

# Differential miRNA Expression in Oral Cancer Oncosomes: A Pilot *In Vitro* Study

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Original Research Article

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## ABSTRACT

**Aims:** Exosomes are small membranous secreted vesicles (30-120 nm) believed to function as intercellular messengers delivering their cargo of RNA and protein to target cells. While many cells secrete exosomes, cancer cells have been found to produce higher numbers of exosomes than normal cells. Cancer specific exosomes, also termed oncosomes, transport intercellular bioactive molecules including proteins, lipids, and microRNAs (miRNA), the latter of which are discarded into the extracellular environment via exosomes. These bioactive molecules can modulate oral squamous cell carcinomas (OSCC) disease progression *in vivo*. To date, only one study had

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demonstrated the secretion of oncosomes from cultured OSCC cells, therefore the objective of this study is to determine if intact oncosomes can be isolated from oral cancer cells.

**Study Design:** This is an observational laboratory-based study of human oral cancer cell cultures.

**Place and Duration of Study:** This study was conducted in the Department of Biomedical Sciences at the University of Nevada, Las Vegas – School of Dental Medicine between May 2016 and May 2017.

**Methodology:** Using a reagent that binds water and forces less-soluble lipid vesicles out of solution, oncosomes from oral cancer cell cultures (SCC4, SCC9, SCC15, SCC25 and CAL27) were collected by low-speed centrifugation. qRT-PCR was performed on RNA isolated from the culture-derived oncosomes for miR-21, miR-365, miR-155 and miR-133a1; all previously identified from cancers of other tissues.

**Results:** Normal, non-cancerous HGF-1 (human gingival fibroblasts) had low (almost) undetectable expression of miR-21, -133, -155, and -365. Oral cancer cell lines (SCC4, SCC9, SCC15, SCC25 and CAL27) had moderate to high expression of at least one microRNA – although this varied significantly by cell line.

**Conclusion:** Exosomes can be successfully isolated from OSCC conditioned media and miRNAs are detectable through TaqMan microRNA assays, with a unique characteristic expression of the miRNAs in the cell lines examined. Although more investigation is needed, potential correlations between miRNA levels and proliferation rates were also observed.

**Keywords:** oral cancer; oncosomes; microRNA.

## ABBREVIATIONS

*Ribonucleic acid (RNA); microRNA (miRNA); oral squamous cell carcinoma (OSCC); Squamous cell carcinoma (SCC); American Type Culture Collection (ATCC); Fetal Bovine Serum (FBS); Phosphate Buffered Saline (PBS); Reverse transcription (RT); Quantitative polymerase chain reaction (qPCR); 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT); deoxyribonucleotide triphosphate (dNTP); dimethyl sulfoxide (DMSO).*

## 1. INTRODUCTION

Exosomes are small membranous secreted vesicles (30-120 nm) believed to function as intercellular messengers delivering their cargo of ribonucleic acids (RNA) and protein molecules to target cells. Exosomes are secreted by numerous types of cells in culture and are found in abundance in body fluids including blood, saliva, urine, amniotic fluid, cerebrospinal fluid, nasal secretions, and breast milk [1]. While many cells secrete exosomes, cancer cells have been found to produce higher numbers of exosomes than normal cells [2]. Cancer specific exosomes, also termed oncosomes, transport of bioactive molecules including proteins, lipids, and microRNAs (miRNA), which are transferred into the extracellular environment via exosomes.

These bioactive molecules have been demonstrated to modulate oral squamous cell carcinomas (OSCC) disease progression *in vivo* [3]. In fact, oncosome miRNA, which are small non-coding RNA only containing on average 22 nucleotides, are now known to be fundamental

regulators of both mRNA and protein expression [1,2]. However, the isolation of intact exosomes is traditionally accomplished using ultracentrifugation, a method that is difficult and requires specialised equipment [4]. One objective of this research project was to determine if intact oncosomes could be isolated from OSCC cells using a reagent that binds water molecules and forces less-soluble lipid vesicles out of solution allowing the oncosomes to be collected by low-speed centrifugation.

To determine if this procedure was possible, it was necessary to focus on a few specific miRNAs for this initial pilot study. While over 750 miRNAs such as miR-30, miR-204, miR-370, miR144 and miR-193 have been found in both oncosomes and exosomes, the expression of four specific miRNAs: miR-365, miR-21, miR-155, and miR-133a-1 have been extracted from multiple types of cancers, including oral cancers [5]. These miRNAs may be responsible for differential induction of mRNA in target cells, including both up- and down-regulation of mRNA transcription and translation.

