

Original Research Article

Zanthoxylum species in Uganda: a novel wound healing alternative

ABSTRACT

Aim: The study evaluated the efficacy and possible mechanism of the stem bark of Zanthoxylum species (plant X) used by communities for wound healing in South Western Uganda.

Study design: Experimental controlled

Place and Duration: Departments of Pharmacy, Pharmacology and Pharmaceutical Sciences, Faculty of Medicine, Mbarara University of Science and Technology between August 2016 and February 2017.

Methodology: Excision wounds were humanely made on the bark of healthy albino rats and then randomly divided into four groups i.e Group 1 (Plant X water extract) n=9, Group 2 (control herbal drug) n=6, Group 3 (distilled water)n=9 and Group 4 (neomycine antibiotic) n=3. Treatments were applied twice a day for 15 days. The wound areas determined at baseline (day 1), then at day 6 and day 15 for each of the animals in groups 1, 2 and 3. Percentage reduction in wound areas was determined on day 6 and 15 and statistically compared. On day 7 the rats in group 4 and three rats randomly picked by a blinded laboratory technician from groups 1 and 3 were humanely sacrificed for histology examination of wound tissues. Phytochemical analysis of the water extract and effect of solvent were also evaluated.

Results: The Plant X water extract was found to significantly reduce wound areas better than distilled water on day 6 and 15 , (55.93±2.845) Vs (35.06±3.508),p=0.0312 and (93.18±1.721) Vs (74.89±5.604), p=0.0097 , and marginally better than herbal control drug on day 6, (55.93±2.845) Vs (39.55 ± 6.524) , p=0.0799. Five alkaloids were identified by Nuclear magnetic Resonance and Mass spectrometry in the plant X water extract as possible active compounds in wound healing. No significant difference was observed in the solvent effects.

Conclusion: Plant x shows great potential for stimulation of collagen formation and promoting natural wound healing mechanisms and therefore offers an alternative for wound treatment.

Key words: Zanthoxylum spp, Plant X, Wounds, Alternative.

1.0 INTRODUCTION

Injury is one of the leading causes of death in children and working-aged adults in almost every country and there are more than five million injury-related deaths every year, as well as a tremendous burden of disability [1]. The injury healing process involves a complex series of interactions between different cell types, cytokine mediators, and the next extracellular matrix [2]. It also occurs naturally in four phases namely; hemostasis (coagulation), inflammation, proliferation and remodeling [3]. The proliferative phase is characterized by angiogenesis, collagen deposition, granulation tissue formation, epithelization and

36 wound contraction. Alterations in any of these steps can lead to healing delay or even the inability to heal
37 completely [4]. Severe injuries lead to formation of visible wounds on the skin or other parts of the body
38 most of which are difficult to heal and even where they are healed, significant scars are left on the
39 affected part of the body.

40 Almost 25 to 40% of the active components of the synthetic allopathic medicine had origins from higher
41 flowering plants of the world and the clues to discover them came from folklore medicines of various
42 cultures [5, 6]. Some of these plants have immense potential in management of wounds especially for
43 people living in resource limited nations [7]. Despite deliberate efforts to treat wounds, some specific
44 ones due to influence of some disease processes like diabetes mellitus, HIV and varicose ulcers among
45 others have been unhealable and have continued to be entry point of disease causing organisms that
46 can eventually lead to amputation or death[8]. According to Sasidharan et al., (2010) nearly 6 million
47 people suffer from chronic wounds worldwide and the prevalence of chronic wounds in the community
48 was reported as 4.5 per 1000 population, whereas that of acute wounds was nearly double, at 10.5 per
49 1,000 population [9]. In an effort to address this challenge, interventions like stem cell treatment have
50 been considered but this is too expensive for the ordinary patients especially in developing countries like
51 Uganda. Also administration of oral and topical antibiotics has been other options but is rarely successful
52 in treating non-healing wounds [10]. In a bid to find a sustainable therapy, locals of Budibugyo (South
53 Western Uganda) have discovered the usefulness of Plant X stem bark powder (suspected to be a
54 *Zanthoxylum* spp, Rutaceae) in management of both acute and chronic wounds. This present research
55 work aimed at validating the wound healing activity of this Ugandan medicinal plant so as to establish a
56 scientific evidence for the observed community use of the plant to treat non healing wounds of various
57 causes.

58 **2.0 MATERIALS AND METHODS**

59 **2.1 Plant Material**

60 The fresh stem bark of Plant X were supplied by the herbalist under Medical Research Center,
61 Wandegeya Kampala and received at Mbarara University Pharmaceutical Analysis Laboratory. The plant
62 material were washed, shade dried and crushed in to a course powder using electric grinder. It was then
63 stored in a well closed and dry container at room temperature till use in the experiment.

