4

5 6 Mini review

THERAPAUTICS

7 8 9

10 . 11 **ABSTRACT**

12

Background and objectives: Designing of effective anticancer-based peptides has been challenging these days probably due to the instability of the peptides in serum as well as low sensitivity and resistance of cancer cells to these peptides.

Methodology: published papers addressing anticancer activity of lactaptin has been review extensively.

Results: Lactaptin, a peptide from proteolytic cleave of kappa casein human milk has been shown to play a number of roles including Program cell death, Genetic material fragmentation, suppression of metastasis, cytotoxicity to the cancer cells as well as caspases activation.

Conclusions: Lactaptin shows promising activity against cancer cells. Accordingly, enhancing lactaptin activity will greatly enhance it therapeutic efficacy and will provide insight into designing of new therapeutics.

13 14

Keywords: Lactaptin, Apoptosis, DNA fragmentation, Cytotoxicity, Caspases

15 16

1. INTRODUCTION

17 18

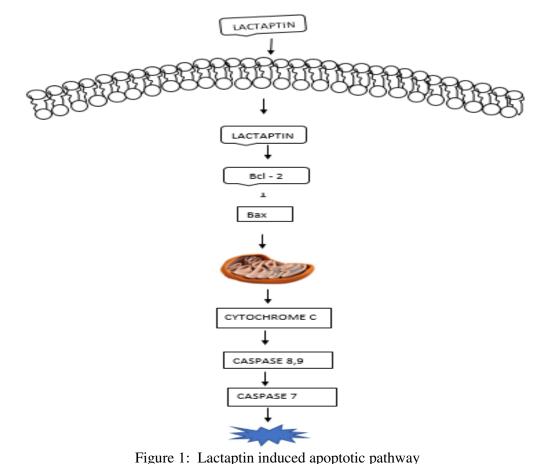
19 Designing of an effective anticancer molecule remains one of the challenges in this century probably 20 due to insensitivity of conventional chemotherapeutics or resistance by the cancer cells [1]. Anticancer 21 molecules that are stable in the serum and are very specific to cancer cells in term of toxicity are very 22 necessary to improve the efficacy of the existing molecules and developed new candidates. Lactaptin, a candidate generated from proteolytic Cleave of human milk kappa-casein has shown a promising 23 24 tumoricidal activity against variety of cancer cells [2,3]. First isolated, purified and characterized by 25 Nekipelaya and colleagues 10 years ago [2], it is made up of 74 amino acids with molecular weight of 8.6 kDa [2]. Several analogs of this peptide have been designed out of which only RL2 with amino 26 27 acids sequences similar to that of lactaptin was able to induce cancer cell death invitro in both human 28 and mice without affecting normal cells. This recombinant peptide refers to as lactaptin analog (RL2), 29 have it tumoricidal activity tested on cultured human cell [4,5]. It demonstrated tumoricidal activity against cancer metastasis of both human and mice cancer cells and also induces apoptosis in the 30 both cells [6,7]. The safety of this peptide has also been investigated, the pharmacokinetics as well as 31 the toxicity of the peptide has been studied [8]. It was shown that lactaptin is safely distributed and 32 biodistribution reduces the concentration available to cancer cell [8, 9]. This bioactive peptide, from 33 human milk has many functions including cancer cell lysis, apoptosis, suppression of metastasis, 34 35 activation of caspases among many others. The purposes of this paper are to discuss the tumoricidal 36 activity of Lactaptin, mechanisms of actions, prospects and challenges.

37

38 2. MECHANISM OF LACTAPTIN INDUCE PROGRAM CELL DEATH

- 39
- 40

The most noted event in the biochemical event of lactaptin induced program cell death were activation of caspases, changes of cytoplasmic membrane and mitochondrial membrane dissipation. It was also 43 found that program cell death was accompanied by released of phosphatidyl serine in the plasma 44 membrane. Koval et.al. reported that the recombinant analogue of lactaptin downregulates Bcl-2 expression and induces program cell death independent of p53 [6,7,11]. Presently, identified 45 pathways of the cell entrance of peptides and proteins into cells include endocytosis facilitated by 46 47 specific cell structures and direct entrance across the cell membrane made possible by properties of 48 these peptides. It has been demonstrated that RL2 gains entry into human neoplastic and non-49 neoplastic cells through binding onto skeletal cell structures. Molecular studies also indicated that Lactaptin analog cross the cell membrane partly through lipid raft mediated dynamin-independent 50 51 pinocytosis and partly through direct penetration across the plasma cell membrane. The activation of 52 the caspases by the peptide [11] might be probably due to the production of reactive oxygen species. 53 Another study of mechanism of action of different peptides has linked the activation of the caspases 54 to production of ROS followed by release of cytochrome C. Although the extrinsic pathway has not 55 been reported as mechanism of action of this peptide, the pathway might also be possible due to 56 receptors of peptides preset at cell membrane. The extrinsic pathway of cell death might start with 57 reception of signals from the peptide onto the death receptor (FADD), the peptide in this case serves 58 as a death receptor ligand equivalent in function to the CD95L and TNFa respectively. Activation of 59 the death receptor lead to the instigation of Pro-caspases 8 and 9 to caspases 8 and 9 (initiation 60 caspases) and activation of caspase 7 which ultimately leads to apoptosis.



