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2
3 **DESIGN AND EVALUATION OF IBUPROFEN SELF NANO-EMULSIFYING**
4 **DRUG DELIVERY SYSTEM**
5

6 **ABSTRACT**

7 **Aim:** The aim of this work was to formulate self-nanoemulsifying drug delivery systems
8 (SNEDDS) for augmenting the biopharmaceutical performance of ibuprofen, a poorly water
9 soluble drug and subsequently evaluate its anti-inflammatory activity.

10 **Methodology:** Pseudoternary phase diagram studies facilitated selection of caprylic/capric
11 glycerides as the oily phase, cremophor EL as surfactants, and polyethylene glycol-400 as the
12 cosurfactant for formulating the SNEDDS. A stable combination from the phase diagram
13 consisting of 27 % *caprylic/capric* glycerides, 58 % cremophor EL and 15 % polyethylene
14 glycol-400 was loaded with ibuprofen and characterized with respect to globule size,
15 polydispersity index (PDI), stability, emulsification time, % drug loading efficiency (DLE),
16 *in vitro* drug release, infinite aqueous dilution, post dilution drug precipitation and *in vivo*
17 anti-inflammatory tests.

18 **Results:** The optimized ibuprofen SNEDDS (ibu-SNEDDS) had a mean globule size of
19 25.23nm, PDI of 0.093, showed excellent emulsification time of 5.0 s, released > 94 % of the
20 drug within 15 min while the pure drug showed only 8.8 % drug release over a period of 1 h,
21 exhibited no phase separation and demonstrated significantly ($P < 0.05$) higher anti-
22 inflammatory effect than the reference drug.

23 **Conclusion:** Our study illustrated the potential use of SNEDDS as a promising nano drug
24 carrier for the efficient delivery of ibuprofen that may solve the low bioavailability, high
25 intra- and intersubject variability frequently associated with the oral delivery of the drug.

26 **Keywords:** ibuprofen, anti-inflammatory, self-nano-emulsifying drug delivery system
27 (SNEDDS), solubility

28

29 INTRODUCTION

30 Drug bioavailability from an oral formulation in the gastrointestinal tract (GIT) is heavily
31 reliant on favourable physiochemical characteristics, including adequate solubility and
32 permeability and resistance to first pass metabolism [1]. A large majority of the newly
33 discovered chemical entities and many existing drug molecules do not meet these criteria [1,
34 2]. Of these limiting factors to oral drug delivery, low water solubility is perhaps the most
35 amenable to resolution based on the use of enabling formulation approaches [3, 4]. In
36 contrast, formulation approaches that markedly enhance intestinal permeability or reduce first
37 pass metabolism, are much less common. Permeation enhancement for oral delivery has met
38 with some moderate successes in early clinical development as described in a recent review
39 by Feeney *et al* [4]. In the case of highly (first pass) metabolised compounds, strategies such
40 as prodrugs, coadministration with inhibitors, or alternative routes of absorption, e.g.
41 pulmonary, nasal and buccal administration are more commonly employed [5]. However, for
42 many compounds with significant permeability or metabolic liabilities, parenteral
43 administration is often required for efficient delivery. For drugs where low aqueous solubility
44 limits absorption, several formulation strategies have been developed and applied to support
45 increases in dissolution rate and/or apparent solubility in the gastrointestinal tract (GIT).
46 These include particle size reduction and nanomilling, salt formation, isolation as a cocrystal
47 or high energy polymorph, the use of surfactants, cyclodextrins, generation of solid
48 dispersions, and formulation in lipid based formulations (LBFs) [2, 4, 6]. Self-
49 nanoemulsifying drug delivery system (SNEDDS) is an oral lipid based formulation. It is a
50 mixture of oil, surfactant and cosurfactant which on gentle agitation in aqueous medium

51 undergo self-emulsification to yield oil-in-water emulsions with droplet sizes of less than or
52 equal to 100 nm [7]. The major advantage of lipid based formulation (LBF), has been in
53 increasing apparent gastrointestinal solubility, it is also becoming increasingly clear that they
54 may provide advantages in permeability and, under some circumstances, in avoiding first
55 pass metabolism [4].

56 LBF confer a range of biopharmaceutical, pharmaceutical and commercial advantages.
57 Pharmaceutically, the ability to process LBF as solutions provides advantage for drugs with
58 inherently low melting points (where solid dose forms may be impractical), for low dose
59 compounds with potential content uniformity issues and for irritant and toxic compounds
60 where dust control is a challenge. Commercially, LBF provide additional patient preference
61 opportunities and in combination with a range of different finished dose forms (softgels, hard
62 capsules or lipid multiparticulates) also provide a platform for evergreening and product life
63 extension [4]. Lipids and many of the other common components of LBF (surfactants and
64 cosolvents), have been described to impact intestinal permeability, both via changes to
65 passive permeability and via inhibition of efflux transporters. Presystemic drug metabolism is
66 also avoided by drugs that are trafficked to the systemic circulation via the intestinal lymph -
67 a process that is supported by coadministration with lipids. Finally, and perhaps most
68 importantly, lipids and LBF significantly enhance the intestinal solubilisation of lipophilic
69 poorly water soluble drugs. This increases exposure and in most cases also attenuates the
70 large positive food effect commonly seen for poorly water soluble drugs after oral
71 administration. These effects stem from integration of poorly water soluble drugs into the
72 lipid digestion/absorption cascade. [8-14].

