

Original Research Article

ANTIBACTERIAL ACTIVITY OF *DACRYODES EDULIS*

METHANOLIC LEAF EXTRACT

Abstract

Dacryodes edulis is used for the treatment of several ailments such as malaria, oral and ear infection. This study was designed to determine the antibacterial and antioxidant potentials of *Dacryodes edulis* leaf extract as well as its rate of killing some selected bacterial strains.

The leaves were dried in hot-air oven at 40 °C to a constant weight and ground into powder.

A known weight of the powder was soaked in a mixture of methanol and sterile distilled water in ratio 3:2 (v/v) for 96 h, filtered and concentrated. The antimicrobial activity of the

crude extract obtained was determined against panel of bacteria using agar-well diffusion method. The minimum inhibitory concentration, minimum bactericidal concentrations, phytochemical and antioxidant properties of the crude extract were also studied.

The extract showed appreciable antibacterial activities against *Bacillus polymyxa* (LIO), *Bacillus stearothermophilus* (NCIB 8222), *Citrobacter freundii* (LIO), *Corynebacterium pyogenes* (LIO), *Enterococcus faecalis* (NCIB 775), *Escherichia coli* (NCIB 86), *Klebsiella pneumoniae* (NCIB 418), *Micrococcus luteus* (NCIB 196), *Pseudomonas aeruginosa* (NCIB 950), *Shigella sp.* (LIO), *Staphylococcus aureus* (NCIB 8588) at a final concentration of 35 mg/mL. The mean diameter of zones of inhibition exhibited by the extract ranged between 12 mm and 23 mm while that of streptomycin ranged from 13 mm and 28 mm and ampicillin ranged from 18 mm and 29 mm. The MIC exhibited by the extract against susceptible test organisms ranged between 0.27 mg/mL and 4.375 mg/mL while MBC ranged between 0.55 mg/mL and 17.5 mg/mL. The phytochemical analysis revealed the presence of tannins,

25 flavonoids, saponins and alkaloids. The antioxidant property result revealed 50% inhibition
26 of the extract at 0.01045 mg/mL while that of the standard (ascorbic acid) was 0.00288
27 mg/mL. The time kill assay showed that the percentage of the cells killed increased with
28 increasing concentrations of the extract, as well as, contact time intervals.

29 In conclusion, *Dacryodes edulis* leaf extract exhibited appreciable antibacterial activity
30 against some of the test organisms used for this study and this supports its usefulness in
31 folklore remedies.

32 **Keywords:** Alkaloids, Antibacterial, Antioxidant, Killing Rate, Flavonoids, Phytochemical,
33 Saponins, Susceptibility

34 1.0 Introduction

35 Herbal and traditional medicine has been a major aspect of the socio-cultural heritage in
36 Africa for hundreds of years even before the advent of conventional medicine. It was once
37 believed to be primitive and wrongly challenged by foreign religions dating back during the
38 colonial rule in Africa subsequently by the conventional or orthodox medical practitioners
39 [1]. Plant-derived medicines have been part of traditional health care in most parts of the
40 world for thousands of years and there is increasing interest in them as sources in the
41 treatment of diseases [2],[3],[4]. Therefore, plants used as medicine is known as medicinal
42 plants [5].

43 *Dacryodes edulis* known as “Native Pear”, “African Pear” or “Bush Butter Tree” in English
44 belong to the family of Burseraceae (G. Don). *Dacryodes edulis* is a dioecious plant and it is
45 found natively in Angola, Benin, Cameroon, Central African Republic, Congo, Cote d’Ivoire,
46 Democratic Republic of Congo, Equatorial Guinea, Gabon, Ghana, Liberia, Nigeria, Sierra
47 lone, Togo and Uganda [6]. It is a medium-sized, evergreen tree attaining a height of 18-40 m

48 in the forest but not exceeding 12 m in plantations. It is generally branched from low down,
49 with a deep, dense crown. The trunk is rather short, slightly fluted, 50-170 cm in diameter
50 and more or less sinuous [6].

51 The extracts from *Dacryodes edulis* have been found to show various biological activities
52 which include antimicrobial, antioxidant and anti-sickling [7]. The leaf sap is instilled into
53 the ear for ear problems and a decoction of the leaves is prepared as a vapour for feverish
54 stiffness with headache [8]. It was reported by Jiofack *et al.* (2010) [9] that the leaves were
55 made into plaster to treat snakebite in Southwest Cameroon. *Dacryodes edulis* is used
56 traditionally medicine to treat ringworm, wound, scabies, skin diseases and inflammation
57 [10].

