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Original Research Article

Antibacterial Potentiality and Brine Shrimp Lethality Bioassay of the Methanol Extract of *Trema orientalis* Leaves

Abstract

Aim: The aim of this study was to evaluate the antibacterial and cytotoxic activities of methanol extract of *Trema orientalis* leaves.

Materials and Methods: Antibacterial activity of *Trema orientalis* leaves was tested against two Gram positive and seven Gram negative bacteria by disc diffusion assay. The liquid micro dilution assay was used for the determination of the minimum inhibitory concentration (MIC). The cytotoxic activity of methanol extract of *Trema orientalis* leaves was analyzed by the brine shrimp lethality bioassay.

Results: The methanol extract exhibited potent antibacterial activity with the zone of inhibition ranging from 9 ± 0.81 to 14 ± 0.81 mm against both the tested Gram positive and all tested Gram negative bacteria except *Pseudomonas denitrificans* and *Xanthomonas campestris*. Comparatively, higher antibacterial activity was found against Gram negative bacteria in case of *Shigella dysenteriae* and *Salmonella typhi* which showed 14 ± 0.81 mm and 13 ± 0.81 mm zones of inhibition respectively. *Salmonella typhi* showed resistance against reference antibiotics (Tetracycline, Erythromycin, Gentamicin and Ciprofloxacin) but methanol extract of leaves exhibited potent antibacterial activity against *Salmonella typhi*. The MIC values for tested Gram positive bacteria was 10 mg/mL while for Gram negative bacteria were ranged from 1.25 to 20 mg/mL. Methanol extract of *Trema orientalis* leaves showed very low cytotoxicity (LC₅₀, 170.215 µg/mL) in comparison with the standard vincristine sulphate having LC₅₀ value 2.477 µg/mL.

29

30 **Conclusion:** The results suggest that the methanol extract of *Trema orientalis* leaves has
31 potent antibacterial activity with minimum cytotoxicity and could lead to the development of
32 novel broad spectrum antibacterial agent.

33

34 **Keywords:** *Trema orientalis*, Antibacterial activity, Cytotoxicity, Disc diffusion, Minimum
35 inhibitory concentration and Brine shrimp lethality bioassay.

36

37 **1. Introduction**

38 Pathogenic microorganisms, such as bacteria, viruses, parasites and fungi cause infectious
39 diseases, which are considered a major threat to human health because of the unavailability of
40 vaccines, limited chemotherapy and emergence of resistant bacteria against antibiotics[1,2].

41 One half of all death in tropical countries is caused due to infectious diseases and responsible
42 for the second leading cause of death worldwide [3]. Most of the current antibiotics have
43 considerable limitations in terms of antimicrobial spectrum and side effects on the host
44 including allergic reactions, immune-suppression and hypersensitivity [4, 5]. Moreover, their
45 indiscriminate and inappropriate overuse has led to increasing clinical resistance of
46 previously sensitive microorganisms and to the occurrence of uncommon infections [1, 6].

47

48 New and re-emerging infectious diseases are rising very rapidly. Due to these problems,
49 attention is now being given to biologically active compounds isolated from plant species
50 because they offer a new source of antimicrobial drugs and are widely perceived as natural
51 and safe, that is, not toxic [5, 7]. Moreover, plant-based medicines contain diverse chemical
52 structure and novel mechanism of action that work in a way of orchestral ensembles which
53 are able to target many elements of the complex cell signaling pathways [8].It is well known

54 that the bioactive plant extracts are promising sources of majority of drugs. For example,
55 plant-based antibiotics such as Quinine (Cinchona) and berberine (Berberis) are highly
56 effective against *Staphylococcus aureus* and *Escherichia coli* [5]. Therefore, there is an urgent
57 need to search for new and more potent anti-bacterial and bioactive agents that can fight
58 against infectious pathogen.

