

Antibacterial Potentiality and Brine Shrimp Lethality Bioassay of the Methanol Extract of *Trema orientalis* Leaves

Abstract

Aim: The aim of this study was to evaluate the antibacterial and cytotoxic activities of methanol extract of *Trema orientalis* leaves.

Materials and Methods: Antibacterial activity of *Trema orientalis* leaves was tested against two Gram positive and seven Gram negative bacteria by disc diffusion assay. The liquid micro dilution assay was used for the determination of the minimum inhibitory concentration (MIC). The cytotoxic activity of methanol extract of *Trema orientalis* leaves was analyzed by the brine shrimp lethality bioassay.

Results: The methanol extract exhibited potent antibacterial activity with the zone of inhibition ranging from 9 ± 0.81 to 14 ± 0.81 mm against both the tested Gram positive and all tested Gram negative bacteria except *Pseudomonas denitrificans* and *Xanthomonas campestris*. Comparatively, higher antibacterial activity was found against Gram negative bacteria in case of *Shigella dysenteriae* and *Salmonella typhi* which showed 14 ± 0.81 mm and 13 ± 0.81 mm zones of inhibition respectively. *Salmonella typhi* showed resistance against reference antibiotics (Tetracycline, Erythromycin, Gentamicin and Ciprofloxacin) but methanol extract of leaves exhibited potent antibacterial activity against *Salmonella typhi*. The MIC values for tested Gram positive bacteria was 10 mg/mL while for Gram negative bacteria were ranged from 1.25 to 20 mg/mL. Methanol extract of *Trema orientalis* leaves showed very low cytotoxicity (LC₅₀, 170.215 µg/mL) in comparison with the standard vincristine sulphate having LC₅₀ value 2.477 µg/mL.

29 **Conclusion:** The results suggest that the methanol extract of *Trema orientalis* leaves has
30 potent antibacterial activity with minimum cytotoxicity and could lead to the development of
31 novel broad spectrum antibacterial agent.

32

33 **Keywords:** *Trema orientalis*, Antibacterial activity, Cytotoxicity, Disc diffusion, Minimum
34 inhibitory concentration and Brine shrimp lethality bioassay.

35

36 **1. Introduction**

37 Pathogenic microorganisms, such as bacteria, viruses, parasites and fungi cause infectious
38 diseases, which are considered a major threat to human health because of the unavailability of
39 vaccines, limited chemotherapy and emergence of resistant bacteria against antibiotics[1,2].

40 One half of all death in tropical countries is caused due to infectious diseases and responsible
41 for the second leading cause of death worldwide [3]. Most of the current antibiotics have
42 considerable limitations in terms of antimicrobial spectrum and side effects on the host
43 including allergic reactions, immune-suppression and hypersensitivity [4, 5]. Moreover, their
44 indiscriminate and inappropriate overuse has led to increasing clinical resistance of
45 previously sensitive microorganisms and to the occurrence of uncommon infections [1, 6].

46

47 New and re-emerging infectious diseases are rising very rapidly. Due to these problems,
48 attention is now being given to biologically active compounds isolated from plant species
49 because they offer a new source of antimicrobial drugs and are widely perceived as natural
50 and safe, that is, not toxic [5, 7]. Moreover, plant-based medicines contain diverse chemical
51 structure and novel mechanism of action that work in a way of orchestral ensembles which
52 are able to target many elements of the complex cell signaling pathways [8].It is well known
53 that the bioactive plant extracts are promising sources of majority of drugs. For example,

54 plant-based antibiotics such as Quinine (Cinchona) and berberine (Berberis) are highly
55 effective against *Staphylococcus aureus* and *Escherichia coli* [5]. Therefore, there is an urgent
56 need to search for new and more potent anti-bacterial and bioactive agents that can fight
57 against infectious pathogen.

58

59 *Trema orientalis* is a medicinal shrub or tree belonging to the family Ulmaceae. Locally, it is
60 known as charcoal tree or gunpowder tree. It is named Chikan or Jibon in Bengali, Nalita in
61 English and Gio in Hindi. It is a fast growing and evergreen tree and distributed all over the
62 world including Bangladesh. The young leaves are eaten as spinach and **in combination with**
63 **lemon juice**, the leaves maceration are used for the treatment of bronchitis, cough, pneumonia
64 and pleurisy. The infusion is prepared from fruits and flowers of *Trema orientalis* for
65 administration to children as a therapy for pneumonia, pleurisy and bronchitis [9]. The aerial
66 parts, flowers, bark, and seeds of *Trema orientalis* exhibit various pharmacological activities
67 including antidiarrheal, antidiabetic, anti-plasmodial, antimalarial and antioxidant activities
68 [10-14]. These pharmacological effects may be mainly due to the fact that it contains
69 important biologically active compounds such as scopoletin, 3, 4-hydroxybenzoic acid,
70 Epicatechin, lupeol, methylswertianin, catechin, hexacosanoic acid, tannins, saponins,
71 flavonoids, triterpenoid, phytosterols and xanthones [9]. Although there are many literatures
72 reporting the ethno-medicinal values of *Trema orientalis*, there is little scientific proof for
73 further using this plant commercially or in a more effective form. Therefore, an attempt was
74 made to evaluate the antibacterial and cytotoxic activities of the crude methanol extract of
75 *Trema orientalis* leaves to support the pharmacological effects and phytochemical
76 investigation of the plant.

