

Antibacterial Activity and Phytochemical Screening of *Mangifera indica* Stem and Leaf Extracts on Clinical Isolates of Methicillin Resistant *Staphylococcus aureus*

ABSTRACT

Background: Medicinal plants have been sources of a number of important compounds which have been discovered during last century. In the light of their established therapeutic efficacy, the pharmaceutical industries are using crude extracts of medicinal plants for manufacturing drugs.

Aim: The study was conducted to investigate the antimicrobial activities of *Mangifera indica* (leaves and stem) and its major antimicrobial constituents (phytochemicals).

Methodology: The aqueous and chloroform extracts from the leaves and stem of the plant was tested using well diffusion method for their antimicrobial activity against Methicillin resistant *Staphylococcus aureus* (MRSA) isolated from wound, skin and urine samples of patients attending Muhammad Abdullah Wase specialist Hospital, Kano.

Results: The result shows that some extracts were active against the microorganisms tasted. Statistical analysis of the result shows that the extracts demonstrated antibacterial activity against the isolates tested with the highest zone of inhibition of 16mm, 14mm, 17mm, 15mm and 15.00mm for the five isolates used respectively. The chloroform extracts of the plant showed higher zones of inhibition compared to aqueous extracts. Preliminary phytochemical analyses of the plant showed that both stem and leaf extracts contain alkaloids, tannins, terpenoid, Anthraquinones, reducing sugar, amino acid, flavonoids, steroid, saponins, cardiac glycosides, resin and phenols.

Conclusion: *Mangifera indica* can be used as antibacterial agent against Methicillin resistant *Staphylococcus aureus*.

Key words: *Mangifera indica*; phytochemicals; Antibacterial activity; Methicillin resistant *Staphylococcus aureus*; Well diffusion.

1. INTRODUCTION

Plant derived products like gums, oils and extracts have been used for therapeutic purpose before the introduction of modern drugs [1] and continues to provide health coverage for over 80% of the world's population [2]. Serious attention is being given to medicinal plants as evidenced by the

29 recommendation given by the World Health Organization (WHO) in 1970. It gave emphasis on the
30 need to include traditional remedies within national drug policies as these plants serve as the best
31 sources of a variety of drugs? It is important to study plants so that a better understanding of their
32 properties, safety and efficacy is derived for improved benefit. The presence of phytochemical
33 constituents in medicinal plants made them useful for healing as well as for curing of human diseases
34 [3]. Phytochemicals are naturally occurring compounds in the medicinal plants, [4]. Large populations
35 of the world, especially in developing countries depend on the traditional system of medicine to treat
36 variety of diseases [5]. Several hundred genera of plants were utilized traditionally for medicinal
37 purposes. The world health organization [6] reported that 80% of the world population relies chiefly on
38 traditional medicine and a major part of the traditional therapies which involve the use of plant extract
39 and their constituents [7]. Mango (*Mangifera indica*), which belongs to the family Anacardiaceae, is
40 commonly called **Mango** (English), **Mongoro** (Yoruba, Nigeria), Mangolo (Igbo, Nigeria) and Mangoro
41 (Hausa, Nigeria) [8]. It grown naturally or cultivated mainly in tropical and subtropical regions and is
42 one of the most popular edible fruits in the world. In India and Nigeria, the infusion of the leaves singly
43 or combined with leaves of *Citrus sinensis* is used in treating diarrhea, dysentery, gastrointestinal tract
44 disorders, typhoid fever, sore throat and scurvy[9].

45 In the present study, the extracts (Aqueous and chloroform) from leaves and stem of **M. indica** were
46 screened for Antibacterial activity against Methicillin resistant *Staphylococcus aureus*. The
47 phytochemical constituents of the extracts were also determined.

