

2 **Attenuation of TNF- α induced liver injury by**
3 **cinnamon extract in rats.**

4
5 **Abstract**

6 **Background:** Nowadays, alternative medicinal therapy is recommended for protection and
7 management liver diseases. Cinnamon is popular flavoring ingredient, widely used in as additive
8 worldwide.

9 **Objective:** The mechanism of hepato-protective activity of water (WE) or ethanolic extracts (EE) of
10 cinnamon against carbon tetrachloride (CCl₄) induced lipid peroxidation and the hepatic injury was
11 investigated in rats.

12 **Materials and methods:** Four groups of male rats were included in this study, Group1: control,
13 Group II; CCl₄ intoxicated, Group III (CCl₄+ WE) and Group IV (CCl₄+ EE) of cinnamon.

14 **Results and Discussion:** Serum AST and ALT were elevated in rats induced by CCl₄ while oral
15 administrated with 100 mg/kg of either (WE or EE), daily showed improvement in these enzymes.
16 The levels of malondialdehyde (MDA), IL-6 and TNF- α (P<0.001) were elevated in response to CCl₄,
17 while the activities of antioxidant enzymes superoxide dismutase (SOD) and catalase (CAT) were
18 significantly decreased (P<0.001). Results showed that WE or EE improved liver functions and lower
19 hydroxyproline content. The results obtained indicated that EE has potent hepatoprotective action
20 more than water extract against CCl₄ by inhibiting release of inflammatory cytokines that enhance
21 release of proteases and neutrophil that cause liver injury. In addition, these extracts exert a protective
22 effect by lowering MDA level and induce the antioxidants capacity.

23 **Conclusion:** It is concluded that free radical-scavenging polyphenols contents inhibit production of
24 inflammatory mediators and enhancing antioxidant capacity.

25 **Keywords:** Cinnamon, CCl₄, cytokines, antioxidants.

26
27 **Author contribution**

28 This work was carried out in collaboration between all authors. 'Author YAM, TAK, SSM, and KOA
29 designed the study, performed the statistical analysis, wrote the protocol, and wrote the first draft of the
30 manuscript. 'Author WMA, KSB,SSY, managed the analyses of the study. All authors read and approved the
31 final manuscript.

32
33
34 **Introduction**

35 Liver contributes in huge numbers of the physiological process include metabolism,
36 excretion storage, detoxification, and synthesis. Signal transduction is involved in living cell
37 function [Gurdip et al., 2007], development, differentiation, apoptosis and cell death.
38 Signaling molecules including hormones, neurotransmitters and growth factors [Lee et al.,
39 2008]. Signals mediated by a growth factor involve binding to its receptor initiates a process
40 that starts with the binding with membrane or intracellular receptor [8]. The amplified signal
41 is then propagated to the nucleus, resulting in induction or repression of gene expression
42 [Peschel et al., 2006]. Mitochondrial dysfunction was suggested to be related with many
43 chronic diseases. Oxidative damage are often related with path physiology of many diseases
44 Cinnamon is widely used as natural spices taken orally with food and enhance the
45 thermogenesis [Anderson & Broadhurst, 2004; Murcia et al., 2004]. Cinnamon is one of the
46 naturally occurring cannabinoid. The biological effect of Cinnamon due to it stimulate pain
47 receptors and release of analgesic mediators. The second action explanation it to activate

48 catecholamine as adrenaline secretion and sympathomimetic effect and subsequently,
49 increases blood pressure. This indicated that the thermo genesis effect by capsicum is
50 mediated by β -adrenergic stimulation and reduction in energy expenditure.

51 Several studies revealed that supplementation of Cinnamon was effective in the treatment of
52 some disease as improve glucose tolerance in experimental diabetic animals [Murcia et al.,
53 2004]. The metabolic role of Cinnamon is attributed to its role as potentiate the interaction
54 with insulin receptor and improve the action

55 In spite of tremendous advances in modern medicine no effective drugs are available, which
56 stimulate liver functions and offers protection to the liver from the damage or help to
57 regenerate hepatic cells (Chattopadhyay, 2003). In absence of reliable liver-protective drugs
58 in modern medicine, a large number of medicinal preparations are recommended for the
59 treatment of liver disorders (Chatterjee, 2000) and quite often claimed to offer significant
60 relief. The goal of the present study was to explore the mechanism of the antioxidant and
61 hepatoprotective efficacy of water or ethanolic extracts of cinnamon against oxidative stress
62 induced by CCl_4 in rats.

