data, the percentage (%) of mortality of the brine
168 shrimp nauplii was calculated for each concentration using the following formula: %
169 Mortality = $N_t/N_0 \times 100$ (Where N_t = Number of dead nauplii after 24 h incubation; N_0 =
170 Number of total nauplii transferred, i.e., 10). The LC50 (median lethal concentration) was
171 determined from the log concentration versus % mortality.

172

173 2.3.7. Statistical Analysis

174 The experimental results obtained from antibacterial and MIC determination assays were
175 expressed as the mean \pm standard deviation (SD) of three replicates. Correlation/regression
176 analysis determined LC50 values. Microsoft Excel 2010 statistical package was used for all
177 reviews.

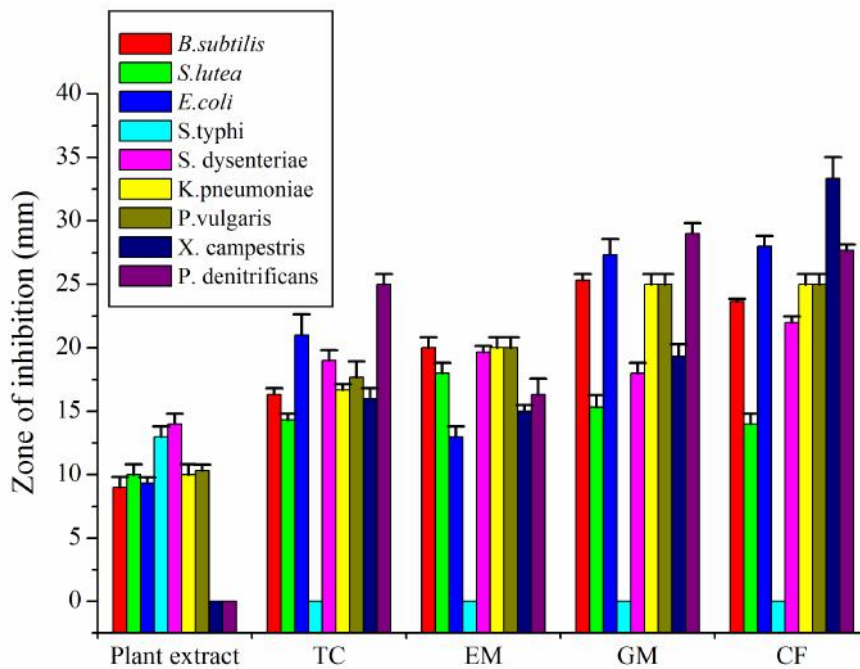
178

179 3. Results

180 3.1. Antibacterial Potentialities of *Trema orientalis* Leaves Extract

181 The antibacterial activities of methanol extract of *Trema orientalis* leaves against the tested
182 bacteria were examined by the occurrence of clear zone of inhibition. The leaves extracted at
183 a concentration of 400 μ g/disc showed significant antibacterial effects against two Gram-
184 positive bacteria (*Bacillus subtilis*, *Sarcina lutea*) and five Gram-negative bacteria
185 (*Escherichia coli*, *Salmonella typhi*, *Shigella dysenteriae*, *Klebsiella pneumonia*, and *Proteus*
186 *vulgaris*) with the zone of inhibition ranging from 9 ± 0.81 to 14 ± 0.81 mm (Figure 1). The
187 highest area of inhibition was found 14 ± 0.81 mm and 13 ± 0.81
188 mm against *Shigella dysenteriae* and *Salmonella typhi* respectively. The inhibition zone was
189 observed 10 ± 0.81 mm against *Klebsiella pneumonia*, *Proteus vulgaris* and *Sarcina lutea*
190 whereas zone of inhibition was 9 ± 0.47 mm against *Bacillus subtilis* and *Escherichia coli*.
191 However, no antibacterial activity was observed against two Gram-negative bacteria,
192 *Xanthomonas campestris* and *Pseudomonas denitrificans* at the used concentration of
193 400 μ g/disc of plant leaves extract. Standard reference antibiotics: Tetracycline,
194 Erythromycin, Gentamicin and Ciprofloxacin were used as positive control showed higher
195 antibacterial activities than the plant goes extract against all the tested bacteria except
196 *Salmonella typhi*. Though *Salmonella typhi* showed resistance against reference antibiotics as
197 a positive control, the methanol extract of *Trema orientalis* leaves exhibited potent inhibition

198 of zone (13 ± 0.81 mm) against *Salmonella typhi* (Figure 1) suggest that it could be a potential
 199 therapeutic drug candidate against *Salmonella typhi*. No region was formed by negative
 200 control.



