

Original Research Article

Selected Wild Plants Ethanol Extracts Bioactivity On

The Coagulation Cascade

ABSTRACT

Aim: As there is little data available on the validity of wild plants use in Palestine for blood disorders, the aim of this study was to determine the anticoagulant properties of *Urtica urens*, *Parietaria judica*, *Satureja thymbra*, *Thymbra spicata*, *Teucrium creticum*, *Verbascum fruticosum*, *Lupinus pilosus*, *Paronychia argentea*, and *Ruta chalepensis*.

Place and Duration of Study: Department of Biotechnology and Biology/ Faculty of Science/ An-Najah National University, between November 2015 and May 2016.

Methodology: Studied plant species ethanol extracts were prepared to final concentrations 12.5, 25 and 50 mg/ml. In vitro PT and aPTT assays were conducted on normal platelet poor plasma blood samples by a digital coagulation analyzer. Statistical analysis of the results conducted using a statistical package SPSS via applying mean values using one-way ANOVA with post-hoc tests.

Results: *Urtica urens* extract prolonged PT at 50 mg/ml, while *T. spicata* at 50 and 25 mg/ml, suggesting their inhibitory effect on the tissue clotting factors, which belong to the extrinsic pathway of the coagulation cascade. *Paronychia argentea* demonstrated a decreasing effect on PT at all studied concentrations recording zero PT, affecting the extrinsic pathway. Furthermore, *U. urens*, *T. spicata*, *P. argentea* and *P. judica* prolonged aPTT at 50 mg/ml due to the inhibition of the contact factors in the intrinsic pathway. The greatest anticoagulation activity was seen in *U. urens* and *T. spicata* as they prolonged both PT and aPTT, so they could have inhibitory effect not only on the clotting factors in the intrinsic and extrinsic pathways, but also, those in the common pathways.

Conclusion: the effective examined plant species provide potential bioactivity from which anticoagulation or anti-bleeding drugs can be exploited.

Keywords: aPTT; PT; Medicinal plants; Coagulation cascade

1. INTRODUCTION

Haemostasis is very in vital under physiological conditions in which the processes of clot formation, anticoagulation and clot dissolution are all in balance. However, the control of such process is very crucial as inappropriate blood clotting is responsible for a large number of deaths [1]. Blood-clotting (coagulation) involves platelets, enzymes and clotting factors among of which the first ones, are regarded as key regulators of both haemostasis and pathogenesis of cardiovascular diseases [2, 3]. On one hand, finding effective treatment for the atherothrombotic diseases, strokes and other cardiovascular diseases is a hot spot of concern [3]. On the other hand, the clinical limitations and adverse side effects associated with the currently used anti-thrombotic agents have fuelled the search for new, safer and effective anti-thrombotic aggregation agents of natural origin. For example, the widely used heparin anticoagulant therapy induces severe side effects over the long term [4, 5]. In addition to that the rising costs of prescription drugs in the maintenance of personal health have increased the interest in medicinal plants. Medicinal plants are rich sources of biologically active compounds [6], which are commonly found at varying concentrations in one or more different parts of a plant. They are either individually or synergistically responsible for the various therapeutic properties of medicinal plants [7]. Therefore, in recent years, naturally occurring chemical substances derived from plants have attracted interest as possible treatments for coagulation disorders and as template molecules for the development of new drugs [8].

The World Health Organization (WHO) estimates that 65-80% of the world's population use traditional medicine as their primary form of health care [9, 10]. While stressing the dangers of using traditional

medicines, it should not be forgotten that a multitude of life-saving drugs have been obtained from plants. As the herbal drugs are wide-spoken as green medicine for their safe and dependable health care paradigms, tremendous efforts have been directed towards the discovery and development of natural products with antiplatelet [11,12], anticoagulant [13,14] and antithrombotic [15] activity of the plants. Ethnobotanical survey revealed that various medicinal plants stood out to manage blood-clotting related diseases [16-18]. Moreover, epidemiologic studies have provided evidence that foods with experimentally proved antithrombotic effect could reduce risk of thrombosis. Some plants or plant parts showing thrombolytic activity have been reported [19, 20]. A recent high demand in both developed and developing countries on natural products and their derivatives has resulted in an upsurge interest in medicinal plants as an alternative medicine [21].

