

Application of multivariate analysis to access selected rice germplasm phenotypic diversity

ABSTRACT

Without variability, it is not possible to conduct a plant breeding program. Germplasm is hence the critical first step in initiating a breeding program. Diversity in *O. glaberrima* accessions is enormous. It needs to be organized and characterized in order to facilitate its use by plant breeders. Two principal components (PRIN1) and (PRIN2) accounted for most of the variability observed in characters measured. PRIN 1 accounted for 56% of the phenotypic and morphological variation. The PRIN 1 was loaded on plant height, number of panicle, biomass wet weight, panicle wet weight and grain yield traits. PRIN 2 accounted for 23% of the variation. PRIN 2 was loaded on biomass wet weight, biomass dry weight, panicle wet weight, panicle dry weight, and harvest index traits. The test for univariate statistics described individual variable to explore pattern of response to variation showed strong statistical significance ($P < 0.001$) on phenotypic differences in all the variables that were measured. These traits are the most important characters contributing to the overall variability. The dendrogram produced grouping that defined nine distinct clusters and minimum genetic distance between clusters varies from 0 to ***. All the selected *Oryza glaberrima* accessions and the *Oryza sativa* were distributed across the nine clusters respectively.

KEY WORDS: Principal Component Analysis, germplasm evaluation, genetic diversity, multivariate.

INTRODUCTION

A huge number of variable are regularly measured by plant breeders of which some may not be of sufficient discriminatory power for germplasm evaluation and characterization of traits (Shaibu and Ogburia, 2002). In such a situation, principal component analysis (PCA) may offer a useful insight in data set to eliminate redundancy. In any crop, germplasm resource not only serves as a valuable source of useful genes but also provides scope for building up a basic population of wide genetic variability. Bringing improvement over existing crop varieties is a continuous process in plant breeding. To achieve this objective, the breeder has to identify diverse parents having high genetic variability for combining desirable characters. Therefore, knowledge of sound genetic diversity is essential for undertaking any recombination breeding program. In earlier days, geographical diversity was considered as a measure of genetic diversity but recently it is observed that genetic materials from same eco geographic origin also possess diverse genetic makeup and it is not uncommon that the genetic materials of different eco geographic origin possess similar genetic architecture due to free and easy material transfer facilities. The usefulness of multivariate analysis for the study of morphologically complex individual and for measuring the degree of divergence

39 between biological populations has been shown in different fields of research. Multivariate
40 statistical techniques which simultaneously analyse multiple measurements on each
41 individual under investigation are widely used in analysis of genetic diversity irrespective of
42 whether it is morphological, biochemical or molecular marker-based and subsequently,
43 classification of germplasm collections. Among the multivariate techniques, principal
44 component analysis (PCA) and cluster analysis had been shown to be very useful in selecting
45 genotypes for breeding program that meet the objective of a plant breeder (Mohammadi and
46 Prasanna, 2003). PCA has been reported to reveal patterns and eliminate redundancy in data
47 sets (Adams, 1995) while morphological and physiological variations routinely occur in crop
48 species. Cluster analysis is commonly used to study genetic diversity and for forming
49 principal subclass for grouping accessions with similar characteristic into one homogenous
50 category. Clustering is also used to summarize information on relationships between objects
51 by grouping similar units so that the relationship may be easily understood for scientific
52 knowledge and advancement.

53 Maji and Shaibu (2012) described that multivariate analysis has been used frequently for
54 genetic diversity analysis in many crops such as barley (*Hordeumvulgare*L.) Sorghum
55 (*Sorghumbicolor*L. Moench), wheat (*Triticum*spp.), peanut (*Arachishypogaea*L.)
56 (Upadhyayaet al., 2009) and vineyard peach (*Prunuspersica*L. Batsch) and in rice (Ganaet
57 al., 2013).Multivariate analysis has been used frequently for genetic diversity analysis in
58 many crops such as finger millet (Dagnachewet al., 2012b) and rice (Ganaet al., 2013). The
59 main objective of any plant scientist is to identify an optimum number of plant characters
60 which are sufficient to elucidate the maximum variability in field crop. This study was
61 undertaken to run a classification analysis on selected *Oryzaglaberrima* accessions and Asian
62 rice varieties by means of descriptive statistic and to understand the association of various
63 characters, PCA and cluster analysis which would enable breeders to classify the available
64 germplasm into distinct groups on the basis of their genetic diversity. However, the results
65 obtained would be valuable in developing an effective breeding.