77 **2. Materials and Methods**

78 **2.1. Chemical and Reagents**

79 All standard antibiotic discs used in this study were purchased from Bio-Rad, USA. Nutrient
80 agar media and Nutrient broth media were obtained from Liofilchem, Italy. Methanol from
81 Merck, Germany and the eggs of Brine shrimp were collected from an aquarium shop
82 (Dhaka, Bangladesh).

83

84 **2.2.Plant Material**

85 The leaves of *Trema orientalis* were collected during the month of January 2015 from
86 Jessore, Bangladesh and was authenticated by botanist. A voucher (DACB 31285) has been
87 deposited in Bangladesh National Herbarium, Mirpur, Bangladesh for further reference. The
88 collected plant leaves were washed with running tap water and dried in shade at room
89 temperature. The air dried leaves were pulverized into fine powder by commercial blender
90 (Philips, South Korea) and stored in sealed container.

91

92 **2.3. Experimental Methods**

93 **2.3.1.Preparation of theExtract**

94 100 g of powder was taken in a 500 ml conical flask added with 350 mL of methanol. The
95 flask was kept for 7 days with continuous shaking at shaking incubator at room temperature.
96 The plant extract was filtered through Whatman no.1 filter paper (Thermo Fisher Scientific,
97 USA.) and then concentrated by using a rotary evaporator (Stuart, UK) and kept at room
98 temperature to evaporate the remaining solvent. After complete evaporation of solvent, only
99 plant's crude extracts were obtained. The amount of crude extracts was 1.0 g which was
100 stored in refrigerator at 4°C in sterile container for further use.

101

102 **2.3.2. Tested Bacterial Preparation**

103 Pure culture of Gram positive bacteria (*Bacillus subtilis* IFO 3026, *Sarcinalutea* IFO 3232)
104 and Gram negative bacteria (*Escherichia coli* IFO 3007, *Proteus vulgaris* MTTC 321,
105 *Klebsiella pneumonia* ATTC 10031, *Xanthomonas campestris* IAM 1671, *Pseudomonas*
106 *denitrificans* KACC 32026) were used in this study and obtained from the Microbiology
107 Laboratory of Department of Biotechnology and Genetic Engineering, Islamic University,
108 Kushtia, Bangladesh. Another two Gram negative bacteria, *Salmonella typhi* and *Shigella*
109 *dysenteriae* were kindly provided by the Microbiology laboratory of Department of
110 Microbiology, Jessore University of Science & Technology. Bacteria were cultured in
111 Nutrient agar media and Nutrient broth media. For antibacterial assay, minimum inhibitory
112 concentration (MIC) determination and the further stock culture preparation, 100 µL of
113 frozen stock culture was inoculated into 125 mL conical flask containing 25 mL of Nutrient
114 broth media and incubated at 37°C with continuous shaking at 250rpm for culturing the
115 bacteria until mid-log phase of absorbance at 600 nm reached at 0.4 by using UV
116 spectrophotometer (Oasis scientific Inc., USA) for bacterial broth culture.

117

118 **2.3.3. Disc Preparation**

119 The Whatman No. 1 filter paper discs (6 mm diameter) were transferred to a small vial and
120 autoclaved at 15 lb/inch² pressure for 15 minutes at 121°C. The discs were completely dried
121 in drying oven at 60°C. 400 mg of crude methanol extract of *Trema orientalis* was dissolved
122 into 10 mL of methanol and each disc was impregnated with 10 µL of 40 mg/mL
123 (400µg/disc) of *Trema orientalis* leaves extract. The discs were completely air dried in the
124 laminar flow cabinet and used for antibacterial assay. Blank discs (negative controls)
125 impregnated with 10 µL of methanol.

126

127 **2.3.4. Antibacterial Activity Assay**

128 Antibacterial activity of crude extract was tested by the disc diffusion method [15]. The
129 prepared discs were placed on nutrient-agar-medium plate spread with 100 μ L of tested
130 bacterial broth culture and the plates were incubated at 37°C for 24h. Standard reference
131 antibiotics Tetracycline (30 μ g/disc), Erythromycin (15 μ g/disc), Gentamicin (10 μ g/disc)
132 and Ciprofloxacin (5 μ g/disc) were used as positive control to ensure the activity of standard
133 antibiotic against the test organisms. The blank discs were used as negative control. After
134 incubation, the culture plates were examined and the inhibition zones formed around each
135 disk were measured in millimeter scale as previously described [16]. Each assay in this
136 experiment was replicated three times.