More specifically, one of the most widespread and over-expressed miRNAs in human malignancies has been identified as miR-21, an inhibitor of PTEN and pro-apoptotic genes, thereby allowing cell survival [6]. In addition, although miR-155 plays a vital role in various physiological and pathological processes such as hematopoietic lineage differentiation, immunity, inflammation, and cardiovascular diseases - some evidence also demonstrates miR-155 as overexpressed in a variety of malignant tumors as well as having a role in the spread of viral infections [7]. In contrast, miR-133 is expressed in muscle tissue and appears to repress the expression of non-muscle genes as well as SRF transcription factor and cyclin D2, which may modulate, in part, cell cycle progression [8]. Finally, although the role of miR-365 in cancer cells is still controversial, recent evidence may suggest an alternative, protective role that may inhibit growth, invasion and metastasis of malignant melanomas – although these functions are not well understood [9].

Squamous cell carcinomas (SCC) of the tongue represent a significant proportion of solid cancers that are diagnosed in late (metastasis forming) stages, therefore the ability to identify any prognostic indicators such as the production and presence of specific miRNA-containing oral oncosomes detectable from saliva could greatly improve detection and screening measures and help to improve clinical outcomes. To date, only one study had demonstrated the secretion of oncosomes from cultured OSCC cells, therefore the objective of this study is to determine if intact oncosomes can be isolated from oral cancer cells *in vitro*.

## 2. METHODOLOGY

### 2.1 Cell Culture

Cell lines that were available for this study (already purchased and cultured) included the human oral squamous cell carcinoma lines, SCC-4 (CRL-1624), SCC-9 (CRL-1629), SCC-15 (CRL-1623), SCC-25 (CRL-1628), and CAL-27 (CRL-2095) were originally obtained from the American Type Culture Collection (ATCC: Manassas, VA). The normal human gingival fibroblast cell line HGF-1 (CRL-2014) was also previously obtained from ATCC and used as a normal control for comparison. All cell lines were maintained in Dulbecco's Modified Eagles' Medium (Hyclone, Logan, UT) containing 4.0 mM L-glutamine, 4.5 g/L glucose, and 110 mg/L

sodium pyruvate. Medium was supplemented with penicillin (100 units/ml) and streptomycin (100 ug/ml) both from Hyclone and 10% fetal bovine serum (FBS). Cells were cultured in 75 cm<sup>2</sup> or 25 cm<sup>2</sup> BD Falcon tissue-culture treated flasks (Bedford, MA) and grown at 37°C and 5% CO<sub>2</sub> in humidified incubators. Cell cultures were passaged during log phase growth at approximately 80% confluence at a sub-cultivation ratio of 1:3.

### 2.2 Exosome Isolation

Intact exosomes were isolated from cell culture. Cells were cultured in T75 flasks and switched to medium containing exosome depleted FBS and incubated for 24 hours before harvest. The conditioned medium was centrifuged at 2000 x g for 30 minutes to remove cells and debris. The medium was decanted and mixed with 0.5 volumes of Total Exosome Isolation reagent (Life Technology) before refrigerating overnight. To pellet the exosomes, the conditioned medium was centrifuged at 10,000 x g for one hour at 4°C. The pellet was resuspended in 200µL of 1X Phosphate Buffered Saline (PBS).

### 2.3 Exosome RNA Extraction

One volume of 2X Denaturing solution (Life Technology) was added to the resuspended exosomes before incubated on ice for 5 minutes. One volume of Acid-Phenol: Chloroform was added and the solution centrifuged at 4°C for 5 minutes at 10 x g. The upper aqueous phase was carefully removed and 1.25 volumes of 100% ethanol added. The sample was pipetted into a filter and centrifuged at 10 x g for 15 seconds. The filter was then washed with miRNA Wash Solution 1 (Life technology) and centrifuged for 15 seconds, this same process was repeated twice more with Wash Solution 2/3. The filter apparatus was then centrifuged on its own for 15 seconds to remove any residual fluid from the filter. The filter was then placed into a new collection tube and 100 µL of heated RNase water was applied to the filter before being centrifuged for 30 seconds. The raw RNA was contained in the flow-through.