58 The aim of this study was to screen qualitatively for the chemical groups and also to evaluate
59 the *in vitro* antibacterial and antioxidant properties of the leaf extract of *Dacryodes edulis*
60 from Nigeria.

61 **2.0 Materials and Methods**

62 **2.1 Microorganisms**

63 Microorganisms used for this study were obtained from Department of Microbiology,
64 Obafemi Awolowo University, Ile-Ife, Osun State, Nigeria. These organisms include typed
65 cultures of National Collection of Industrial Bacteria (NCIB) and Locally Isolated Organisms
66 (LIO).

67 **2.2 Preparation of microorganisms used**

68 The bacterial test organisms used for this experiment were first reactivated in nutrient broth
69 and incubated at 37 °C for 18-24 h in an incubator. After 18 h of incubation, the test
70 organisms were standardized to 0.5 McFarland standard.

71 **2.3 Plant specimen**

72 The leaf of *Dacryodes edulis* used for this experiment was collected from Opa Area of Ile-Ife,
73 Osun State.

74 **2.4 Drying and extraction of leaf of *Dacryodes edulis***

75 *Dacryodes edulis* leaf was dried in hot-air oven at 40 °C until a constant weight of the sample
76 was obtained and later ground into fine powder. Exactly 1000 g of the powered sample was
77 soaked in methanol/sterile distilled water in the ratio 3:2 (v/v) and this was left on the
78 laboratory bench for 96 h with occasional agitation during this period.

79 The mixture was later filtered into a sterile flask using glass wool and funnel. The filtrate
80 collected was concentrated *in vacuo* in a rotary evaporator and then lyophilized. The resultant
81 powder was kept in air-tight bottle and stored at 4 °C in the refrigerator for further use.

82 **2.5 Antibacterial susceptibility testing of the crude extract of leaf of *Dacryodes edulis***
83 **against some selected bacterial test organisms**

84 The crude extract was screened for antibacterial activities using agar-well diffusion method
85 on Muller-Hinton agar medium as described by [11]. With the aid of an inoculating loop, a
86 loop full of the standardized broth culture of the test organism was added to 18 mL of sterile
87 molten Muller-Hinton agar medium which had already been cooled down to 45 °C. This was
88 well mixed and poured into a sterile Petri dish and allowed to set. The required numbers of
89 wells were bored into the medium using 6 mm sterile cork borer. The wells were made about
90 5 mm to the edge of the plate and labelled accordingly. Reconstituted extract solution (35
91 mg/mL concentration) was filled up into the wells. Streptomycin and ampicillin were used as
92 positive controls at a concentration of 1 mg/mL. The plates were allowed to stand for one
93 hour and thereafter, the plates were incubated upright at 37 °C for 24 h. The relative

94 susceptibility of the test organisms to the extract as indicated by clear zones of growth
95 inhibition around the wells was measured and recorded in millimetres.

96 **2.6 Determination of minimum inhibitory concentrations (MICs)**

97 The MICs of the extract was done using method described by Akinpelu and Kolawole, (2004)
98 [12]. Two-fold dilutions of the plant extract were prepared and 2 mL of different
99 concentration of the solution were added to 18 mL of pre-sterilized molten nutrient agar in
100 McCartney bottle at a temperature of 45 °C to a series of concentrations between 0.137
101 mg/mL and 35 mg/mL. The media were then poured into sterile Petri dishes and allowed to
102 set. The surface of the nutrient agar plate was allowed to dry before streaking with
103 standardized 18 h old broth culture of the susceptible bacterial strains. All the plates were
104 then labelled accordingly and incubated at 37 °C for 24 h and then examined for the presence
105 or absence of growth of the test organisms. The lowest concentrations preventing the growth
106 of the test organisms were taken as the minimum inhibitory concentrations (MICs) of the
107 extract.

108 **2.7 Determination of minimum bactericidal concentrations (MBCs)**

109 Samples from plates with no apparent growth in the MIC assay plate were sub-cultured onto
110 freshly prepared sterile nutrient agar plates without extract inclusion and then incubated at 37
111 °C for 48 h. The lowest concentration of the extract that did not show any growth on a new
112 set of nutrient plate was taken as the minimum bactericidal concentration of the extract.

113 **2.8 Phytochemical analysis of the leaf extract of *Dacryodes edulis***

114 A small portion of the dried extract was used for phytochemical screening following the
115 methods of Harbourne (1998) and Trease and Evans (2002) [13][14].