59

60 *Trema orientalis* is a medicinal shrub or tree belonging to the family Ulmaceae. Locally, it is
61 known as charcoal tree or gunpowder tree. It is named Chikan or Jibon in Bengali, Nalita in
62 English and Gio in Hindi. It is a fast growing and evergreen tree and distributed all over the
63 world including Bangladesh. The young leaves are eaten as spinach and **in combination with**
64 **lemon juice**, the leaves maceration are used for the treatment of bronchitis, cough, pneumonia
65 and pleurisy. The infusion is prepared from fruits and flowers of *Trema orientalis* for
66 administration to children as a therapy for pneumonia, pleurisy and bronchitis [9]. The aerial
67 parts, flowers, bark, and seeds of *Trema orientalis* exhibit various pharmacological activities
68 including antidiarrheal, antidiabetic, anti-plasmodial, antimalarial and antioxidant activities
69 [10-14]. These pharmacological effects may be mainly due to the fact that it contains
70 important biologically active compounds such as scopoletin, 3, 4-hydroxybenzoic acid,
71 Epicatechin, lupeol, methylswertianin, catechin, hexacosanoic acid, tannins, saponins,
72 flavonoids, triterpenoid, phytosterols and xanthones [9]. Although there are many literatures
73 reporting the ethno-medicinal values of *Trema orientalis*, there is little scientific proof for
74 further using this plant commercially or in a more effective form. Therefore, an attempt was
75 made to evaluate the antibacterial and cytotoxic activities of the crude methanol extract of
76 *Trema orientalis* leaves to support the pharmacological effects and phytochemical
77 investigation of the plant.

78 **2. Materials and Methods**

79 **2.1. Chemical and Reagents**

80 All standard antibiotic discs used in this study were purchased from Bio-Rad, USA. Nutrient
81 agar media and Nutrient broth media were obtained from Liofilchem, Italy. Methanol from
82 Merck, Germany and the eggs of Brine shrimp were collected from an aquarium shop
83 (Dhaka, Bangladesh).

84

85 **2.2.Plant Material**

86 The leaves of *Trema orientalis* were collected during the month of January 2015 from
87 Jessore, Bangladesh and was authenticated by botanist. A voucher (DACB 31285) has been
88 deposited in Bangladesh National Herbarium, Mirpur, Bangladesh for further reference. The
89 collected plant leaves were washed with running tap water and dried in shade at room
90 temperature. The air dried leaves were pulverized into fine powder by commercial blender
91 (Philips, South Korea) and stored in sealed container.

92

93 **2.3. Experimental Methods**

94 **2.3.1.Preparation of theExtract**

95 100 g of powder was taken in a 500 ml conical flask added with 350 mL of methanol. The
96 flask was kept for 7 days with continuous shaking at shaking incubator at room temperature.
97 The plant extract was filtered through Whatman no.1 filter paper (Thermo Fisher Scientific,
98 USA.) and then concentrated by using a rotary evaporator (Stuart, UK) and kept at room
99 temperature to evaporate the remaining solvent. After complete evaporation of solvent, only
100 plant's crude extracts were obtained. The amount of crude extracts was 1.0 g which was
101 stored in refrigerator at 4°C in sterile container for further use.

102

103 **2.3.2. Tested Bacterial Preparation**

104 Pure culture of Gram positive bacteria (*Bacillus subtilis* IFO 3026, *Sarcinalutea* IFO 3232)
105 and Gram negative bacteria (*Escherichia coli* IFO 3007, *Proteus vulgaris* MTTC 321,
106 *Klebsiella pneumonia* ATTC 10031, *Xanthomonas campestris* IAM 1671, *Pseudomonas*
107 *denitrificans* KACC 32026) were used in this study and obtained from the Microbiology
108 Laboratory of Department of Biotechnology and Genetic Engineering, Islamic University,
109 Kushtia, Bangladesh. Another two Gram negative bacteria, *Salmonella typhi* and *Shigella*
110 *dysenteriae* were kindly provided by the Microbiology laboratory of Department of
111 Microbiology, Jessore University of Science & Technology. Bacteria were cultured in
112 Nutrient agar media and Nutrient broth media. For antibacterial assay, minimum inhibitory
113 concentration (MIC) determination and the further stock culture preparation, 100 µL of
114 frozen stock culture was inoculated into 125 mL conical flask containing 25 mL of Nutrient
115 broth media and incubated at 37°C with continuous shaking at 250rpm for culturing the
116 bacteria until mid-log phase of absorbance at 600 nm reached at 0.4 by using UV
117 spectrophotometer (Oasis scientific Inc., USA) for bacterial broth culture.