201

202 **Figure 1.** Effect of methanol extract of *Trema orientalis* leaves on two Gram-positive and
 203 seven Gram-negative bacteria. Values are represented as mean \pm SD (n=3). SD, Standard
 204 deviation; TC, Tetracycline; EM, Erythromycin; GM, Gentamycin; CF, Ciprofloxacin.

205

206 **3.2. Minimum Inhibitory Concentration**

207 The lowest concentration of methanol extract which prevents the visible growth of bacterium
 208 is the minimum inhibitory concentration. The MIC values of crude extract of *Trema*
 209 *orientalis* leaves were found ranging from 1.25 to 20 mg/mL (Table 1). The best MIC was
 210 1.25 mg/mL against *Escherichia coli*, *Salmonella typhi* and *Shigella dysenteriae* as this
 211 concentration completely inhibited the growth of these bacteria. The least efficacy was shown
 212 against *Proteus vulgaris* and *Klebsiella pneumonia* which was inhibited at 20 mg/mL

213 concentration. The reasonable MIC value was demonstrated against Gram-positive bacteria
 214 (*Bacillus subtilis* and *Sarcina lutea*) which were inhibited at 10 mg/mL concentration.

215

216 **Table 1:** Minimum inhibitory concentration of methanol extract of *Trema orientalis* leaves.

Tested bacteria	Minimum Inhibitory Concentration (mg/mL)						
	20	10	5	2.5	1.25	0.625	0.312
	Number of bacterial colonies survived at above concentration						
<i>Bacillus subtilis</i>	0	0	7±.816	17± 1.63	54±3.74	96±3.26	121±3.74
<i>Sarcina lutea</i>	0	0	12±2.16	43±4.32	66±4.39	107±4.08	144±6.53
<i>Escherichia coli</i>	0	0	0	0	0	47±3.74	133±3.74
<i>Salmonella typhi</i>	0	0	0	0	0	77±3.26	124±4.32
<i>Shigella dysenteriae</i>	0	0	0	0	0	55±4.08	112±4.08
<i>Klebsiella pneumoniae</i>	0	5±0.81	16±2.16	52±2.82	79±4.08	123±3.26	223±2.94
<i>Proteus vulgaris</i>	0	23±2.94	54±3.74	77±2.44	91±2.44	155±4.08	175±4.54

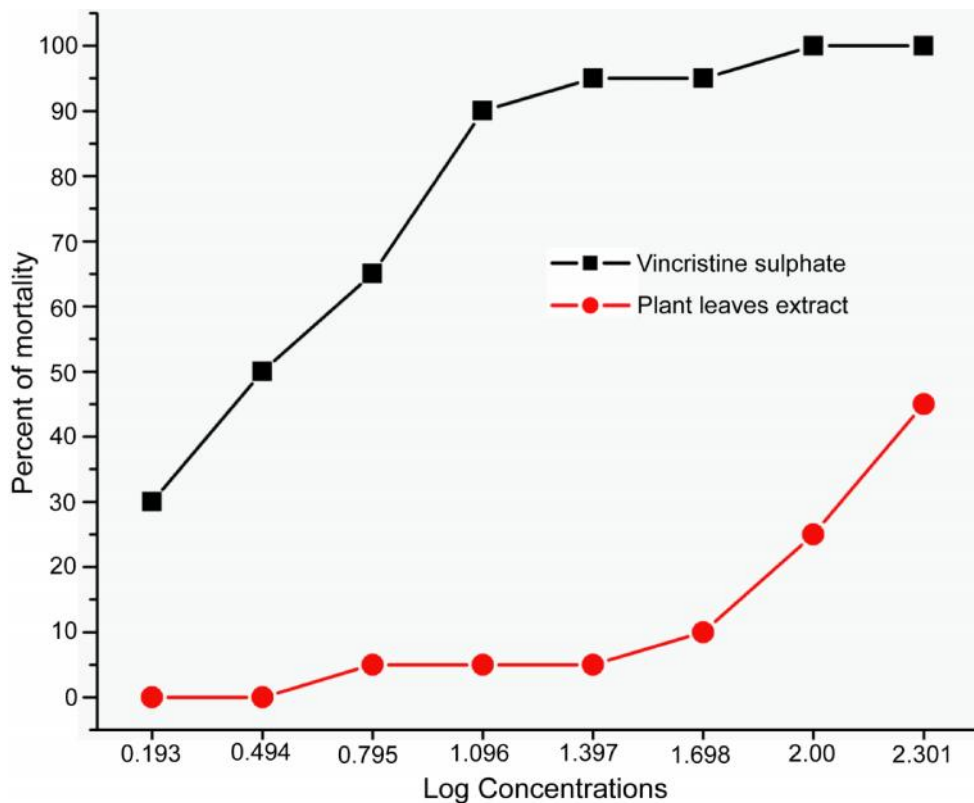
217 Values are represented as mean ± SD (n=3). SD, Standard deviation.

