

TRANSCRIPTION FACTOR 7 LIKE 2 (TCF7L2) VARIATION CONTRIBUTES TO VEGF ALTERATIONS IN DIABETIC RETINOPATHY

ABSTRACT

Diabetic retinopathy (DR) is a multifactorial disease which causes blindness among people with Diabetes worldwide. It has complex pathophysiology linked to various genetic variations. TCF7L2 (Transcription factor 7 like 2) is considered as one of the most important candidate genes which plays a major role in hyperglycemia and neovascularization. Neovascularization is one of the clinical symptoms of DR found to be associated with upregulation of vascular endothelial growth factor (VEGF) as established by numerous published articles. The purpose of this review is to highlight the role of TCF7L2 polymorphisms in the development of DR via alterations in VEGF expression levels. We used available published data to explain the association of TCF7L2 polymorphisms with DR. We concluded that genetic studies reports revealed TCF7L2 polymorphisms might be associated with DR development.

Keywords: DM, DR, NPDR, PDR, TCF7L2, wnt, polymorphism

ABBREVIATIONS

DM: Diabetes Mellitus

DR: Diabetic Retinopathy

NPDR: Non-proliferative DR

PDR: Proliferative DR

TCF7L2: Transcription factor 7 like 2

VEGF: Vascular endothelial growth factor

1. INTRODUCTION

Diabetes mellitus (DM) is a metabolic disorder in which chronic hyperglycemia result to defect in insulin secretion or action leading to acute and chronic complications[1]. The complication usually manifest in the form of damage to vascular system of the body and are less common in DM patients with controlled hyperglycemia [2]. Uncontrolled hyperglycemia causes impaired metabolism which may result in oxidative stress, increased lipolysis (breakdown of lipids), elevated ketone bodies and increased gluconeogenesis[3, 4]. These factors affect the body tissues and subsequently they can cause both morphological and functional defects in organs such as the heart, kidneys, liver and eye[5]. Diabetic Retinopathy (DR) is a diabetic complication which causes morphological damage to the eye leading to visual impairment and blindness[6]. DR clinically leads to retinal ischemia accompanied by hemorrhages, destruction of the pericytes referred to as microaneurysms, hard exudates, cotton wool spots, intraretinal microvascular abnormalities and neovascularization[7–9]. The early stage of DR is characterized by vascular permeability; this condition is called non-proliferative diabetic retinopathy (NPDR) whereas progression of the NPDR results in abnormal growth of the retinal blood vessels leading to the

neovascularization that is a major symptom in the advanced stage of DR, which is known as proliferative diabetic retinopathy (PDR) [10] (Fig. 1). Recent researches have shown abnormalities in the expression of glucagon like peptide-1 (GLP-1); an incretin hormone lead to hyperglycemia and vascular endothelial growth factor (VEGF), which subsequently lead to neovascularization that might be triggered as a result of variations in the gene of a transcription factor in the wnt signaling pathway referred to as transcription factor 7 like 2 (*TCF7L2*) [11–15].

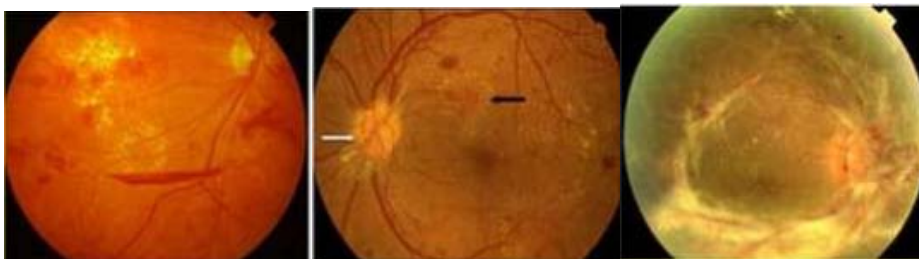
2. TRANSCRIPTION FACTOR 7 LIKE 2 (TCF7L2)

Transcription factor 7 like 2 (*TCF7L2*) also called transcription factor 4 (Tcf4) is a member of T-cell factor (Tcf)/Lymphoid enhancer factor (Lef) transcription factor family[16]. *TCF7L2* gene spans 17 exons on chromosome 10q25.3 which encodes for a transcription factor involved in the Wnt signaling pathway [16, 17](Fig 2). Several single nucleotide polymorphisms (SNPs) including rs7903146 and rs12255372 in the intron region of the *TCF7L2* were identified and found to have association with metabolic disorders including T2DM. The rs7903146 is a nucleotide change from C to T at position 112998590 in the fourth intron of *TCF7L2*, whereas rs12255372 is a change in nucleotide at position 113049143 in the fifth intron from G to T (Fig 3).