66 MATERIALS AND METHODS

67 Plant materials, data collection and experimental design

68 The study was conducted to characterize 30 rice germplasm accessions consisted of 22
69 selected *Oryzaglaberrima* and 8 *Oryzasativa* lines obtained from AfricaRice gene bank were
70 evaluated under lowland conditions at AfricaRice lowland breeding station, at the
71 International Institute of Tropical Agriculture, Ibadan, Nigeria. IITA is located at (Latitude7°
72 3 “N and Longitude 3° 45 “E in the forest-savannah agro ecology). These lines were

evaluated in replicated trials in alpha lattice design during the dry season of 2014. The 21 day old seedlings were transplanted into a well levelled puddled field of 1.2m² plot at spacing of 20cm between rows and hills consisting two row plots of 3m length. Ten morphological traits were measured including days to flowering, plant height, number of tiller, number of panicle, biomass wet weight, biomass dry weight, panicle wet weight, panicle dry weight, harvest index and grain yield.

DATA ANALYSIS

Univariate statistics, standard deviation and probability level for each one of the 10 characters were calculated. Clustering of genotypes into similar groups was performed using Ward's hierarchical algorithm based on squared Euclidean distances. For the ten agro-morphological characters, the data were standardized to have a mean of zero and a variance of one prior to squared Euclidean distance calculation.

The principal component analysis method explained by Harman (1976) was followed in the extraction of the components and determined of the percentage variability explained by each component, so identify the patterns of morphological variation. The univariate descriptive statistics, cluster and principal component analysis were performed using the SAS version 9.2 software (SAS Institute, 1996, Cary, NC) of ten morphological traits in thirty rice genotypes.

RESULTS AND DISCUSSIONS

1- Univariate statistics:

The test for univariate statistics described individual variable to explore pattern of response to variation showed strong statistical significantly ($P < 0.001$) on phenotypic differences in all the variables that were measured (Table 1). This indicates that, the quantitative traits may be modified variously by the environmental conditions and each contributing such a small amount of phenotype that their individual effects cannot be detected by Mendelian methods but by only statistical methods. High variability observed for most of the traits indicated the possibility of improvement of these traits by direct selection. These traits should be given prominence in rice improvement.

Table 1: Univariate test statistics of some phenotypic characters identified by principal component analysis clusters.

Variable	Total standard deviation	Pooled standard deviation	Between standard deviation	R-Square	F Value	Pr> F
Days to flowering (days)	347.8	124.3	370.4	0.99	1743.2	<.0001
Plant height (cm)	14.1	11.9	8.5	0.32	10.1	<.0001
Number of tillers (#)	44.2	8.4	46.3	0.96	598.4	<.0001
Number of panicle (#)	121.9	28.9	126.4	0.94	373.0	<.0001
Biomass wet weight (gm ⁻²)	114.8	91.6	76.5	0.39	13.7	<.0001
Biomass dry weight (gm ⁻²)	188.8	139.0	139.6	0.48	19.8	<.0001
Panicle wet weight (gm ⁻²)	60.89	52.8	34.4	0.28	8.35	<.0001
Panicle dry weight (gm ⁻²)	57.3	54.4	22.9	0.14	3.5	<.0001
Harvest index (%)	63.3	35.9	56.0	0.69	47.6	<.0001
Grain yield (t ha ⁻¹)	195.3	79.2	191.1	0.84	114.1	<.0001

2- Cluster analysis:

Similarity indices and pattern of relationships among the *O. glaberrima* and *O. sativa* from clusters and principal component analysis are useful in evaluating the potential breeding value of both the checks and *O. glaberrima* lines. The checks - FARO 52, TOX 4004, NERICA-L25, NERICA-U4, IR77298-14.1-2-B-10, IR74371-1-1, CG 14 and Apo were distributed across six clusters. In Figure (1), the dendrogram produced grouping that defined nine distinct clusters and minimum genetic distance between clusters varies from 0 to (***). The nine, first and second clusters consisted of one (TOG5396), two (TOG7442 and TOG7996) and five (TOG6617, TOG6519, TOG13647, TOG12349 and TOG7400) genotypes belongs to *Oryzaglaberrima*, respectively. The fifth cluster composed of three genotypes belongs to *O. sativa* check i.e., CG14, IR77298 and IR74371. Similarly, the seventh cluster comprised of three genotypes, but one genotype (FARO52) belong to *O. sativa* check and two genotypes (TOMOYANG and RAM133) belongs to *Oryzaglaberrima*. While, the third, fourth, sixth and eighth clusters composed of four genotypes. The four genotypes belong to *Oryzaglaberrima* viz., TOG13429, TOG6335, TOG6379 and TOG6408 in third cluster. During the fourth and sixth clusters, one genotype belong to *O. sativa* check (NERICA-L25 and TOX4004, respectively) and three genotypes belongs to *Oryzaglaberrima* i.e., (TOG6399, TOG9119 and TOG6410) and (A2-123, TOG6311 and TOG6520), respectively. However, the NERICA-U4 and Apo genotypes belongs to *O. sativa* check and the IRAT109 and TOG5534 genotypes belongs to *Oryzaglaberrima* at the eighth cluster.