218

219 **3.3. Cytotoxic Activity of Methanol Extract of *Trema orientalis* leaves**

220 The percent mortality of brine shrimp nauplii at different concentrations of plant extracts and
 221 vincristine sulfate as a positive control are shown in Figure 2. It is clear that percent mortality
 222 of brine shrimp nauplii is proportional to the concentration of extracts. The mortality rate was
 223 increased with the extract concentration increased. As shown in Table 2, the methanol extract
 224 of *Trema orientalis* leaves demonstrated an LC50 value of 170.215 µg/mL whereas

225 vincristine sulphate showed the LC50 value of 2.477 $\mu\text{g}/\text{mL}$. This indicates that the plant
226 goes extract has much higher LC50 value compared to that of vincristine sulphate. The crude
227 methanol extracts resulted in LC50 values greater than 100 $\mu\text{g}/\text{mL}$ were considered non-toxic
228 in the brine shrimp lethality assay [18], support the notion that the methanol extract of *Trema*
229 *orientalis* leaves is non-toxic for host and had the potential for further investigation. There
230 was no mortality in the negative control groups indicating the test as a valid one and the
231 results obtained are only due to the activity of the tested agents.
232



233
234 **Figure 2.** Brine shrimp lethality for methanol extract of *Trema orientalis* leaves and
235 vincristine sulphate from the linear correlation between log concentrations versus %
236 mortality.

237
238

239 **Table 2.** The cytotoxicity of methanol extracts of *Trema orientalis* leaves and vincristine
240 sulphate on brine shrimp nauplii.

Sample	LC ₅₀ (µg/mL)	Regression equation	R ²
Plant extract	170.215	y=29.851x + (-16.592)	0.785
Vincristine sulphate	2.477	y=33.004x+36.976	0.838

241

242

243 **4. Discussion**

244 Infectious diseases are the second leading cause of death worldwide. Recently, the
245 emergences of antibiotic-resistant infections are rising very rapidly, which are the major
246 threat to human health as well as an economic burden on country's healthcare system,
247 patients and families. The effectiveness of many conventional antibiotics is being endangered
248 by the rapid emergence of microbial resistance to current therapeutic agents because of their
249 overuse, misuse, and a lack of new drug development by the pharmaceutical industry [19-21].
250 Plant-derived natural secondary metabolites represent a potential source of antimicrobial
251 agents which have the different mode of action than a conventional drug. Acceptance of
252 medicines from natural plant product as an alternative form of a healthcare system is
253 increasing because they are serving as promising sources of the novel antibiotic prototype
254 that could be of clinical importance to improve health care [22, 23]. Therefore, we
255 investigated the *Trema orientalis* leaves for its antibacterial and cytotoxic activities. The
256 present study showed that the methanol extract of *Trema orientalis* leaves at a concentration
257 of 400 µg/disc has potent antibacterial activity against both the Gram-positive (*Bacillus*
258 *subtilis* and *Sarcina lutea*) and Gram-negative bacteria (*Escherichia coli*, *Salmonella typhi*,
259 *Shigella dysenteriae*, *Klebsiella pneumoniae* and *Proteus vulgaris*). The activity exhibited by