137

138 **2.3.5. Determination of Minimum Inhibitory Concentration (MIC)**

139 Minimum inhibitory concentration (MIC) of methanol extract of *Trema orientalis* was
140 determined by a two-fold serial dilution method as previously described [17]. The methanol
141 crude extract of *Trema orientalis* leaves was dissolved in Nutrient broth medium in an
142 eppendorf tube (Watson Co. Ltd., Japan) to achieve a concentration of 40 mg/mL. The
143 solution of eppendorf tube was serially diluted to obtain 20, 10, 5, 2.5 and 1.25 mg/mL of
144 concentrations. The 0.5 mL of bacterial broth culture of each tested bacteria was transferred
145 to each eppendorf tube. Thus, the total amount of solution in each eppendorf tube was 1 mL.
146 The control tubes contain 0.5 mL bacterial broth cultures with 0.5 mL nutrient broth media.
147 The solution of all eppendorf tubes were mixed properly by vortexing and incubated at 37°C
148 for 24 h with continuous shaking at 250 rpm. After incubating 24 h, 100 μ L of solution from
149 each eppendorf tube were spread over the nutrient-agar-media plate. The plates were
150 incubated at 37°C for 16 h for bacterial growth and the number of colony was counted for
151 MIC determination.

152

153 **2.3.6. Brine Shrimp Lethality Bioassay**

154 Brine shrimp lethality bioassay is the most convenient system for preliminary assessment of
155 cytotoxicity of plant extracts. The brine shrimp lethality bioassay of the methanol extract of
156 *Trema orientalis* leaves was evaluated as previously described procedure against *Artemia*
157 *salina* as a test organism to monitor the cytotoxicity of a compound [18]. The eggs of Brine
158 shrimp (*Artemia salina*) were collected from an aquarium shop (Dhaka, Bangladesh) and
159 incubated for 28°C with constant oxygen supply and hatched for two days to provide a large
160 number of larvae called nauplii. The different concentrations of crude extract were prepared
161 by dissolving them in DMSO (not more than 50 µL in 5 mL solution) plus seawater (3.8%
162 NaCl in water) to attain serial dilution from 200-1.562 µg/mL. The standard vincristine
163 sulphate was used as a positive control. The varying concentration of solution of vincristine
164 sulphate was prepared by serial dilution into DMSO to attain serial dilution from 200-1.562
165 µg/mL. A vial containing 50 µL of DMSO diluted to 5 mL simulated seawater used as a
166 control. Twenty mature shrimps were placed into each of the experimental vials. After 24 h,
167 the vials were inspected using a magnifying glass, and the number of surviving nauplii in
168 each vial was counted. From this data, the percentage (%) of mortality of the brine shrimp
169 naupili was calculated for each concentration using the following formula: % Mortality =
170 $N_t/N_0 \times 100$ (Where N_t = Number of dead nauplii after a 24 h incubation; N_0 = Number of
171 total nauplii transferred i.e., 10). The LC50 (median lethal concentration) was determined
172 from the log concentration versus % mortality.

173

174 **2.3.7. Statistical Analysis**

175 The experimental results obtained from antibacterial and MIC determination assays were
176 expressed as mean ± standard deviation (SD) of three replicates. LC50 values were

177 determined by correlation/regression analysis. Microsoft Excel 2010 statistical package was
178 used for all analyses.

