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<u>Original Research Article</u> Attenuation of TNF-α induced liver injury by

cinnamon extract in rats.

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3

5 Abstract

6 *Background:* Nowadays, alternative medicinal therapy are recommended for protection and 7 management liver diseases. Cinnamon is popular flavoring ingredient, widely used in as additive 8 worldwide.

9 *Objective:* The mechanism of hepato-protective activity of water (WE) or ethanolic extracts (EE) of 10 cinnamon against carbon tetrachloride (CCl₄) induced lipid peroxidation and hepatic injury was 11 investigated in rats.

Materials and methods: Four groups of male rats were included in this study, Group1: control,
 Group II; CCl₄ intoxicated, Group III (CCl₄+ WE) and Group IV (CCl₄+ EE) of cinnamon.

Results and Discussion: Serum AST and ALT were elevated in rats induced by CCl_4 while oral

administrated with 100 mg/kg of either (WE or EE), daily showed improvement in these enzymes. The levels of MDA ,IL-6 and TNF- α (P<0.001) were elevated in response to CCl₄ , while the activities of antioxidant enzymes superoxide dismutase (SOD) and catalase (CAT) were significantly decreased (P<0.001) . Results showed that WE or EE improved liver functions and lower hydroxyproline content. The results obtained indicated that EE have potent hepatoprotective action

19 hydroxyproline content. The results obtained indicated that EE have potent hepatoprotective action 20 more than water extract against CCl_4 by inhibiting release of inflammatory cytokines that enhance

release of proteases and neutrophil that cause liver injury. In addition, these extracts exert a protective

effect by lowering MDA level and induce the antioxidants capacity.

23 Conclusion: It is concluded that, free radical-scavenging polyphenols contents inhibit production of

24 inflammatory mediators and enhancing antioxidant capacity.

- 25 *Keywords*: Cinnamon, CCl₄, cytokines, antioxidants.
- 26

27 Author contribution

28 Thiswork was carried out in collaboration between all authors. 'Author YAM, TAK, SSM, KOA and

designed the study, performed the statistical analysis, wrote the protocol, and wrote the first draft of the manuscript. 'Author WMA, KSB,SSY, managed the analyses of the study. All authors read and approved the final manuscript.

32 33

34 Introduction

35 Liver contribute in huge numbers of physiological process include metabolism, excretion storage, detoxification and synthesis.Signal transduction is involved in living cell function 36 37 [Gurdip et al., 2007], development, differentiation, apotosis and cell death. Signaling molecules including hormones, neurotransmitters and growth factors [Lee et al., 2008]. 38 39 Signals mediated by a growth factor involve binding to its receptor initiates a process that 40 starts with the binding with membrane or intracellular receptor [8]. The amplified signal is 41 then propagated to the nucleus, resulting in induction or repression of gene expression 42 [Peschel et al., 2006]. Mitochondrial dysfunction was suggested to be related with many 43 chronic diseases. Oxidative damage are often related with path physiology of many diseases

Cinnamon is widely used as natural spices taken orally with food and enhance the thermogenesis [Anderson & Broadhurst, 2004; Murcia et al., 2004]. Cinnamon is one of the naturally occurring cinnaboid. The biological effect of Cinnamon due to it stimulate pain

47 receptors and release of analgesic mediators .The second action explanation it to activate

48 catecholamine as adrenaline secretion and sympathomimitic effect and subsequently, 49 increases blood pressure. This indicated that the thermo genesis effect by capsicum is 50 mediated by β -adrenergic stimulation and reduction in energy expenditure.

51 Several studies revealed that, supplementation of Cinnamon were effective in treatment of

- 52 some disease as improve glucose tolerance in experimental diabetic animals [Murcia et al.,
- 53 2004]. The metabolic role of Cinnamon is attributed to its role as potentiate the interaction54 with insulin receptor and improve action

55 In spite of tremendous advances in modern medicine no effective drugs are available, which 56 stimulate liver functions and offers protection to the liver from the damage or help to 57 regenerate hepatic cells (Chattopadhyay, 2003). In absence of reliable liver-protective drugs 58 in modern medicine, a large number of medicinal preparations are recommended for the 59 treatment of liver disorders (Chatterjee, 2000) and quite often claimed to offer significant 60 relief. The goal of the present study was to explore the mechanism of the antioxidant and 61 hepatoprotective efficacy of water or ethanolic extracts of cinnamon against oxidative stress 62 induced by CCl₄ in rats.

63

64 Experimental design

65 Animals

66 Sixty male rats weighing (100-120 grams) were included in current study .The animals were 67 kept at 27 ± 2 °C. Standard diet and water are given *ad libitum*.