73 Ibuprofen, a propionic acid derivative, is a non-steroidal anti-inflammatory drug (NSAID). It
74 is used in the management of mild to moderate pain and inflammation in conditions such as
75 dysmenorrhoea, headache including migraine, postoperative pain, dental pain,

76 musculoskeletal and joint disorders such as ankylosing spondylitis, osteoarthritis, and
77 rheumatoid arthritis including juvenile idiopathic arthritis, peri-articular disorders such as
78 bursitis and tenosynovitis, and soft-tissue disorders such as sprains and strains. It is also used
79 to reduce fever. Ibuprofen is also used as an alternative to indomethacin in the treatment of
80 patent ductus arteriosus. The drug is practically insoluble in water and poorly absorbed from
81 the gastrointestinal tract following oral administration leading to correspondingly low
82 bioavailability [15, 16]. The aim of this study is to develop a SNEDDS that will have
83 ibuprofen intact in a solubilized form thereby culminating in invents that may maintain lumen
84 solubility and enhanced consistent absorption profile. The presence of surfactants in the
85 formulation may additionally provide a permeability-enhancement effect in the gut lumen.

86 **MATERIAL AND METHODS**

87 **Material**

88 Ibuprofen (ibu) was kindly provided as a gift sample by Pal Pharmaceutical Nigeria Ltd,
89 Cremophor EL (PEG-35-castor oil) by Gattefosse, France. Sesame oil (from Sigma Chemical
90 Co., USA) Caprylic/Capric Triglyceride (GTCC) (from Aeco Group Limited, China),
91 polyethylene glycol-400 (BDH Chemicals Ltd Poole England) were used as procured.
92 Malvern Zetasizer ZS90 (M/s Malvern Instruments, Worcestershire, UK). All other reagents
93 and solvents were of analytical grade.

94 **Methods**

95 **Solubility studies**

96 The solubility of ibuprofen in the oil, the various surfactants and co-surfactants was
97 determined. Briefly, an excess quantity of ibuprofen was added to the oil, various surfactants
98 and co-surfactants respectively and vortex-mixed for 15 min. Each suspension was
99 subsequently centrifuged. The resulting supernatant was filtered, diluted appropriately with

100 simulated intestinal fluid without enzyme (SIF). The solubilized fraction of ibuprofen in the
101 solubility samples was assayed by spectrophotometric method at the wavelength of 221 nm.

102 **Construction of pseudoternary phase diagrams**

103 Sesame oil or labrafac CC was the oil phase, Cremophor EL was the surfactant and
104 polyethylene glycol-400 was the co-surfactant. The phase titration studies were carried out by
105 water titration method for constructing the pseudoternary phase diagrams employing lipid
106 and surfactant/co-surfactant mixtures (Smix) in the ratios ranging between 1:9 and 4:1. The
107 Smix ratios of 1:0, 1:1, 2:1, 3:1 and 4:1 were explored to delineate the boundaries of
108 nanoemulsion region [17, 20]. At each ratio, the mixtures were visually observed for different
109 phases, i.e., micro/nanoemulsion, micro/nanogel, emulsion and emulgel, respectively. A
110 completely transparent appearance of the liquid system was taken up as the
111 micro/nanoemulsion, while its semisolid gel like consistency was taken up as the
112 micro/nanogel. Likewise, a liquid with milky appearance was treated as an emulsion, while
113 its semisolid form with gel like consistency was taken up as emulgel [20, 21]. The amount of
114 water at which transparency-to-turbidity transition occurs was derived from the weight
115 measurements. The results were then plotted on a pseudo-ternary phase diagram using
116 SigmaPlot 13.0 software to demarcate the nanoemulsification region. No attempts were made
117 to completely identify the other regions of the phase diagrams. Based on the results,
118 appropriate percentage of oil, surfactant and co-surfactant was selected, correlated in the
119 phase diagram and were used for preparation of SNEDDS containing ibuprofen.

120 **Formulation of ibuprofen SNEDDS**

121 Based on the stable batches obtained from the demarcated nano-emulsifying region,
122 appropriate oil, surfactant and cosurfactant were selected and used in the preparation of nano-
123 emulsifying drug delivery system containing ibuprofen. Ibuprofen was dissolved in the
124 appropriate oil in a water bath at $50\text{ }^{\circ}\text{C} \pm 5\text{ }^{\circ}\text{C}$ with frequent shaking. After complete

125 dissolution, the surfactant and cosurfactant were added and vortexed. The resultant Ibuprofen
 126 SNEDDS (ibu-SNEDDS) formulations were stored for further studies. Placebo formulations
 127 were also prepared in a similar manner without the addition of ibuprofen. The compositions
 128 of the developed ibu-SNEDDS are shown in Table 1.

129 **Table 1: Composition of the developed ibu-SNEDDS**

Components	Composition (mg)			
	A1	A2	A3	A4
Ibuprofen	400	400	400	400
Sesame oil	216	238.4	-	-
Labrafac CC	-	-	216	238.4
Cremophor EL	464	449.6	464	449.6
PEG-400	120	112	120	112

130 **Characterization of the Ibuprofen-SNEDDS**

131 *Phase separation and drug precipitation*

132 Two (2) mL samples of each of the formulation was diluted to 10 mL and 100 mL with
 133 distilled water respectively at room temperature (28 ± 3 °C), stored for a period of 24 h and
 134 observed afterwards for phase separation and drug precipitation.