The results of cluster analysis suggested that there is genetic diversity among these genotypes for studied traits. The standalone of one of the selected *O. glaberrima* accession may have some underlying unique features not found in other groups (Figure 1). This work has demonstrated the level of similarity between and within the selected lines. Hybridization among these genotypes provided more possibility to having more phenotypic diversity and could be used in breeding programs to achieve maximum heterosis and improvement of studied traits in rice.

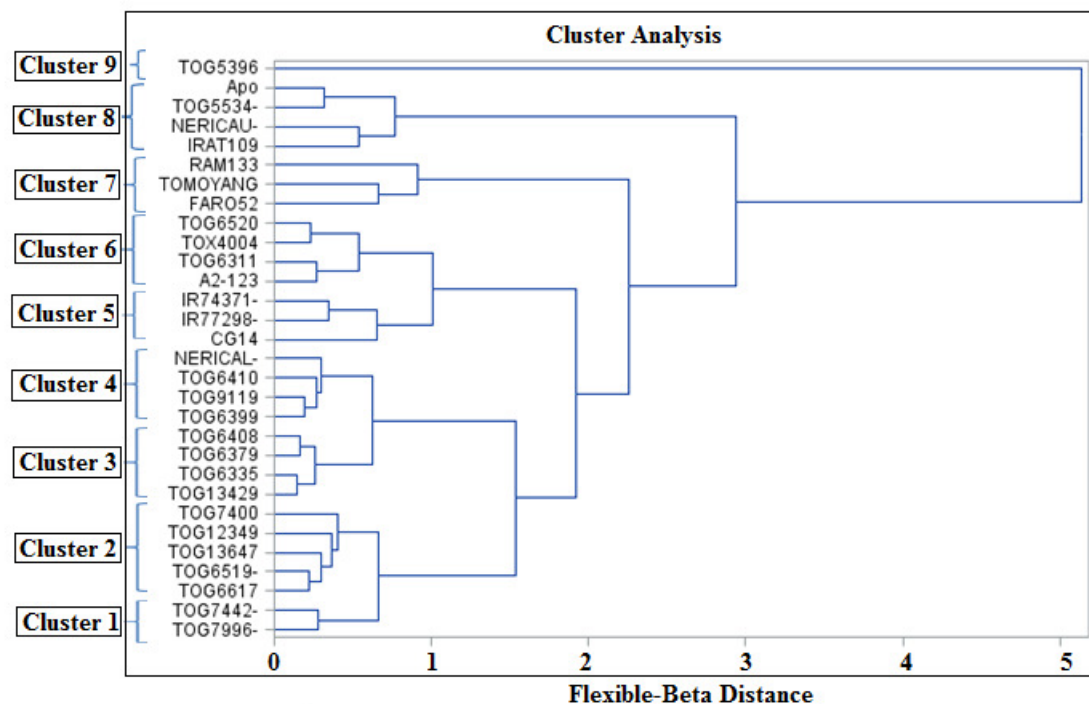


Figure 1. Clustering pattern and dendrogram of selected *O. glaberrima* and *O. sativa* lines.

3- Principal component analysis:

The ten phenotypic traits measured, ten principal component axes were revealed from PCA analysis. Two principal components (PRIN1) and (PRIN2) accounted for most of the variability observed in traits measured (Table 2). The PRIN 1 accounted for 56% of the total phenotypic and morphological variation. The PRIN 1 was loaded on plant height, number of panicle, biomass wet weight, panicle wet weight and grain yield traits. The PRIN 2 accounted for 23% of the total variation. The PRIN 2 was loaded on biomass wet weight, biomass dry weight, panicle wet weight, panicle dry weight and harvest index traits. The most important descriptions were those associated with grain yield based on the analysis of natural

variability in phenotypic traits measured in rice (Table 2). Days to flowering, number of tillers, biomass dry weight and harvest index traits negatively contributed in the first principal component axes (PRIN1), and only days to flowering negatively contributed to the total variation in the second principal component axes (PRIN2).