118

119 **2.3.3. Disc Preparation**

120 The Whatman No. 1 filter paper discs (6 mm diameter) were transferred to a small vial and
121 autoclaved at 15 lb/inch² pressure for 15 minutes at 121°C. The discs were completely dried
122 in drying oven at 60°C. 400 mg of crude methanol extract of *Trema orientalis* was dissolved
123 into 10 mL of methanol and each disc was impregnated with 10 µL of 40 mg/mL
124 (400µg/disc) of *Trema orientalis* leaves extract. The discs were completely air dried in the
125 laminar flow cabinet and used for antibacterial assay. Blank discs (negative controls)
126 impregnated with 10 µL of methanol.

127

128 **2.3.4. Antibacterial Activity Assay**

129 Antibacterial activity of crude extract was tested by the disc diffusion method [15]. The
130 prepared discs were placed on nutrient-agar-medium plate spread with 100 μ L of tested
131 bacterial broth culture and the plates were incubated at 37°C for 24h. Standard reference
132 antibiotics Tetracycline (30 μ g/disc), Erythromycin (15 μ g/disc), Gentamicin (10 μ g/disc)
133 and Ciprofloxacin (5 μ g/disc) were used as positive control to ensure the activity of standard
134 antibiotic against the test organisms. The blank discs were used as negative control. After
135 incubation, the culture plates were examined and the inhibition zones formed around each
136 disk were measured in millimeter scale as previously described [16]. Each assay in this
137 experiment was replicated three times.

138

139 **2.3.5. Determination of Minimum Inhibitory Concentration (MIC)**

140 Minimum inhibitory concentration (MIC) of methanol extract of *Trema orientalis* was
141 determined by a two-fold serial dilution method as previously described [17]. The methanol
142 crude extract of *Trema orientalis* leaves was dissolved in Nutrient broth medium in an
143 eppendorf tube (Watson Co. Ltd., Japan) to achieve a concentration of 40 mg/mL. The
144 solution of eppendorf tube was serially diluted to obtain 20, 10, 5, 2.5 and 1.25 mg/mL of
145 concentrations. The 0.5 mL of bacterial broth culture of each tested bacteria was transferred
146 to each eppendorf tube. Thus, the total amount of solution in each eppendorf tube was 1 mL.
147 The control tubes contain 0.5 mL bacterial broth cultures with 0.5 mL nutrient broth media.
148 The solution of all eppendorf tubes were mixed properly by vortexing and incubated at 37°C
149 for 24 h with continuous shaking at 250 rpm. After incubating 24 h, 100 μ L of solution from
150 each eppendorf tube were spread over the nutrient-agar-media plate. The plates were
151 incubated at 37°C for 16 h for bacterial growth and the number of colony was counted for
152 MIC determination.