116

117

118 2.8.1 Test for alkaloids

119 Exactly 0.5 g of the plant extract was dissolved in 5 mL of 1% HCl separately on a steam
120 bath. One millilitre of the filtrate was treated with a few drops of Meyer's reagent. Also,
121 another 1 mL of filtrate was treated again with a few drops of Wagner's reagent. Turbidity or
122 precipitation with these reagents was taken as an evidence for the presence of alkaloids.

123 2.8.2 Test for tannins

124 Exactly 0.5 g of the plant extract was stirred with 10 mL sterile distilled water. This was
125 filtered using a sterile filter paper and ferric chloride reagent was added to the filtrate. The
126 presence of blue-black or green precipitate indicates the presence of tannin in the extract.

127 2.8.3 Test for flavonoids

128 Exactly 0.1 g of the extract was dissolved in 2 mL of ethylacetate in a test tube. The tube was
129 vigorously shaken to ensure proper dissolution. The solution was decanted into another test
130 tube in order to remove the particles. Few drops of dilute ammonia were then added to the
131 filtrate. Since the liquids were immiscible, there was a formation of an alkaline layer. The test
132 was observed for a light brown colour change of the alkaline layer below the aqueous layer,
133 which indicated the presence of flavonoids.

134 2.8.4 Test for saponins

135 Exactly 1.0 g of the extract was dissolved in 5 mL of distilled water. Few drops of Fehling's
136 solution were added to the extract solution. Presence of green colour indicated the presence
137 of saponins.

138 2.9 Antioxidant activity of leaf extract of *D. edulis***139 2.9.1 2, 2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging assay**

The hydrogen radical scavenging properties of the extract from the plant sample was determined by the stable radical DPPH (2, 2-diphenyl-1-picrylhydrazyl radical) method proposed by Blois (1958) [15] as described by Brand-Williams *et al.* (1995) [16]. The reaction of DPPH with an antioxidant compound which can donate hydrogen leads to its reduction and there is a colour change from deep violet to light yellow, this change in colour is measured spectrophotometrically at a wavelength of 517 nm.

To 1 mL of varying concentration of the extract sample and ascorbic acid serving as standard was added to 1 mL 0.3 mM DPPH in methanol respectively and allowed to react. The mixture was vortexed and incubated in the dark for 30 min and the absorbance was measured at 517 nm against a DPPH negative control containing 1 mL of methanol in place of the extract sample. The percentage inhibition of the DPPH scavenging activity was calculated using the equation below:

$$\text{DPPH \% Inhibition} = \{1 - (A_{517 \text{ nm sample}} / A_{517 \text{ nm control}})\} \times 100$$

Where;

$A_{517 \text{ nm sample}}$ is the absorbance of the sample (extract/standard) at 517 nm.

$A_{517 \text{ nm control}}$ is absorbance of the negative control at 517 nm.

Sample concentration providing 50% inhibition (IC_{50}) was calculated from the graph by plotting inhibition percentage against extract concentration.

2.10 Rate of killing of *D. edulis* methanolic extract

The rate of kill was determined using the method described by Odenholt *et al.* (2001) [17] with some modifications. Experiment was performed using the extract on the viability of *Escherichia coli* representing Gram-negative and *Staphylococcus aureus* representing Gram-

positive organisms. Viable count of the test organisms was initially determined. A 0.5 mL volume of known cell density (by viable counts of 10^6 cfu/mL) from each test organism suspension was added to 4.5 mL of different concentration of the extract. The suspension was thoroughly mixed and held at room temperature (28-30 °C) and the killing rate was determined over a period of 2 h. Exactly 0.5 mL of each suspension was withdrawn at the appropriate time intervals and transferred to 4.5 mL nutrient broth recovery medium containing 3% “Tween 80” to neutralize the carry-over effects of the antimicrobial compounds from the test suspensions. The suspension was shaken properly then serially diluted up to 10^{-5} in sterile physiological saline. Exactly 0.5 mL of the final dilution of the test organism was transfer into pre-sterile nutrient agar at 45 °C and plated out. The plates were allowed to set and incubated upside down at 37 °C for 72 h. Control experiment which was set up without the inclusion of antimicrobial agent (i.e extract). Viable counts were made in triplicates for each sample. Depression in the viable counts indicated killing by the antimicrobial agent.

3.0 Results and Discussion

3.1 Results

The extract obtained from *D. edulis* leaves was brownish yellow in colour with a yield of 62 g of extract from 1000 g of the powdered leaves of *D. edulis* that was soaked.

The extract of *D. edulis* at a concentration of 35 mg/mL was found to be active against eleven out of fourteen of the **bacterial test organisms** used for this study with varying level of activities (Table 1).