63

64 **Experimental design**

65 **Animals**

66 Sixty male rats weighing (100-120 grams) were included in the current study. The animals
67 were kept at 27 ± 2 °C. Standard diet and water are given *ad libitum*.

68

69 **Preparation of cinnamon extract**

70 The cinnamon powder was obtained from the local market at Jeddah, Saudi Arabia.

71 All reagents and solvents used in this study were punched from Aldrich Company until
72 otherwise stated. The dried powder was defatted with petroleum ether (100 grams in 200 ml
73 ether). The defatted material was extracted with 95% ethanol and then vacuum dried. One
74 part of powder was extracted in boiling water then filtered and vacuum dried.

75 **Determination of total phenolics**

76 The content of phenolics in cinnamon extracts was determined according to the method
77 described by Negi & Jayaprakasha (2003). The cinnamon extracts (100 mg) was dissolved in
78 a 10 ml of a mixture of methanol: water (6:4 v/v). Cinnamon extracts (equivalent to 100 mg)
79 in 0.2ml was mixed with 1.0 ml of ten-fold diluted Folin-Ciocalteu reagent. The absorbance
80 was measured at 765nm after 30 minutes.

81

82 **Hepatotoxicity and treated groups.**

83 Animals were divided into four groups ($n = 15$). Group I (control). Group II (CCl_4) rats were
84 injected a single dose of CCl_4 in corn oil (1ml/kg B.W, s.c.) (Avijeet et al.,2008). Groups III
85 and IV were administered orally by gastric tube 100 mg/kg of aqueous or ethanolic extracts,
86 respectively, in the form of aqueous suspension once daily for 7 days, then animals were
87 administered a simultaneously single dose of CCl_4 (1ml /kg B.W, s.c.) Blood was collected,
88 serum was separated at 3500 rpm for 10 min.

89 **Serum biochemical assay**

90 Serum enzymes aspartate aminotransferase (AST) and serum glutamate pyruvate
91 transaminase (ALT) were determined by commercial kits according to (Reitman & Frankel,
92 1957).

93 **Estimation of MDA, SOD, and CAT in liver tissue**

94 Liver homogenates (5% w/v) were prepared in cold 50mM potassium phosphate buffer (pH
95 7.4) using glass homogenizer in ice. The cell debris was removed by centrifugation at 5000
96 rpm for 15 at 4°C using a refrigerated centrifuge. The clear portion for the estimation of

malondialdehyde (MDA) (Yagi and Rastogi, 1979), superoxide dismutase (SOD) (Kakkar et al., 1972) and catalase activities (Smna, 1972)

Assay of liver Hydroxyproline

According to Patiyal & katoch , (2006). Briefly, liver sections (0.2g) was hydrolyzed (in 6 mol/L HCl at 100 °C for 2 hours).Samples were incubated for 10 min in 0.05 mol/L chloramine-T at room temperature, followed by 15 min at 65 °C. The absorbance at 570 nm and resulting values compared to a Hydroxyproline standard curve. The Hydroxyproline content was expressed as ug /grams liver tissue.

Assay of inflammatory mediators (TNF α and IL-6)

The levels of inflammatory mediators (TNF α and IL-6) in liver homogenate were determined as described (De Laurentiis et al.,2010), using a specific rat ELISA. The ELISA kits were obtained from BD Biosciences, Pharmingen, San Diego, CA, USA. Determination of TNF- α and IL-6 were performed according to the manufacturer's instructions. ELISA reader. The levels of TNF- α and IL-6 were expressed as pg/mg protein.

Histopathological studies

Sections were prepared and then stained with hematoxylin and eosin dye.

Statistical analysis

Statistical analysis was performed on a PC using SPSS, V.13, (special package for social sciences). Data are presented as arithmetic mean \pm S.D., The difference among means has been analyzed by one-way ANOVA. A value of $P < 0.05$ was considered as statistically significant.

Results and discussion

Phenolic compounds of cinnamon extracts.