In general, we are interested in keeping only those principal components (PRIN1 and PRIN2) whose eigenvalues are greater than 1. The eigenvalues for PRIN1 and PRIN2 were 5.65 and 2.31, respectively. Components with an eigenvalue of less than 1 account for less variance than did the original variable (which had a variance of 1), and so are of little use. Hence, the point of principal components analysis is to redistribute the variance in the correlation matrix (using the method of eigenvalue decomposition) to redistribute the variance to first components extracted (Table 2). The scree plot graphs showed the eigenvalue against the number of component (Figure 2). The values in the two columns of the Table 2 showed two principal component axes. From the third component, it showed that the line is almost flat (Figure 1), meaning that each successive component is accounting for smaller and smaller amounts of the total variance.

Table 2: Eigenvector (“Weight”) and Eigen value (“Load”) of the correlation matrix and their contribution to total variation in rice.

Descriptor variables	PRIN 1	PRIN 2
Days to flowering (days)	-0.39	-0.01
Plant height (cm)	0.23	0.01
Number of tiller (#)	-0.41	0.04
Number of panicle (m^{-2})	0.40	0.03
Biomass wet weight (gm^{-2})	0.28	0.27
Biomass dry weight (gm^{-2})	-0.28	0.43
Panicle wet weight (gm^{-2})	0.18	0.53
Panicle dry weight (gm^{-2})	0.01	0.60
harvest index (m^{-2})	-0.35	0.30
Grain yield (gm^{-2})	0.39	0.11
Eigen value	5.65	2.31
Proportion	0.56	0.23