135 *Assessment of emulsification time*

136 Aliquot (1) mL portion of each formulation was introduced into a beaker containing 250 mL
 137 of distilled water, maintained at 37 ± 1 °C under continuous stirring at 50 rpm. The time
 138 required to obtain a completely uniform cloudy/turbid dispersion was recorded as the
 139 emulsification time.

140 The tendency to form an emulsion was judged as ‘good’ when droplets spread easily in water
 141 and formed fine cloudy/turbid/milky dispersion, and it was judged ‘bad’ when there was poor

142 or no dispersion with immediate coalescence of oil droplets, especially when stirring was
143 stopped [22].

144 ***Centrifugation studies***

145 After 100-fold dilution with distilled water, 5 mL sample of each formulation was transferred
146 into a glass test tube and centrifuged at 4,000 rpm for 5 min in a laboratory centrifuge.
147 Thereafter, the samples were checked for physical instability, such as phase separation and
148 drug precipitation.

149 ***Loading efficiency***

150 About 1 g of each formulation was dissolved in 100 mL of 0.1N NaOH and filtered via a
151 whatman filter paper. The filtered solution was appropriately diluted and assayed for drug
152 content by spectrophotometric method at λ_{\max} of 221 nm.

153 ***Globule size determination***

154 Aliquot (1 mL) of each formulation (batches which did not exhibit phase separation or drug
155 precipitation i.e. A1, A3 and A4) was diluted 100-fold in distilled water, followed by gentle
156 mixing. The resultant mixture was then subjected to globule size analysis and polydispersity
157 index (P.I.) using a Malvern Zetasizer ZS90 (M/s Malvern Instruments, Worcestershire, UK).

158 ***Release rate determination***

159 A drug release study was carried out on the selected formulation (batch A3). The basket
160 method was adopted for this experiment. The studies were performed by dialysis bag method
161 in 500 mL of simulated gastric fluid (SGF) without pepsin (pH 1.2) for 1 h. Formulation
162 containing 400 mg of ibuprofen was filled into dialysis bags and subjected to drug release
163 studies. The drug release studies were also carried out for the pure drug for comparative
164 evaluation of the dissolution performance. The dissolution medium temperature was
165 maintained at $37\text{ }^{\circ}\text{C} \pm 1\text{ }^{\circ}\text{C}$ while the rotation speed was set at 100 rpm. Aliquots (5 mL) were
166 withdrawn at predetermined time interval, namely 5, 10, 15, 20, 30, 40, 50 and 60 min,

167 followed by replenishment with an equal volume of fresh dissolution medium. The drug
168 content was analyzed by spectrophotometric method at λ_{max} of 221 nm.

169 ***Stability studies***

170 The selected formulation (batch A 3) was stored for 6 weeks under refrigeration ($4 - 8 \pm 2$
171 $^{\circ}\text{C}$), ambient room temperature ($27 - 30 \pm 2$ $^{\circ}\text{C}$) and high temperature (45 ± 2 $^{\circ}\text{C}$) and
172 evaluated for pH, drug content, drug precipitation and emulsification time.

173 ***Anti-inflammatory studies***

174 The anti-inflammatory activity of the selected ibu-loaded SSEDDS (batch A3) was carried
175 out using the rat paw oedema test method [23]. All experimental protocols were in
176 accordance with the Ahmadu Bello University Zaria Committee on Animal Use and Care.
177 The phlogistic agent employed in the study was fresh undiluted egg albumin [22]. Adult
178 Wistar rats of either sex (weighing between 180 to 200 g) randomly divided into various
179 groups ($n = 5$ per group) as depicted in Table 4 were used for the study. The rats were fasted
180 and deprived of water for 12 h before the experiment. The deprivation of water was to ensure
181 uniform hydration and to minimize variability in oedematous response [23]. Group 1 was
182 administered distilled water and served as control. Group 2 was administered pure sample of
183 ibuprofen (6 mg/kg) dispersed in distilled water. Group 3 received placebo SNEDDS while
184 group 4 was administered ibu-SNEDDS (batch A3) with equivalent of 6 mg/kg ibuprofen
185 orally using a 1 mL syringe. Thirty minutes post treatment oedema was induced by injection
186 of 0.1 ml of fresh undiluted egg-albumin into the sub plantar region of the left hind paw of
187 each rat. The paw diameter was measured with the aid of a Vernier caliper 1, 2, 3, 4, 5 h after
188 the injection of the egg albumin. The percentage inhibition of paw edema was calculated by
189 the formula [24].

$$190 \text{ \% inhibition of paw oedema} = \frac{V_c - V_t}{V_c} \times 100 \quad (1)$$