Micrococcus luteus, *Bacillus polymxa*, *Enterococcus faecalis*, *Escherichia coli*, *Klebsiella pneumoniae*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Shigella* sp., *Bacillus stearothermophilus*, *Citrobacter freundii* and *Corynebacterium pyogenes* were susceptible to

the extract. The zones of inhibition exhibited by the extract against the test organisms ranged between 12 mm and 23 mm. *Bacillus polymyxa*, *Micrococcus luteus*, *Pseudomonas aeruginosa* and *Staphylococcus aureus* showed the lowest zone of inhibition of 12 mm while *Klebsiella pneumoniae* showed the highest zone of inhibition with 23 mm. The MIC exhibited by the extract against *Pseudomonas aeruginosa*, *Shigella* sp. and *Klebsiella pneumoniae* was 0.27 mg/mL, *Bacillus stearothermophilus* was 0.55 mg/mL, while that of *Enterococcus faecalis* was 1.09 mg/mL (Table 2). On the other hand, MIC exhibited against *Bacillus polymyxa*, *Micrococcus luteus*, *Staphylococcus aureus*, *Escherichia coli*, *Citrobacter freundii* was 2.19 mg/mL while *Corynebacterium pyogenes* exhibited 4.375 mg/mL (Table 2).

Table 1: Antibacterial susceptibility test of *D. edulis* leaf extract against some selected bacterial test organisms

Microorganisms	Zones of Inhibition (mm)*			
	<i>D. edulis</i> (35 mg/mL)	Strep (1 mg/mL)	Amp (1 mg/mL)	M/W (1:1) (v/v)
<i>Enterococcus faecalis</i> (NCIB 775)	14	19	18	0
<i>Escherichia coli</i> (NCIB 86)	16	0	26	0
<i>Bacillus polymyxa</i> (LIO)	12	23	0	0
<i>Micrococcus luteus</i> (NCIB 196)	12	14	28	0
<i>Pseudomonas aeruginosa</i> (NCIB 950)	12	31	0	0
<i>Staphylococcus aureus</i> (NCIB 8588)	12	15	23	0
<i>Klebsiella pneumoniae</i> (NCIB 418)	23	21	0	0
<i>Shigella</i> sp. (LIO)	13	13	20	0
<i>Bacillus stearothermophilus</i> (NCIB 8222)	14	20	28	0
<i>Citrobacter freundii</i> (LIO)	13	0	22	0
<i>Corynebacterium pyogenes</i> (LIO)	19	21	27	0
<i>Bacillus cereus</i> (NCIB 6344)	0	13	0	0
<i>Clostridium sporogenes</i> (NCIB 582)	0	0	29	0
<i>Bacillus subtilis</i> (NCIB 3610)	0	28	0	0

198 Keys:

199 mm*: Mean of three replicates.

200 LIO: Locally Isolated Organism.

201 NCIB: National Collection for Industrial Bacteria.

202 M/W: Methanol/Sterile distilled water.

203 **Table 2: Minimum inhibitory concentration (MIC) and Minimum bactericidal**
 204 **concentration (MBC) exhibited by *D. edulis* leaf extract against susceptible**
 205 **bacterial test organisms**

Microorganisms	MIC (mg/mL)	MBC (mg/mL)
<i>Enterococcus faecalis</i> (NCIB 775)	1.09	2.19
<i>Escherichia coli</i> (NCIB 86)	2.19	4.375
<i>Bacillus polymyxa</i> (LIO)	2.19	8.75
<i>Micrococcus luteus</i> (NCIB 196)	2.19	8.75
<i>Pseudomonas aeruginosa</i> (NCIB 950)	0.27	0.55
<i>Staphylococcus aureus</i> (NCIB 8588)	2.19	4.375
<i>Klebsiella pneumoniae</i> (NCIB 418)	0.27	0.55
<i>Shigella</i> sp. (LIO)	0.27	0.55
<i>Bacillus stearothermophilus</i> (NCIB 8222)	0.55	2.19
<i>Citrobacter freundii</i> (LIO)	2.19	4.375
<i>Corynebacterium pyogenes</i> (LIO)	4.375	17.5

206 **Keys:**

207 NCIB: National Collection of Industrial Bacteria.

208 LIO: Locally Isolated Organism.

209 Table 2 shows the minimum bactericidal concentration of the leaf of *Dacryodes edulis*
 210 against bacterial isolates. The MBC exhibited by the extract against *Pseudomonas*
 211 *aeruginosa*, *Shigella* sp. and *Klebsiella pneumoniae* was 0.55 mg/mL, *Bacillus*
 212 *stearothermophilus* and *Enterococcus faecalis* was 2.19 mg/mL, while that of *Staphylococcus*

213 *aureus* and *Citrobacter freundii* was 4.375 mg/mL. On the other hand, MBC exhibited
214 against *Bacillus polymyxa* and *Micrococcus luteus* was 8.75 mg/mL. Finally, the MBC
215 exhibited against *Escherichia coli* and *Corynebacterium pyogenes* was 17.5 mg/mL.