Results obtained showed that total phenolic of water extract was 11.5% while ethanol extract was 15.5%. The hepatoprotective effect of water or ethanol extracts on CCl₄-intoxicated rats are shown in Table 1. In the CCl₄ intoxicated group serum AST and ALT were significantly increased as compared to control group ($p < 0.001$). The elevated activities of serum AST and ALT were significantly reduced in the animals groups treated with either water or ethanolic extracts. Treatment with ethanolic extract showed highly significant activity ($P < 0.001$) than water extract. So, the ethanol extract treated group was superior to the water extract. Results obtained revealed an increase in the level of liver MDA in CCl₄- intoxicated rats compared to control group. Treatment with extracts significantly prevented this raise in levels. The activities of SOD and CAT have significantly reduced in the CCl₄-intoxicated group, while it was significantly elevated in pretreated groups with either extract. The ethanolic extract has shown more protective than water extract. As a result of CCl₄ administration, inflammation increases and the release of cytokines like IL-6 and TNF stimulated. Data in table 2 showed that the serum level of IL-6 and TNF were markedly increased as a result of CCl₄ hepatotoxicity compared with the normal control group ($P < 0.01$). Rats given water or ethanol extract diets showed a significantly lower IL-6 and TNF levels ($P < 0.05$) compared with untreated animals.

Cinnamomum Verum belongs to the family Lauraceae and possesses significant anti allergic, anti ulcerogenic, antipyretic and anesthetic activities (Kurokawa & Shiraki, 1998). The bark yields an essential oil containing cinnamaldehyde and eugenol. Several biological activities

146 such as peripheral vasodilator, antitumor, antifungal, cytotoxic and ant mutagenic activities
147 have been attributed to cinnamaldehyde (Shaughnessy& DeMarini, 2001).

148 Carbon tetrachloride (CCl₄) is being used extensively to investigate hepatoprotective activity
149 on various experimental animals (Bhathal et al., 1983). The free radical scavenging activity
150 of water or ethanolic extracts of cinnamon were evaluated.

151 Serum AST and ALT activities were used as a marker of liver damage. CCl₄ produces an
152 experimental damage (James & Pickering, 1976). The toxic metabolite CCl₃ radical is
153 produced by cytochrome p₄₅₀ which further reacts with oxygen to give trichloromethyl
154 peroxy radical.

155 Thabrew et al., (1987) found that serum transaminases return to normal with the healing of
156 hepatic parenchyma and regeneration of hepatocytes. The ethanolic extract induced
157 suppression of the increased ALT and AST activities.

158 CCl₄ produces free radical that not only directly cause damage to tissues, but also initiate
159 inflammation. Kupffer cells produce subsequently proinflammatory cytokines and activate
160 other non-parenchymal cells involved in liver inflammation. TNF- α is produced by resident
161 macrophages after CCl₄ administration and subsequently stimulates the release of cytokines
162 from macrophages and induces phagocyte oxidative metabolism and NO production (Morio
163 et al., 2001). NO is a highly reactive oxidant and it can augment oxidative stress by reacting
164 with ROS and forming peroxynitrite (Rodenas et al., 1995). Another mediator of CCl₄-
165 induced hepatic inflammation which is induced by pro-inflammatory cytokines, leading to the
166 formation of proinflammatory substrates from arachidonic acid (Planaguma et al., 2005). We
167 observed increases in the serum level of TNF- α and IL-6, which were attenuated by
168 cinnamon extracts.

169 In this study, WE or EE treated rats showed a significant restore liver functions compared to
170 CCl₄ group. Also, lowering collagen precipitation and hydroxyproline content that were
171 observed in CCl₄ rats. The histopathological examination of liver tissue support this
172 observation and showed that these extracts have a significant antifibrotic action as indicated
173 by the disappearance of collagen accumulation.

174 Reduced lipid peroxidation was revealed by a significant decrease in MDA level in water or
175 ethanol extracts pretreated groups with simultaneously a significant elevation in SOD and
176 CAT activities. Results obtained showed that ethanolic extract was more potent antioxidant
177 than water extract. The antioxidant properties of cinnamon extracts are attributable to the
178 ability of its phenolic constituents to quench reactive oxygen species. In conclusion, this
179 study suggests that ethanolic extract of cinnamon has a potent hepatoprotective activity in
180 CCl₄-induced liver injury in rats.

181 **Conclusion:** These observations were documented by biochemical results that supporting the
182 potential clinical use of cinnamon in the treatment of some hepatic disorders. Further studies
183 will be carried out to determine the types of phenol compounds that attributed to its
184 antioxidant property.