68

69 **Preparation of cinnamon extract**

70 Cinnamon powder was obtained from the local market at Jeddah, Saudi Arabia.

All reagents and solvents used in this study were punched from Aldrich Company until otherwise stated. The dried powder was defatted with petroleum ether (100 grams in 200 ml ether). The defatted material was extracted with 95% ethanol and then vacuum dried. One part of powder was extracted in boiling water then filtered and vacuum dried.

74 part of powder was extracted in bonning water then find

75 **Determination of total phenolics**

The content of phenolics in cinnamon extracts was determined according to the method described by Negi & Jayaprakasha (2003). The cinnamon extracts (100 mg) was dissolved in

a 10 ml of mixture of methanol: water (6:4 v/v). Cinnamon extracts (equivalent to 100 mg) in

0.2ml was mixed with 1.0 ml of ten-fold diluted Folin-Ciocalteu reagent. The absorbance was
 measured at 765nm after 30 minutes.

81

82 Hepatotoxicity and treated groups.

Animals were divided into four groups (n = 15). Group I (control). Group II (CCl4) rats were injected single dose of CCl₄ in corn oil (1ml/kg B.W, s.c.) (Avijeet et al.,2008). Groups III and IV were administered orally by gastric tube 100 mg/kg of aqueous or ethanolic extracts, respectively, in the form of aqueous suspension once daily for 7 days, then animals were administered simultaneously single dose of CCl₄ (1ml /kg B.W, s.c.) Blood was collected, serum was separated at 3500 rpm for 10 min.

89 Serum biochemical assay

90 Serum enzymes aspartate aminotransferase (AST) and serum glutamate pyruvate 91 transaminase (ALT) were determined according to (Reitman & Frankel, 1957).

92 Estimation of MDA, SOD and CAT in liver tissue

23 Liver homogenates (5% w/v) were prepared in cold 50mM potassium phosphate buffer (pH

- 94 7.4) using glass homogenizer in ice. The cell debris was removed by centrifugation at 5000
- 95 rpm for 15 at 4^oC using refrigerated centrifuge. The clear portion for the estimation of
- 96 malondialdehyde (MDA) (Yagi and Rastogi, 1979), superoxide dismutase (SOD) (Kakkar et
- al., 1972) and catalase activities (Smna, 1972)

98 Assay of liver Hydroxyproline

According to Patiyal & katoch , (2006). Briefly, liver sections (0.2g) was hydrolyzed (in 6 mol/L HCl at 100 °C for 2 hours).Samples were incubated for 10 min in 0.05 mol/L chloramine-T at room temperature, followed by 15 min at 65 °C. The absorbance at 570 nm and resulting values compared to a Hydroxyproline standard curve. The Hydroxyproline content was expressed as ug/grams liver tissue.

104

105 Assay of inflammatory mediators (TNFα and IL-6)

106 The levels of inflammatory mediators (TNF α and IL-6) in liver homogenate were determined 107 as described (De Laurentiis et al.,2010), using a specific rat ELISA. The ELISA kits were 108 obtained from BD Biosciences, Pharmingen, San Diego, CA, USA. Determination of TNF- α 109 and IL-6 were performed according to the manufacturer's instructions. ELISA reader. The 110 levels of TNF- α and IL-6 were expressed as pg/mg protein.

111

112 Histopathological studies

113 Sections were prepared and then stained with hematoxylin and eosin dye.

114 Statistical analysis

115 Statistical analysis was performed on a PC using SPSS, V.13, (special package for social 116 sciences). Data are presented as arithmetic mean \pm S.D., The difference among means has 117 been analyzed by one-way ANOVA. A value of P < 0.05 was considered as statistically 118 significant.

119

120 Results and discussion

122 Phenolic compounds of cinnamon extracts.

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124 Results obtained showed that, total phenolic of water extract was 11.5% while ethanol extract 125 was 15.5%. The hepatoprotective effect of water or ethanol extracts on CCl₄-intoxicated rats 126 are shown in Table 1. In the CCl₄ intoxicated group serum AST and ALT, were significantly 127 increased as compared to control group (p < 0.001). The elevated activities of serum AST and 128 ALT were significantly reduced in the animals groups treated with either water or ethanolic 129 extracts. Treatment with ethanolic extract showed highly significant activity ($P \le 0.001$) than 130 water extract. So, the ethanol extract treated group was superior to the water extract. 131 Results obtained revealed an increase in the level of liver MDA in CCl₄- intoxicated rats 132 compared to control group. Treatment with extracts significantly prevented this raise in 133 levels. The activities of SOD and CAT have significantly reduced in CCl₄-intoxicated group, 134 while it was significantly elevated in pretreated groups with either extracts. Ethanolic extract 135 has shown more protective than water extract. As a result of CCl₄ administration, 136 inflammation increases and the release of cytokines like IL-6 and TNF stimulated. Data in 137 table 2 showed that the serum level of IL-6 and TNF were markedly increased as a result of 138 CCl_4 hepatotoxicity compared with the normal control group (P < 0.01). Rats given water or 139 ethanol extract diets showed a significantly lower IL-6 and TNF levels (P < 0.05) compared 140 with untreated animals.