Genome-wide association studies (GWAS) reported a relationship between a common micro-satellite region (DG10S478) in intron 3 of the *TCF7L2* gene and T2DM [7, 8]. In addition, several studies identified other polymorphisms of *TCF7L2* gene associated to T2DM, amongst which are rs7903146 (C/T) and rs12255372 (G/T) [21].



Normal Background DR Moderate NPDR



Severe NPDR PDR neovascularization Fibrovascular membranes

Figure 1. Fundus images of DR progression. This progression develops from background DR (mild DR) a form of NPDR to Fibrovascular membrane a form of PDR, as the DR progresses new vessels are formed. Gradual loss in the red colour and change in vein size occur with progression of DR. This image is adopted from El-bab et al. with modification[6].

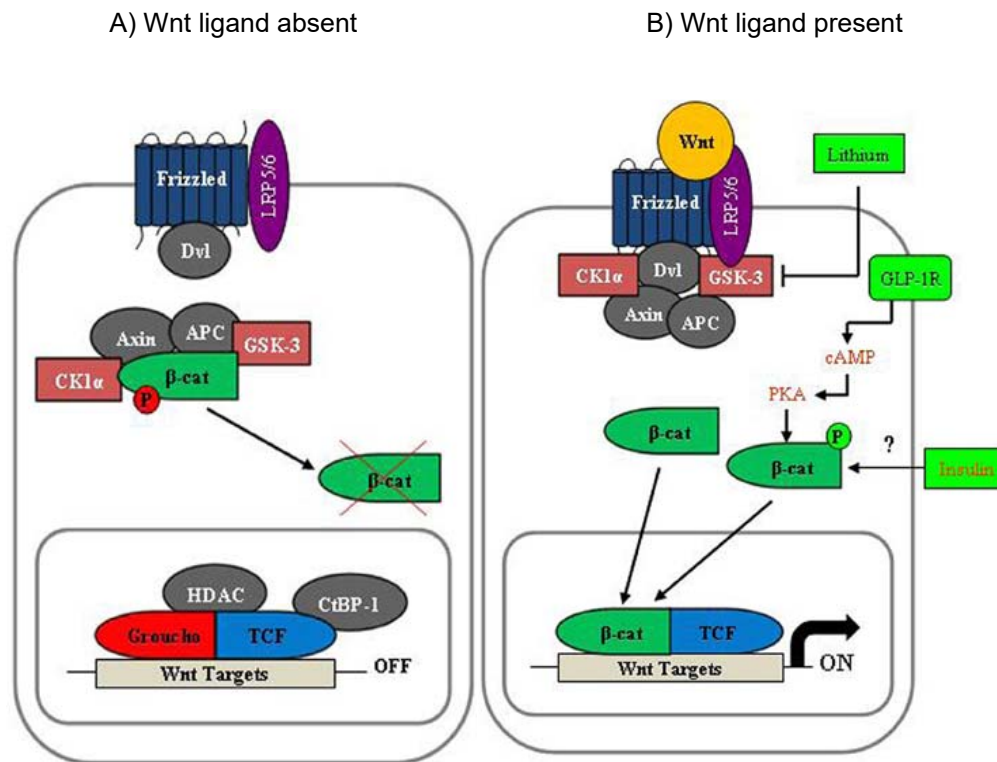


Figure 2. Canonical Wnt signaling pathway A) Shows β catenin regulation in the absence of wnt ligands. The destruction complex containing CK 1 α , axin, APC, and GSK-3 phosphorylates β catenin and mark it for proteosomal degradation B) Shows the β catenin stimulation in presence of wnt ligands which prevents interaction between the destruction complex and β catenin and subsequently promote wnt targeted gene expression. This image is adopted from Chiang et al.[12].