153

154 **2.3.6. Brine Shrimp Lethality Bioassay**

155 Brine shrimp lethality bioassay is the most convenient system for preliminary assessment of
156 cytotoxicity of plant extracts. The brine shrimp lethality bioassay of the methanol extract of
157 *Trema orientalis* leaves was evaluated as previously described procedure against *Artemia*
158 *salina* as a test organism to monitor the cytotoxicity of a compound [18]. The eggs of Brine
159 shrimp (*Artemia salina*) were collected from an aquarium shop (Dhaka, Bangladesh) and
160 incubated for 28°C with constant oxygen supply and hatched for two days to provide a large
161 number of larvae called nauplii. The different concentrations of crude extract were prepared
162 by dissolving them in DMSO (not more than 50 µL in 5 mL solution) plus seawater (3.8%
163 NaCl in water) to attain serial dilution from 200-1.562 µg/mL. The standard vincristine
164 sulphate was used as a positive control. The varying concentration of solution of vincristine
165 sulphate was prepared by serial dilution into DMSO to attain serial dilution from 200-1.562
166 µg/mL. A vial containing 50 µL of DMSO diluted to 5 mL simulated seawater used as a
167 control. Twenty mature shrimps were placed into each of the experimental vials. After 24 h,
168 the vials were inspected using a magnifying glass, and the number of surviving nauplii in
169 each vial was counted. From this data, the percentage (%) of mortality of the brine shrimp
170 naupili was calculated for each concentration using the following formula: % Mortality =
171 $N_t/N_0 \times 100$ (Where N_t = Number of dead nauplii after a 24 h incubation; N_0 = Number of
172 total nauplii transferred i.e., 10). The LC50 (median lethal concentration) was determined
173 from the log concentration versus % mortality.

174

175 **2.3.7. Statistical Analysis**

176 The experimental results obtained from antibacterial and MIC determination assays were
177 expressed as mean \pm standard deviation (SD) of three replicates. LC50 values were
178 determined by correlation/regression analysis. Microsoft Excel 2010 statistical package was
179 used for all analyses.

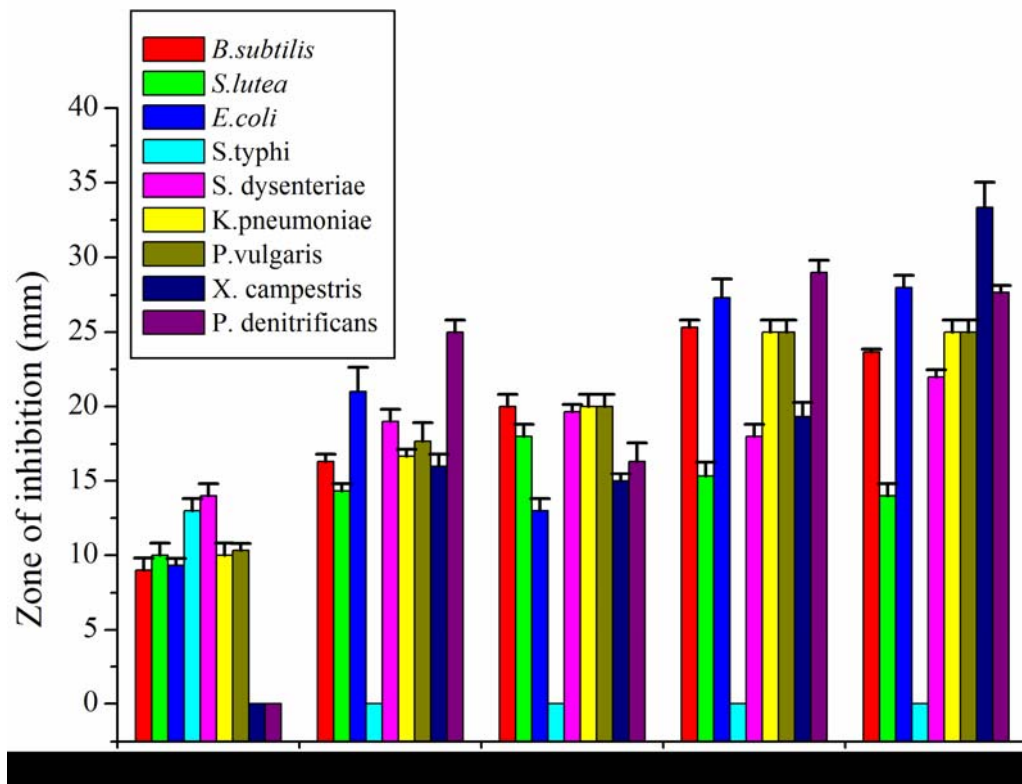
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181 3. Results