191 Statistical analysis

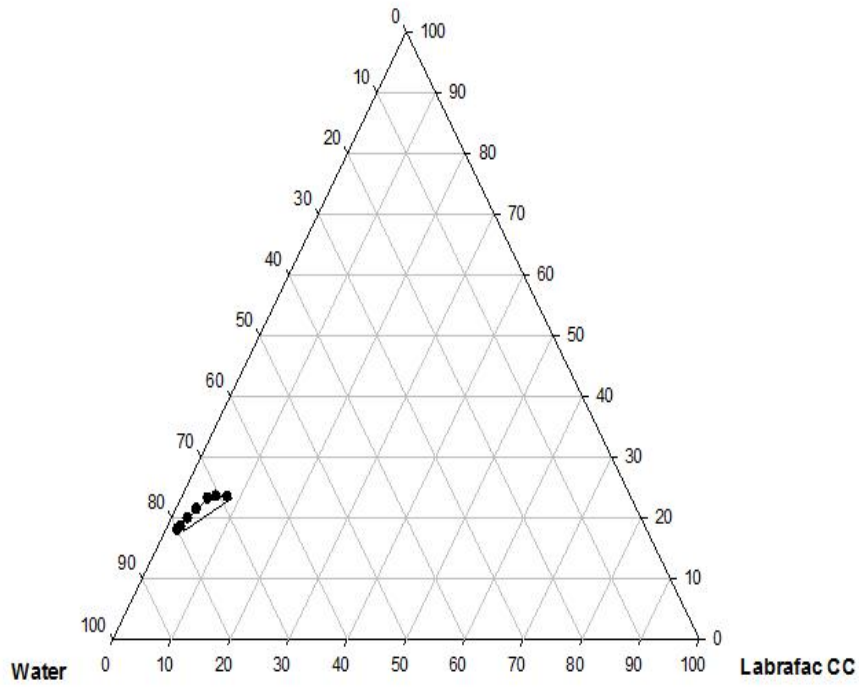
192 The data generated from the various determinations were analyzed using SPSS 20.0 software
193 (SPSS, Chicago, IL, USA) and are presented as mean \pm standard deviation (SD). The
194 differences between the data sets were determined using T-test and $p < 0.05$ was considered
195 statistically significant.

196 RESULTS AND DISCUSSION**197 Results****198 *Pseudo-ternary phase diagram***

199 Mixtures that exhibited phase separation or could not form transparent systems were
200 discarded. On the other hand those mixtures that produced transparent systems were noted
201 and a pseudoternary phase diagram plotted. The area of nanoemulsion existence is depicted in
202 Figure 1 and 2 with the delineated outline. The maximum field of self-microemulsion was
203 obtained with a surfactant - cosurfactant mixture ratio of 4:1.

204

Cremophor EL/PEG-400 (4:1 surfactant mixture)

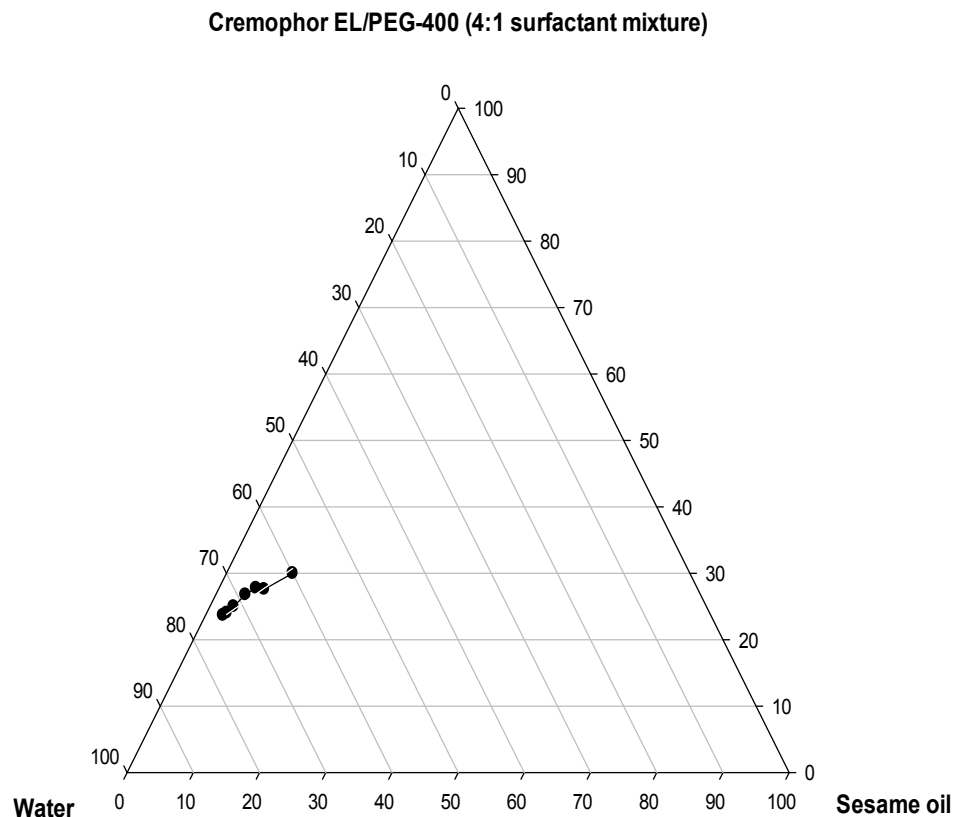


205

206 **Fig. 1: Pseudo-ternary phase diagram for cremophor EL/PEG-400 (4:1), labrafac CC**
 207 **and water**

208

209



210

211 **Fig. 2: Pseudo-ternary phase diagram for cremophor EL/PEG-400 (4:1), sesame oil and**
 212 **water**

213 *Emulsification time, phase separation, drug precipitation and loading efficiency*

214 Batch A2 exhibited drug precipitation upon storage for three (3) months and was therefore

215 dropped. It also exhibited phase separation, batch A1, A3 and A4 however past both test.