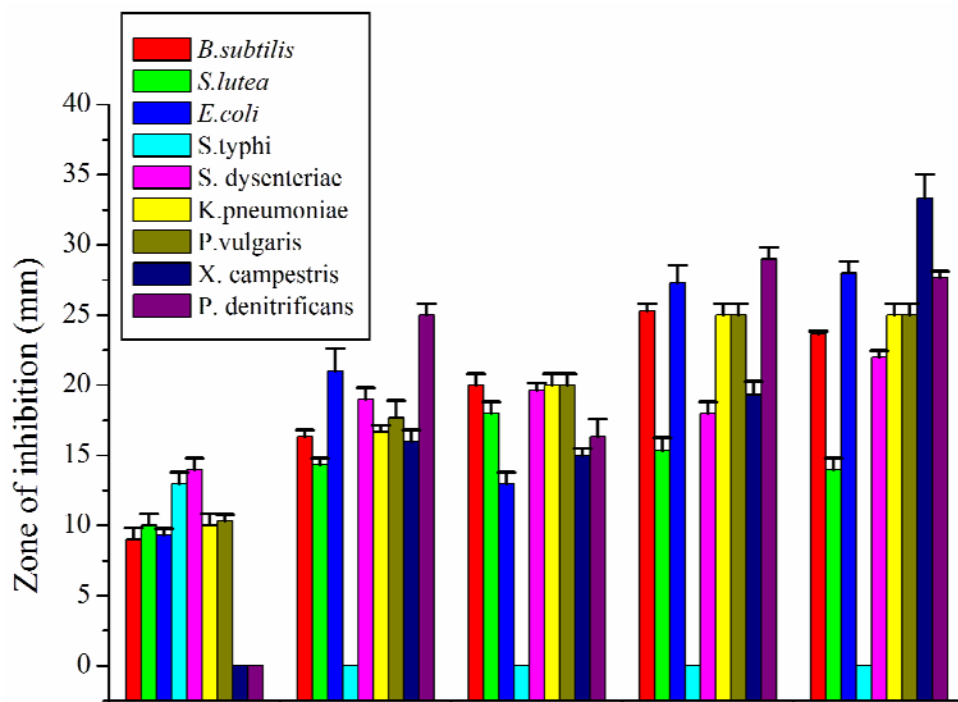
179

180 3. Results

181 3.1. Antibacterial Potentialities of *Trema orientalis* Extract

182 The antibacterial activities of methanol extract of *Trema orientalis* leaves against the tested
183 bacteria were examined by the occurrence of clear zone of inhibition. The leaves extracted at
184 a concentration of 400 µg/disc showed significant antibacterial effects against two Gram
185 positive bacteria (*Bacillus subtilis*, *Sarcina lutea*) and five Gram negative bacteria
186 (*Escherichia coli*, *Salmonella typhi*, *Shigella dysenteriae*, *Klebsiella pneumoniae*, and
187 *Proteus vulgaris*) with the zone of inhibition ranging from 9 ± 0.81 to 14 ± 0.81 mm (Figure 1).
188 The highest zone of inhibition was found 14 ± 0.81 mm and 13 ± 0.81
189 mm against *Shigella dysenteriae* and *Salmonella typhi* respectively. The inhibition zone was
190 observed 10 ± 0.81 mm against *Klebsiella pneumoniae*, *Proteus vulgaris* and *Sarcina lutea*
191 whereas zone of inhibition was 9 ± 0.47 mm against *Bacillus subtilis* and *Escherichia coli*.
192 However, no antibacterial activity was observed against two Gram negative bacteria,
193 *Xanthomonas campestris* and *Pseudomonas denitrificans* at the used concentration of
194 400 µg/disc of plant leaves extract. Standard reference antibiotics: Tetracycline,
195 Erythromycin, Gentamicin and Ciprofloxacin were used as positive control showed higher
196 antibacterial activities than the plant leaves extract against all the tested bacteria except
197 *Salmonella typhi*. Though *Salmonella typhi* showed resistance against reference antibiotics as
198 positive control, methanol extract of *Trema orientalis* leaves exhibited strong zone of
199 inhibition (13 ± 0.81 mm) against *Salmonella typhi* (Figure 1) suggest that it could be a

200 potential therapeutic drug candidate against *Salmonella typhi*. No zone was formed by
 201 negative control.



202

203 **Figure 1.** Effect of methanol extract of *Trema orientalis* leaves on two Gram positive and
 204 seven Gram negative bacteria. Values are represented as **mean \pm SD (n=3)**.SD, Standard
 205 deviation; TC, Tetracycline; EM, Erythromycin; GM, Gentamycin; CF, Ciprofloxacin.

206

207 3.2. Minimum Inhibitory Concentration

208 The lowest concentration of methanol extract which prevents visible growth of bacterium is
 209 the minimum inhibitory concentration. The MIC values of crude extract of *Trema orientalis*
 210 leaves were found ranging from 1.25 to 20 mg/mL (Table 1). The best MIC was 1.25 mg/mL
 211 against *Escherichia coli*, *Salmonella typhi* and *Shigella dysenteriae* as this concentration
 212 completely inhibited the growth of these bacteria. The least efficacy was shown against
 213 *Proteus vulgaris* and *Klebsiella pneumoniae* which was inhibited at 20 mg/mL concentration.

214 The moderate MIC value was shown against Gram positive bacteria (*Bacillus subtilis* and
 215 *Sarcina lutea*) which were inhibited at 10 mg/mL concentration.

216

217 **Table 1:** Minimum inhibitory concentration of methanol extract of leaves of *Trema*
 218 *orientalis*.