Several studies in Palestine have been published concerning many plant extracts biological active properties such as antibacterial, antitumor, antifungal and antioxidant of wild plants [22]. As the intrinsic and extrinsic blood coagulation pathways can be detected using various in vitro assays, including prothrombin time, partial thromboplastin time and thrombin time [23]. A study considered the anticoagulation effect of the wild plants in Palestine was conducted on *Viscum album* extracts from olive and almond host plants. It showed significant prolongation effect on PT and aPTT [24].

As there is little data available on the validity of wild plants use in Palestine for blood disorders or wound healing, the aim of this study was to determine the anticoagulant properties of *Urtica urens* L., *Parietaria judica* L. (Urticaceae), *Satureja thymbra* L., *Thymbra spicata* L., *Teucrium creticum* L. (Lamiaceae), *Verbascum fruticosum* Post (Scrophulariaceae), *Lupinus pilosus* L. (Fabaceae), *Paronychia argentea* Lam. (Caryophyllaceae), and *Ruta chalepensis* L. (Rutaceae). The anticoagulant activity in this study was performed directly on samples of human blood by determining the PT and the aPTT, which are one of the most important tests to monitor coagulation and anticoagulant effects.

Those plant species under investigation were selected for this research as many studies indicated directly or indirectly their bioactivity on the coagulation cascade, as well as due to ethnobotanical observations. For example, studies have reported that *Satureja hortensis* has blood anticoagulant activity by having inhibitory effect on blood platelet adhesion, aggregation and secretion, which might be the reason for its traditional use in treating cardiovascular and blood clot problems [25]. Therefore, the Savory species in Palestine (*S. thymbra*) was thought might have similar effect as they belong to the same genus. From the same point of view *Urtica urens* was investigated in this study as another species *U. dioica* was recorded to be the source of agglutinin protein (UDA), which causes nonspecific agglutination of erythrocytes [26]. Moreover, the presence of essential oils as Thyme which may inhibit blood clotting in *Thymbra spicata* [27] was the motive for its selection in this study. In addition to that, native people use this plant in lowering blood pressure. The other plant species under investigation in this research were used as medicinal plants by native people. Such as, *Parietaria judica*, which is known by blood herb as it is used to treat wounds. In addition, *Ruta chalepensis* is recorded as possible apportion factor among pregnant women, eliciting its prospect effect on the circulatory system. This coincided with its use in traditional medicine in menstrual problems as was proved to have antiplatelet activities [28]. The remaining examined plant species were chosen due to their other recorded bioactivities [22]. In this sense, it was expected if those plant species were examined may lead to the emergence of novel potential natural products effective on the coagulation cascade.

2. MATERIAL AND METHODS

2.1. Plant Materials

The wild plant species (*Urtica urens* L., *Satureja thymbra* L., *Verbascum fruticosum* Post, *Thymbra spicata* L., *Lupinus pilosus* L., *Teucrium creticum* L., *Paronychia argentea* Lam., *Parietaria judica* L. and *Ruta chalepensis* L.) were collected from different locations in West Bank, Palestine. The collected plant species were identified by Ghadeer Omar, Department of Biology & Biotechnology, An-Najah National University; Palestine. Representative plant specimens of the nine plant species under investigation were collected, pressed till drying, treated chemically, mounted on herbarium sheets and provided with voucher numbers, and then they were deposited at An-Najah National University herbarium. The aerial parts of plant materials were washed, air dried, ground into powder using grinder and stored at room temperature until they were used.

2.2. Plant Extraction Procedure

Ten grams of each plant powder were soaked in 100 ml of 70% ethanol for one week at room temperature with interval shaking. Then the mixtures were centrifuged for 5 min at 5000 rpm. The supernatants were evaporated by a rotary evaporator. The obtained powder of each of the plant species was dissolved in 1% dimethyl sulfoxide (DMSO) to a final concentrations equal to 50, 25, 12.5 mg/ml.

2.3. Blood Sample Preparation

Five healthy volunteers were asked to give blood samples. The volunteers were not under any medication and not smokers. The citrated blood samples were prepared as the following; each blood sample was centrifuged at 3000 rpm for 15 min to obtain the Platelets Poor Plasma (PPP) [29]. All samples were subjected to PT and aPTT assays within 2 hours after blood collection. The clotting time for both tests was recorded by a digital coagulation analyzer (Coa DATA 4004, LAberBioMedical Technologies, Germany). All measurements were carried out in triplicate. The negative control for PT and aPTT assays was 1% DMSO.