48 **2. MATERIALS AND METHODS**

49 **2.1 Plant materials**

50 The plant materials used in this study consisted of the leaves, stem bark and root of *Mangifera indica*
51 **plant** which was collected from Bayero University, Kano old campus. Botanical Identification and
52 Authentification of the plant materials was done at Herbarium unit by a staff of the department of plant
53 Biology, Bayero University, Kano with the following Voucher specimen number: BUKHAN 0348.
54 Voucher specimens were deposited there for future reference. The samples were washed with water
55 and removed dust **and** rinsed with distilled water. Sample was air dried for two-weeks and pulverized
56 into powder form using sterile mortar and pestle in the laboratory as described by Mukhtar and Tukur

57 [10]. The powder sample was bagged in a black polythene bag and stored in air tight container for
58 further work.

59 **2.2 Test organisms**

60 Clinical isolates of *Staphylococcus aureus* were obtained from the laboratory of Muhammad Abdullah
61 Wase Specialist hospital Kano for further experiment. Identification and characterization of the
62 isolates was conducted there by using three procedures namely Gram staining, cultural
63 characterization using selective or indicative media and biochemical characterization. Methicillin
64 resistant *S. aureus* were determined by using Oxacillin 10µg sensitivity disc. The pure isolates of each
65 of the test organism were inoculated in sterile slants containing Nutrient agar and transported to the
66 department of Microbiology Kano University of Science and Technology, Wudil and refrigerated at
67 4°C before use.

68 **2.3 Preparation of extracts**

69 The bioactive components were extracted using the methods of Akerele *et al.* [11] with slight
70 modification. Two hundred and fifty milliliters (250 ml) each of chloroform and water were added unto
71 25g portions of leaves and stem bark powder in separate sterile conical flasks and allowed to soak at
72 ambient temperature for 7 days. The extracts were then filtered using Whatman no. 1 filter paper and
73 the filtrates concentrated at 70°C using a rotary evaporator [11]. The solid residues obtained were
74 reconstituted in DMSO at stock concentration, stored in the refrigerator at 4°C until used.

75 **2.4 Determination of phytochemical constituents**

76 The freshly prepared extracts were subjected to standard phytochemical analyses for different
77 constituents such as tannins, alkaloids, flavonoids, anthraquinones, glycosides, saponins and phenols
78 as described by Jigna *et al.* [12].

79 **2.5 Assay for antibacterial activity**

80 The antibacterial screening was carried out using the agar diffusion method as described by Lino and
81 Deogracious [13]. The test bacteria isolates were first inoculated into tubes of nutrient broth
82 separately and incubated at 37°C for 24 h. Each of the cultures was then adjusted to 0.5 McFarland
83 turbidity standards and inoculated (0.1 ml each) onto Mueller Hinton agar (MHA, Oxoid) plates. A
84 sterile cork borer was then used to make five wells (6 mm diameter each) for different concentrations

of the extract on each of the plates containing cultures of the different test isolate. The different concentrations of 0.1 ml of 30, 60, 90 and 120mg/ml of the extract were then introduced into four wells using sterile **Pasteur** pipettes. The fifth wells contain the standard antibiotics Gentamicin (62.5 mg/ml) which was used as positive control. The culture plates were allowed to stand on the working bench for 30 min for pre diffusion and were then incubated at 37°C for 24 h. After 24 h, antibacterial activity was determined by measurement of diameter zones of inhibition (mm) (against the test isolate) around each of the extracts and the antibiotic [13].

2.6 STATISTICAL ANALYSIS

The data was analyzed using One-Way Anova and the statistical program SPSS 21.0(Statistical Package for the Social Sciences). The results were presented as the means \pm standard deviation. Significance level for the differences was set at $p < 0.05$.

3. RESULTS AND DISCUSSION

Results of preliminary phytochemical screening of the leaf and stem of *Mangifera indica* are presented in Table 1. Results showed the presence of alkaloids, **anthraquinones**, **xanthoprotein**, flavonoids, resins, saponin, amino acid, tannin and **cardiac** glycoside while steroid were absent in the leaf. On the **other** hand, **anthraquinones** was absent in the stem extract.

Table 2 shows the results of antibacterial effects of extracts of the plant leaf and stem against the test isolates. Results showed that the activity of the extracts against the test bacteria increased with increase in the concentration with the chloroform stem extracts demonstrating higher activity (17 mm, 120 mg/ml,) than water extracts (15 mm, 50 mg/ml).