64 **2.2 Preparation of the Extracts**

65 A portion (500g) of the raw powder was hot macerated using distilled water. The mixture was filtered
66 using a muslin cloth followed by Whatman's filter papers. The filtrate was evaporated using rotary
67 evaporator (RV 10 D S99) at 40°C and low pressures followed by oven drying at 50°C to obtain a
68 constant weight extract. The phytochemical groups were analyzed using the methods previously
69 described [11]. The remaining raw powder portion (500g) was serially extracted as follows: It was first
70 subjected to Petroleum ether using Soxhlet apparatus, the resulting residue was cold macerated in
71 ethanol (96%) and finally hot maceration in distilled water. The different filtrates were treated as for water
72 extract to obtain dry fractions of petroleum ether, ethanol and water.

73 **2.3 Preparation of the treatments and controls**

74 A 5% extract solution of each extract was made by dissolving 5g in 100mls of its extraction solvent for
75 application on the excision wounds. A herbal drug for wound treatment on the Ugandan market used a
76 positive control was previously tested for wound healing while distilled water served as a blank control in
77 wound treatment phase[12].

78 **2.4 Creation of wounds and application of treatments**

79 Fifty seven (57) inbred Wistar albino rats (150 – 200g) of either sex and of approximately the same age
80 were obtained from the same colony at the animal research facility of department of Pharmacology,
81 Mbarara University of Science and Technology. They were housed in clean cages with access to clean
82 water and standard laboratory pellet diet *ad libitum* throughout the experimentation period as per National
83 Institutes of Health (NIH) guidelines for animal handling in teaching and research.

84 Excision wounds were created on the barks of rats after shaving and application of analgesia and local
85 anesthesia using diclofenac, lignocaine and adrenaline by injection into the site for wound excision [13].
86 The animals were then randomly picked without replacement by a blinded Laboratory Technician and
87 placed into groups: group 1 (n=9) for plant X aqueous extract, group 2 (n=6) for the herbal wound healing
88 drug, group 3 (n=9) for the distilled water, group 4 (n=3) for the neomycine group, group 5 (n=10) for the
89 petroleum fraction, group 6 (n=10) for the ethanol fraction and group 7 (n=10) for the distilled water
90 fraction group. The fresh wounds were left for 24hours before starting the topical applications of the
91 treatments twice a day for 15 days. The wound diameters were measured using a digital Vernier caliper in

92 diagonal way as 'a' and 'b' which were used to determine wound area (mm²) using the formula ($\pi a*b$)/4
 93 for each animal at baseline and then at day 6 and day 15. Wound contraction was calculated as
 94 percentage of the reduction in wound area of the day of measurement from the baseline value i.e.
 95 Percentage of wound contraction = [(Initial wound area – Specific day wound area) / Initial wound area] x
 96 100 equation [6]. A sample of plant X powder was sent to department of Chemistry, Wits University South
 97 Africa for active compound identification using Nuclear Magnetic Resonance and Mass spectroscopy.

98 **2.5 Statistical Analysis**

99 The data obtained was analyzed using GraphPad Prism software version 7.03. One way ANOVA was
 100 used for determining the statistical significant difference in the group means. The inter group significance
 101 was analyzed using Turkey's multiple comparison test and a *P* value < .05 was considered to be
 102 statistically significant. All the values are presented as Mean ± SEM with their corresponding P values.

103 **3.0 RESULTS AND DISCUSSION**

104 The crude aqueous extract revealed the presence of various phytochemical groups with alkaloids being
 105 abundant as shown in table I.

106

107 Table I: Phytochemical groups identified in the crude aqueous stembark extract of Plant X.

Phytochemical group	Presence	Phytochemical group	Presence
Terpenoids	+	Saponins	+
Tannins	+	Anthroquinone glycosides	+
Flavonoids	-	Alkaloids	++
Amino acids	+	Phenols	+
Glycosides	+	Steroids	+

108 Presence (+) Absent (-) Abundant (++)

109 Plant X crude water extract demonstrated better wound size reduction effect than the control treatments
 110 (Table 2).