2.4. Prothrombin Time (PT) Assay

For *in vitro* PT assay, 50 μ l normal citrated (PPP) was incubated with 50 μ l from each plant extract at different concentrations (50, 25 and 12.5 mg/ml) for 5 min at 37°C. Clotting time was immediately recorded after the addition of 100 μ l PT reagent (Hemostat thromboplastin-SI. Human, Germany).

2.5. Activated Partial Thromboplastin Time (Aptt) Assay

For *in vitro* aPTT assay, 50 μ l normal citrated (PPP) was incubated with 50 μ l from each plant extract at different concentrations (50, 25 and 12.5 mg/ml) for 2 min at 37°C. Then 50 μ l aPTT reagent (Human, Germany) was added and incubated for further 3 min at 37°C. The aPTT clotting time was immediately recorded after the addition of 100 μ l calcium chloride solution (Human, Germany).

2.6. Statistical Analysis

Statistical analysis of the PT and aPTT results was conducted using a statistical package SPSS via applying mean values using one-way ANOVA with post-hoc tests to determine if there was a significant difference among the different studied wild plant species and among the different concentrations of each examined plant species extract relative to the control. P value < 0.05 was considered to be significant.

3. RESULTS

The recorded PT values of the examined volunteers blood samples showed no individual variations ($p < .01$). As a result they would be considered as representative blood samples for this study. Their PT values were examined under the effect of different plant ethanol extracts at different concentrations (12.5, 25 and 50 mg/ml). Results revealed significant anticoagulation effect of some of the examined plant species at 50 and 25 mg/ml ($p = .05$). *Urtica urens* and *T. spicata* prolonged the PT significantly ($p = .05$) relative to the control at 50 mg/ml, while *T. spicata* has had anticoagulation effect at 25 mg/ml ($p = .05$). Their effect was concentration dependent ($p = .05$). However, this dose dependent behavior was also observed in *R. chalepensis* and *S. thymbra* ($p = .05$) in spite of the absence of significant anticoagulation effect of examined concentrations.

But *P. argentea*, showed an opposite effect by decreasing the PT up to a point causing immediate blood clotting at all examined concentrations.

The detailed PT results of all effective and non-effective studied plant species are represented in Table 1. and Figure 1.

Table 1. The effect of the studied plant species extracts at different concentrations (50, 25 & 12.5 mg/ml) on PT values of five human platelets poor plasma samples.

Plant Species	Voucher number	Concentration (mg/ml)	PT (s)	*P value	**P value
<i>Lupinus pilosus</i>	659	50	20.12	.571	
		25	15.3	.865	.144
		12.5	13.24	1	
<i>Parietaria judaica</i>	1692	50	15.87	.985	
		25	12.9	1	.505
		12.5	12.5	.878	
<i>Paronychia argentea</i>	1206	50	0	.002	
		25	0	.002	.001
		12.5	0	.002	
<i>Ruta chalepensis</i>	1088	50	16.01	.982	
		25	13.27	1	.035
		12.5	12.95	1	
<i>Satureja thymbra</i>	1365	50	26.43	.065	
		25	17.24	.425	.001
		12.5	14.82	1	
<i>Teucrium creticum</i>	1531	50	19.99	.589	
		25	14.7	.951	.097
		12.5	13.01	1	
<i>Thymbra spicata</i>	551	50	33.13	.002	
		25	22.24	.011	.034
		12.5	13.68	1	
<i>Urtica urens</i>	1759	50	38.41	.001	
		25	17.1	.455	0.001
		12.5	13.89	1	
<i>Verbascum fruticosum</i>	763	50	22.5	.291	
		25	16.3	.644	.609
		12.5	13.44	.203	

*p value = .05 was significant among the different studied plant species relative to the control (blood sample without plant extract). **p value = .05 was significant among the different concentrations of each examined plant species.