216 They all had emulsification time less than 10 s. The loading efficiency was between 96-98 %.

217 The results are as presented in Table 2.

218

219

220

221 **Table 2: Results of emulsification time, phase separation, drug precipitation and**
 222 **loading efficiency assessment of the developed ibu-SNEDDS**

Sample	Emulsification time (sec)	Phase separation	Drug precipitation	Loading efficiency
A1	8.0±0.04	No	No	97.0
A2	8.5±0.01	Yes	Yes	96.0
A3	5.0±0.05	No	No	96.0
A4	7.0±0.03	No	No	98.0

223 ***Mean globule size determination and polydispersity index***

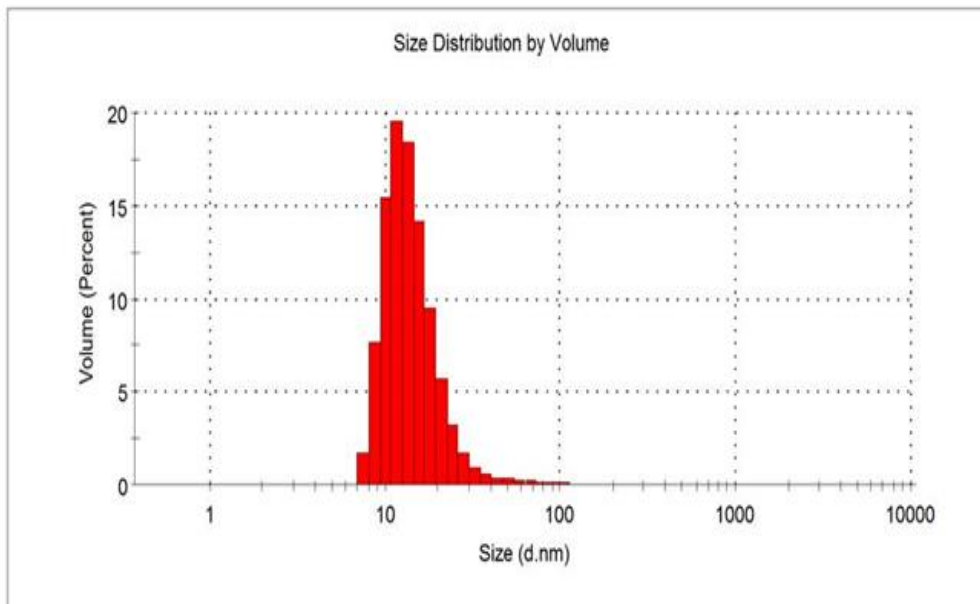
224 Figure 3, 4 and 5 provide a graphical presentation of the results of mean globule size (Z) and
 225 polydispersity index (PDI) of the formulation. The mean globule size of batch A1, A3 and A4
 226 was found to be 40.36, 25.23 and 22.18 nm respectively, all less than 100 nm, typical of
 227 SNEDDS. The polydispersity index which describes the degree of uniformity in droplet size
 228 was 0.495, 0.093 and 0.143 respectively.

229

230

Results

	Size (d.n...	% Volume:	St Dev (d.n...
Z-Average (d.nm): 40.36	Peak 1: 1527	0.0	276.9
Pdl: 0.495	Peak 2: 15.26	99.9	11.16
Intercept: 0.948	Peak 3: 4402	0.1	1210
Result quality Good			

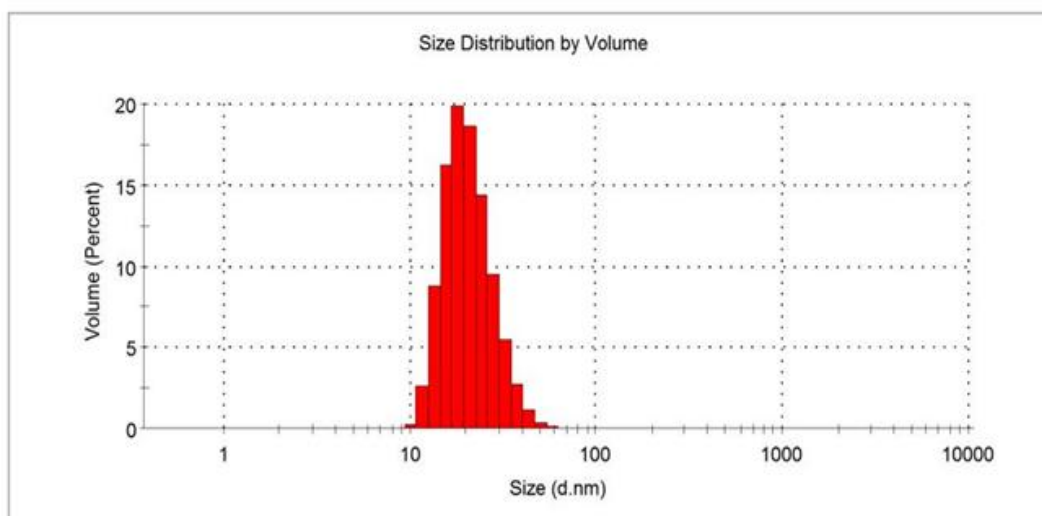


231

232 **Figure 3: Graphical presentation of globule size (Z) and polydispersity index (PDI) of**
 233 **batch A1**

Results

	Size (d.n...	% Volume:	St Dev (d.n...
Z-Average (d.nm): 25.23	Peak 1: 21.31	100.0	6.740
Pdl: 0.093	Peak 2: 0.000	0.0	0.000
Intercept: 0.956	Peak 3: 0.000	0.0	0.000
Result quality Good			



234

235 **Figure 4: Graphical presentation of globule size (Z) and polydispersity index (PDI) of**
236 **batch A3**