185

186 Consent is not applicable.

187

188 **Ethical Approval:**

189 As per international standard or university standard was written ethical approval has been
190 collected and preserved by the author(s).

191

192

193 **References**

194

- 195 Anderson, R. A., & Broadhurst, C. L. (2004). Isolation and characterization of polyphenol
196 type-A polymers from cinnamon with insulin-like biological activity. *Journal of Agricultural*
197 *and Food Chemistry*, 52, 65–70.
198
- 199 Avijeet J, Manish S, Lokesh D, Anurekha J, Rout S, Gupta V, Krishna L (2008). Antioxidant
200 and hepatoprotective activity of ethanolic and aqueous extracts of *Momordica dioica* Roxb.
201 Leaves. *Journal of Ethnopharmacology* 115 61–66
202
- 203 Bhathal, P.S., Rose, N.R., Mackay, I.R., Whittingham, S., (1983). Strain differences in mice
204 in carbon tetrachloride-induced liver injury. *British Journal of Experimental Pathology* 64,
205 524–533.
206
- 207 Chatterjee, T.K., (2000). *Medicinal Plants with Hepatoprotective Properties in Herbal*
208 *Opinions*, vol. III. Books and Allied (P) Ltd., Calcutta, 135.
209
- 210 Chattopadhyay, R.R., (2003). Possible mechanism of hepatoprotective activity of
211 *Azadirachta indica* leaf extract. Part II. *Journal of Ethnopharmacology* 89, 217–219.
212
- 213 De Laurentiis A, Fernandez-Solari J, Mohn C, Burdet B et al., “The hypothalamic
214 endocannabinoid system participates in the secretion of oxytocin and tumor necrosis factor-
215 alpha induced by lipopolysaccharide,” *Journal. Neuroimmunol.* Vol. 221, no. 1-2, pp. 32–41,
216 2010.
- 217 Gurdip S , Sumitra M , DeLampasona M , Cesar A.N. Catalan (2007) .A comparison of
218 chemical, antioxidant and antimicrobial studies of cinnamon leaf and bark volatile oils,
219 oleoresins and their constituents *Food and Chemical Toxicology* 45. 1650–1661
220
- 221 James GW, Pickering RW. (1976) The protective effect of a novel compound RU-18492 on
222 galactosamine induced hepatotoxicity in rats. *Drug Research* 26, 2197–2199.
223
- 224 Kakkar, P., Das, B., Visvanathan, P.N., (1972). A modified spectrophotometric assay of
225 superoxide dismutase. *Indian Journal of Biochemistry* 197, 588–590.
- 226 Kim WR, Kremers WK. Benefits of "the benefit model" in liver trans-plantation. *Hepatology*
227 2008; 48:697–698.
- 228 Kurokawa, M., Kumeda, C. A., Yamamura, J., Kamiyama, T., & Shiraki, K. (1998).
229 Antipyretic activity of cinnamyl derivatives and related compounds in influenza virus infected
230 mice. *European Journal of Pharmacology*, 348, 45–51.
231
- 232 Lan Su , Jun-Jie Yin , Denys Charles , Kequan Zhou , Jeffrey Moore Liangli . Total phenolic
233 contents, chelating capacities, and radical-scavenging properties of black pepper, corn,
234 nutmeg, rosehip, cinnamon and oregano leaf *Food Chemistry* 100 (2007) 990–997
- 235 Lee WM, Squires RH , Nyberg SL, Doo E, Hoofnagle JH. Acute liver failure: summary of a
236 workshop. *Hepatology* 2008; 47:1401–1415.
- 237 Morio, L.A., Chiu, H., Sprowles, K.A., Zhou, P., Heck, D.E., Gordon, M.K., Laskin, D.L.,
238 2010. Distinct roles of tumor necrosis factor-alpha and nitric oxide in acute liver