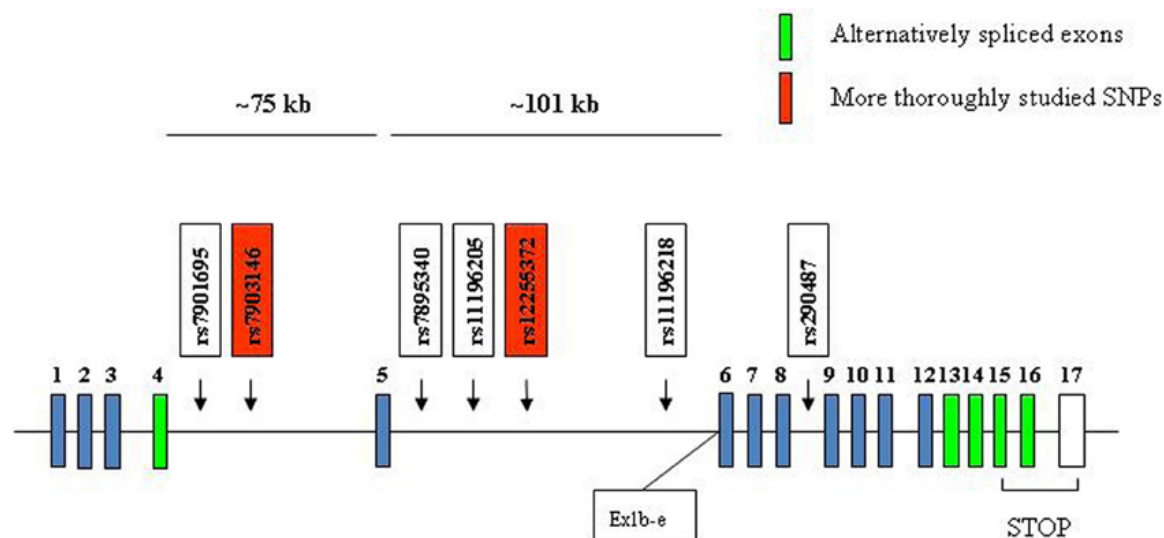


Figure 3. *TCF7L2* gene structure. The *TCF7L2* is located in chromosome 10q25.3. The blue coloured bar-shape are exons whereas the green coloured bar-shape indicate exons that undergo alternative splicing, STOP is the region where transcription of *TCF7L2* gene terminate, whereas The two single nucleotide polymorphisms (SNPs) in red are the most studied. This image is adopted from Ip et al.[22].

3. ROLE OF *TCF7L2* POLYMORPHISMS IN UPREGULATION OF VEGF LINKED DR

Genetic variation of rs7903146 (c.382-41435C>T) and rs12255372 (c.482+9017G>T) were successfully linked to T2DM in various ethnic groups [1, 13, 18, 20, 22–25]. But it is still not clear if *TCF7L2* genetic variant is related to DR [26, 27].

Although the exact mechanism of *TCF7L2* in DR development is not clearly established. However, polymorphisms in the *TCF7L2* might be associated with DR via Wnt targeted genes, several studies have reported the association of these genes with DR in different cohorts i.e. Chinese, Japanese, Indian and American population VEGF [15, 28, 29], ICAM-1 [30, 31] and eNOS[28, 32]. VEGF is a vasoactive factor and a mediator of vascular leakage; it is partly responsible for the collapse of the inner blood retinal barrier. which is upregulated in the retina in DR[29]. VEGF expression is increased in the neovascular membranes of diabetic patients with DR [33]. VEGF antagonists have been found useful in the treatment of DR[34]. The VEGF family is part of the platelet-derived growth factor (PDGF) supergene family members which consist of VEGF α , VEGF β , VEGF γ , VEGF δ , VEGF ϵ and PlGF (placental growth factor)[6, 35, 36]. VEGF α has been studied extensively and reported to play critical role in both vasculogenesis and neovascularization [37-42]. Investigation on PDR shows the relationship between *TCF7L2* and VEGF α [28].