### 2.4 TaqMan MicroRNA Assays

Reverse Transcriptions (RT) carried out were 15 µL reactions with a master mix consisting of 100 mM deoxyribonucleotide triphosphate (dNTPs), 10X reverse Transcription Buffer, RNase Inhibitor, and MultiScribe Reverse Transcriptase added to that mixture was 3 µL of the miR

specific primer. The thermal cycler was set to 16°C for 30 minutes, 42°C for another 30 minutes, then 85°C for five minutes before cooling to 4°C. Next a quantitative polymerase chain reaction (qPCR) was carried out using 20 µL reactions consisting of 20x TaqMan Small RNA assay, TaqMan Universal PCR Master Mix II and 1.33 µL of the corresponding product from the RT-PCR reaction. The thermal conditions for the reactions were 50°C for 2 minutes, 95°C for 10 minutes, then 40 cycles of 95°C for 15 seconds and 60°C for 60 seconds. Standard curves were made doing a five-fold serial dilution of CAL-27 cDNA for miR-365, miR-21 and miR-155 while standard curves for miR-133a was done with SCC-9 cDNA.

Thermo Scientific Verso 1-Step RT-PCR: Reverse transcription and PCR were carried out together in a single 25 µL reaction containing 2.5 µL of exosomal RNA, the desired forward and reverse primers, RT enhancer, 2X RT-PCR buffer, and versa reverse transcriptase enzyme. Thermal conditions for the reactions followed 50°C for 30 minutes, 95°C for 2 minutes then 35 cycles of 95°C for 20 seconds, 55°C for 30 seconds and a 72°C extension for 1 minute.

## 2.5 Vybrant MTT Cell Proliferation Assay

Cells were plated at a density of  $1 \times 10^4$  cells/well in triplicate on a 96 well plate. One plate was incubated for 24 hours, the second for 48 hours, and the third for 72 hours. When harvesting, first, all medium was replaced with 100 µL of fresh, phenol red-free medium. Then, 10 mM 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide or MTT stock solution was added to each well. A negative control was added to 6 wells containing just the medium alone. The plate was incubated at 37°C for 4 hours. After incubation, all but 25 µL of medium was removed from the cells before 50 µL of dimethyl sulfoxide (DMSO) was added to each well. The plate was then incubated again for 10 minutes before the absorbance reading at 540nm.

## 3. RESULTS

All cell lines were incubated in complete media supplemented with exosome-depleted FBS, which resulted in visible exosome precipitation pellets for exosomal RNA isolation and subsequent one-step RT-PCR experiments (Fig. 1). TaqMan MicroRNA Assays for miR-365, miR-21, miR-155, and miR-133 were performed and standard curves for each of the miRNAs were generated from cDNAs, which allowed relative quantitation of each of the target miRNAs among these cell lines (Table 1).