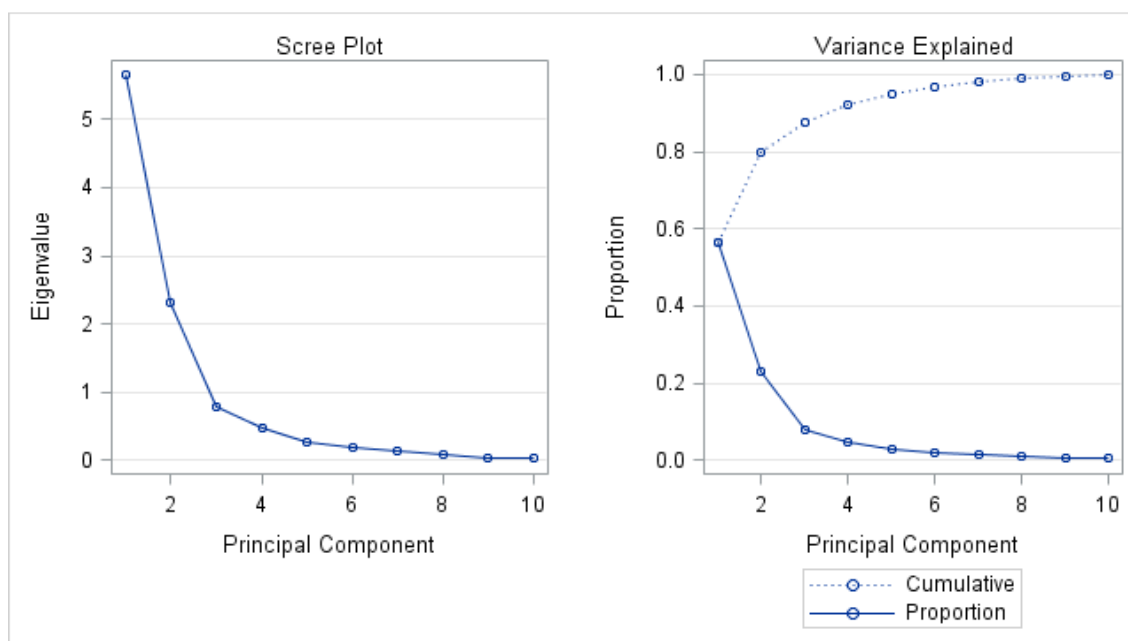


Figure 2: Scree plot and variance explained by the principal component analysis

The evaluation of germplasm frequently comprises measurement of phenotypic characters or agronomic traits of interests, such as resistance to pests and diseases and tolerance to physiological stresses that are influenced by environment which they grow. The data measuring different characters are most or often required by plant breeders. According to Ng and Padulosi (1992) and (Peters and Williams, 1984), breeders continually working and looking for variation in germplasm for use in crop improvement. Therefore, continuous evaluation of germplasm should be done to broaden the genetic base of the species and identification of additional genes of interest or alternative source that control a particular attribute or trait for use in crop improvement. As demonstrated in this study, genetic variability in crop species should be exploited further to identify traits of interest so as to develop new rice varieties with high stability to resist or tolerate adverse environments and biotic conditions demonstrated by Gana, (2006). Awopetu and Gana (1997) studied the genetic relationship of some rice varieties and observed that origin, habitat and breeding background contributed to variation in the rice population. Therefore this study is conducted to explain variation among the selected *Oryzaglaberrima* accessions *Oryzasativavarieties* and identify traits that contribute to variability in this population and for their possible exploitation in breeding programs. However, consistent effort is therefore required to

evaluate rice germplasm collection and breeding rice adaptable to harsh African environment
high grain yield.

CONCLUSION

Variation in plant phenotypic traits among rice accessions is a common phenomenon. Without variability, it is not possible to conduct a plant breeding program. Germplasm is hence the critical first step in initiating a breeding program. Diversity in *O. glaberrima* accessions is enormous. It needs to be organized and characterized in order to facilitate its use by plant breeders. There is need to understand how this tremendous variation originates and the manner in which it is organized or classified. We therefore need to know the sources to which breeders may go to find *O. glaberrima* germplasm with enormous variation to initiate their breeding programs, and the rationale for selecting materials for breeding and improving rice adaptable to African environment.

REFERENCES

1. Shaibu, A.A and Ogburia M.N (2002). Structural organization and morphological diversity of somaclonal variation in triploid plantain (*Musa* Sp. AAB group) landraces. *The Nigerian Journal of Research and Production* vol. 1, 31-37
2. Adams MW (1995). An estimate of homogeneity in crop plants with special reference to genetic vulnerability in dry season. *Phseolus vulgaris. Euphytica*, 26: 665-679.
3. Mohammadi SA, Prasanna BM (2003). Analysis of genetic diversity in crop plants- salient statistical tools and considerations. *Crop Sci.* 43, 1235-1248.
4. Maji, AT and Shaibu, AA (2012). Application of principal component analysis for rice germplasm characterization and evaluation. *J. Plant Breeding and Crop Sci.* Vol. 4, 87-93.
5. Upadhyaya HD, Gowda CLL, Pundir RPS, Reddy VG, Singh S (2006). Development of core subset of finger millet germplasm using geographical origin and data on 14 quantitative traits. *Gen. Res. Crop Evo.* 53: 679-685.
6. Gana AS, Shaba SZ, Tsado EK (2013). Principal component analysis of morphological traits in thirty-nine accessions of rice (*Oryza sativa* L.) grown in a rainfed lowland ecology of Nigeria. *J. Plant Breed. Crop Sci.* Vol. 5, 120-126.
7. Dagnachew Lule, Kassahun Tesfaye, Masresha Fetene and Santie De Villiers (2012b). Multivariate Analysis for Quantitative Traits in Finger Millet (*Eleusine coracana* subsp. *coracana*) Population Collected from Eastern and

- 227 Southeastern Africa: Detection for Patterns of Genetic Diversity. *Int. J.Agric. Res.* 7
 228 (6): 303-314.
 229
- 230 8. Jobson JD (1992). Applied multivariate data analysis. Volume II: Categorical and
 231 Multivariate Methods. New York: Springer Verlag, USA.
 232
- 233 9. Harman HH (1976), Modern factor analysis. 3rd ed. University of Chicago Press,
 234 Chicago, 376.
 235
- 236 10. SAS Institute (1996), SAS/STAT software: Users guide, version 9.2. SAS Inst., Cary,
 237 NC.
 238
- 239 11. Ng NQ, Padulosi S (1992). Constraints in the accessibility and use of germplasm
 240 collection. In Biotechnology enhances research on tropical crops in Africa edited by
 241 Thottappily, L. M. Monti D. R. Mohan Raj. A. W. Moore IITA Ibadan.
 242
- 243 12. Peters JP, Williams JT (1984). Towards better use of gene bank with special reference
 244 to information. FAO/IBPGR Plant genetic resources Newsletter 60: 22-31.
 245
- 246 13. Gana AS (2006). Variability studies of the response of rice varieties to biotic and
 247 abiotic stresses. Unpublished Ph.D Thesis, University of Ilorin.
 248
- 249 14. Awopetu JA, Gana, AS (1997). A Numerical Analysis of Genetic relationship within
 250 rice accessions. Nig. J. Gene X11:1-8.