239 injury induced by carbon tetrachloride in mice. *Toxicol. Appl. Pharmacol.* 172, 44–51.
240
241 Murcia, M. A., Egea, I., Romojaro, F., Parras, P., Jimenez, A. M., & Martinez-Tome, M.
242 (2004). Antioxidant evaluation in dessert spices compared with common food additives.
243 Influence of irradiation procedure. *Journal of Agricultural and Food Chemistry*, 52, 1872–
244 1881.
245
246 Mancini-Filho, J., & Van-Koijj, A. (1998). Antioxidant activity of cinnamon (*Cinnamomum*
247 *Zeylanicum*, Breyne) extracts. *Bollettino Chimico Farmaceutico*, 137, 443–447.
248
249 Planaguma, A., Claria, J., Miquel, R., Lopez-Parra, M., Titos, E., Masferrer, J.L., Arroyo,
250 V., Rodes, J., 2005. The selective cyclooxygenase-2 inhibitor SC-236 reduces liver fibrosis
251 by mechanisms involving non-parenchymal cell apoptosis and PPARgamma activation.
252 *FASEB J.* 19, 1120–1122.
253
254 Peschel, W., Sanchez-Rabaneda, F., Dickmann, W., Plesehen, A., Gartiza, I., Jimenez, D.,
255 Lamuela-Raventos, R., Buxaderas, S., Codina, C., (2006). An Industrial approach in the
256 search of natural antioxidants from vegetables and fruit wastes. *Food Chemistry* 97, 137–
257 150.
258
259 Reitman, S., Frankel, S., 1957. A colorimetric method for the determination of serum
260 glutamate oxaloacetate transaminase. *American Journal of Clinical Pathology* 28, 53–56.
261 Rekka, E., Kourounakis, P.
262
263 Rodenas, J., Mitjavila, M.T., Carbonell, T., 1995. Simultaneous generation of nitric oxide and
264 superoxide by inflammatory cells in rats. *Free Radic. Biol. Med.* 18, 869–875.
265
266 Shahidi, F., Janitha, P.K., Wanasundara, P.D., (1992). Phenolic antioxidants. *Critical*
267 *Reviews in Food Science and Nutrition* 32, 67–103.
268
269 Shaughnessy, D. T., Setzer, R. W., & DeMarini, D. M. (2001). The antimutagenic effect of
270 vanillin and cinnamaldehyde on spontaneous mutation in *Salmonella* TA 104 is due to a
271 reduction in mutations at GC but not AT sites. *Mutation Research*, 480/481, 55–69.
272
273 Smna, K.A., 1972. Colorimetric assay of catalase. *Analytical Biochemistry* 47, 389–394.
274
275 Thabrew, M.I., Joice, P.D.T.M., Rajatissa, W.A., (1987). Comparative study of efficacy of
276 *Paetta indica* and *Osbeckia octandra* in the treatment of liver dysfunction. *Planta Medica* 53,
277 239–241.
278
279 Vekiari, S. A., Oreopoulou, V., Tzia, C., & Thomopoulos, C. D. (1993). Oregano flavonoids
280 as lipid antioxidants. *Journal of the American Oil Chemists_ Society*, 70, 483–487.
281
282 Yagi, K., Rastogi, R., (1979). Assay for lipid peroxides in animal tissues by thiobarbituric
283 acid reaction. *Analytical Biochemistry* 95, 351–358.
284
285
286

287 Table (1): Serum aminotransferase enzymes (ALT and AST), lipid peroxide product (Malendialdlyde)
 288 and antioxidant enzyme activities; superoxide dismutase (SOD), catalase, and of all studied groups
 289 (Mean±SD).

Animal groups	Normal Control	CCl ₄ group	WE+ CCl ₄	EE+ CCl ₄
Parameters	group			
Serum ALT IU/ml	28.4±4.56	64.0±7.86	37.9±7.14	31.6±5.08
P ₁ value	---	P<001	<0.001	<0.001
P ₂ value	---	---	<0.001	<0.05
P ₃ -value	---	---	---	<0.05
Serum AST IU/ml	32.9±4.56	74.0±7.86	36.8±7.14	31.0±5.08
P ₁ value	---	P<001	<0.001	<0.001
P ₂ value	---	---	<0.001	<0.05
P ₃ -value	---	---	---	<0.05
MDA (mmol/mg/protein) Mean±SD	3.31± 0.14	8.14± 0.57	3.94± 0.27	3.30± 0.32
P ₁ value	---	<0.001	<0.001	<0.001
P ₂ value	---	---	<0.001	<0.01
P ₃ -value	---	---	---	<0.05
SOD (MU/mg protein) Mean±SD	316.8± 13.8	209.5± 34.0	312.7± 25.8	289.3± 23.2
P ₁ value	---	<0.001	N.S	<0.001
P ₂ value	---	---	<0.001	0.01
P ₃ -value	---	---	---	0.01
(nmol/min/mg protein) Catalase Mean±SD	8899± 2667.7	3192.0± 146.6	7582.6± 1482.5	5281.3± 935.3
P ₁ value	---	<0.001	N.S	<0.001
P ₂ value	---	---	<0.001	0.01
P ₃ -value	---	---	---	0.05