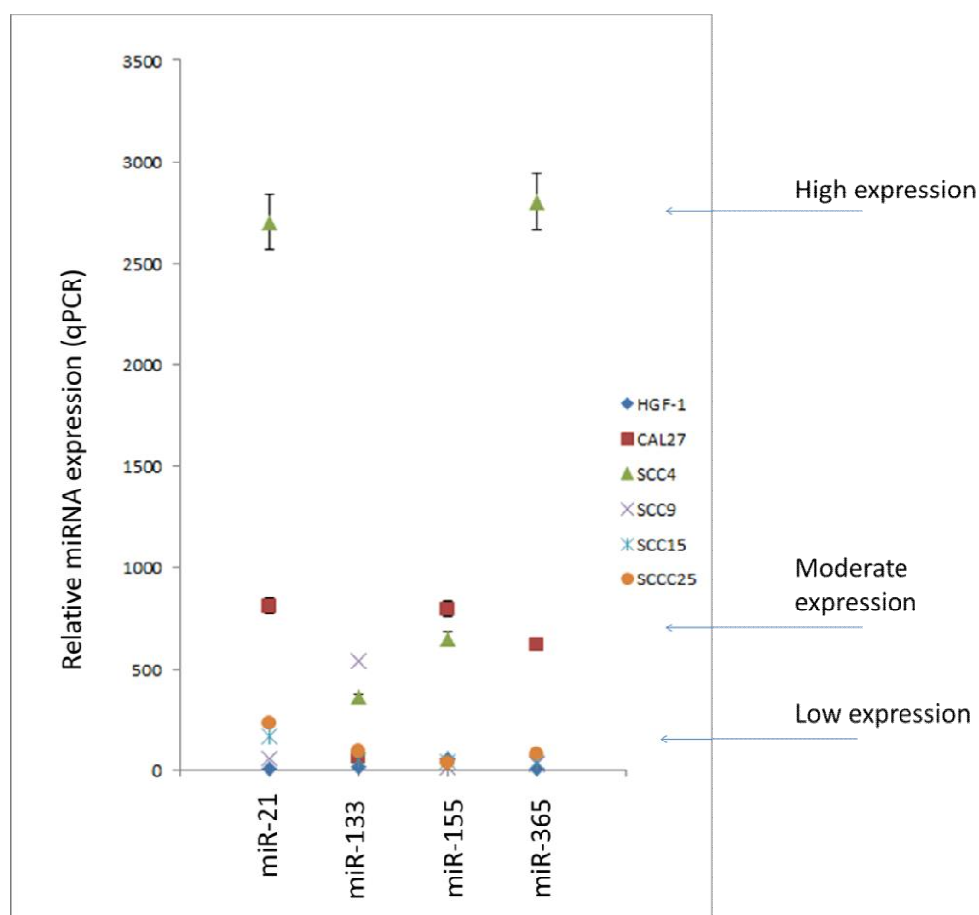
Normal HGF cells contained very low levels of the four target miRNAs with cycle threshold-exceeding 30 cycles. The OSCC cell line, CAL-27 demonstrated moderate levels of all the examined miRNAs with the exception of miR-133 which was 8-fold less abundant. The expression profile of the miRNAs in the OSCC cell line, SCC-9, was remarkably different demonstrating low levels of miR-365, miR-155, and miR-21 with moderate levels of miR-133.

Both SCC-15 and SCC-25 contained approximately 4-fold more miR-21 than the other miRNAs examined, although most levels were comparatively low. SCC-4 cells contained the highest levels of the miRNAs examined with very high levels of miR-365 and miR-21 and moderate expression of miR-155 and miR-133.

These quantitative data derived from the qPCR were compiled and microRNA levels categorised with the relative-fold expression (RFE) as Low (RFE < 100), Moderate (RFE =100-1000), and High (RFE > 1000) (Table 2). These data demonstrated that each oral cancer cell line expressed moderate or high levels of at least one microRNA, with four of the five expressing moderate or high levels of miR-21. Two oral cancers (SCC-4, SCC-9) expressed moderate levels of miR-133, while two cell lines (SCC-4,

**Table 1. Cell line characteristics**

Cell line	Tissue of origin	Phenotype
HGF-1 (CRL-2014)	Gingival fibroblast	Normal
SCC-4 (CRL-1624)	Tongue, epithelial	Squamous cell carcinoma
SCC-9 (CRL-1629)	Tongue, epithelial	Squamous cell carcinoma
SCC-15 (CRL-1623)	Tongue, epithelial	Squamous cell carcinoma
SCC-25 (CRL-1628)	Tongue, epithelial	Squamous cell carcinoma
CAL-27 (CRL-2095)	Tongue, epithelial	Squamous cell carcinoma



**Fig. 1. Relative miRNA expression of oral cancers.** Quantitative qPCR from exosome-derived cDNAs allowed relative quantitation of miR-21, miR-133, miR-155 and miR-365. Low-level expression was observed in HGF-1 (normal) cells, with differential expression observed among the oral cancer cell lines. High-level expression of miR-21 and miR-365 was observed among SCC-4 cells, which exhibited moderate expression of miR-133 and miR-155. CAL-27 cells exhibited moderate expression of all microRNAs except miR-133. Moderate expression of miR-133 was observed among SCC-9 cells

**Table 2. Quantification of differential miRNA-containing oral cancer exosomes**

	miR-21	miR-133	miR-155	miR-365
HGF-1 (normal)	LOW (RFE = 9)	LOW (RFE=13)	LOW (RFE=57)	LOW (RFE=9)
SCC-25	<b>MOD</b> (RFE=230)	LOW (RFE=10)	LOW (RFE=40)	LOW (RFE=75)
SCC-15	<b>MOD</b> (RFE=160)	LOW (RFE=50)	LOW (RFE=30)	LOW (RFE=75)
SCC-9	LOW (RFE=40)	<b>MOD</b> (RFE=500)	LOW (RFE=30)	LOW (RFE=20)
SCC-4	<b>HIGH</b> (RFE=2800)	<b>MOD</b> (RFE=400)	<b>MOD</b> (RFE=600)	<b>HIGH</b> (RFE=2800)
CAL-27	<b>MOD</b> (RFE=800)	LOW (RFE=30)	<b>MOD</b> (RFE=800)	<b>MOD</b> (RFE=650)

CAL-27) expressed moderate levels of miR-155. Finally, two oral cancers expressed moderate or high levels of miR-365 (SCC-4, CAL-27).