Table 1: phytochemical analysis of leaf and stem extract of *Mangifera indica*

Phytochemical	Leaves	Stem bark
Reducing sugar	+	+
Tannins	+	+
Anthraquinones	+	-
Steroids	-	+
Terpenoids	+	+
Saponins	+	+
Flavonoids	+	+

Alkaloids	+	+
Phenol	+	+
Xanthoprotein	+	+
Cardiac glycoside	+	+

109 Key: (+) Present, (-) Absent

110 **Table 2: Antibacterial activity of the extracts**

111

ISOLATES	CONCENTRATION (mg/ml)/ ZONE OF INHIBITION (mm)					EXTRACTS
	120	90	60	30	CONTROL	
<i>Isolate 1</i>	14	13	07	06	19	LAE
	11	09	08	06		LCE
	16	12	10	06		SAE
	15	13	09	06		SCE
<i>Isolate 2</i>	06	06	06	06	06	LAE
	13	10	09	09		LCE
	06	06	06	06		SAE
	14	12	10	06		SCE
<i>Isolate 3</i>	10	08	07	06	17	LAE
	13	12	09	06		LCE
	17	13	10	06		SAE
	14	13	10	08		SCE
<i>Isolate 4</i>	13	13	08	06	15	LAE
	15	14	12	10		LCE
	12	12	11	06		SAE
	13	13	11	08		SCE
<i>Isolate 5</i>	12	10	10	06	17	LAE
	12	09	09	08		LCE
	13	12	08	06		SAE
	15	12	11	09		SCE

112

113

LAE= Leaf aqueous extract, LCE=Leaf chloroform extract, SAE= Stem aqueous extract,

SCE=Stem chloroform extract

In the present research, *M. indica* leaf, and stem were screened for phytochemical analysis and antibacterial activities of the extracts against methicillin resistant *Staphylococcus aureus*. On phytochemical screening, the leaf extract possesses the following phytochemicals reducing sugar, tannin, anthraquinones, terpenoid, saponin, flavonoids, phenol, xanthoprotein, however, steroid is absent. The stem bark extract possessed the following phytochemicals; terpenoid, saponin, flavonoids, alkaloid, phenol, xanthoprotein and cardiac glycoside while anthraquinones is absent. The result of phytochemical screening from this study shows similarities to several studies conducted by many researchers in an attempt to determine phytochemical constituents of different part of *M. indica*. The result was in conformity with that of by Doughari, and Manzara, [14] on *In vitro* antibacterial activity of crude leaf extracts of *M. indica*, the preliminary phytochemical analysis revealed the presence of tannins, glycosides, saponins and phenols. Another experiment conducted to determine the phytochemical constituents in *Mangifera indica* by Sanwaral and Susish [15] showed the presence of alkaloid, flavonoids, tannins, saponins, glycosides and anthraquinones. The above finding supported the result of the present study. This result is also supported by a study conducted by Aiyelaagbe and Osamudiamen [16].

The result of this study shows that the *M. indica* extract (chloroform and aqueous) possesses antibacterial activity against MRSA. The results of antibacterial activity of the extracts of *M. indica* from this study were in conformity with several studies conducted by many researchers on antibacterial activity of *M. indica*. Experiment conducted by Chidozie *et al.* [17] on antibacterial activity of crude extract of *M. indica* shows that it is highly effective against some pathogenic bacteria namely *Salmonella typhi*, *Escherichia coli*, *Staphylococcus aureus*, *Proteus vulgaris*, *Shigella spp*, on the other hand, the extract found to be non effective against *Streptococcus faecalis*. The aqueous and ethanol extract of leaves and stems of mango at 50 and 25 mg/ml has been found sufficient activity against bacteria; *Staphylococcus aureus*, *Streptococcus pyogenes*, *Streptococcus pneumoniae*, *Pseudomonas aeruginosa*, *Enterococcus faecalis* [18]. The above work supported this research that *M. indica* extract possessed antibacterial agent. This work is also in conformity with that of Vega-vega *et al.* [19] who found the antibacterial ability of extract against *Salmonella enterica*, *Listeria*

monocytogenes and *Escherichia coli*. Sahrawat *et al.* [20] also determines antibacterial activities of *M. indica* leaf on methanol, ethanol and benzene extract were studied against bacteria some as *Proteus vulgaris*, *Pseudomonas fluorescens*, *Shigella flexneri*, *Klebsiella pneumonia* and *Salmonella typhi* at 100 µl/ml concentration. Antibacterial activity of mango extracts upon gram-positive, gram-negative bacteria was also demonstrated [21] and it is thought that the antibacterial activity of mango extract is due to the presence of tannin and mangiferin.