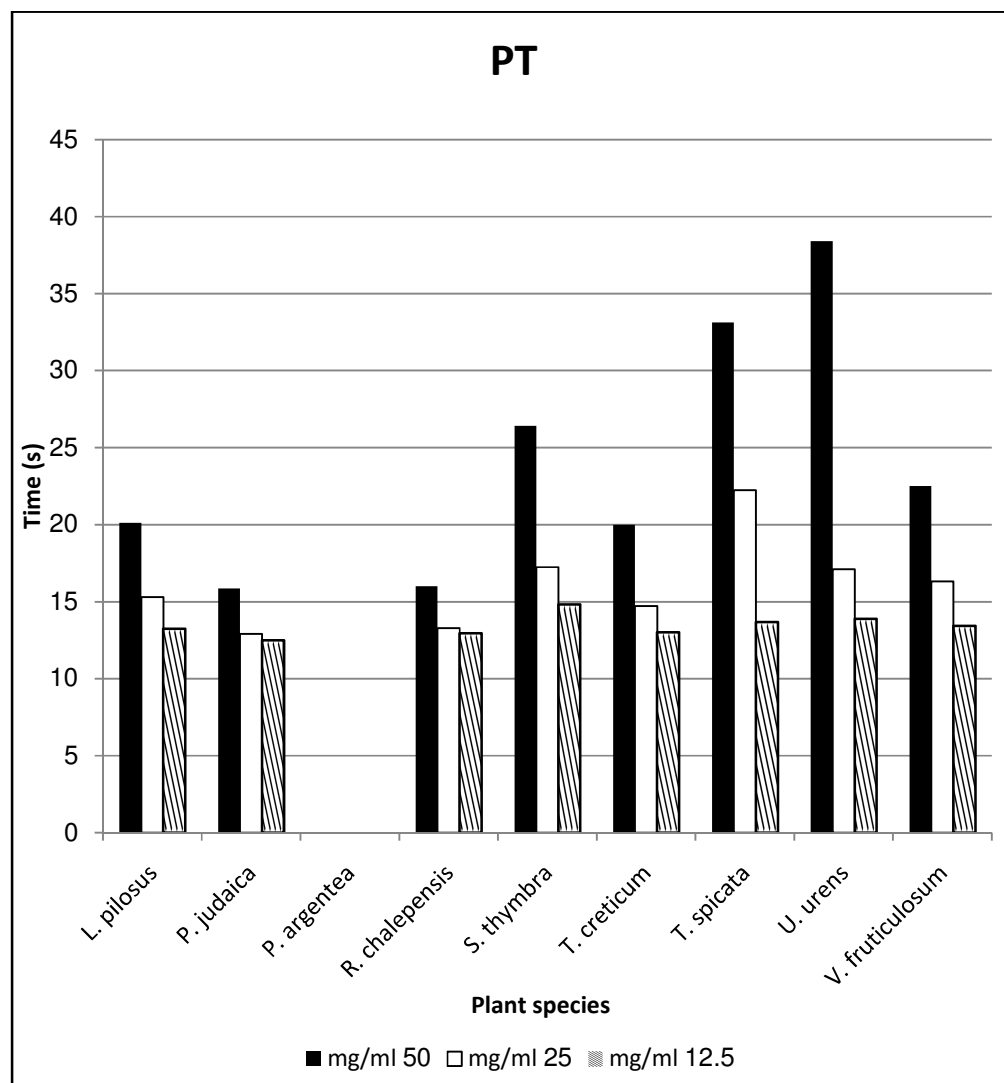


Fig. 1. The effect of the studied plant species extracts on PT values of five human platelets poor plasma samples at 50, 25 & 12.5 mg/ml concentrations.

Also the examined blood samples were considered as representative for the aPTT assay as they have had no individual variations in their recorded aPTT values ($p = .998$). *Urtica urens*, *T. spicata*, *P. argentea* and *P. judica* increased the aPTT at 50 mg/ml ($p = .05$). Among of them, *P. argentea* achieved similar significant anticoagulation effect ($p = .05$) at all other studied concentrations (12.5 and 25 mg/ml). The recorded anticoagulation effects were concentration dependent in *U. urens*, *T. spicata* and *P. judica* ($p = .05$). Detailed results of aPTT of all examined plant species extracts are shown in Table 2 and Figure 2.

Table 2. The effect of the studied plant species extracts at different concentrations (50, 25 & 12.5 mg/ml) on aPTT values of five human platelets poor plasma samples.