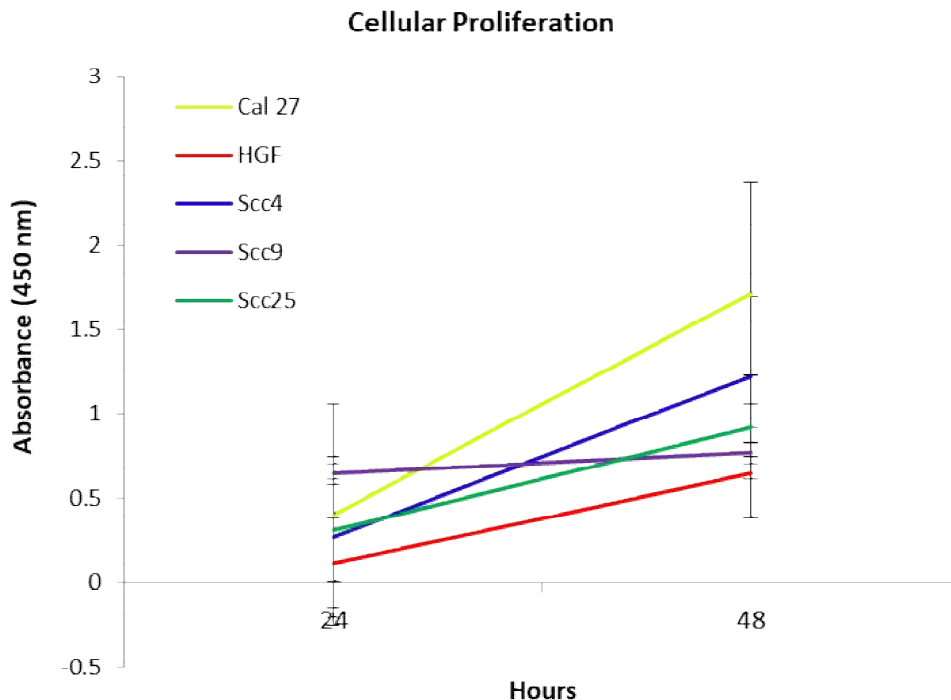
In addition, cellular proliferation was measured for all cell lines to determine if any associations might exist between growth rate and miRNA expression (Fig. 2). These data revealed that the cell lines exhibiting the highest RFE levels of miRNAs were also the most rapidly dividing cell lines (CAL-27, SCC-4). Moreover, the cell line with the slowest relative growth overall (HGF-1) was also the only culture that exhibited low miRNA expression for all the microRNAs examined. The other cell lines with moderate expression (SCC-9, SCC25) were also found to exhibit more modest growth in comparison with the CAL-27 and SCC-4 cell lines.

#### 4. DISCUSSION

MicroRNAs hold great promise as potential biomarker and prognostic indicators for various

cancers [11,12]. In addition, a better understanding of the molecular basis of miRNA expression as they relate to tumorigenesis and metastatic potential is necessary to advance the potential of miRNAs for improving clinical outcomes and for use as anti-cancer agents. [13,14].

Of the miRNAs examined in this study, miR-21 has been the most extensively investigated to date. miR-21 acts as an oncogene by decreasing expression of PTEN ultimately promoting cell cycle progression and proliferation and inhibiting apoptosis via an AKT-dependent pathway [15,16]. Many independent studies have demonstrated overexpression of miR-21 in biopsies of head and neck tumors [10,17,18]. In addition, some studies have reported that miR-21 expression may be an independent prognostic factor indicating poor patient survival in tongue squamous cell carcinomas [19-21]. In the present study, miR-21 was found to be overexpressed in four of the five OSCC cell lines examined.