182 3.1. Antibacterial Potentialities of *Trema orientalis* Extract

183 The antibacterial activities of methanol extract of *Trema orientalis* leaves against the tested
184 bacteria were examined by the occurrence of clear zone of inhibition. The leaves extracted at
185 a concentration of 400 μ g/disc showed significant antibacterial effects against two Gram
186 positive bacteria (*Bacillus subtilis*, *Sarcina lutea*) and five Gram negative bacteria
187 (*Escherichia coli*, *Salmonella typhi*, *Shigella dysenteriae*, *Klebsiella pneumoniae*, and
188 *Proteus vulgaris*) with the zone of inhibition ranging from 9 ± 0.81 to 14 ± 0.81 mm (Figure 1).
189 The highest zone of inhibition was found 14 ± 0.81 mm and 13 ± 0.81
190 mm against *Shigella dysenteriae* and *Salmonella typhi* respectively. The inhibition zone was
191 observed 10 ± 0.81 mm against *Klebsiella pneumoniae*, *Proteus vulgaris* and *Sarcina lutea*
192 whereas zone of inhibition was 9 ± 0.47 mm against *Bacillus subtilis* and *Escherichia coli*.
193 However, no antibacterial activity was observed against two Gram negative bacteria,
194 *Xanthomonas campestris* and *Pseudomonas denitrificans* at the used concentration of
195 400 μ g/disc of plant leaves extract. Standard reference antibiotics: Tetracycline,
196 Erythromycin, Gentamicin and Ciprofloxacin were used as positive control showed higher
197 antibacterial activities than the plant leaves extract against all the tested bacteria except
198 *Salmonella typhi*. Though *Salmonella typhi* showed resistance against reference antibiotics as
199 positive control, methanol extract of *Trema orientalis* leaves exhibited strong zone of

200 inhibition (13±0.81mm) against *Salmonella typhi* (Figure 1) suggest that it could be a
 201 potential therapeutic drug candidate against *Salmonella typhi*. No zone was formed by
 202 negative control.



203
 204 **Figure 1.** Effect of methanol extract of *Trema orientalis* leaves on two Gram positive and
 205 seven Gram negative bacteria. Values are represented as mean ± SD (n=3).SD, Standard
 206 deviation; TC, Tetracycline; EM, Erythromycin; GM, Gentamycin; CF, Ciprofloxacin.

207
 208 **3.2.Minimum Inhibitory Concentration**

209 The lowest concentration of methanol extract which prevents visible growth of bacterium is
 210 the minimum inhibitory concentration. The MIC values of crude extract of *Trema orientalis*
 211 leaves were found ranging from 1.25 to 20 mg/mL (Table 1). The best MIC was 1.25 mg/mL
 212 against *Escherichia coli*, *Salmonella typhi* and *Shigella dysenteriae* as this concentration
 213 completely inhibited the growth of these bacteria. The least efficacy was shown against
 214 *Proteus vulgaris* and *Klebsiella pneumoniae* which was inhibited at 20 mg/mL concentration.

215 The moderate MIC value was shown against Gram positive bacteria (*Bacillus subtilis* and
 216 *Sarcina lutea*) which were inhibited at 10 mg/mL concentration.

217

218 **Table 1:** Minimum inhibitory concentration of methanol extract of leaves of *Trema*
 219 *orientalis*.

220

Tested bacteria	Minimum Inhibitory Concentration (mg/mL)						
	20	10	5	2.5	1.25	0.625	0.312
	Number of bacterial colonies survived at above concentration						
<i>Bacillus subtilis</i>	0	0	7±.816	17± 1.63	54±3.74	96±3.26	121±3.74
<i>Sarcina lutea</i>	0	0	12±2.16	43±4.32	66±4.39	107±4.08	144±6.53
<i>Escherichia coli</i>	0	0	0	0	0	47±3.74	133±3.74
<i>Salmonella typhi</i>	0	0	0	0	0	77±3.26	124±4.32
<i>Shigella dysenteriae</i>	0	0	0	0	0	55±4.08	112±4.08
<i>Klebsiella pneumoniae</i>	0	5±0.81	16±2.16	52±2.82	79±4.08	123±3.26	223±2.94
<i>Proteus vulgaris</i>	0	23±2.94	54±3.74	77±2.44	91±2.44	155±4.08	175±4.54