In the study conducted by Majourie [22], the result shows that different extracts of *M. indica* had different compounds with antibacterial activity. This suggests that the antibacterial activity could be due to different classes of compounds. Some of the classes of compounds identified in the crude extract, such as alkaloids and terpenoid, have been reported to possess antibacterial activity [22]. The study conducted by Doughari and Manzara, [14] reveals that the active components of leaves of *M. indica* L. which were extracted using cold water and organic solvents (acetone and methanol) and were tested against *Staphylococcus aureus*, *Streptococcus pyogenese*, *Streptococcus pneumoniae*, *Bacillus cereus*, *Escherichia coli*, *Pseudomonas aeruginosa*, *Proteus mirabilis*, *Salmonella typhi* and *Shigella flexneri* using the agar well (cup plate) diffusion method. Both the acetone and methanol extracts inhibited the growth of gram positive bacteria, with acetone extract exerting more activities on all the gram positive bacteria with zone of inhibition between 15 - 16 mm, and a gram negative bacterium *S. typhi* (14 mm) at 250 mg/ml. Whereas, water extract was not active on any of the bacterial pathogens tested at any of the concentration of the extract used. This also supported the present study.

4. CONCLUSION AND RECOMMENDATION

Results of the study showed that leaf and stem extracts of *Mangifera indica* possessed phytochemical substances that can be used as components of new antimicrobial agents. However, the chloroform extract demonstrated higher antibacterial property against the isolates compared to water. The finding of this study also shows that the stem back extract possessed higher antibacterial agents compared to leaf extract. It is recommended that there is need for further investigations in terms of toxicological studies and purification of active components with the view to using the plant in novel drug development. The study has also justified the traditional usage of this plant as health remedy.

ETHICAL APPROVAL

Ethical approval was obtained from Kano State Hospital Management Board based on the consent of Muhammad Abdullahi Wase Specialist Hospital ethical committee.

REFERENCES

- [1] Lima ME, Cordeiro M, Claudia MY, Marcos EG, Sobra M and Moreno PR. Antimicrobial activity of the essential oil from the specimens of *Pimenta pseudocaryophyllus* (Gomes) L. R. Landrum (Myrtaceae) native from Sao Paulo State. *Brazil Pharmacol.* 2006; 3: 589-593.
- [2] Kafaru E. Immense help formative workshop. In: Khan R, Islam B, Akram M, Shakil S, Ahmad A, Ali MS, Sadiqui M, Khan AU. (2008). Antimicrobial Activity of Five Herbal Extracts against Multi Drug Resistant (MDR) strains of Bacteria and Fungus of Clinical Origin. *Molecules* 1994; 13
- [3] Nostro A, Germano MP, D'angelo V, Mariano A and Lanattel MA. Extraction method and bioautography for evaluation of medicinal plant antimicrobial activity. *Letter in Appl. Microbiol.* 2000; 30: 379
- [4] Abdul Wadood, Ghufuran M, Babar Jamal SB, Naeem M, Khan A, Ghaffar R. et al. Phytochemical Analysis of Medicinal Plants Occurring in Local Area of Mardan. *Biochemistry & Analy. Biochem.* 2003; 2: 144. DOI 10.4172/2161-1009.1000144
- [5] McGaw LJ, Jager AK and Staden JV. Antibacterial, anti-helminthes and anti-amoebic activity in South Africa medicinal plants. *J. Ethno.* 2000; 72 : 247 – 263
- [6] World Health Organization (WHO). 2004 Use of antimicrobials outside human medicine and result and antimicrobial resistance in humans. World Health Organization 2002. Archived from the Original on 13 May, 2004
- [7] Ahmed I and Beg AZ. Antimicrobial and phytochemical studies on 45 Indian Medicinal plants against multi-drug resistance human pathogens. *J Ethnopharm.* 2003; 74: 113-123.
- [8] Emeruwa AC. The conservation of medicinal plants. *J. Nat. Prods.* 1991; 45 (2): 123-127.
- Haslam E. Plant Polyphenols- *Vegetable Tannins Revisited*. Cambridge University Press; Cambridge, U.K. 1989
- [9] Lakshminarayana G, Chandrasekhara Rao T, Ramalinga swamy PA. Varietal variations in contents, characteristic and composition of mango seeds and fat, *J. of the American oil chem. society* 1983; 60: 88-89
- [10] Mukhtar MD and Tukur A. In-vitro screening activity of *Pistia stratiutes* extract. *NISED Journal* 1999; 1 (1): 5 – 6