219

Tested bacteria	Minimum Inhibitory Concentration (mg/mL)						
	20	10	5	2.5	1.25	0.625	0.312
	Number of bacterial colonies survived at above concentration						
<i>Bacillus subtilis</i>	0	0	7± .816	17± 1.63	54±3.74	96±3.26	121±3.74
<i>Sarcina lutea</i>	0	0	12±2.16	43±4.32	66±4.39	107±4.08	144±6.53
<i>Escherichia coli</i>	0	0	0	0	0	47±3.74	133±3.74
<i>Salmonella typhi</i>	0	0	0	0	0	77±3.26	124±4.32
<i>Shigella dysenteriae</i>	0	0	0	0	0	55±4.08	112±4.08
<i>Klebsiella pneumoniae</i>	0	5±0.81	16±2.16	52±2.82	79±4.08	123±3.26	223±2.94
<i>Proteus vulgaris</i>	0	23±2.94	54±3.74	77±2.44	91±2.44	155±4.08	175±4.54

220 Values are represented as mean ± SD (n=3).

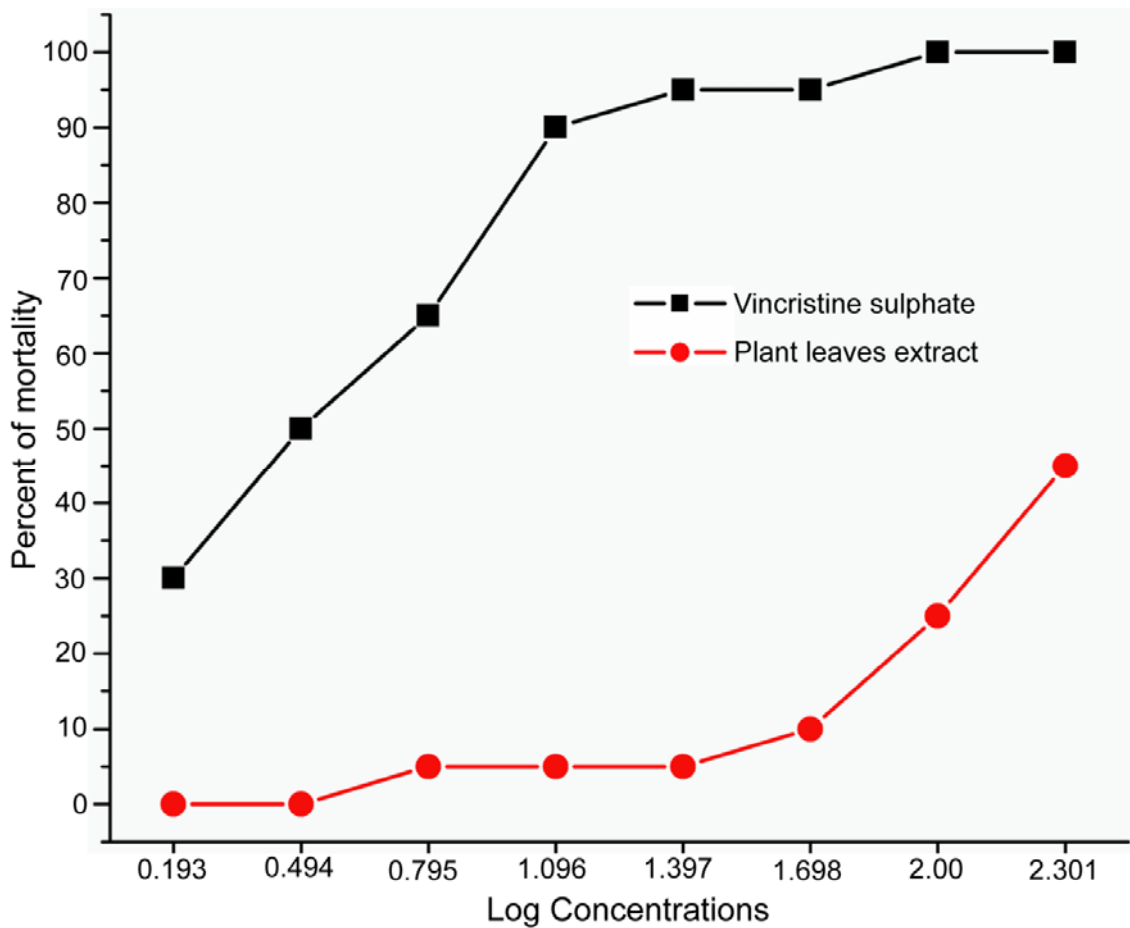
221

222 3.3.Cytotoxic Activity of Methanol Extract of *Trema orientalis*

223 The percent mortality of brine shrimp naupli at different concentrations of plant extracts and
 224 vincristine sulfate as positive control are shown in Figure 2. It is clear that percent mortality
 225 of brine shrimp naupli is proportional to the concentration of extracts. The mortality rate was
 226 increased with the extract concentration increased. As shown in Table 2, methanol extract of
 227 *Trema orientalis* demonstrated a LC50 value of 170.215 µg/mL whereas vincristine sulphate
 228 showed the LC50 value of 2.477 µg/mL. This indicates that the plant leaves extract has much
 229 more higher LC50 value compared to that of vincristine sulphate. The crude methanol

230 extracts resulted in LC50 values greater than 100 µg/mL were considered non-toxic in the
231 brine shrimp lethality assay [19], support the notion that the methanol extract of *Trema*
232 *orientalis* leaves is non-toxic for host and had the potential for further investigation. There
233 was no mortality in the negative control groups indicating the test as a valid one and the
234 results obtained are only due to the activity of the tested agents.