216 The phytochemical analysis of the leaf extract of *D. edulis* revealed the presence of alkaloids,
217 saponins, flavonoids and tannins as presented in Table 3 below.

218 **Table 3: Phytochemical components revealed by *D. edulis* leaf extract**

Phytochemicals	Result
Alkaloids	Positive
Flavonoids	Positive
Tannins	Positive
Saponins	Positive

219

220 **Figures 1a and 1b shows** result of the antioxidant property of the extract at different
221 concentrations in addition to comparison with the standard (ascorbic acid) at initial
222 concentration of 5 mg/mL. The 50% inhibition concentration for the extract was observed at
223 0.01045 mg/mL while that of the standard (ascorbic acid) was at 0.00288 mg/mL.

224

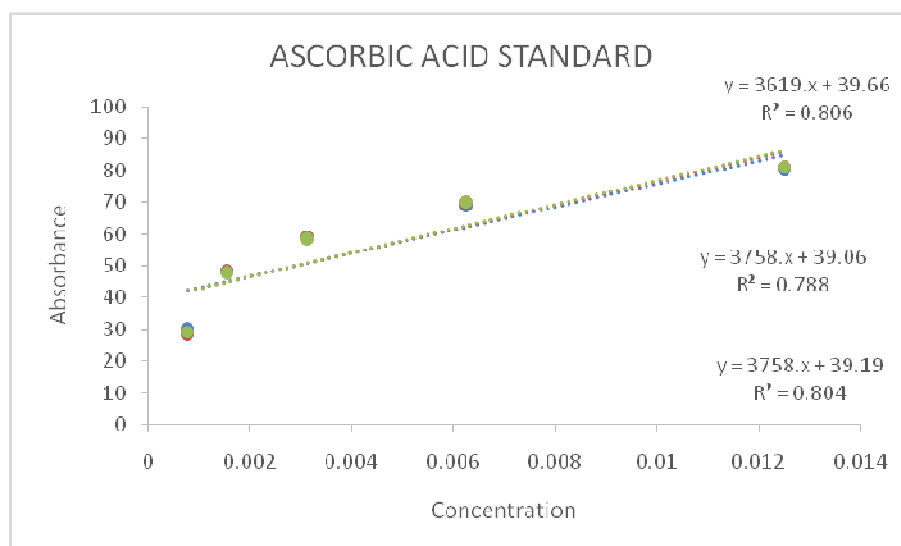


Figure 1a: The graph of concentration against absorbance for the standard (ascorbic acid).

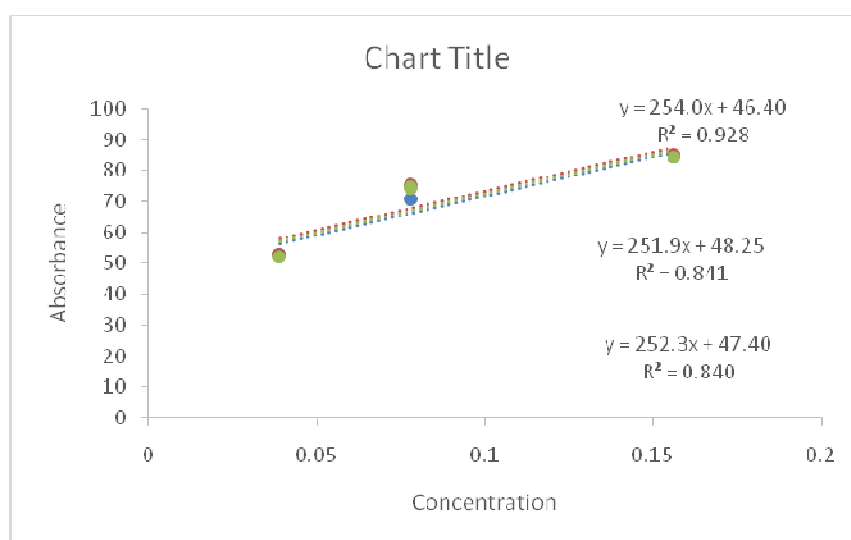
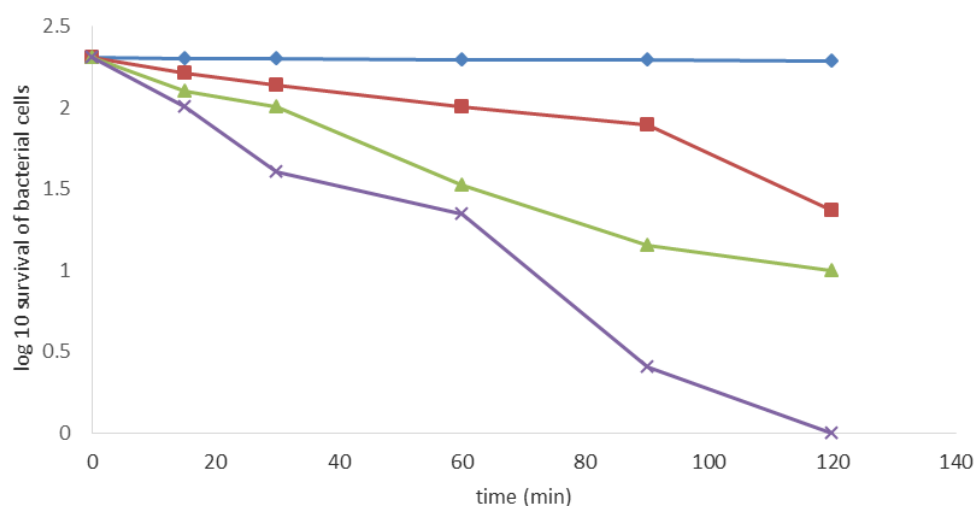