221 Values are represented as mean ± SD (n=3).

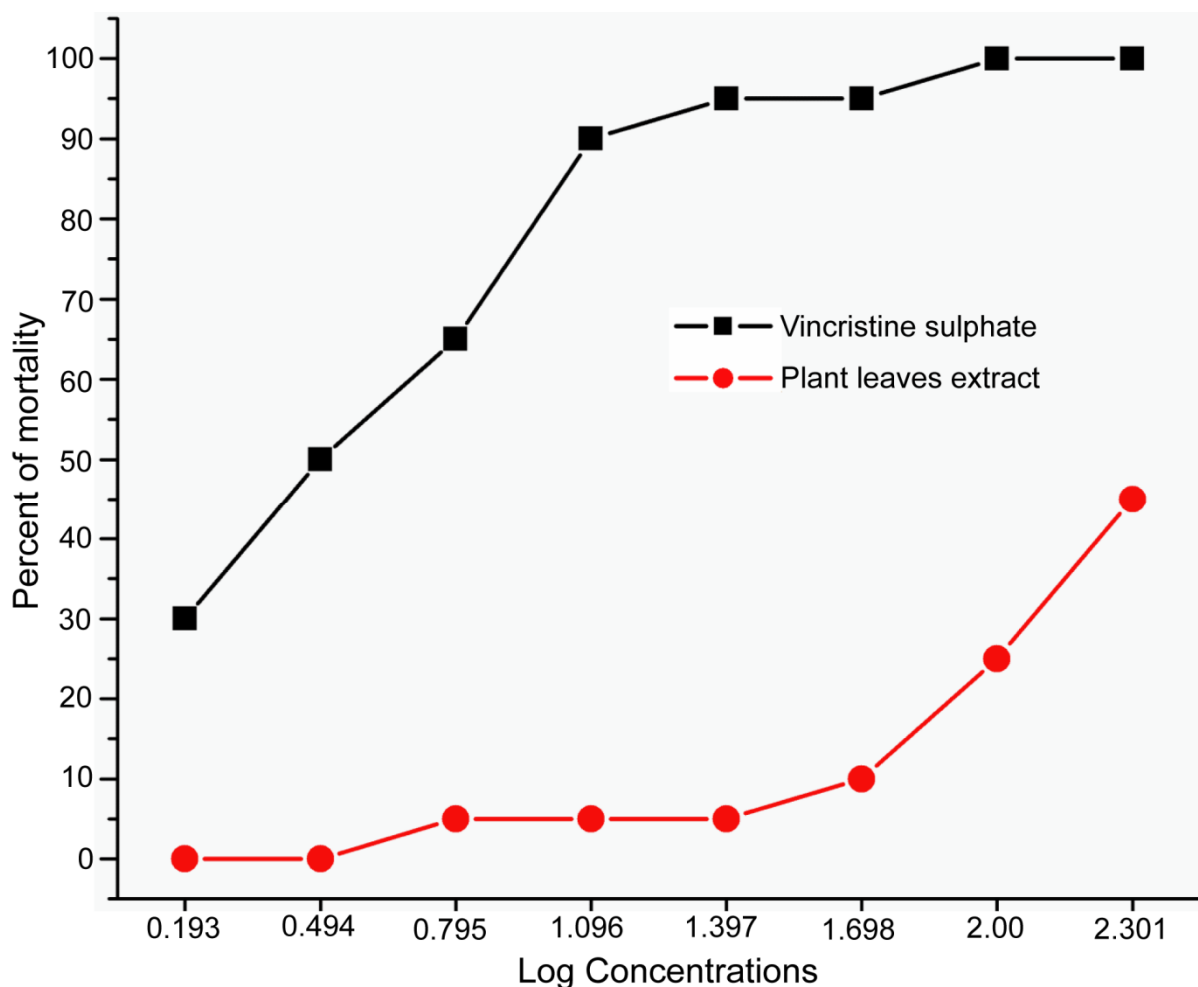
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223 3.3.Cytotoxic Activity of Methanol Extract of *Trema orientalis*