235



236

237 **Figure 2.** Brine shrimp lethality for methanol extract of *Trema orientalis* leaves and
238 vincristine sulphate from linear correlation between log concentrations versus % mortality.

239

240

241

242

243 **Table 2.** The cytotoxicity of methanol extract of *Trema orientalis* leaves and vincristine
244 sulphate on brine shrimp nauplii.
245

Sample	LC ₅₀ (µg/mL)	Regression equation	R ²
Plant extract	170.215	y=29.851x + (-16.592)	0.785
Vincristine sulphate	2.477	y=33.004x+36.976	0.838

246

247 **4. Discussion**

248 Infectious diseases are the second leading cause of death worldwide. Recently, the emergence
249 of antibiotic-resistant infection are rising very rapidly, which are major threat to human
250 health as well as economic burden on country's healthcare system, patients and families. The
251 effectiveness of many conventional antibiotics is being endangered by the rapid emergence of
252 microbial resistance to current therapeutic agents because of their overuse, misuse, and a lack
253 of new drug development by the pharmaceutical industry [20-22]. Plant-derived natural
254 secondary metabolites represent a potential source of antimicrobial agents which have
255 different mode of action than conventional drug. Acceptance of medicines from natural plant
256 product as an alternative form of healthcare system is increasing because they are serving as
257 promising sources of novel antibiotic prototype that could be of clinical importance to
258 improve health care [23, 24]. Therefore, we investigated the *Trema orientalis* leaves for its
259 antibacterial and cytotoxic activities. The present study showed that the methanol extract of
260 *Trema orientalis* leaves at a concentration of 400 µg/disc has potent antibacterial activity
261 against both the Gram positive (*Bacillus subtilis* and *Sarcina lutea*) and Gram negative
262 bacteria (*Escherichia coli*, *Salmonella typhi*, *Shigella dysenteriae*, *Klebsiella pneumoniae* and
263 *Proteus vulgaris*). The activity exhibited by the methanol extract of *Trema orientalis* leaves
264 may be attributed to the presence of some bioactive compounds in *Trema orientalis* leaves
265 and these findings are in agreement with the previous reports that the alkaloids, phenolics,

266 triterpenoids, glycosides, tannins, saponins, *flavonoids*, *steroids* etc. are the major bioactive
267 molecules in *Trema orientalis* which have enormous potential to inhibit microbial pathogens
268 [25,26]. As shown in figure 1, methanol extract of *Trema orientalis* leaves exhibited similar
269 extent of antibacterial potentiality against both the Gram positive and Gram negative bacteria
270 indicates its broad spectrum antibacterial activity. This activity may be caused by the various
271 polar and non-polar bioactive constituents present in methanol extract of *Trema orientalis*
272 leaves because they may be acted either individually or combined to penetrate the outer
273 phospholipidic membrane of Gram negative bacteria and peptidoglycane layer of Gram
274 positive bacteria to inhibit or kill both the Gram positive and Gram negative bacteria. Indeed,
275 methanol is an amphiphilic compound that is able to extract more of the extractives of polar
276 molecules and also non-polar ones [4]. It has been reported that organic solvents use has
277 varying abilities to extract bioactive substances from medicinal plant. These observations
278 may be attributed to two reasons: firstly, the nature of biological active components whose
279 activity can be enhanced in the presence of methanol; secondly, the stronger extraction
280 capacity of methanol could have produced greater number of active constituents responsible
281 for antibacterial activity [27]. The results of the minimum inhibitory concentration showed
282 that the antibacterial activity of the methanol extract of *Trema orientalis* leaves is
283 concentration dependent. It is remarkable that *Salmonella typhi* showed antibiotic resistance
284 against all the tested commercial antibiotics that were used as positive control but *Trema*
285 *orientalis* leaves extract showed potent antibacterial effect against *Salmonella typhi*. This is
286 the most significant part of this study and indicate the necessity of natural plant products to
287 combat against growing resistance of bacteria. *Salmonella typhi* is a type of multi-drug
288 resistance (MDR) strain. In all MDR strains so far examined, multiple resistances have been
289 encoded by plasmids of the H1 incompatibility group [28]. Since the methanol extract of
290 *Trema orientalis* is found to exhibit anti-bacterial activity, the magnitude of toxicity of *Trema*