Plant Species	Voucher number	Concentration (mg/ml)	aPTT (s)	*P value	**P value
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		50	71.59	.968	
<i>Lupinus pilosus</i>	659	25	32.28	1	.142
		12.5	27.56	1	
		50	206.23	.015	
<i>Parietaria judaica</i>	1692	25	43.07	1	.004
		12.5	31.59	1	
		50	400	.001	
<i>Paronychia argentea</i>	1206	25	388.52	.001	.174
		12.5	289.37	.001	
		50	36.6	1	
<i>Ruta chalepensis</i>	1088	25	28.95	1	.398
		12.5	28.78	1	
		50	154.38	.362	
<i>Satureja thymbra</i>	1365	25	126.68	.329	.377
		12.5	30.93	1	
		50	23.84	1	
<i>Teucrium creticum</i>	1531	25	28.3	1	.24
		12.5	26.71	1	
		50	375.59	.001	
<i>Thymbra spicata</i>	551	25	177.02	.179	.002
		12.5	31.08	1	
		50	343.37	.001	
<i>Urtica urens</i>	1759	25	193.05	.11	.017
		12.5	65.52	.861	
		50	140.05	.237	
<i>Verbascum fruticosum</i>	763	25	29.42	1	.099
		12.5	27.84	1	

*p value = .05 was significant among the different studied plant species relative to the control (blood sample without plant extract). **p value = .05 was significant among the different concentrations of each examined plant species extract.

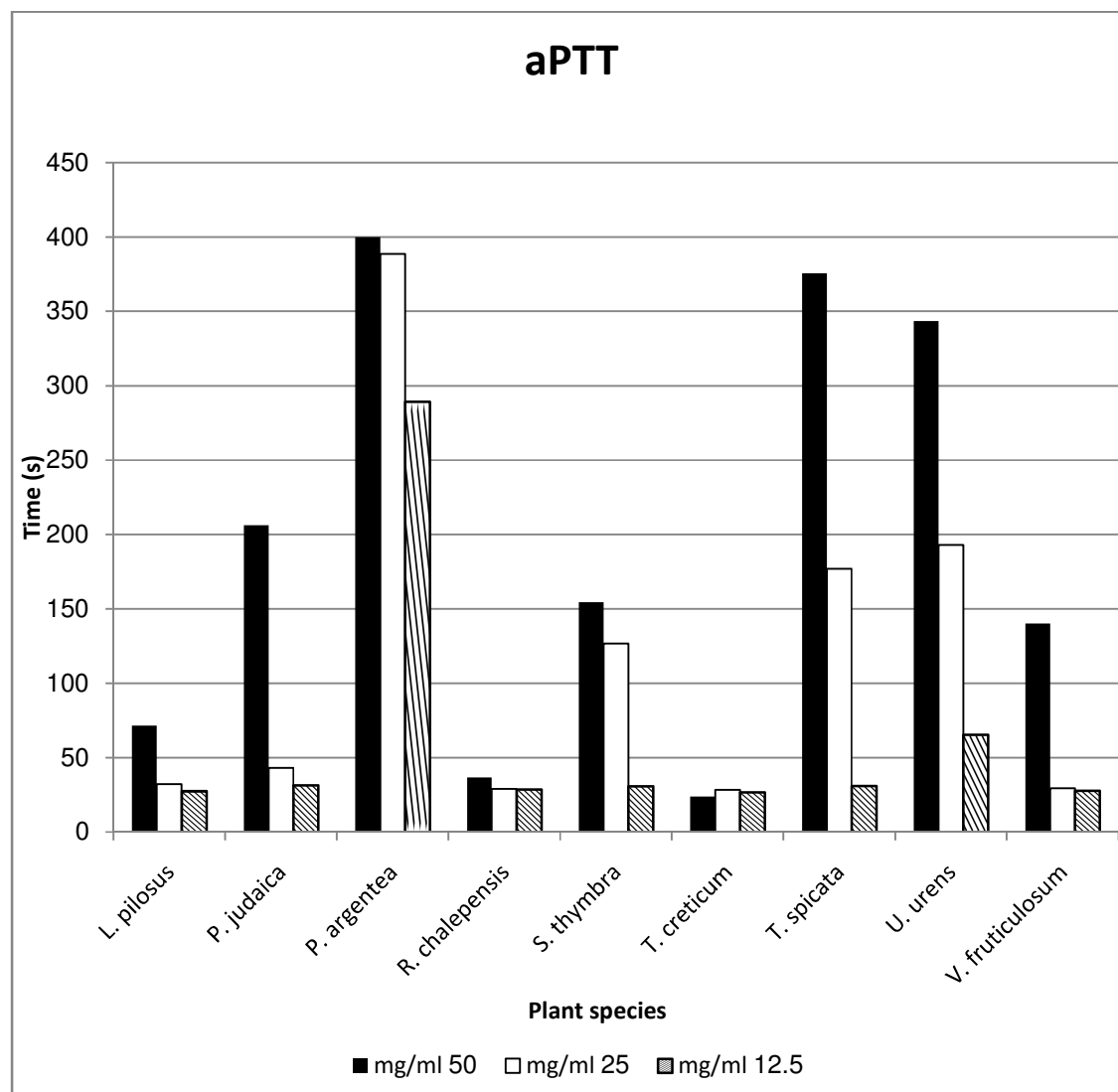


Fig. 2. The effect of the studied plant species extracts on aPTT values of five human platelets poor plasma samples at 50, 25 & 12.5 mg/ml concentrations.