Figure 1b: The graph of concentration against absorbance for *D. edulis* extract.

The extent and rate of killing of *E. coli* by methanolic extract at 1 x MIC, 2 x MIC and 3 x MIC concentrations are as shown in Figure 2. The percentage of the organisms killed by the extract at 1 x MIC in 15 minutes was 6.2% while the percentage of the cells killed at 30 minutes rose to 10.7%. After 60 minutes of contact time with this extract, the percentage of the organisms killed was 21.9%. When the contact time was later increased to 90 minutes,

235 34.7% of the test organisms were killed and this rose to 48.3% after 120 minutes of contact
236 time.

237 When the MIC of the extract was doubled, the percentage of the cells killed was 20.3% at 15
238 minutes of contact time. At 30 minutes of contact time, 39.2% of the test organisms were
239 killed and when the contact time was increased to 60 minutes, the percentage of the cells
240 killed was 45.8%. When the contact was increased to 90 minutes, the percentage of the
241 organisms killed has increased to 59.6%, while it rose to 67.9% after 120 minutes of contact
242 time. The extent and rate of killing of the organism by the extract at 3 x MIC followed the
243 same trend as the concentrations of the extract increased with increase in time, the percentage
244 of the organisms killed also increased.

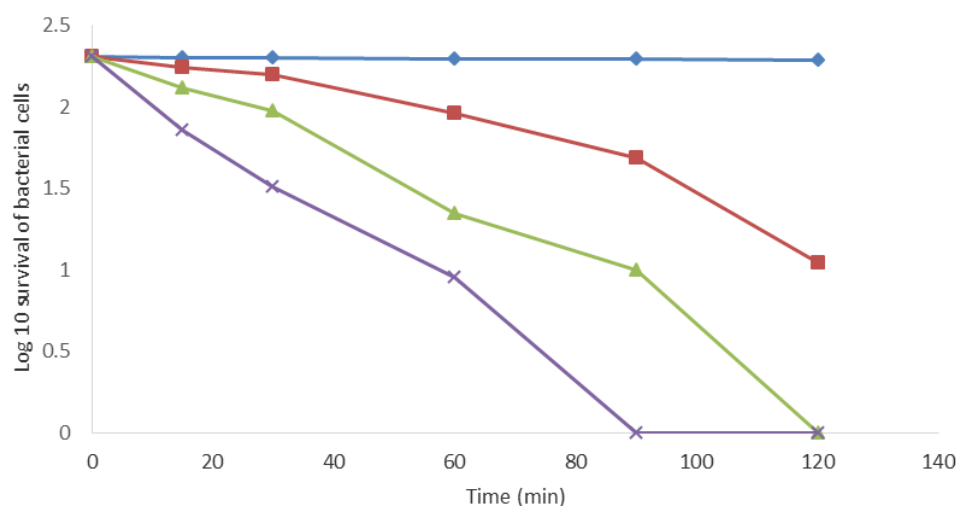


245

246 **Figure 2: The extent and rate of killing of *E. coli* cells by methanolic crude extract at 1**
247 **x MIC (—■—), 2 x MIC (—▲—), 3 x MIC (—×—) and control (—◆—).**

248 Each point represents the log₁₀ of mean survival of bacterial cells at a particular time interval
249 in the presence of the extract.