291 *orientalis* extract is safe or acceptable at the therapeutic doses must be considered. Plant
292 samples with a lower LC50 value are considered more toxic in nature. Extracts are
293 considered non-toxic if the LC50 is greater than 100 µg/mL in the brine shrimp lethality
294 assay [19]. Therefore cytotoxic assay was conducted in this study to determine the toxicity
295 profile of methanol extract of *Trema orientalis* leaves through the brine shrimp lethality
296 bioassay. Results of brine shrimp cytotoxicity were shown in Table 2 where the LC₅₀ value is
297 170.215µg/mL. This indicates that methanol extract of *Trema orientalis* leaves is not toxic for
298 host and can be a good source of potential antibacterial agents. Although several researches
299 reported the antibacterial activity of leaves and stalk [25], seed [26], and bark [29] extracts of
300 *Trema orientalis*, to the best of our knowledge, no detailed scientific proof for anti-bacterial
301 and cytotoxic activities of *Trema orientalis* leaves available yet for further using this plant for
302 the development of potential new drugs or use in a more effective form. The findings of this
303 study indicates that the extract could be used against infections caused by the tested bacteria
304 and showed a good correlation between the reported uses of *Trema orientalis* in traditional
305 medicine against infectious diseases.

306

307 **5. Conclusion**

308 The findings of the present study has revealed that the methanol extract of *Trema orientalis*
309 leaves has great antibacterial potentiality due to the presence of the compounds with high
310 antibacterial properties that can be a source of natural antibacterial agents in developing new
311 drugs as an alternative to synthetic bactericides. The cytotoxic activity exhibited by the plant
312 leaves was within the permissible limit. Isolation and characterization of the active
313 compounds could lead to a better understanding of the antibacterial mechanism for potential
314 drug candidates for the infectious diseases in future.

315

316 **Acknowledgments**

317 This research was funded by a research grant from Department of Genetic Engineering &
318 Biotechnology, Jessore University of Science & Technology, Jessore-7408, Bangladesh
319 (grant number GEBT/JUST/2016-4829)

320

321 **Conflict of Interest Statement**

322 All the authors declared no conflicts of interest with regard to this study.

323

324 **Authors' contributions**

325 NB and MMK collected the plant and carried out the laboratory work. MMR designed the
326 experiments, performed statistical analysis, wrote the manuscript and supervised the work.

327 MJU also performed statistical analysis and contributed to critical reading of the manuscript.

328 All authors read and approved the final manuscript.

329

330 **References**

- 331 1. Assob JCN, Kamga HLF, Nsagha DS, Njunda AL, Nde PF and Asongalem EA, et al.
332 Antimicrobial and toxicological activities of five medicinal plant species from
333 Cameroon traditional medicine. BMC Complement. Altern. Med. 2011;11:70.
334 doi:10.1186/1472-6882-11-70.
- 335 2. Ameya G, Manilal A and Merdekios B. *In vitro* antibacterial activity and
336 phytochemical analysis of *Nicotiana tabacum* L. extracted in different organic
337 solvents. Open Microbiol. J. 2017;11:352-359.

- 338 3. Olajuyigbe OO, Afolayan AJ. Synergistic Interactions of methanolic extract of *Acacia*
339 *mearnsii* De Wild. with antibiotics against bacteria of clinical relevance. Int. J. Mol.
340 Sci. 2012;13(7):8915-8932.
- 341 4. Londonkar RL, MadireKattegouga U, Shivsharanappa K, Hanchinalmath JV.
342 Phytochemical screening and *in vitro* antimicrobial activity of *Typha angustifolia*
343 Linn leaves extract against pathogenic gram negative microorganisms. J. Pharm. Res.
344 2013; 6(2):280-283.
- 345 5. Bibi Y, Nisa S, Chaudhary FM and Zia M. Antibacterial activity of some selected
346 medicinal plants of Pakistan. BMC Complement. Altern. Med. 2011;11:52.
347 doi:10.1186/1472-6882-11-52.
- 348 6. Ventola CL. The antibiotic resistance crisis: Part 1: causes and threats. Pharm.
349 Ther.2015;40(4):277-283.
- 350 7. Maiyo ZC, Ngure RM, Matasyoh JC and Chepkorir R: Phytochemical constituents
351 and antimicrobial activity of extracts of three *Amaranthus* plant species. Afr. J.
352 Biotechnol.2010;9(21):3178-3182.
- 353 8. Lalrinzuali K, Vabeiryureilai M and Jagetia GC. Investigation of the anti-
354 inflammatory and analgesic activities of ethanol extract of stem bark of *Sonapatha*
355 *Oroxylum indicum in vivo*. Int. J. Inflamm. 2016. doi.org/10.1155/2016/8247014.
- 356 9. Adinortey MB, Galyuon IKand Asamoah NO. *Trema orientalis* Linn. Blume: A
357 potential for prospecting for drugs for various uses. Pharmacogn. Rev.
358 2013;7(13):67-72.
- 359 10. Sayeed MA, Jainul MA, Azam S, Babar ZM and Azad AK. *In vivo* antidiarrheal
360 activity of methanolic extract of *Trema oreintalis* leaves. Ph. OL 2017;2:187-192.