4. DISCUSSION

The extrinsic (tissue factors) and/or the intrinsic (contact factors) pathway constitute the coagulation cascade revealing into blood haemostasis [30]. The standard clotting times for these two pathways are between 12.5 and 13.7 seconds for PT and between 31 and 39 seconds for aPTT [31]. Therefore, in comparison to the standard values of PT and aPTT, it is apparent that some of the examined plants in this study have had anticoagulation or coagulation effect on the examined blood samples.

Urtica urens extract prolonged PT at 50 mg/ml, while *T. spicata* one was at both 50 and 25 mg/ml. These out findings suggest that both plant extracts may have an inhibitory effect on the tissue clotting factors, which belong to the extrinsic pathway of the coagulation cascade [32, 33].

Absence of significant anticoagulation effect on the examined blood samples was observed by *S. thymbra* and *R. chalepensis* extracts. Nevertheless, they performed a concentration dependent effect at all examined concentrations on the PT, as the statistical analysis of the recorded data among the different

examined concentrations was significant ($p = 0.05$). This suggests their possible anticoagulation effect at higher concentrations.

On the contrary for those plants, *P. argentea* demonstrated a marked decreasing effect on the PT at all its studied concentrations. It caused direct immediate clotting of the examined blood samples recording zero PT. As a result, a proposed presence of tissue factors like substances or activators to tissue thromboplastin could explain the thrombotic effect of *P. argentea* on the extrinsic pathway of the coagulation cascade [33, 34]. This out finding confirms the folkloric antihemorrhagic therapy use of some plants by herbalist [35].

Furthermore, *U. urens*, *T. spicata*, *P. argentea* and *P. judica* prolonged the aPTT relative to the control significantly ($p = 0.05$). This anticoagulation bioactivity was recorded at 50 mg/ml. The observed anticoagulation effect could be referred to the inhibition of the contact factors in the intrinsic pathway of the coagulation cascade [36], including the factors XII, XI, IX, and VIII [32, 33]. The obtained data in this study coincide with previous literatures in that some plant extracts have anticoagulation effect via increasing the aPTT [33, 37]. Among the previously mentioned effective plant species extracts on aPTT, only *P. argentea* did not show significant dose dependent manner ($p > 0.05$).

The greatest anticoagulation activity was seen in *U. urens* and *T. spicata* as they prolonged both PT and aPTT, indicating that both plant species could have inhibitory effect not only on the clotting factors in the intrinsic and extrinsic pathways, but also, those in the common pathways of the coagulation cascade. Therefore, the factors X, V, II and I could be susceptible for inhibition [36]. In contrast, the common pathway in the coagulation cascade is not vulnerable to *P. argentea* extract as it increased aPTT and decreased PT.

In overall the recorded results go along with literature in that biological substances in some plants (for example polysaccharide) can cause activation or inhibition to both the intrinsic and the extrinsic pathways [37]. In addition to that, many phytochemical substances attenuated the coagulation cascade and can inhibits or decrease the activity of tissue factors or thrombin [34].

As anticoagulant medicines can reduce the mortality and hospitalization, which result from the cardiovascular thrombotic diseases, screening of some medicinal wild plants in this study was conducted to provide scientific validation to their traditional medicinal use. Moreover, it could also lead to the discovery of new pharmacologically active drugs. The fundamental results of this research that some of the studied plant species extracts posses potent anticoagulation properties, which deduce the use of herbal preparations before undergoing any surgical procedure should be rather ceased. That can be referred to their possible anticoagulation effect, which may increase the risk of bleeding episodes. Moreover, in spite of the apparent anticoagulation effect of the examined plants *in vitro* as was proven in this study, it is not conclusive as *in vivo* studies will be required to determine their true physiological effect in combination to their cytotoxicity detection.