250 The extent and rate of killing of *Staph. aureus* by methanolic extract at 1 x MIC, 2 x MIC and
 251 3 x MIC concentrations are as shown in Figure 3. The percentage of the organisms killed by
 252 the extract at 1 x MIC in 15 minutes was 9.1% while the percentage of the cells killed at 30
 253 minutes rose to 17.5%. After 60 minutes of contact time with this extract, the percentage of
 254 the organisms killed was 29.3%. When the contact time was later increased to 90 minutes, the
 255 percentage of the cells killed was 35.7% and this rose to 53.4% after 120 minutes of contact
 256 time. When the MIC of the extract was doubled, the percentage of cells killed was 21.4% at
 257 15 minutes of contact time. At 30 minutes of contact time, the percentage of cells killed
 258 increased to 35.3%; and when the contact increased to 60 minutes, the cells killed was 52.8%.
 259 At 90 minutes contact time, the percentage of the organisms killed has increased to 74.7%
 260 while it rose to 100.0% after 120 minutes of contact time. The extent and rate of killing of the
 261 extract at 3 x MIC followed the same trend with the previous tests.



262

263 **Figure 3: The extent and rate of killing of *Staph. aureus* cells by methanolic crude**
 264 **extract at 1 x MIC (—■—), 2 x MIC (—▲—), 3 x MIC (—×—) and control (—◆—).**

265 Each point represents the log₁₀ of mean survival of bacterial cells at a particular time interval
 266 in the presence of the extract.

3.2 Discussion

The antibacterial activity of the leaf extract of *Dacryodes edulis* was determined against fourteen microorganism comprising of both Gram positive and Gram negative bacteria. The result of this investigations shows that the extract exhibited *in vitro* antibacterial activity against eleven out of fourteen of the tested bacterial isolates at a final concentration of 35 mg/mL (Table 1). The test organisms inhibited by the extract were *Escherichia coli*, *Bacillus polymyxa*, *Enterococcus faecalis*, *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Shigella* sp., *Bacillus stearothermophilus*, *Klebsiella pneumoniae*, *Citrobacter freundii*, *Micrococcus luteus* and *Corynebacterium pyogenes* (Table 1). The zones of inhibitions exhibited by the extract against the susceptible bacteria ranged between 12 mm and 23 mm while the zone of inhibition exhibited by streptomycin against the bacterial isolates ranged between 13 mm and 28 mm and that of ampicillin was between 18 mm and 29 mm (Table 1).

The minimum inhibitory concentrations (MICs) exhibited by the extract against the susceptible test organisms ranged between 0.27 mg/mL for *Pseudomonas aeruginosa*, *Shigella* sp. and *Kleb. pneumoniae*, 0.55 mg/mL for *B. stearothermophilus* and 1.09 mg/mL for *E. faecalis*. On the other hand, the MIC of 2.19 mg/mL was observed for *Bacillus polymyxa*, *Micrococcus luteus*, *Staph. aureus*, *E. coli*, *Citrobacter freundii*, while it was 4.375 mg/mL for *Corynebacterium pyogenes* (Table 2). The minimum bactericidal concentrations exhibited by the extract against the susceptible test organisms ranged between 0.55 mg/mL for *Ps. aeruginosa*, *Shigella* sp. and *Kleb. pneumoniae*, 2.19 mg/mL for *B. stearothermophilus* and *E. faecalis* and 4.375 mg/mL for *Staph. aureus* and *Citrobacter freundii*, 8.75 mg/mL for *B. polymyxa* and *Micrococcus luteus* and 17.50 mg/mL was observed for *E. coli* and *Corynebacterium pyogenes* (Table 2).