111 Table 2: Percentage wound reduction effect of Plant X compared with controls.

Time	Percentage Mean ± SEM, n=6		
	Distilled water	Plant X. extract	P-values
Day 6	35.06 ± 3.508	55.93 ± 2.845	0.0312*
Day 15	74.89 ± 5.604	93.18 ± 1.721	0.0097**
Day 6	Distilled water	Control herbal drug	P-values
	35.06 ± 3.508	39.55 ± 6.524	

Day 15	74.89 ± 5.604	86.75 ± 2.498	0.0784	112
	Plant X extract	Control herbal drug		
Day 6	55.93 ± 2.845	39.55 ± 6.524	0.0799	
Day 15	93.18 ± 1.721	86.75 ± 2.498	0.4228	

113

114 There was no statistical significant difference between the healing effects produced by ethanol, petroleum
 115 ether, ethanol and aqueous fractions (Table 3).

116

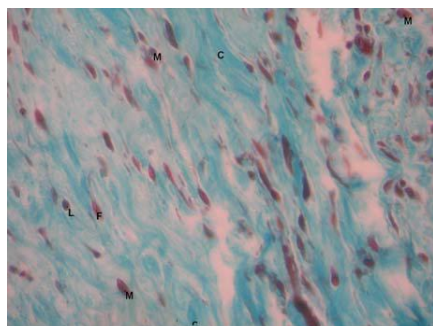
117 Table 3: Comparison of wound area reduction effect of plant X extracted in various solvents

% wound reduction (mean ± SEM, N=10)			
	Petroleum ether fraction	Ethanol fraction	P-value
Day 6	13.03 ± 1.988	1.675 ± 6.18	0.5434
Day 15	70.7 ± 5.579	56.09 ± 3.893	0.3274
	P. ether fraction	Aq. fraction	
Day 6	13.03 ± 1.988	11.76 ± 10.82	0.9926
Day 15	70.7 ± 5.579	57.03 ± 9.794	0.3742
	Ethanol fraction	Aq. fraction	
Day 6	1.675 ± 6.18	11.76 ± 10.82	0.5974
Day 15	56.09 ± 3.893	57.03 ± 9.794	0.9948

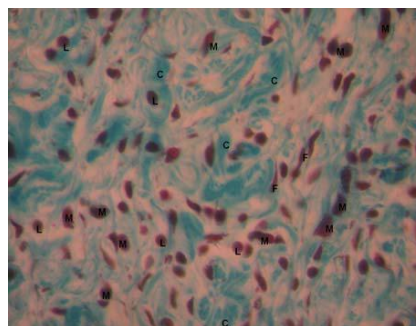
118

119 Histology analysis revealed that Plant X stimulated collagen formation more than the control treatments
 120 (neomycin or water) and the wound tissue had fewer inflammatory cells indicative of better healing effects
 121 and possible anti-inflammatory effects (Figure 1-3).

122



123

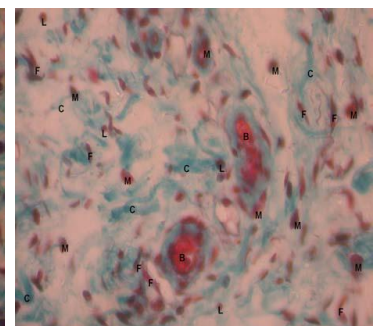


124

125

126

127



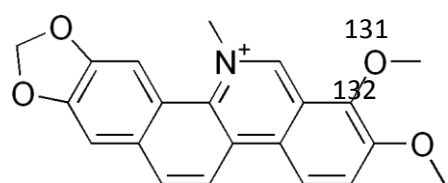
128 Figure 1: Plant X treated

Figure 2: Neomycine treated

Figure 3: Water treated

129 M= Macrophages; L= Lymphocytes; C= Collagen fibers; F= Fibroblasts; B=Blood vessel

130 Five previously known alkaloids were detected in the plant X 130 samples (Figures 4 to 8).



133
134

135 Figure 4: Chelerythrine

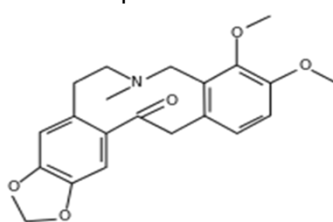


Figure 5: Fagarine

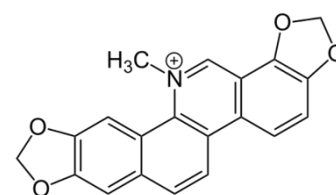
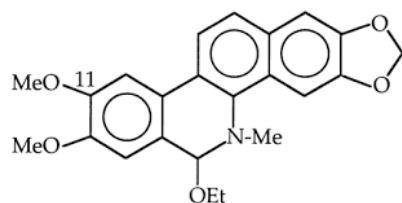