The expression of *VEGF α* increases with increase in expression of *TCF7L2*; which might be as a result of rs7903146 (c.382-41435C>T), several studies have reported rs7903146 and rs12255372 (c.482+9017G>T) to be in linkage disequilibrium[19, 34, 35], thus showing that both rs7903146 and rs12255372 might play crucial role in the upregulation of *VEGF α* . There are two binding sites in the *VEGF α* promoter region linked to *TCF7L2*, which may implicate in increased expression of *VEGF α* transcription through *TCF7L2* binding, therefore, genetic polymorphism may lead to elevated *TCF7L2* levels which result in over expression of *VEGF α* ; related to derangement of retinal vessels and neovascularization [28]. We believe the mechanism revealing the association of *TCF7L2* polymorphism to *VEGF α* is applicable to other genes expressed by *TCF7L2* in the wnt signaling pathway.

CONCLUSION

In addition to confirming the association of *TCF7L2* gene variants to DM, *TCF7L2* SNPs might play a role in the development of DM complications including DR via upregulation of *VEGF*. Studies are required to establish the relationship between *TCF7L2* polymorphisms with other diseases associated with neovascularization such as Age related macular degeneration, diabetic macular edema, and corneal neovascularization etc.

REFERENCES

1. Dalhat MH, Bashiru I, Bello HJ, Saidu Y, and Abbas AY. Association of Transcription Factor 7 Like 2 (*TCF7L2*) rs12255372 (G/T) Gene Polymorphism and Type 2 Diabetes Mellitus. *Journal of Advances in Biology & Biotechnology*. 2017; 15 (4): 1–7.
2. Maji D. Prevention of microvascular and macrovascular complications in diabetes mellitus,” *Journal of the Indian Medical Association*. 2004; 102(8): 426–436.
3. Cade WT. Diabetes-related microvascular and macrovascular diseases in the physical therapy setting. *Physical therapy*. 2008; 88 (11):1322–1335.
4. Cunha-Vaz JG. Pathophysiology of diabetic retinopathy. *The British journal of ophthalmology*. 1978; 62(6):351–355.
5. Fowler MJ. Microvascular and Macrovascular Complications of Diabetes, *Clinical Diabetes*. 2008; 26(2):77–82.
6. El-Bab MF, Shawky N, Al-Sisi A, Akhtar M. Retinopathy and risk factors in diabetic patients from Al-Madinah Al-Munawarah in the Kingdom of Saudi Arabia. *Clinical ophthalmology (Auckland, N.Z.)*. 2012; 6:269-276.
7. Alami FM, Ahmadi M, Bazrafshan H, Tabarraei A, Khosravi A, Tabatabaiefar MA, Samaei NM. Association of the *TCF7L2* rs12255372 (G/T) variant with type 2 diabetes mellitus in an Iranian population. *Genetics and Molecular Biology*. 2012;35(2):413–417.
8. Ola MS. Edited by Mohammad Shamsul Ola. Croatia: aneza Trdine 9, 51000 Rijeka, Croatia. 2012: 249-331.
9. Kowluru RA, Zhong Q, Santos JM. Matrix metalloproteinases in diabetic retinopathy: potential role of MMP-9., *Expert opinion on investigational drugs*. 2012; 21(6):797–805.