224 The percent mortality of brine shrimp naupli at different concentrations of plant extracts and
 225 vincristine sulfate as positive control are shown in Figure 2. It is clear that percent mortality
 226 of brine shrimp naupli is proportional to the concentration of extracts. The mortality rate was
 227 increased with the extract concentration increased. As shown in Table 2, methanol extract of
 228 *Trema orientalis* demonstrated a LC50 value of 170.215 µg/mL whereas vincristine sulphate
 229 showed the LC50 value of 2.477 µg/mL. This indicates that the plant leaves extract has much
 230 more higher LC50 value compared to that of vincristine sulphate. The crude methanol

231 extracts resulted in LC50 values greater than 100 µg/mL were considered non-toxic in the
232 brine shrimp lethality assay [19], support the notion that the methanol extract of *Trema*
233 *orientalis* leaves is non-toxic for host and had the potential for further investigation. There
234 was no mortality in the negative control groups indicating the test as a valid one and the
235 results obtained are only due to the activity of the tested agents.

236



237

238 **Figure 2.** Brine shrimp lethality for methanol extract of *Trema orientalis* leaves and
239 vincristine sulphate from linear correlation between log concentrations versus % mortality.

240

241

242

243

244 **Table 2.** The cytotoxicity of methanol extract of *Trema orientalis* leaves and vincristine
 245 sulphate on brine shrimp nauplii.
 246

Sample	LC ₅₀ (µg/mL)	Regression equation	R ²
Plant extract	170.215	y=29.851x + (-16.592)	0.785
Vincristine sulphate	2.477	y=33.004x+36.976	0.838

247

248 **4. Discussion**

249 Infectious diseases are the second leading cause of death worldwide. Recently, the emergence
 250 of antibiotic-resistant infection are rising very rapidly, which are major threat to human
 251 health as well as economic burden on country’s healthcare system, patients and families. The
 252 effectiveness of many conventional antibiotics is being endangered by the rapid emergence of
 253 microbial resistance to current therapeutic agents because of their overuse, misuse, and a lack
 254 of new drug development by the pharmaceutical industry [20-22]. Plant-derived natural
 255 secondary metabolites represent a potential source of antimicrobial agents which have
 256 different mode of action than conventional drug. Acceptance of medicines from natural plant
 257 product as an alternative form of healthcare system is increasing because they are serving as
 258 promising sources of novel antibiotic prototype that could be of clinical importance to
 259 improve health care [23, 24]. Therefore, we investigated the *Trema orientalis* leaves for its
 260 antibacterial and cytotoxic activities. The present study showed that the methanol extract of
 261 *Trema orientalis* leaves at a concentration of 400 µg/disc has potent antibacterial activity
 262 against both the Gram positive (*Bacillus subtilis* and *Sarcina lutea*) and Gram negative
 263 bacteria (*Escherichia coli*, *Salmonella typhi*, *Shigella dysenteriae*, *Klebsiella pneumoniae* and
 264 *Proteus vulgaris*). The activity exhibited by the methanol extract of *Trema orientalis* leaves
 265 may be attributed to the presence of some bioactive compounds in *Trema orientalis* leaves
 266 and these findings are in agreement with the previous reports that the alkaloids, phenolics,