Figure 6: Sanquinarine

136



137

138 Figure 7: Dihydrionitidine

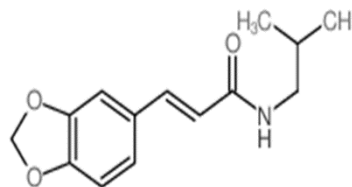


Figure 8: Trans fagaramide

139

140 4.0 DISCUSSION

141 Wound healing is a complex process involving a series of physiological and biochemical changes, but
 142 these steps can be shortened by herbs which possess antiseptic, antioxidant and anti-inflammatory
 143 activities [14]. Our findings indicate that the aqueous extract significantly had better wound healing effect
 144 than the herbal drug on the market by day 6. This could be attributed to the phytochemical groups like
 145 phenols and alkaloids previously implicated in wound healing [15] and identified in the plant X extract.
 146 According to Sunita et al., (2017), the wound healing potential of natural phytomedicines can be
 147 explained by the presence of saponins (anti-oxidant and antimicrobial activity), tannins (antimicrobial) and
 148 triterpenoids (promotes wound contraction and the rate of epithelization)[16] and these group of
 149 compounds were present in the plant X extract (table 1). Five alkaloids (Figures 4-8) were also identified
 150 in the aqueous extract and some of these compounds are known to have unique effects important for
 151 wound healing particularly Sanquinarine is a tissue regenerator with anti-inflammatory effects [17, 18].
 152 Chelerythrine has antimicrobial and antitumor properties [19].

153 The histology pictures (Figures 1, 2 and 3) indicate significant differences in collagen formation (blue
 154 color) with the plant X extract having the highest accumulation compared to the neomycin and water

155 treated groups. This could be attributed to amino acids in the plant extract (Table 1) as source of hydroxy
156 proline which is a constituent of collagen [20] or it could due to other compounds in the extract causing
157 induction of collagen formation by a mechanism that remains to be established. The low levels of
158 infiltration by inflammation promoting cells in the histology picture of Plant X treated wounds indicates the
159 possible anti-inflammatory effects of the plant X. The histology also shows abundant tissues of well-
160 organized building materials like fibroblasts, grand substance and fibrous tissue associated with
161 excessive angiogenesis all seen in the plant X extract group indicating that plant X appears to accelerate
162 the natural wound mechanism. This is further demonstrated by the shorter time taken for the wounds to
163 heal again in plant X treated groups (Table 2).

164 **5.0 CONCLUSION**

165 Plant x shows great potential for use in stimulation of collagen formation, shortening wound healing time
166 and promoting natural wound healing mechanisms that mimic stem cell stimulation. This mechanism
167 offers great hope for a cheaper alternative for healing of difficult to heal wounds and needs further
168 exploration for possible development into a drug for wider clinical application as a low cost alternative.

169

170 **ETHICAL APPROVAL**

171 The Ethical clearance TREC007/17 was obtained from THETA Uganda Research Ethics Committee,
172 accredited by Uganda National Council for Science and Technology with a focus on traditional medicine
173 research approval and the study was conducted in accordance with the national and international
174 institutional rules concerning animal experiments and biodiversity rights. The experimental animals were
175 humanely treated throughout the study and at the end of the study were sacrificed under general
176 anesthesia and incinerated.

177 **REFERENCES**

178 [1] Mock C, Quansah R, Krishnan R, Arreola-Risa C, Rivara F. Strengthening the prevention and care of
179 injuries worldwide. *Lancet*. 2004; 363(9427):2172-9. DOI: 10.1016/S0140-6736(04)16510-0