**Fig. 2. Proliferation results of HGF-1, CAL-27, SCC-4, SCC-9, and SCC-25. Examination of cellular growth demonstrated some cells with rapid growth (CAL-27, SCC-4), some with more moderate growth (SCC-9, SCC-25), with the slowest growth observed among the normal human gingival fibroblast (HGF-1)**

Interestingly, few studies have evaluated miR-133 in oral or head and neck cancers [10,22]. These studies found miR-133 levels to be downregulated in recurrent head and neck tumors but may be otherwise highly expressed. This study demonstrated low RFE for miR-133 in three of the five oral cancers evaluated, with CAL-27 (the fastest growing) exhibiting among the lowest expression. However, the low levels observed among the normal and slower growing cancers may not directly compare with studies of other cancers, which may suggest further research into these observations is warranted.

More research has evaluated miR-155 and the role in oral cancer disease progression [23-25]. These findings have revealed miR-155 expression is correlated with poor clinical prognosis and disease progression [26,27]. The results of this study demonstrated that the most rapidly growing cell lines (CAL-27, SCC-4) exhibited the highest RFE of miR-155, which may suggest these findings support previous observations in other oral tumors.

Finally, the exact role of miR-365 in head and neck cancers remains unknown. Research suggests miR-365 may inhibit growth, invasion, and metastasis of some cancers, such as lung cancer [28]. However, other studies have suggested miR-365 may be associated with poor outcomes and survival in pancreatic cancers [29]. The results of this study demonstrated that only the rapidly dividing cell lines CAL-27 and SCC-4 had a moderate or high expression of miR-365, while the relatively slower-growing cell lines all exhibited low expression. This may suggest that miR-365 functions in oral cancers in a similar manner to pancreatic cancers – although more research into these observations is clearly warranted.

Although these results clearly demonstrated novel data regarding miRNA expression among oral cancer oncosomes, several limitations should also be considered when evaluating this information. First, this study involved two commercially available oral cancer cell lines, due to funding and other limitations. Future studies might include additional oral cancer cell lines in order to confirm these initial findings. In addition, no primary tumor isolates were available to the study authors, which may further limit the clinical inferences that could be drawn from these data. Finally, additional diversity of miRNA screening may be possible in future studies, as the functions and structures of new miRNAs are

identified and characterised that may potentially influence cellular behaviours and phenotypes.

## 5. CONCLUSIONS

To date, only one study had demonstrated the secretion of oncosomes from cultured OSCC cells [30]. This study demonstrated that exosomes can be successfully isolated from OSCC conditioned media and that microRNA expression can be quantified and evaluated using TaqMan microRNA assays. While more investigation will be needed, potential correlations between miRNA levels and proliferation rates have been observed that suggest miR-21, miR-155 and miR-365 may be useful biomarkers to evaluate potential growth and proliferation of oral cancers.

## CONSENT AND ETHICAL APPROVAL

It is not applicable.

## COMPETING INTERESTS

Authors have declared that no competing interests exist.

## REFERENCES

1. Principe S, Hui AB, Bruce J, Sinha A, Liu FF, Kislinger T (2013) Tumor-derived exosomes and microvesicles in head and neck cancer: implications for tumor biology and biomarker discovery. *Proteomics* 13 (10-11):1608-1623. doi:10.1002/pmic.201200533
2. Taylor DD, Gercel-Taylor C (2008) MicroRNA signatures of tumor-derived exosomes as diagnostic biomarkers of ovarian cancer. *Gynecologic oncology* 110 (1):13-21. doi:10.1016/j.ygyno.2008.04.033
3. Hannafon BN, Ding WQ (2013) Intercellular Communication by Exosome-Derived microRNAs in Cancer. *International journal of molecular sciences* 14 (7):14240-14269. doi:10.3390/ijms140714240
4. Zhang Z, Wang C, Li T, Liu Z, Li L (2014) Comparison of ultracentrifugation and density gradient separation methods for isolating Tca8113 human tongue cancer cell line-derived exosomes. *Oncology letters* 8 (4):1701-1706. doi:10.3892/ol.2014.2373