141

142 Cinnamomum verum belongs to the family Lauraceae and possesses significant anti allergic,
143 anti ulcerogenic, antipyretic and anaesthetic activities (Kurokawa & Shiraki, 1998). The bark
144 yields an essential oil containing cinnamaldehyde and eugenol. Several biological activities

such as peripheral vasodilator, antitumor, antifungal, cytotoxic and ant mutagenic activities

- such as peripheral vasodilator, antitumor, antifungal, cytotoxic and ant mutagenic activities
- has been attributed to cinnamaldehyde (Shaughnessy& DeMarini, 2001).

147 Carbon tetrachloride (CCl₄) is being used extensively to investigate hepatoprotective activity
148 on various experimental animals (Bhathal et al., 1983). The free radical scavenging activity
149 of water or ethanolic extracts of cinnamon were evaluated.

Serum AST and ALT activities were used as a marker of liver damage. $CC1_4$ produces an experimental damage (James & Pickering, 1976). The toxic metabolite $CC1_3$ radical is produced by cytochrome p_{450} which further reacts with oxygen to give trichloromethyl peroxy radical.

Thabrew et al., (1987) found that serum transaminases return to normal with the healing of hepatic parenchyma and regeneration of hepatocytes. The ethanolic extract induced suppression of the increased ALT and AST activities.

157 CCl₄ produces free radical that not only directly cause damage to tissues, but also initiate 158 inflammation. Kupffer cells produce subsequently proinflammatory cytokines, and activate 159 other non-parenchymal cells involved in liver inflammation. TNF- α is produced by resident 160 macrophages after CCl_4 administration and subsequently stimulates the release of cytokines 161 from macrophages and induces phagocyte oxidative metabolism and NO production (Morio 162 et al., 2001). NO is a highly reactive oxidant and it can augramsent oxidative stress by 163 reacting with ROS and forming peroxynitrite (Rodenas et al., 1995). Another mediator of 164 CCl₄- induced hepatic inflammation which is induced by pro-inflammatory cytokines, 165 leading to formation of proinflammatory substrates from arachidonic acid (Planaguma et al., 166 2005). We observed increases in the serum level of TNF- α and IL-6, which were attenuated 167 by cinnamon extracts.

168 In this study WE or EE treated rats showed a significant restore liver functions compared to 169 CCl4 group. Also, lowering collagen precipitation and hydroxyproline content that were 170 observed in CCl4 rats. The histopathological examination of liver tissue support this 171 observation and showed that these extract have a significant antifibrotic action as indicated

172 by the disappearance of collagen accumulation.

Reduced lipid peroxidation was revealed by significant decrease in MDA level in water or ethanol extracts pretreated groups with simultaneously a significant elevation in SOD and CAT activities. Results obtained showed that, ethanolic extract was more potent antioxidant than water extract. The antioxidant properties of cinnamon extracts are attributable to the ability of its phenolic constituents to quench reactive oxygen species. In conclusion, this study suggests that ethanolic extract of cinnamon has a potent hepatoprotective activity in CCl4-induced liver injury in rats.

180 **Conclusion:** These observations were documented by biochemical results that supporting 181 the potential clinical use of cinnamon in the treatment of some hepatic disorders. Further 182 studies will be carried out to determine the types of phenol compounds that attributed to its 183 antioxidant property.

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- 283 Table (1): Table (1): Serum aminotransferase enzymes (ALT and AST), lipid peroxide product
- 284 (Malendialdlyde) and antioxidant enzyme activities; superoxide dismutase (SOD), catalase, and of all
- studied groups (Mean<u>+</u>SD).