- 180 [2] Surana SS, Kumar SR. Wound healing activity of methanol extract of leaves of *Machlus macrantha*
181 Nees. Int. J. Pharm. Phytopharmacol Res. 2013; 3 (3): 200-202.
- 182 [3] Heil N, Bravo K, Montoya A, Robledo S, Osorio E. Wound healing activity of *Ullucus tuberosus*, an
183 Andean tuber crop. Asian Pac J Trop Biomed. 2017; 7(6): 538-543.
184 <http://dx.doi.org/10.1016/j.apjtb.2017.05.007>
- 185 [4] Simin B, Saeed H, Abdollah GP, Faham K, Elham MK, Parisa H.S. Wound healing activity of extract
186 from *Thymus daenensis* in burn wound model: An experimental animal study. Acta Poloniae
187 Pharmaceutica - Drug Research. 2016; 73(6):1615-1622.
- 188 [5] Agyare C, Boakye YD, Bekoe EO, Hensel A. Review: African medicinal plants with wound healing
189 properties. J.Ethno- pharmacol. 2016; 177:85–100.
- 190 [6] Subhashini S, Arunachalam KD. Investigations on the phytochemical activities and wound healing
191 properties of *Adhatoda vasica* leave in Swiss albino mice. African Journal of Plant Science. 2011;
192 5(2):133-145.
- 193 [7] Raina R, Prawez S, Verma PK, Pankaj NK. Medicinal Plants and their Role in Wound Healing.
194 VetScan. 2008; 3(1): 1-7
- 196 [8] Judy Harker. Wound healing complications associated with lower limb. World Wide Wounds.2006.
197 Revision 1. Accessed on 13th September 2017. Available:
198 [http://www.worldwidewounds.com/2006/september/Harker/Wound-Healing-Complications-Limb-](http://www.worldwidewounds.com/2006/september/Harker/Wound-Healing-Complications-Limb-Amputation.html)
199 [Amputation.html](http://www.worldwidewounds.com/2006/september/Harker/Wound-Healing-Complications-Limb-Amputation.html), Last Modified: Friday, 29-Sep-2006 15:28:16 BST.
- 200 [9] Sasidharan S, Rajoo N, Rathinam YL, Rajoo A. Wound Healing Potential of *Elaeis guineensis* Jacq
201 Leaves in an Infected Albino Rat Model. Molecules. 2010; 15:3186-3199, DOI:
202 10.3390/molecules15053186
- 203 [10] Wright J.B, Lam K, Burrell RE. Wound management in an era of increasing bacterial antibiotic
204 resistance : A role for topical silver treatment. Am J Infect Control. 1998; 26(6):572-7
- 206 [11] Nayak BS, Sandiford S. Maxwell A. Evaluation of the Wound-healing Activity of Ethanolic Extract of
207 *Morinda citrifolia* L . Leaf. eCAM. 2009; 6(3):351–356 DOI:10.1093/ecam/nem127
- 208 [12] Ogwang PE, Nambatya GN, Nyafuono J, Agwaya M, Omujal F, Tumusiime et al. Preclinical efficacy
209 and safety of herbal formulation for management of wounds. African Health Sciences. 2011; 11(3): 524 –
210 529
- 211 [13] Sutar IP, Koca U, Esra KA, Yilmazer D, Alper M. Assessment of Wound Healing Activity of the

212 Aqueous Extracts of *Colutea cilicica* Boiss. Bal. Fruits and Leaves. Evidence-Based Complementary and
213 Alternative Medicine. 2011, Article ID 758191, 7 pages DOI:10.1093/ecam/nep190

214
215 [14] Renu S, Vipin M, Shiv Kumar P, Manoj M. Evaluation of wound healing activity of ethanolic extract of
216 *ocimum basilicum* and *aegle marmelos* leaves in male albino rats, Int. J. Drug Res. Tech. 2012; 2 (2):198-
217 202.

218
219 [15] Regina CLA, Ana PLV, Simone F, Heloísa P, Daniela SC. Phytochemical Composition, Antioxidant
220 Activity, and the Effect of the Aqueous Extract of Coffee (*Coffea arabica* L.) Bean Residual Press Cake
221 on the Skin Wound Healing, Oxidative Medicine and Cellular Longevity. 2016: Article ID 1923754.
222 <http://dx.doi.org/10.1155/2016/1923754>

223
224 [16] Sunita T, Isha SK, Anil KS, Santosh KV. Pharmacological evaluation of wound healing activity of
225 herbal based formulation "Amree Plus®" in diabetic rats, The Journal of Phytopharmacology. 2017;
226 6(1):1-10.

227
228 [17] Xiaofeng N, Ting F, Weifeng L (2012). The anti-inflammatory effects of sanguinarine and its
229 modulation of inflammatory mediators from peritoneal from macrophages. European Journal of
230 Pharmacology. 2012; 689(1–3): 262-269. <https://doi.org/10.1016/j.ejphar.2012.05.039>

231
232 [18] Tsukamoto H, Seiji K, Yoshiki M, Arisa Y. Evaluation of Anticancer Activities of
233 Benzo[c]phenanthridine Alkaloid Sanguinarine in Oral Squamous Cell Carcinoma Cell Line. Anticancer
234 Research. 2011; 31(6):2841-2846.

235 [19] Fang M, Xin-Juan Y, Le Z, Hai-Jun H, Feng Z, Xu-Dong D et al. Structural modification of
236 sanguinarine and chelerythrine and their antibacterial activity. Natural Product Research. 2011; 25(9):
237 863-875. <http://dx.doi.org/10.1080/14786419.2010.482055>

238 [20] Ramchandra G, Ashish G, Prabhakar S, Prakash P. Wound healing and antioxidant effect of leaves
239 on incision *Calliandra haematocephala* and excision wound models. Asian Journal of Pharmacy and
240 Pharmacology. 2016; 2(2):34-39.

241