10. Alghadyan AA. Diabetic retinopathy – An update, *Saudi Journal of Ophthalmology*.2011;25(2): 99–111.
11. Freeman JS. Role of the incretin pathway in the pathogenesis of type 2 diabetes mellitus.*Cleveland Clinic Journal of Medicine*. 2009; 76(5):12–19.
12. Chiang Y-TA., Ip W, Jin T. The role of the Wnt signaling pathway in incretin hormone production and function. *Frontiers in physiology*. 2012;(3): 273.
13. Ciccacci C, Di Fusco D, Cacciotti L, Morganti R, D'Amato C, Novelli G, Sangiuolo F, Spallone V, Borgiani P. TCF7L2 gene polymorphisms and type 2 diabetes: Association with diabetic retinopathy and cardiovascular autonomic neuropathy. *Acta Diabetologica*. 2013;5(50): 789–799.
14. Migliorini A, Lickert H. Beyond association: A functional role for Tcf7l2 in β -cell development. *Molecular metabolism*. 2015;5(4):365–366.
15. Lyssenko V, Lupi R, Marchetti P, Del Guerra S, Orho-Melandar M, Almgren P, Sjögren M, Ling C, Eriksson KF, Lethagen AL, Mancarella R, Berglund G, Tuomi T, Nilsson P, Del Prato S, Groop L. Mechanisms by which common variants in the TCF7L2 gene increase risk of type 2 diabetes. *The Journal of clinical investigation*. 2007; 117(8):2155–2163.
16. Groop L. Open chromatin and diabetes risk. *Nature Publishing Group*. 2010; 10(3):190–192.
17. Buraczynska M, Zukowski P, Ksiazek P, Kuczmazewska A, Janicka J, Zaluska W. Transcription factor 7-like 2 (TCF7L2) gene polymorphism and clinical phenotype in end-stage renal disease patients. *Molecular Biology Reports*. 2014;6(41):4063–4068.
18. Grant SFA, Thorleifsson G, Reynisdottir I, Benediktsson R, Manolescu A, Sainz J, Helgason A, Stefansson H, Emilsson V, Helgadottir A, Styrkarsdottir U, Magnusson KP, Walters GB, Palsdottir E, Jonsdottir T, Gudmundsdottir T, Gylfason A, Saemundsdottir J, Wilensky RL, Reilly MP, Rader DJ, Bagger Y, Christiansen C, Gudnason V, Sigurdsson G, Thorsteinsdottir U, Gulcher JR, Kong A, Stefansson K. Variant of transcription factor 7-like 2 (TCF7L2) gene confers risk of type 2 diabetes. *Nature genetics*. 2006; 38(3):320–323.
19. Buraczynska M, Swatowski A, Markowska-Gosik D, Kuczmazewska A, Ksiazek A. Transcription factor 7-like 2 (TCF7L2) gene polymorphism and complication/comorbidity profile in type 2 diabetes patients. *Diabetes research and clinical practice*. 2011;3(93):390–395.
20. Nanfa D, Sobngwi E, Atogho-Tiedeu B, Noubiap JJN, Donfack OS, Mofo EPM, Guewo-Fokeng M, Nguimmo Metsadjio A, Ndonwi Ngwa E, Pokam Fosso P, Djahmeni E, Djokam-Dadjeu R, Evehe MS, Aminkeng F, Mbacham WF, Mbanya JC. Association between the TCF7L2 rs12255372 (G/T) gene polymorphism and type 2 diabetes mellitus in a Cameroonian population: a pilot study.

- 174 *Clinical and translational medicine*. 2015;(4):17.
- 175 21. Dalhat MH, Bello HJ, brahim B, and Labbo A. Association of rs7903146 TCF7L2 (C/T) Gene
176 Polymorphism and Type 2 Diabetes Mellitus in Pakistani Population. *Journal of Applied Life*
177 *Sciences International*. 2017; 14(4):1–7.
- 178 22. Ip W, Chiang Y-TA, Jin T. The involvement of the wnt signaling pathway and TCF7L2 in diabetes
179 mellitus: The current understanding, dispute, and perspective. *Cell & bioscience*. BioMed Central.
180 2012;2(1):28.
- 181 23. Ip W, Chiang Y, Jin T. The involvement of the wnt signaling pathway and TCF7L2 in diabetes
182 mellitus: The current understanding, dispute, and perspective, *Cell & Bioscience*.2012. 2(1);28.
- 183 24. Bodhini D, Radha V, Dhar M, Narayani N, Mohan V. The rs12255372(G/T) and rs7903146(C/T)
184 polymorphisms of the TCF7L2 gene are associated with type 2 diabetes mellitus in Asian Indians.
185 *Metabolism: clinical and experimental*. 2007; 56(9):1174–1178.
- 186 25. Javadi MA, Katibeh M, Rafati N, Dehghan MH, Zayeri F, Yaseri M, Sehat M, Ahmadi H.
187 Prevalence of diabetic retinopathy in Tehran province: a population-based study. *BMC*
188 *ophthalmology*. 2009;9:12.
- 189 26. Sudchada P, Scarpace K. Diabetic Retinopathy : a Systematic Review,” *Genetics and Molecular*
190 *Research*.2014;13(3):5865–5872.
- 191 27 Luo J, Zhao L, Chen AY, Zhang X, Zhu J, Zhao J, Ouyang H, Luo H, Song Y, Lee J, Patel SH,
192 Shaw PX, Sadda S, Zhuo Y, Rosenfeld MG, Zhang K. TCF7L2 Variation and Proliferative Diabetic
193 Retinopathy. *Diabetes*. 2013;7(62):2613–2617.
- 194 28. Suganthalakshmi B, Anand R, R. Kim, R. Mahalakshmi, Karthikprakash S, Namperumalsamy P,
195 Sundaresan P. Association of VEGF and eNOS gene polymorphisms in type 2 diabetic
196 retinopathy. *Molecular vision*. 2006;12:336–341.
- 197 29. Awata T, Inoue K, Kurihara S, Ohkubo T, Watanabe M, Inukai K, Inoue I, Katayama S. A common
198 polymorphism in the 5'-untranslated region of the VEGF gene is associated with diabetic
199 retinopathy in type 2 diabetes. *Diabetes*. 2002; 51(5):1635–1639.
- 200 30. Sun H, Cong X, Sun R, Wang C, Wang X, Liu Y. Association between the ICAM-1 K469E
201 polymorphism and diabetic retinopathy in Type 2 diabetes mellitus: A meta-analysis. 2014: 1-6.
- 202 31. Vinita K, Sripriya S, Prathiba K, Vaitheeswaran K, Sathyabaarathi R, Rajesh M, Amali J,
203 Umashankar V, Kumaramanickavel G, Pal SS, Raman R, Sharma T, SNDREAMS project. ICAM-1
204 K469E polymorphism is a genetic determinant for the clinical risk factors of T2D subjects with
205 retinopathy in Indians: a population-based case-control study. *BMJ open*.2012;2(4):1-8.
- 206 32. Verma QUA, Han P.-Y., Nakagawa T, Johnson RJ, Grant MB, Campbell-Thompson M, Jarajapu
207 YPR, Lei B, Hauswirth WW. Diabetic eNOS-knockout mice develop accelerated
208 retinopathy.*Investigative ophthalmology & visual science*.2010;51(10):5240–5246.
- 209 33. Wu G. *Diabetic Retinopathy: The Essentials*. Lippincott Williams & Wilkins.2012: 50-400.