As the extraction of different ingredients from medicinal plant materials involves the use of various solvents based on their ability to extract bioactive compounds of different solubility and polarities [38], the use of other extract types of the non-effective plant species in this study may reveal different effects, which were not obtained here. As well as, when applied to the recorded effective ones may also show different bioactivity on the different blood parameters.

4. CONCLUSION

In conclusion, the effective examined plant species provide potential bioactivity from which anticoagulation or anti-bleeding drugs can be exploited. Therefore, further analysis is recommended for the identification and extraction of the effective ingredients in the effective recorded examined plant species. Moreover, the evaluation of the active plant extracts bioactivity mechanism on the coagulation cascade is suggested. In addition, it is recommended to identify the specific clotting factors in the coagulation cascade that are most sensitive to the action of these plants.

CONSENT

As per international standard or university standard written patient consent has been collected and preserved by the authors.

REFERENCES

1. Elliot WH, Elliot DC. Biochemistry and Molecular Biology. 3ed ed. Oxford: Oxford University Press, UK; 2005.
2. Bakdash N, Williams MS. Spatially distinct production of reactive oxygen species regulates platelet activation. *Free Radic Biol Med*. 2008; 45(2): 158–166.
3. Fabre JE, Gurney ME. Limitations of current therapies to prevent thrombosis: a need for novel strategies. *Mol Biosyst*. 2010; 2:305–315.
4. Freedman M. Pharmacodynamics, Clinical indications and adverse effects of heparin. *J Clin Pharmacol*. 1992; 32: 584–596.
5. Pawlaczyk I, Czerchawski L, Pilecki W, Lamer-zarawska E, Gancarz R. Polyphenolic-polysaccharide compounds from selected medicinal plants of Astraceae and Rosaceae families: Chemical characterization and blood anticoagulant activity. *Carbohydr Polym*. 2009; 77: 568–575.
6. Palombo EA. Phytochemicals from traditional medicinal plants used in the treatment of diarrhoea: modes of action and effects on intestinal function. *Phytother Res*. 2006; 20(9):717–24.
7. Purohit S, Vyas S. Medicinal Plant Cultivation: A Scientific Approach. India: Agrobios; 2004. 624 p.
8. Lee W, Yang EJ, Ku SK, Song KS, Bae JS. Anticoagulant activities of oleanolic acid via inhibition of tissue factor expressions. *KSBMB*; 2012. 390-395 p.
9. Scott G, Springfield EP, Coldrey N. A pharmacognostical study of 26 South African plant species used as traditional medicines. *Pharm Biol*. 2004; 42: 182–213.
10. Stafford GI, Jager AK, van Staden J. Effect of storage on the chemical composition and biological activity of several popular South African plants. *J Ethnopharmacol*. 2005; 97: 107–115.
11. Demrow HS, Slane PR, Folts JD. Administration of wine and grape juice inhibits *in vivo* platelet activity and thrombosis in stenosed canine coronary arteries. *Circulation*. 1995; 91(4):1182–1188.
12. Briggs WH, Folts JD, Osman HE. Administration of raw onion inhibits platelet-mediated thrombosis in dogs. *J Nutr*. 2001; 131(10):2619–2622.
13. Leta GC, Mourão PA, Tovar AM. Human venous and arterial glycosaminoglycans have similar affinity for plasma low-density lipoproteins. *Biochim Biophys Acta*. 2002; 1586 (3):243–253.
14. Zhiguang Li, Wang H, Jiazeng L, Guangshen Z, Cunji G. Basic and clinical study on the antithrombotic mechanism of glycosaaminoglycan extracted from sea cucumber. *Chin Med J*. 2000; 113(8):706–711.
15. Rajapakse N, Jung WK, Mendis E, Moon SH, Kim SK. A novel anticoagulant purified from fish protein hydrolysate inhibits factor Xllaand platelet aggregation. *Life Sci*. 2005; 76(22):2607–2619.
16. Drew AK, Meyers SP. Safety issues in herbal medicine: implications for the health professions. *Med J Aust*. 1997; 166: 538–541.
17. Talalay P, Talalay P. The importance of using scientific principles in the development of medicinal agents from plants. *Acad Med*. 2001; 76: 238–247.
18. Hoareau L, Da Silva EJ. Medicinal plants: a re-emerging aid. *Electronic J Biotechnol*. 1999; 2: 296–300.
19. Yamamoto J, Yamada K, Naemura A, Yamashita T, Arai R. Testing various herbs for antithrombotic effect. *Nutrition*. 2005; 21(5):580–587.
20. Prasad S, Kashyap RS, Deopujari JY, Purohit HJ, Taori GM, Dagainawala HF. Effect of *Fagonia Arabica* (Dhamasa) on in vitro thrombolysis. *BMC Complem Altern M*. 2007; 7:36.
21. Hanson BA. Understanding Medicinal Plants—Their Chemistry and Therapeutic Action. New York: Haworth Herbal Press; 2005. 307 p.
22. Omar G, Abdallah LA, Ismail S, Al masri MY. Screening Of Selected Medicinal Wild Plant Extracts Antibacterial Effect as Natural Alternatives. *Int J Ind Med P*. 2013; 46 (2): 1299–1304.