237

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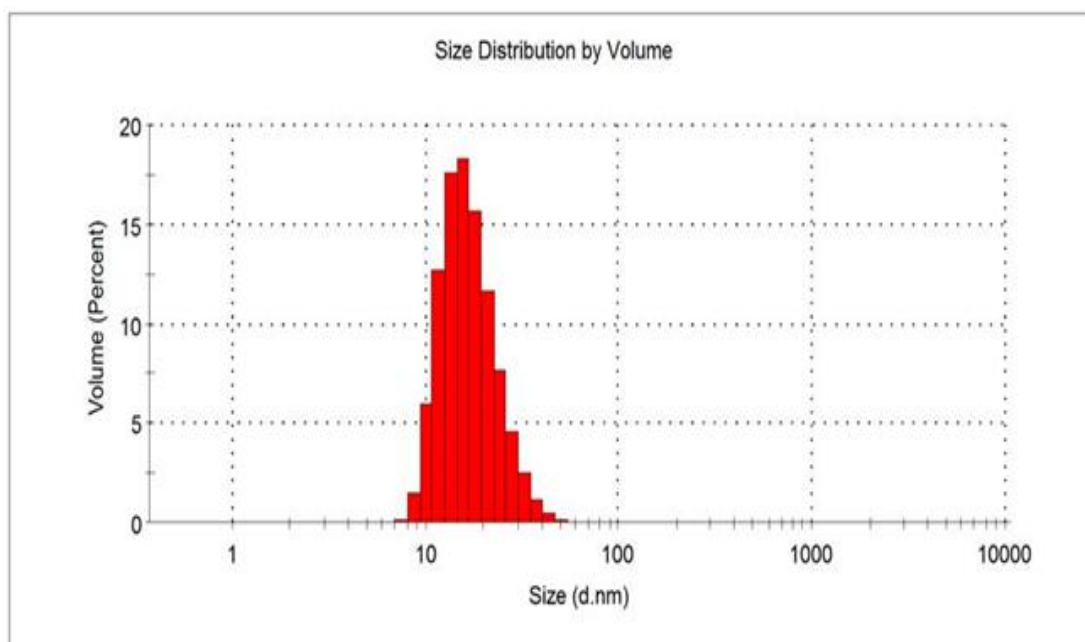
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242

Results

	Size (d.n...	% Volume:	St Dev (d.n...
Z-Average (d.nm): 22.18	Peak 1: 17.50	100.0	6.254
Pdl: 0.143	Peak 2: 4948	0.0	906.5
Intercept: 0.966	Peak 3: 0.000	0.0	0.000
Result quality Good			

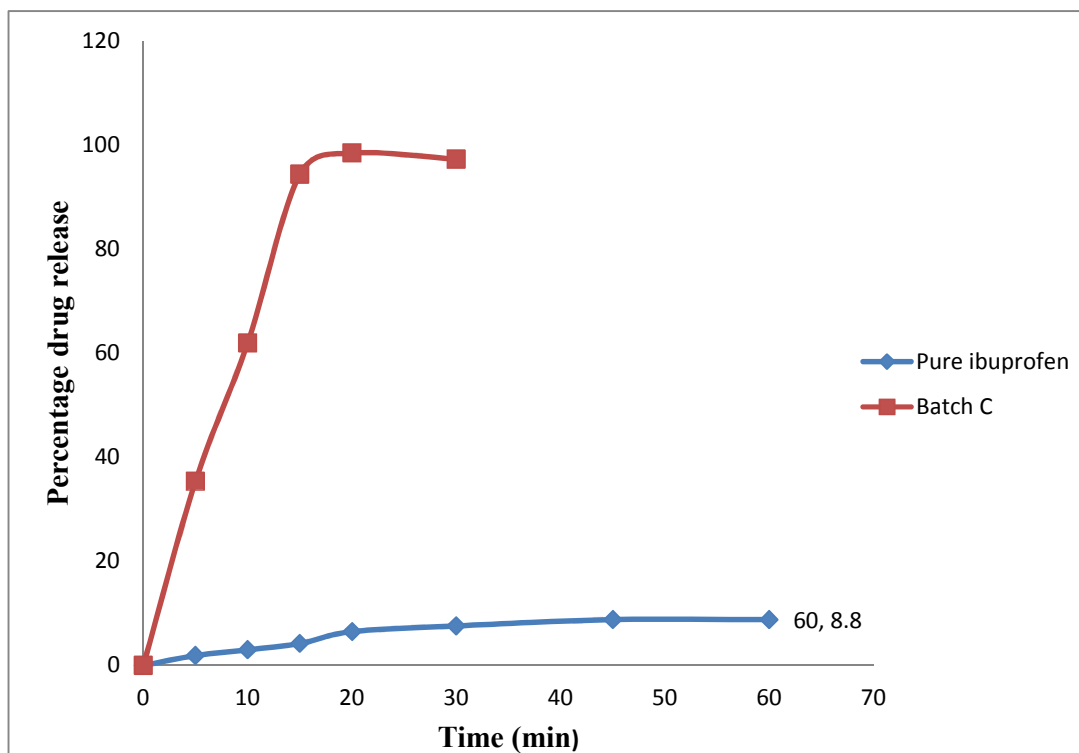


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244 **Figure 5: Graphical presentation of globule size (Z) and polydispersity index (PDI) of**
 245 **batch A4**

246 *Release rate determination*

247 The ibu-SNEDDS formulation showed marked improvement in the drug release rate
 248 compared to the pure drugs as shown in the Figure 6. The pure drug showed only 8.8 %
 249 release over a period of 60 min while about 94 % of the drug was released from the
 250 developed ibu-SNEDDS within 15 min.