5. Mathivanan S, Fahner CJ, Reid GE, Simpson RJ (2012) ExoCarta 2012: database of exosomal proteins, RNA and lipids. *Nucleic acids research* 40 (Database issue):D1241-1244. doi:10.1093/nar/gkr828
6. Meng F, Henson R, Wehbe-Janek H, Ghoshal K, Jacob ST, Patel T (2007) MicroRNA-21 regulates expression of the PTEN tumor suppressor gene in human hepatocellular cancer. *Gastroenterology* 133 (2):647-658. doi:10.1053/j.gastro.2007.05.022
7. Faraoni I, Antonetti FR, Cardone J, Bonmassar E (2009) miR-155 gene: a typical multifunctional microRNA. *Biochimica et biophysica acta* 1792 (6):497-505. doi:10.1016/j.bbadis.2009.02.013
8. Liu N, Bezprozvannaya S, Williams AH, Qi X, Richardson JA, Bassel-Duby R, Olson EN (2008) microRNA-133a regulates cardiomyocyte proliferation and suppresses smooth muscle gene expression in the heart. *Genes & development* 22 (23):3242-3254. doi:10.1101/gad.1738708
9. Bai J, Zhang Z, Li X, Liu H (2015) MicroRNA-365 inhibits growth, invasion and metastasis of malignant melanoma by targeting NRP1 expression. *International journal of clinical and experimental pathology* 8 (5):4913-4922
10. Courthod G, Franco P, Palermo L, Pisconti S, Numico G (2014) The role of microRNA in head and neck cancer: current knowledge and perspectives. *Molecules* 19 (5):5704-5716. doi:10.3390/molecules19055704
11. Jiang S, Hu C, Liu P, Lu M. Tumor-derived exosomes in cancer metastasis risk diagnosis and metastasis therapy. *Clin Transl Oncol*. 2018 Jul 26. doi: 10.1007/s12094-018-1918-0. [Epub ahead of print] Review. PMID: 30051211
12. Wang D, Yang S, Wang H, Wang J, Zhang Q, Zhou S, He Y, Zhang H, Deng F, Xu H, Zhong S, Fu L, Tang J. The progress of circular RNAs in various tumors. *Am J Transl Res*. 2018 Jun 15;10(6):1571-1582. eCollection 2018. Review. PMID: 30018701
13. Dragomir M, Calin GA. Circular RNAs in Cancer - Lessons Learned From microRNAs. *Front Oncol*. 2018 May 28;8:179. doi: 10.3389/fonc.2018.00179. eCollection 2018. Review. PMID: 29911069
14. Nebbioso A, Tambaro FP, Dell'Aversana C, Altucci L. Cancer epigenetics: Moving forward. *PLoS Genet*. 2018 Jun 7;14(6):e1007362. doi: 10.1371/journal.pgen.1007362. eCollection 2018 Jun. Review. PMID: 29879107
15. Liu H, Cheng L, Cao D, Zhang H. Suppression of miR-21 Expression Inhibits Cell Proliferation and Migration of Liver Cancer Cells by Targeting Phosphatase and Tensin Homolog (PTEN). *Med Sci Monit*. 2018 May 29;24:3571-3577. doi: 10.12659/MSM.907038. PMID: 29807978
16. Wang P, Guan Q, Zhou D, Yu Z, Song Y, Qiu W. miR-21 Inhibitors Modulate Biological Functions of Gastric Cancer Cells via PTEN/PI3K/mTOR Pathway. *DNA Cell Biol*. 2018 Jan;37(1):38-45. doi: 10.1089/dna.2017.3922. Epub 2017 Nov 29. PMID: 29185784
17. Sannigrahi MK, Sharma R, Panda NK, Khullar M. Role of non-coding RNAs in head and neck squamous cell carcinoma: A narrative review. *Oral Dis*. 2017 Sep 21. doi: 10.1111/odi.12782. [Epub ahead of print] Review. PMID: 28941018
18. Tseng HH, Tseng YK, You JJ, Kang BH, Wang TH, Yang CM, Chen HC, Liou HH, Liu PF, Ger LP, Tsai KW. Next-generation Sequencing for microRNA Profiling: MicroRNA-21-3p Promotes Oral Cancer Metastasis. *Anticancer Res*. 2017 Mar;37(3):1059-1066. PMID: 28314265
19. Li J, Huang H, Sun L, Yang M, Pan C, Chen W, Wu D, Lin Z, Zeng C, Yao Y, Zhang P, Song E (2009) MiR-21 indicates poor prognosis in tongue squamous cell carcinomas as an apoptosis inhibitor. *Clinical cancer research : an official journal of the American Association for Cancer Research* 15 (12):3998-4008. doi:10.1158/1078-0432.CCR-08-3053
20. Troiano G, Mastrangelo F, Caponio VCA, Laino L, Cirillo N, Lo Muzio L. Predictive Prognostic Value of Tissue-Based MicroRNA Expression in Oral Squamous Cell Carcinoma: A Systematic Review and Meta-analysis. *J Dent Res*. 2018 Jul;97(7):759-766. doi: 10.1177/0022034518762090. Epub 2018 Mar 13. PMID: 29533734
21. Supic G, Zeljic K, Rankov AD, Kozomara R, Nikolic A, Radojkovic D, Magic Z. miR-183 and miR-21 expression as biomarkers of progression and survival in tongue