260 the methanol extract of *Trema orientalis* leaves may be attributed to the presence of some
261 bioactive compounds in *Trema orientalis* leaves and these findings are in agreement with the
262 previous reports that the alkaloids, phenolics, triterpenoids, glycosides, tannins, saponins,
263 *flavonoids, steroids* etc. are the major bioactive molecules in *Trema orientalis* which have
264 enormous potential to inhibit microbial pathogens [24, 25]. As shown in Figure 1, the
265 methanol extract of *Trema orientalis* leaves exhibited similar extent of antibacterial
266 potentiality against both the Gram positive and Gram-negative bacteria indicates its broad
267 spectrum antibacterial activity. This activity may be caused by the various polar and non-
268 polar bioactive constituents present in the methanol extract of *Trema orientalis* leaves
269 because they may be acted either individually or combined to penetrate the outer
270 phospholipidic membrane of Gram-negative bacteria and peptidoglycan layer of Gram-
271 positive bacteria to inhibit or kill both the Gram positive and Gram-negative bacteria. Indeed,
272 methanol is an amphiphilic compound that can extract more of the extractives of polar
273 molecules and also non-polar ones [4]. It has been reported that organic solvents use has
274 varying abilities to extract bioactive substances from the medicinal plant. These observations
275 may be attributed to two reasons: firstly, the nature of biologically active components whose
276 activity can be enhanced in the presence of methanol; secondly, the stronger extraction
277 capacity of methanol could have produced the greater number of active constituents
278 responsible for antibacterial activity [26]. The results of the minimum inhibitory
279 concentration showed that the antibacterial activity of the methanol extract of *Trema*
280 *orientalis* leaves is concentration dependent. It is remarkable that *Salmonella typhi* showed
281 antibiotic resistance against all the tested commercial antibiotics that were used as a positive
282 control, but *Trema orientalis* leaves extract showed the potent antibacterial effect against
283 *Salmonella typhi*. This is the most significant part of this study and indicates the necessity of
284 natural plant products to combat against growing resistance of bacteria. *Salmonella typhi* is a

285 type of multi-drug resistance (MDR) strain. In all MDR strains so far examined, multiple
286 resistances have been encoded by plasmids of the H1 incompatibility group [25]. Since the
287 methanol extract of *Trema orientalis* leaves is found to exhibit antibacterial activity, the
288 magnitude of toxicity of *Trema orientalis* leaves extract is safe or acceptable at the
289 therapeutic doses must be considered. Plant samples with a lower LC50 value are considered
290 more toxic. Extracts are considered non-toxic if the LC50 is greater than 100 µg/mL in the
291 brine shrimp lethality assay [18]. Therefore cytotoxic assay was conducted in this study to
292 determine the toxicity profile of methanol extract of *Trema orientalis* leaves through the
293 brine shrimp lethality bioassay. Results of brine shrimp cytotoxicity were shown in Table 2,
294 where the LC₅₀ value is 170.215 µg/mL. This indicates that methanol extract of *Trema*
295 *orientalis* leaves is not toxic for host and can be a good source of potential antibacterial
296 agents. Although several researchers reported the antibacterial activity of leaves and stalked
297 [24], seed [27], and bark [28] extracts of *Trema orientalis*, to the best of our knowledge, no
298 detailed scientific proof for anti-bacterial and cytotoxic activities of *Trema orientalis* leaves
299 available yet for further using this plant for the development of potential new drugs or use in
300 a more effective form. The findings of this study indicate that the extract could be used
301 against infections caused by the tested bacteria and showed a good correlation between the
302 reported uses of *Trema orientalis* in traditional medicine against infectious diseases.

303

304 **5. Conclusion**

305 The findings of the present study has revealed that the methanol extract of *Trema orientalis*
306 leaves has great antibacterial potentiality due to the presence of the compounds with high
307 antibacterial properties that can be a source of natural antibacterial agents in developing new
308 drugs as an alternative to synthetic bactericides. The cytotoxic activity exhibited by the plant
309 leaves was within the permissible limit. Isolation and characterisation of the active

310 compounds could lead to a better understanding of the antibacterial mechanism for potential
311 drug candidates for the infectious diseases in future.

312

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315 Biotechnology, Jessore University of Science & Technology, Jessore-7408, Bangladesh
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317

318 **Conflict of Interest Statement**

319 All the authors declared no conflicts of interest concerning this study.

320

321 **Authors' contributions**

322 NB and MMK collected the plant and carried out the laboratory work. MMR designed the
323 experiments, performed statistical analysis, wrote the manuscript and supervised the work.
324 MJU also performed statistical analysis and contributed to critical reading of the paper. All
325 authors read and approved the final manuscript.

326

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