290

291 WE. Water extract

EE. Ethanol extract

292 P₁ – comparison to normal control

P₃- water E versus ethanol E

293 P₂ comparison to CCl₄ intoxicated group

N.S= non significan

294

295 Table (2): Serum IL6, TNF- α and liver hydroxyproline content in the different studied groups
 296 (Mean±SD)

Parameters Groups	IL-6 (ng/mgprotein)	TNF-α (ng/mgprotein)	Hydroxyproline (µg/g protein)
Normal Mean ± SE	220 ± 38	0.13 ± 0.013	73 ± 1.6
CCl ₄ group Mean ± SE P ¹ value	1102 ± 106 <0.0001*	2.54 ± 0.122 <0.001*	304 ± 16.5 <0.001*

CCl ₄ +WE Mean ± SE P ¹ value P ² value	687 ± 75 <0.001* <0.000*	0.92 ± 0.05 0.001* 0.000*	122 ± 4.3 <0.001* <0.000*
CCl ₄ +EE Mean ± SE P ¹ value P ² value	390 ± 60 <0.001* <0.000*	0.82 ± 0.05 <0.001* <0.000*	122 ± 4.3 <0.001* <0.000*

297
298

299

WE. Water extract

EE. Ethanol extract

300

P₁ – comparison to normal control

P₃- water E versus ethanol E

301

P₂ comparison to CCl₄ intoxicated group

N.S= non significant

302

303

Fig (1) Light micrographs of mice liver treated with CCl₄, without and with

304

pretreatment with either water or ethanol extract of cinnamon compared with normal

305

control. Representative sections from (a) normal control liver (b) rats intoxicated

306

with CCl₄, showing extensive hepatocellular necrosis; and (c) pretreated treated with

307

water E and (d) pretreated with ethanol extract showing absence of hepatocellular

308

necrosis, magnification X 250.

309

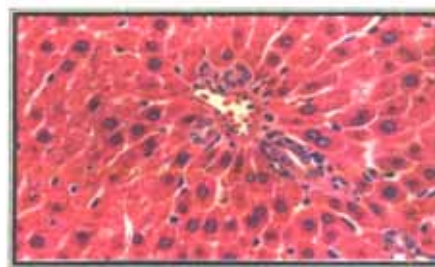
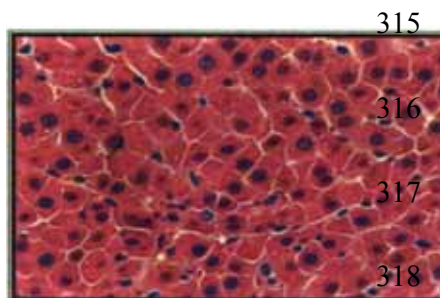
310

311

312

313

314



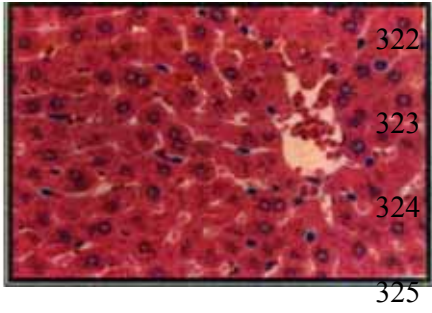
319

320

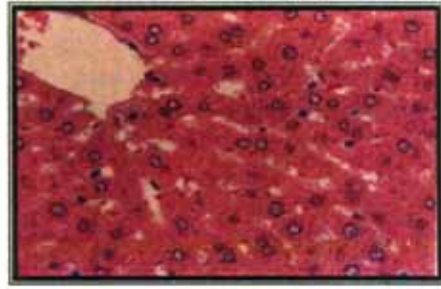
a) control group

b) CCl₄ group

321



326 c) WE+ CCl₄



d) EE+ CCl₄