- 361 11. Jiji KN, Pramod C, Prasad BS, Muralidharan DRP. Evaluation of antidiabetic
362 activity of ethanolic extract of *Trema orientalis* (L.) Blume leaves. IOSR-JPBS
363 2016;11(5):17-26.
- 364 12. Olanlokun JO, David OM and Afolayan AJ. *In vitro* antiplasmodial activity and
365 prophylactic potentials of extract and fractions of *Trema orientalis* (Linn.) stem
366 bark. BMC Complement. Altern. Med. 2017;17:407. DOI 10.1186/s12906-017-
367 1914-x.
- 368 13. Oyebola OE, Morenikeji OA and Ademola IO. *In vivo* antimalarial activity of
369 aqueous leaf and bark extracts of *Trema orientalis* against *Plasmodium berghei* in
370 mice. J. Parasit. Dis. 2017;41(2):398-404.
- 371 14. Uddin SN. Antioxidant and antibacterial activities of *Trema orientalis* Linn: an
372 indigenous medicinal plant of Indian subcontinent. Orient. Pharm. Experiment.
373 Med. 2008 8(4):395-399.
- 374 15. Bauer AW, Kirby WMM, Sherris JC and Turck M. Antibiotic susceptibility testing
375 by a standardized single disk method. Am. J. Clin. Pathol. 1966;45(4):493-6.
- 376 16. Rahman MM, Rahman MM, Akhter S, Jamal MAHM, Pandeya DR and Haque
377 MA, et al. Control of coliform bacteria detected from diarrhea associated patients
378 by extracts of *Moringa oleifera*. Nepal Med. Coll. J. 2010;12(1):12-19.
- 379 17. Chandrasekaran M and Venkatesalu V. Antibacterial and antifungal activity of
380 *Syzygium jambolanum* seeds. J. Ethno. Pharmacol. 2004;91:105-108.
- 381 18. Meyer BN, Ferrigni NA, Putnam JE, Jacobsen LB, Nichols DE and McLaughlin
382 JL. Brine Shrimp: A Convenient General Bioassay for Active Plant Constituents. J.
383 Med. Plants Res.1982;45:31-34.
- 384 19. Naz R, Ayub H, Nawaz S, Islam ZU, Yasmin T and Bano A, et al. Antimicrobial
385 activity, toxicity and anti-inflammatory potential of methanolic extracts of four

- 386 ethnomedicinal plant species from Punjab, Pakistan. BMC Complement. Altern.
387 Med.2017;17:302. doi:10.1186/s12906-017-1815-z.
- 388 20. Gould IM and Bal AM. New antibiotic agents in the pipeline and how they can
389 overcome microbial resistance. Virulence 2013;4(2):185-191.
- 390 21. Sengupta S, Chattopadhyay MK and Grossart HP. The multifaceted roles of
391 antibiotics and antibiotic resistance in nature. Front. Microbiol.2013;4:47.
- 392 22. Manyi-Loh C, Mamphweli S, Meyer E and Okoh A. Antibiotic use in agriculture
393 and its consequential resistance in environmental Sources: potential public health
394 implications. Molecules 2018;23:795; doi:10.3390/molecules23040795.
- 395 23. Koduru S, Grierson DS and Afolayan AJ. Antimicrobial activity of *Solanum*
396 *aculeastrum*. Pharm. Biol. 2006;44:283-286.
- 397 24. Okeke MI, Iroegbu CU, Eze EN, Okoli AS and Esimone CO. Evaluation of
398 extracts of the root of *Landolphia owerrience* for antibacterial activity. J.
399 Ethnopharmacol. 2001;78:119-127.
- 400 25. Akin-Osanaiye BC, Gabriel AF, Omoniyi AO and Ezeani SC. Scientific approach
401 on the antimicrobial potentials of bioactive phytochemicals of *Trema orientalis*
402 leaves and stalk. Europ. Acad. Res.2016;3(12):12972-12981.
- 403 26. Akin-Osanaiye BC and Rukayyah A. Phytochemical analysis, anti-microbial
404 screening and anti-oxidant activity of the seed of *Trema orientalis*. Acad. J. Sci.
405 2014;3(1):211-217.
- 406 27. Bhattacharjee I, Chatterjee SK, Chatterjee S and Chandra G. Antibacterial
407 potentiality of *Argemone mexicana* solvent extracts against some pathogenic
408 bacteria. Mem. Inst. Oswaldo. Cruz. Rio de Janeiro 2006;101(6):645-648.
- 409 28. Alim S, Bairagi N, Shahriyar S, Kabir MM and Rahman MH. *In vitro* antibacterial
410 potential of *Bixa orellana* L.against some pathogenic bacteria and comparative

411 investigation on some standard antibiotics. J. Pharmacogn. Phytochem.
412 2016;5(2):178-181.

413 29. Rout J, Sajema AL, Nathb M and Sengupta M. Antibacterial efficacy of bark
414 extracts of an ethnomedicinal plant *Trema orientalis* Blume. Curr. Trends
415 Biotechnol. Pharm. 2012;6 (4):464-471.

416