251

252 **Figure 6: Plot representing percent drug released from pure ibuprofen and Batch A3**253 ***Stability studies***

254 During the 12 weeks of stability study, none of the stored batch samples showed any change
 255 in color or appearance under all storage conditions. No significant difference was found
 256 between the emulsification time of freshly prepared and stored samples. No drug
 257 precipitation was observed with any batch under all storage condition. However, there was a
 258 decreases in drug content by 3.3 % between samples stored at refrigerated or ambient
 259 temperature and elevated temperature as presented in Table 3.

260

261

262

263 **Table 3: Results of drug content, emulsification time, phase separation and drug**
 264 **precipitation assessment of the six (6) weeks old loaded SEDDS at stored under**
 265 **refrigeration, ambient temperature and elevated temperature**

Storage condition	Sample	Drug content (%)	Emulsification time (sec)	Phase separation	Drug precipitation
Refrigeration	A3	95.7	5.0±0.02	No	No
Ambient temperature (27-30±2 °C)	A3	96.0	5.0±0.30	No	No
Elevated temperature (45 ± 2 °C)	A3	92.7	6.0±0.90	No	No

266 ***Anti-inflammatory studies***

267 The results of anti-inflammatory studies are as shown in Table 4. Results showed that the
 268 developed ibu-SNEDDS exerted significantly ($P < 0.05$) higher anti-inflammatory activity
 269 than the reference ibuprofen powder ($P = 0.01$) and blank formulation (placebo) ($P = 0.00$).

270 **Table 4: Anti-inflammatory properties of ibu-SNEDDS**

S/No	Treatment	Percentage decrease in paw oedema					
		1 h	2 h	3 h	4 h	5 h	Mean
1	Aqueous artemether dispersion	2.8	16	17.2	18.6	23.6	15.6
2	Placebo SNEEDS	0.19	0.03	0.19	0.13	0.17	0.14
3	Ibu-SNEDDS	36.6	43.7	48	51.7	59.1	47.8
Aqueous artemether and ibu-SNEDDS		T-Test	Statistic 7.443	P -value 0.01***			
Placebo SNEDDS and ibu-SNEDDS		T-Test	Statistic 18.938	P -value 0.00***			

***indicates significant difference at 1% level of error

271 **Discussion**

272 ***Pseudo-ternary phase diagram***

273 This present study involved the use of pre-concentrates consisting oil, surfactants and
 274 cosurfactants and the pseudoternary diagram was only used to select the appropriate oil,

275 surfactant and co-surfactant mixtures. Phase diagram makes it easy to find out the
276 concentration range of components for the existence range of nanoemulsions. The integral
277 properties of the oil and surfactants largely determined the nature of the plot [7]. The largest
278 field of SNEDDS was obtained when labrafac CC was used as oil phase. The delineated area
279 in the phase diagram indicates the nanoemulsion existence region. The compositions of the
280 developed ibu-SNEDDS were selected from within the delineated area. The selected
281 SNEDDS yielded nanoemulsion that could withstand accelerated stress tests such as storage
282 at elevated temperature, refrigeration and centrifugation at 4000 rpm. This preconcentrate
283 would readily form microemulsion in the body on dilution with physiological fluids. These
284 systems often require high surfactant concentrations in order to provide very low interfacial
285 tension ($\leq 10^{-3}$ mN/m) and sufficient interfacial coverage to microemulsify entire oil and
286 water phases [25, 26]. The ease and degree of surface tension lowering was increased at high
287 Smix content. In order to reduce the interfacial tension to significantly low levels, a co-
288 surfactant was combined with the surfactant.

289 ***Emulsification time, phase separation, drug precipitation and loading efficiency***

290 The rate of emulsification is an important index for the assessment of the efficacy of
291 emulsification. The importance of this is that the formulation should disperse completely and
292 quickly when subjected to aqueous dilution under mild agitation [24, 27, 28]. All the batches
293 exhibited prompt and fast emulsification with the highest been 8.5 s. This indicates that the
294 formulations will disperse promptly upon contact with aqueous medium under mild agitation.
295 Phase separation and drug precipitation is a huge threat to the stability of the formulations
296 [7]. Since the formation of nanoemulsion from SNEEDS is a spontaneous process, the
297 formulation should possess considerable stability against creaming, cracking and
298 precipitation. All except batch A2 demonstrated stability (absence of phase separation and
299 drug precipitation) after storage for 48 h and after appropriate dilutions. In addition, absence

300 of drug precipitation or phase separation upon centrifugation further confirmed stability. This
301 confirmed high degree of physical stability and robust nature of the prepared formulations.
302 The observed drug precipitation in batch A2 indicates that the formulation have low drug
303 loading capability. The batch contained relatively high percentage of oil and sesame oil as it
304 oily phase, this may be responsible for it low drug loading capability. The batches had
305 loading efficiency values that fell within 96 to 98 %. This means that the drug was well
306 encapsulated within the oil droplets.

307 *Mean globule size determination and polydispersity index*

308 All the formulations exhibited globule size in the nanometric range. Batch with labrafac CC
309 had smaller globule size (25.23 nm and 22.18 nm) and PDI (0.093 and 0.143 respectively)
310 than the formulation containing sesame oil as the oily phase which had a droplet size of 40.36
311 nm and a PDI of 0.495. This result is in consonant with the report that labrafac CC has a
312 relatively shorter triglyceride chain, which is the reason behind the smaller mean droplet size
313 of microemulsions formulated with it [29, 30]. Droplet size distribution following self-
314 nanoemulsification is a critical factor to evaluate a self-nanoemulsion system. Droplet size is
315 thought to have an effect on drug absorption. The smaller the droplet size, the larger the
316 interfacial surface area will be provided for drug absorption [31 - 34]. Besides, larger sizes
317 may be predisposed to early drug precipitation prior to absorption. Polydispersity is the ratio
318 of standard deviation to the mean droplet size and is inversely proportional to droplet size
319 uniformity; the higher the polydispersity the lower the uniformity of droplet size [7].