267 triterpenoids, glycosides, tannins, saponins, *flavonoids*, *steroids* etc. are the major bioactive
268 molecules in *Trema orientalis* which have enormous potential to inhibit microbial pathogens
269 [25,26]. As shown in figure 1, methanol extract of *Trema orientalis* leaves exhibited similar
270 extent of antibacterial potentiality against both the Gram positive and Gram negative bacteria
271 indicates its broad spectrum antibacterial activity. This activity may be caused by the various
272 polar and non-polar bioactive constituents present in methanol extract of *Trema orientalis*
273 leaves because they may be acted either individually or combined to penetrate the outer
274 phospholipidic membrane of Gram negative bacteria and peptidoglycane layer of Gram
275 positive bacteria to inhibit or kill both the Gram positive and Gram negative bacteria. Indeed,
276 methanol is an amphiphilic compound that is able to extract more of the extractives of polar
277 molecules and also non-polar ones [4]. It has been reported that organic solvents use has
278 varying abilities to extract bioactive substances from medicinal plant. These observations
279 may be attributed to two reasons: firstly, the nature of biological active components whose
280 activity can be enhanced in the presence of methanol; secondly, the stronger extraction
281 capacity of methanol could have produced greater number of active constituents responsible
282 for antibacterial activity [27]. The results of the minimum inhibitory concentration showed
283 that the antibacterial activity of the methanol extract of *Trema orientalis* leaves is
284 concentration dependent. It is remarkable that *Salmonella typhi* showed antibiotic resistance
285 against all the tested commercial antibiotics that were used as positive control but *Trema*
286 *orientalis* leaves extract showed potent antibacterial effect against *Salmonella typhi*. This is
287 the most significant part of this study and indicate the necessity of natural plant products to
288 combat against growing resistance of bacteria. *Salmonella typhi* is a type of multi-drug
289 resistance (MDR) strain. In all MDR strains so far examined, multiple resistances have been
290 encoded by plasmids of the H1 incompatibility group [28]. Since the methanol extract of
291 *Trema orientalis* is found to exhibit anti-bacterial activity, the magnitude of toxicity of *Trema*

292 *orientalis* extract is safe or acceptable at the therapeutic doses must be considered. Plant
293 samples with a lower LC50 value are considered more toxic in nature. Extracts are
294 considered non-toxic if the LC50 is greater than 100 µg/mL in the brine shrimp lethality
295 assay [19]. Therefore cytotoxic assay was conducted in this study to determine the toxicity
296 profile of methanol extract of *Trema orientalis* leaves through the brine shrimp lethality
297 bioassay. Results of brine shrimp cytotoxicity were shown in Table 2 where the LC₅₀ value is
298 170.215µg/mL. This indicates that methanol extract of *Trema orientalis* leaves is not toxic for
299 host and can be a good source of potential antibacterial agents. Although several researches
300 reported the antibacterial activity of leaves and stalk [25], seed [26], and bark [29] extracts of
301 *Trema orientalis*, to the best of our knowledge, no detailed scientific proof for anti-bacterial
302 and cytotoxic activities of *Trema orientalis* leaves available yet for further using this plant for
303 the development of potential new drugs or use in a more effective form. The findings of this
304 study indicates that the extract could be used against infections caused by the tested bacteria
305 and showed a good correlation between the reported uses of *Trema orientalis* in traditional
306 medicine against infectious diseases.

307

308 **5. Conclusion**

309 The findings of the present study has revealed that the methanol extract of *Trema orientalis*
310 leaves has great antibacterial potentiality due to the presence of the compounds with high
311 antibacterial properties that can be a source of natural antibacterial agents in developing new
312 drugs as an alternative to synthetic bactericides. The cytotoxic activity exhibited by the plant
313 leaves was within the permissible limit. Isolation and characterization of the active
314 compounds could lead to a better understanding of the antibacterial mechanism for potential
315 drug candidates for the infectious diseases in future.

316

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319 Biotechnology, Jessore University of Science & Technology, Jessore-7408, Bangladesh
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321

322 **Conflict of Interest Statement**

323 All the authors declared no conflicts of interest with regard to this study.

324

325 **Authors' contributions**

326 NB and MMK collected the plant and carried out the laboratory work. MMR designed the
327 experiments, performed statistical analysis, wrote the manuscript and supervised the work.

328 MJU also performed statistical analysis and contributed to critical reading of the manuscript.

329 All authors read and approved the final manuscript.

330

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