202 [11] Akerele JO, Obasuyi O, Ebomoyi MI, Oboh IE, Uwumarongie OH. Antimicrobial activity of
 203 ethanol extract and fractions of the seeds of *Garcinia kola* Heckel (Guttiferae). *Africa J. Biotechnol.*
 204 2008; 7(2): 169-172.

205 [12] Jigna P, Sumitra C. In-vitro antimicrobial activities of extracts of *Launaea procumbens* Roxb.
 206 (Labiateae), *Vitis vinifera* L. (Vitaceae) and *Cyperus rotundus* L. (Cyperaceae). *Afri. Jornal Biomed.*
 207 Res. 2006. 9(2): 89-93.

208 [13] Lino A and Deogracios O. The in-vitro antibacterial activity of *Annona senegalensis*, *Securidacca*
 209 *longipendiculata* and *Steanotaenia araliacea*- Ugandan Medicinl plants. *Afri. Health Sci.*2006; 6(1):
 210 31-35.

211 [14] Doughari JH and Manzara S. In vitro antibacterial activity of crude leaf extracts of *Mangifera*
 212 *indica* Linn. *A. J. of Microbiol. Research.* 2008; 2:067-072.

213 [15] Sanwaral A and Sushil K. Antibacterial activity of *Mangifera indica* leaves against drug resistant
 214 bacterial strain. *Intern. journal of advance research.* 2013; 1 (6): 82-86

215 [16] Aiyelaagbe OO and Osamudiamen PM. Phytochemical Screening for Active Compouds in
 216 *Mangifera indica* leaves from Ibadan, Oyo State. *Plant Sciences Research, Ibadan.* 2009; 2(1): 11-13

217 [17] Chidozie VN, Adoga GI, Chukwu OC, Chukwu ID and Adekeye AM. Antibacterial And
 218 Toxicological Effects of the Aqueous Extract Of *Mangifera Indica* Stem Bark On Albino Rats. 2014.

219 [18] Shabani Z, Sayadi A. The Antimicrobial in Vitro Effects of Different Concentrations of Some Plant
 220 Extracts Including Tamarisk, March, Acetone and Mango. *Journal of Appl. Pharm. Science.* 2014; (5):
 221 75-79.

222 [19] Vega-Vega V, Silva-Espinoza BA, Cruz-Valenzuela MR, Bernal-Mercado AT, Gonzalez Aguilar
 223 GA, Ruiz-Cruz S et al. Antimicrobial and antioxidant properties of by product extracts of mango fruit.
 224 *Journal of Applied Botany and Food Quality;* 2013; 86: 205-211

225 [20] Sahrawat A, Pal S, Shahi SK. Antibacterial activity of *Mangifera indica* (mango) leaves against
 226 drug resistant bacterial strains. *International Journal of Advanced Research* 2013; 1 (6):82-86.

227 [21] Savikin K, Menkovic N, Zdunic G, Stevic T, Radanovic D and Jankovic T. Antimicrobial activity of
 228 *Gentiana lutea* L. extracts. *Naturforsch* 2009; 64:339-342.

229 [22] Marjorie MC. Plants products as antimicrobial agents. *Clin Microbiology. Rev* 1999; 12 (4): 564-
 230 582