| Animal groups | Normal Control | CCI ₄ group | WE+ CCI ₄ | EE+ CCI ₄ |
|--------------------------------|----------------------|------------------------|------------------------|-----------------------|
| Parameters | group | | | |
| Serum ALT | | | | |
| IU/ml | 28.4 <u>+</u> 4.56 | 64.0 <u>+</u> 7.86 | 37.9 <u>+</u> 7.14 | 31.6 <u>+</u> 5.08 |
| P ₁ value | | P<001 | < 0.001 | < 0.001 |
| P ₂ value | | | < 0.001 | < 0.05 |
| P ₃ -value | _ | | | <0.05 |
| Serum AST | | | | |
| IU/ml | 32.9 <u>+</u> 4.56 | 74.0 <u>+</u> 7.86 | 36.8 <u>+</u> 7.14 | 31.0 <u>+</u> 5.08 |
| P ₁ value | | P<001 | < 0.001 | < 0.001 |
| P ₂ value | | | < 0.001 | < 0.05 |
| P ₃ -value | | | | <0.05 |
| MDA (mmol/mg/protein) | | | | |
| Mean <u>+</u> SD | 3.31 <u>+</u> 0.14 | 8.14 <u>+</u> 0.57 | 3.94 <u>+</u> 0.27 | 3.30 <u>+</u> 0.32 |
| P ₁ value | | < 0.001 | < 0.001 | < 0.001 |
| P ₂ value | | | < 0.001 | < 0.01 |
| P ₃ -value | | | | <0.05 |
| SOD (MU/mg protein) | | | | |
| Mean <u>+</u> SD | 316.8 <u>+</u> 13.8 | 209.5 <u>+</u> 34.0 | 312.7 <u>+</u> 25.8 | 289.3 <u>+</u> 23.2 |
| P ₁ value | | < 0.001 | N.S | < 0.001 |
| P ₂ value | | | < 0.001 | 0.01 |
| P ₃ -value | | | | 0.01 |
| (nmol/min/mg protein) Catalase | | | | |
| Mean <u>+</u> SD | 8899 <u>+</u> 2667.7 | 3192.0 <u>+</u> 146.6 | 7582.6 <u>+</u> 1482.5 | 5281.3 <u>+</u> 935.3 |
| P ₁ value | | < 0.001 | N.S | < 0.001 |
| P ₂ value | | | < 0.001 | 0.01 |
| P ₃ -value | | | | 0.05 |

286

WE. Water extract

EE. Ethanol extract

288 P_1 – comparison to normal control P_3 - water E versus ethanol E

- 289 P_2 comparison to CCl_4 intoxicated group N.S= non significan
- 290

291 Table (2): Serum IL6, TNF- α and liver hydroxyproline content in the different studied groups

292 (Mean<u>+</u>SD)

| Parameters Groups | IL-6 (ng/mgprotein) | TNF-α (ng/mgprotein) | Hydroxyproline (µg/g protein) |
|--|------------------------|-------------------------------|----------------------------------|
| Normal Mean ± SE | 220 ± 38 | 0.13 ± 0.013 | 73 ± 1.6 |
| $\begin{array}{c} \text{CCI}_4 \text{ group} \\ \text{Mean} \pm \text{SE} \\ \text{P}^1 \text{ value} \end{array}$ | 1102 ± 106 <0.0001* | $2.54 \pm 0.122 \\ < 0.001^*$ | 304 ± 16.5 <0.001* |

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| $\begin{array}{c} \text{CCI}_4 + \text{WE} \\ \text{Mean} \pm \text{SE} \\ \text{P}^1 \text{ value} \\ \text{P}^2 \text{ value} \end{array}$ | $687 \pm 75 \\ < 0.001^* \\ < 0.000^*$ | $0.92 \pm 0.05 \\ 0.001^{*} \\ 0.000^{*}$ | $122 \pm 4.3 \\ < 0.001^* \\ < 0.000^*$ |
|--|--|---|---|
| $\begin{array}{c} \text{CCI}_4 + \text{EE} \\ \text{Mean} \pm \text{SE} \\ \text{P}^1 \text{ value} \\ \text{P}^2 \text{ value} \end{array}$ | $390 \pm 60 \\ < 0.001^{*} \\ < 0.000^{*}$ | 0.82 ± 0.05 < 0.001^{*} < 0.000^{*} | $122 \pm 4.3 \\ < 0.001^{*} \\ < 0.000^{*}$ |

EE. Ethanol extract

293 294

WE. Water extract

- 296 P_1 comparison to normal control P_3 water E versus ethanol E297 P_2 comparison to CCl₄ intoxicated groupN.S= non significan
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Fig (1) Light micrographs of mice liver treated with CC14, without and with pretreatment with either water or ethanol extract of cinnamon compared with normal control. Representative sections from (a) normal control liver (b) rats intoxicated with CC1₄, showing extensive hepatocellular necrosis; and (c) pretreated treated with water E and (d) pretreated with ethanol extract showing absence of hepatocellular necrosis, magnification X 250.

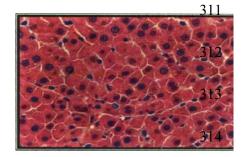
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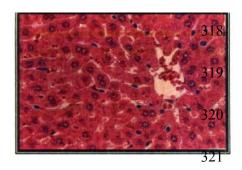
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a) control group

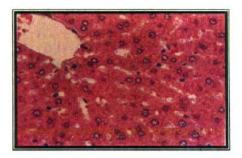


b) CCl₄ group

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322 c) WE+ CCl₄



d) EE+ CCl₄