- carcinoma patients. *Clin Oral Investig*. 2018 Jan;22(1):401-409. doi: 10.1007/s00784-017-2126-y. Epub 2017 Jun 14. PMID: 28616749
22. Citron F, Armenia J, Franchin G, Polesel J, Talamini R, D'Andrea S, Sulfaro S, Croce CM, Klement W, Otasek D, Pastrello C, Tokar T, Jurisica I, French D, Bomben R, Vaccher E, Serraino D, Belletti B, Vecchione A, Barzan L, Baldassarre G. An Integrated Approach Identifies Mediators of Local Recurrence in Head and Neck Squamous Carcinoma. *Clin Cancer Res*. 2017 Jul 15;23(14):3769-3780. doi: 10.1158/1078-0432.CCR-16-2814. Epub 2017 Feb 7. PMID: 28174235
  23. Fu S, Chen HH, Cheng P, Zhang CB, Wu Y. MiR-155 regulates oral squamous cell carcinoma Tca8113 cell proliferation, cycle, and apoptosis via regulating p27Kip1. *Eur Rev Med Pharmacol Sci*. 2017 Mar;21(5):937-944. PMID: 28338203
  24. Zeng Q, Tao X, Huang F, Wu T, Wang J, Jiang X, Kuang Z, Cheng B. Overexpression of miR-155 promotes the proliferation and invasion of oral squamous carcinoma cells by regulating BCL6/cyclin D2. *Int J Mol Med*. 2016 May;37(5):1274-80. doi: 10.3892/ijmm.2016.2529. Epub 2016 Mar 16. PMID: 26986233
  25. Baba O, Hasegawa S, Nagai H, Uchida F, Yamatoji M, Kanno NI, Yamagata K, Sakai S, Yanagawa T, Bukawa H. MicroRNA-155-5p is associated with oral squamous cell carcinoma metastasis and poor prognosis. *J Oral Pathol Med*. 2016 Apr;45(4):248-55. doi: 10.1111/jop.12351. Epub 2015 Aug 26. PMID: 26307116
  26. Jamali Z, Asl Aminabadi N, Attaran R, Pournagiazar F, Ghertasi Oskouei S, Ahmadpour F. MicroRNAs as prognostic molecular signatures in human head and neck squamous cell carcinoma: a systematic review and meta-analysis. *Oral Oncol*. 2015 Apr;51(4):321-31. doi: 10.1016/j.oraloncology.2015.01.008. Epub 2015 Feb 9. Review. PMID: 25677760
  27. Manikandan M, Deva Magendhra Rao AK, Rajkumar KS, Rajaraman R, Munirajan AK. Altered levels of miR-21, miR-125b-2\*, miR-138, miR-155, miR-184, and miR-205 in oral squamous cell carcinoma and association with clinicopathological characteristics. *J Oral Pathol Med*. 2015 Nov;44(10):792-800. doi: 10.1111/jop.12300. Epub 2014 Dec 8. PMID: 25482863
  28. Zhan B, Lu D, Luo P, Wang B. Prognostic Value of Expression of MicroRNAs in Non-Small Cell Lung Cancer: A Systematic Review and Meta-Analysis. *Clin Lab*. 2016 Nov 1;62(11):2203-2211. doi:10.7754/Clin.Lab.2016.160426. Review. PMID: 28164670
  29. Ebrahimi S, Hosseini M, Ghasemi F, Shahidsales S, Maftouh M, Akbarzade H, Parizadeh SA, Hassanian SM, Avan A. Circulating microRNAs as Potential Diagnostic, Prognostic and Therapeutic Targets in Pancreatic Cancer. *Curr Pharm Des*. 2016;22(42):6444-6450. doi:10.2174/1381612822666160817095047. Review. PMID: 27539232
  30. Petersen B, Kingsley K. Differential expression of miR-21, miR-133 and miR-155 from exosome fractions isolated from oral squamous cell carcinomas in vitro. *J Med Discov*. 2016, 1(1): jmd16010.