23. Gou YL, Sze A, Rowlands DK, Chung YW, Chan HC. Effects of bakfoong pill on blood coagulation and platelet aggregation. *Biol Pharm Bull.* 2003; 26: 241–246.
24. Abulhasan M, Jaradat N, Abu-Hasan N, Almasri M, Abu Taha A, Rabbaa A, Natsheh N, Shalalfeh S, Najib M. Bioactivity of *Viscum album* extracts from olive and almond host plants in Palestine. *Phcog J.* 2014; 6 (2): 117–123.
25. Mihajilov-Krstev T, Radnović D, Kitić D, Stojanović-Radić Z, Zlatković B. Antimicrobial activity of *Satureja hortensis* L. essential oil against pathogenic microbial strains. *Archives of Biological Sciences.* 2010; 62 (1):159-66.
26. Kavalali GM, editor. *Urtica: The genus Urtica.* CRC Press; 2004.
27. Kiliç T. Analysis of essential oil composition of *Thymbra spicata* var. *spicata*: antifungal, antibacterial and antimycobacterial activities. *Zeitschrift Für Naturforschung C.* 2006 ; 61(5-6): 324-8.
28. Shehadeh MB, Afifi FU, Abu-Hamdah SM. Platelet aggregation inhibitors from aerial parts of *Ruta chalepensis* grown in Jordan. *Integrative Medicine Insights.* 2007; 2:35
29. Saluja H, Dehane V, Mahindra U. Platelet-Rich fibrin: A second generation platelet concentrate and a new friend of oral and maxillofacial surgeons. *Ann Maxillofac Surg.* 2011; 1(1): 53–57.
30. Chistokhodova N, Nguyen C, Calvino T, Kachirskaia I, Cunningham G, Miles D H. Antithrombin activity of medicinal plants from Central Florida. *J Ethnopharmacol.* 2002; 81: 277–280.
31. Lentner C. *Geigy Scientific Tables Vol 3 Physical chemistry, composition of blood, hematology, somatometric data.* 8th ed. Basle, Switzerland: Ciba-Geigy Limited; 1984. 234 p.
32. Adams RL, Bird RJ. Coagulation cascade and therapeutics update:Relevance to nephrology. Part 1: Overview of coagulation, thrombophilias and history of anticoagulants. *J Nephrol.* 2009; 14: 462–470.
33. Cordier W, Cromarty AD, Botha E, Steenkamp V. Effects of selected South African plant extracts on haemolysis and coagulation. *Hum Exp Toxicol.* 2012; 31(3): 250–257.
34. Cordier W, Steenkamp V. Herbal remedies affecting coagulation: A review. *Pharm Biol.* 2011; 50(4):1–10.
35. Dandjesso C, Klotoé JR, Dougnon TV, Sègbo J, Atègbo J-M, Gbaguidi F, Fah L, Fanou B, Loko F, Dramane K. Phytochemistry and hemostatic properties of some medicinal plants sold as anti-hemorrhagic in Cotonou markets (Benin). *Indian J Sci Technol.* 2012; 5(8):3105–3109.
36. Hood JL, Eby CS. Evaluation of a Prolonged Prothrombin Time. *Clin Chem.* 2008; 54(4):765–769.
37. Mengome LE, Voxeur A, Akue JP, Lerouge P, Ngouamizokou A, Moutsimbi RM. Analysis of Toxicity and Hemostatic Properties of Polysaccharides from Plants Endemic to Gabon. *BJPT.* 2014; 5(6): 186–193.
38. Liu RH. Health benefits of fruit and vegetables are from additive and synergistic combinations of phytochemicals. *Am J Clin Nutr.* 2003; 78: 517S–520S.