320 Based on the results of the above investigations, batch A3 was chosen as the optimum
321 formulation on the basis of possession of minimal globule size and emulsification time (i.e.,
322 necessary for faster solubilization and absorption of drugs) [20].

323 *Release rate determination*

324 *In vitro* release studies are performed to determine the rate at which the drug in a formulation
325 is released into the dissolution medium and to also have an idea about the self-emulsification
326 efficiency of the developed system. There was a marked improvement in the drug release rate
327 from the optimized ibu-SNEDDS as compared to the pure drug. This confirmed that the
328 optimized formulation is markedly better than the pure drug. Over 94 % of the drug was
329 released within 15 min for the optimized formulations, while pure drug showed only 8.8 %
330 release over a time period of 1 h. At 20 min, 98.5 % of the drug was released from the ibu-
331 SNEDDS while the pure drug only showed 6.5 % drug release representing about 15-fold
332 increase over the pure sample. Significant improvement in dissolution rate indicated
333 improved solubilization of the drug in the aqueous media ostensibly owing to spontaneous
334 emulsification of the lipidic and emulsifying agents to produce the ultrafine emulsions by
335 micellar solubilization [20, 35]. The developed SNEDDS is expected to quickly present
336 ibuprofen in solubilized form in gastric fluids after ingestion and would provide large
337 interfacial area for ibuprofen absorption.

338 ***Stability studies***

339 During the 12 weeks of stability study, there was no change in any of the physical parameters
340 - phase separation, drug precipitation, appearance and smell of the developed ibu-SNEDDS
341 under all storage conditions. This indicates the stability of the formulation. Also, there was no
342 significant difference in the ibuprofen content at zero time and through the 12 - week stability
343 study period under refrigeration and ambient storage conditions. This indicates that ibuprofen
344 is chemically and physically stable in the formulation. For samples stored at elevated
345 temperature, there was about 3.3 % decrease in drug content when compared with the drug
346 content at time zero, this is expected since the rate of degradation is markedly influenced by

347 temperature. At high temperatures, reactions may take place which are not significant at
348 normal temperatures [36].

349 *Anti-inflammatory studies*

350 Ibuprofen is a known poorly soluble drug that may suffer from inconsistent bioavailability
351 owing to inconsistent dissolution and absorption. It is well established that dissolution is the
352 rate limiting step to absorption [24]. The improved aqueous solubility was a key factor to the
353 improved bioavailability and consequently anti-inflammatory activity. Poor drug dissolution
354 in the gastrointestinal tract (GIT) was probably responsible for the observed relatively low
355 anti-inflammatory activity of the reference drug. Other than poor water solubility, some drugs
356 are known to be susceptible to the degradation effect of stomach acid [7, 37]. SNEDDS
357 emulsify into nano droplets that offer gastro-protection to the entrapped drug solution and
358 thus prevent contact between the drug and stomach acid. This may also have contributed to
359 the observed increased in anti-inflammatory activity of the ibu-SNEDDS. Lipid base
360 formulations have been widely reported to promote lymphatic drug transport. Drug transport
361 via the lymphatic system avoids first pass effect and may consequently result to increased
362 plasma concentration and faster on set of action [2, 4, 6]. The higher anti-inflammatory
363 activity of ibu-SNEDDS is a combined result of the nanosize of the nanoemulsion, increase
364 dissolution rate of ibuprofen which would ease prompt absorption likely enhancement in
365 bioavailability due to the lipidic nature of the formulation, protection of the drug from the
366 acidic environment of stomach the stomach.

367 **Conclusions**

368 SNEDDS containing poorly water-soluble drug, ibuprofen, was prepared and optimized by
369 using *in vitro* parameters like globule size, polydispersity index and emulsification time. The
370 components and their ratio ranges for the formulation were determined by pseudo-ternary

371 phase diagram construction. The optimum formulation contains Labrafac CC as oil phase,
372 Cremophor EL as a surfactant, and PEG-400 as cosurfactant. The formulation consisted of 27
373 % caprylic/capric glycerides, 58 % cremophor EL and 15 % polyethylene glycol-400, yielded
374 SNEDDS with a globule size of 25.23 and a PDI 0.093, and had sufficient drug loading and
375 rapid self-emulsification in aqueous media. This optimized SNEDDS showed good *in vitro*
376 release with about 15-fold increase over the pure ibuprofen sample. *In vivo* anti-inflammatory
377 efficacy results showed that the developed ibu-SNEDDS exerted significantly ($P < 0.05$)
378 higher anti-inflammatory activity than the reference ibuprofen powder. Our study illustrated
379 the potential use of SNEDDS as a promising nano drug carrier for the efficient delivery of
380 ibuprofen.

381 **ETHICAL APPROVAL**

382 The authors declare that "Principles of laboratory animal care" (NIH publication No. 85-23,
383 revised 1985) were followed. All experiments have been examined and approved by the
384 Ahmadu Bello University Zaria, Nigeria, Committee on Animal Use and Care.

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