290 The phytochemical analysis of the leaf extracts of *D. edulis* revealed the presence of
 291 alkaloids, flavonoids, tannins and saponins (Table 3). These phytochemical compounds are
 292 known to be biologically active and thus contributed to the antibacterial and antioxidant
 293 properties of *D. edulis*. Studies have shown that saponins although non-toxic can generate
 294 adverse physiological responses in animal that consume them. They exhibit cytotoxic effect
 295 and growth inhibition against a variety of cell which make them have anti-inflammatory and
 296 anti-cancer properties. They also show tumour inhibiting activity in animals [18]. Alkaloid
 297 which was revealed in the extract of *Dacryodes edulis* have been widely studied for their
 298 potential use in elimination of human cancer cell lines [19]. Pure isolate alkaloids and their
 299 synthetic derivatives have been used as analgesic, anti-spasmodic and bactericidal agent [20].
 300 Flavonoid also detected in the leaf of *Dacryodes edulis* are known to possess antibacterial,
 301 anti-inflammatory, anti-allergic, anti-mutagenic, antiviral, anti-thrombotic, vasodilatory and
 302 anti-oxidant properties [21]. Flavonoids ability of scavenging hydroxyl radicals, super oxide
 303 anions and lipid peroxyradicals highlight many of the flavonoids health promoting functions
 304 in organisms, which are important for prevention of diseases associated with oxidative
 305 damage of membrane, proteins and DNA [22]. Flavonoids in human diet may reduce risk of
 306 various cancer as well as preventing menopause symptoms [21]. Tannins present in the
 307 *Dacryodes edulis* have been studied to be one of the major phytochemical found in many
 308 higher plants. Tannins have a characteristic strange smell and bitter taste. Tannins could bind
 309 to proteins through the effective formation of strong complexes with proteins and other
 310 macromolecules. Thus, they could have a major impact on animal nutrition, including
 311 inhibition of growth rate digestive enzymes [23]. Tannins have been implicated with various
 312 pharmaco-therapeutic effects [24]. Tannins are useful as an anti-inflammatory agent and in
 313 the treatment of burns and other wounds based on their anti-hemorrhagic and antiseptic
 314 potentials [25]. Tannins-rich remedies are used as antihelminthics, antioxidants,

315 antimicrobials, antivirals, in cancer chemotherapy and to chelate dietary iron [26] [27] [28]
 316 [29]. Results observed in this study support the usefulness of *Dacryodes edulis* in folklore
 317 remedies.

318 The result of the antioxidant property test of the leaf of *Dacryodes edulis* showed that the
 319 inhibition of DPPH scavenging radicals increases with increasing concentrations of the
 320 extract (Figure 2). This is an indication that this extract is rich in flavonoids which is an anti-
 321 stress and anti-ageing. *Dacryodes edulis* may be useful in production of anti-ageing drugs.
 322 And it has been reported that *D. edulis* is rich in vitamins A, B and E which are useful against
 323 skin wrinkling [30]. The results of the findings support the usefulness of *Dacryodes edulis* in
 324 folklore remedies.

325 *Klebsiella pneumoniae*, *Citrobacter freundii*, *Shigella* sp., *Enterococcus faecalis* are
 326 pathogens that are responsible for various ailment in man. These pathogens were susceptible
 327 to the leaf of *Dacryodes edulis*. This is an indication that drugs formulated from this plant
 328 may be useful in eradicating diseases caused by these pathogens.

329 The leaf extract of *Dacryodes edulis*, compared favourably with streptomycin and/or
 330 ampicillin used as standard control in this work. For example, the zone of inhibition exhibited
 331 against *Enterococcus faecalis* was 14 mm for the extract and 19 mm, 18 mm for streptomycin
 332 and ampicillin respectively. Also, *Micrococcus luteus* showed a zone of 12 mm relative to the
 333 streptomycin standard, which showed a zone of 14mm and lastly, *Corynebacterium pyogenes*
 334 with a zone of 19 mm relative to both streptomycin and ampicillin standard which showed
 335 zones of 21 mm and 27 mm respectively. In some cases where the streptomycin and
 336 ampicillin standard did not show any activity, the extract did. For example, where the
 337 streptomycin showed no inhibition against *E. coli* but the extract inhibited the growth of this

organism. Again, ampicillin was not able to inhibit the growth of *Pseudomonas aeruginosa* but the extract did.

The test organisms comprised of both Gram positive and Gram negative and the extract was sensitivity against both groups which indicates that *D. edulis* leaf extract has a broad spectrum activity. Drug formulated from this plant extract may go a long way in combating wide range of these pathogenic bacteria.

The extract exhibited appreciable kill rate against *Staph. aureus* and *E. coli*. As the concentrations of the extract increased with increase in contact time, there appears to be an increase in populations of test bacteria killed (Figures 2 - 3). This is an indication of monophasic effect exhibited by the extract which was similar to the effect reported by Akinpelu *et al.* (2008)[31]. The ability of this extract to eliminate the test organisms within the shortest time is an indication that such extract can be used for the production of antimicrobial compound for the treatment of infections caused by these pathogens.

4.0 Conclusions

The leaf extract of *Dacryodes edulis* was found to possess broad spectrum antibacterial activity. The leaf extract of *Dacryodes edulis* found to contain flavonoids, alkaloids, saponins and tannins which have been confirmed to possess antimicrobial activities, has hence supported the reason for the antibacterial and antioxidant efficacies of this plant.

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