- 210 34. Wu Y, Zuo Y, Chakrabarti R, Feng B, Chen S, Chakrabarti S. ERK5 Contributes to VEGF
211 Alteration in Diabetic Retinopathy. *Journal of ophthalmology*. 2010; 2010:1-11.
- 212 35. Moreno A, Lozano M, Salinas P. Diabetic retinopathy. *Nutrición hospitalaria*. 2013; 28 (2):53–56.
- 213 36. Lois N, McCarter RV, O'Neill C, Medina RJ, Stitt AW. Endothelial progenitor cells in diabetic
214 retinopathy.*Frontiers in endocrinology*. 2014; 5: 44.
- 215 37. Horikoshi M, Hara K, Ito C, Nagai R, Froguel P, Kadowaki T. A genetic variation of the
216 transcription factor 7-like 2 gene is associated with risk of type 2 diabetes in the Japanese
217 population.*Diabetologia*. 2007; 50(4):747–51.
- 218 38. Kang C, Yu H, Yi GS. Finding type 2 diabetes causal single nucleotide polymorphism
219 combinations and functional modules from genome-wide association data.*BMC medical*
220 *informatics and decision making*.2013;13(1):3.
- 221 39. Clifford RL, Deacon K, Knox AJ. Novel Regulation of Vascular Endothelial Growth Factor-A
222 (VEGF-A) by Transforming Growth Factor 1: REQUIREMENT FOR Smads, -CATENIN, AND
223 GSK3. *Journal of Biological Chemistry*. American Society for Biochemistry and Molecular Biology.
224 2008;283(51):35337–35353.
- 225 40. Rangasamy S, McGuire PG, Das A. Diabetic retinopathy and inflammation: novel therapeutic
226 targets. *Middle East African journal of ophthalmology*. 2012;19(1):52–59.
- 227 41. Qazi Y, Maddula S, Ambati BK. Mediators of ocular angiogenesis. *Journal of Genetics*. 2009: 495–
228 515.
- 229 42. Muendlein A, Saely CH, Geller-Rhomberg S, Sonderegger G, Rein P, Winder T, et al. Single
230 nucleotide polymorphisms of TCF7L2 are linked to diabetic coronary atherosclerosis. *PLoS ONE*.
231 2011;6(3):2–8.

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