62

61

63 64

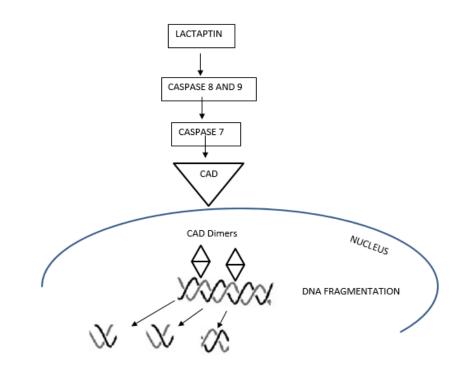
65 66

67 3. DNA FRAGMENTATION

68

69 DNA fragmentation involved the series of event which leads to the damage of genomic DNA. it often occurs following treatment of cancer cells with anti-cancer peptides capable of causing program cell death and subsequently leading to the end of cell's life. Lactaptin analog penetrates the cell through the membrane into the cytoplasm [9] favoring cellular cascades initiating this damage. Although some are seen it as not completely necessary [12, 13], genomic DNA fragmentation differentiated dead cell 74 from a living cell and facilitated dead cell uptake by phagocytosis as well as improving the whole 75 process of apoptosis [13-16]. Particular enzymes have been pinpointed in cleavage to the genomic 76 materials, out of which caspase activated DNase (CAD) located in nucleus has been in the forefront [17-19]. CAD consist of essentially proteolytic enzyme with nuclease activity that break genomic DNA 77 78 in the course of apoptosis and enhances cell diffentiation. They exert it activity through dimerization of 79 it monomers which induces the formation of sharp molecular scissor-like structure that cut double 80 stranded DNA [13]. An important mechanism of inhibition of the activity of this nuclease might be 81 prevention of the formation of this dimer, formation of dimer from CAD and ICAD (inhibitor of CAD) 82 [13] The initial mechanism of CAD activity starts with inactivation of ICAD by proteolytic cleavage and 83 irreversible inactivation of DNA repair mechanism both carried out by caspase 3 which is one of the execution caspases [13]. Lactaptin has been shown to activates caspase 7 [11], which may play the 84 85 same role as caspase 3, thus, lactaptin may causes cleavage of ICAD through activation of caspase 7 86 and inactivate DNA repair mechanism which can ultimately leads to DNA fragmentation. 87

88



89 90

91

92

Fig.2. DNA Fragmentation by CAD Dimers

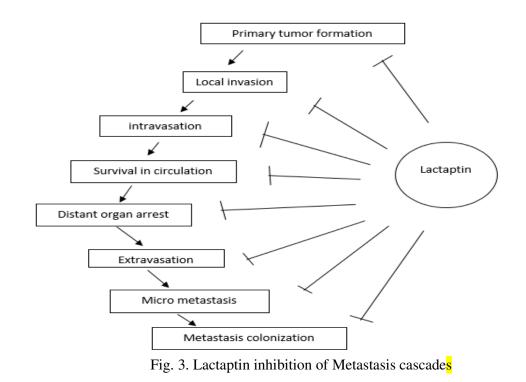
93 4. SUPPRESSION OF METASTASIS

94

95 Metastasis depict many-stage cyto-biological process involving cascades in which transformed cells 96 travels to distance tissues and adapted to the new microenvironment. In metastasis, cancer cell 97 Invade surrounding tissues and blood vessels, travel through the blood stream to reinvade nearby 98 tissues. The transformation of cell' both epigenetic and genetic material has been one of the factors 99 driven this cascade. Lactaptin as an anticancer peptide have been shown to act against varieties of cancer cells and has been shown to inhibit the metastasis of both mice and human cell triggering 100 101 them to apoptosis [6,7]. Quantitative measurement of metastatic tissues shows three times decrease 102 in hepatic metastasis of a mice administer with recombinant lactaptin analog compare with control 103 mice [6,7]. In the same vein, recurrent administration of RL2 significantly extended the life of 104 experimental animal injected intravenously with cancer cells [7]. The peptide essentially delayed 105 tumor in experimental animals against their control counterpart [7]. Inhibition of tumor Metastatic rate 106 of RL2 has been calculated as 43% [7] which indicate the antimetastatic activities of the peptide and the need to modify it and improve it therapeutic applications. 107

108 One good attribute of anti-metastatic candidate is that it must be able to block the proliferation and 109 persistence of cancer cells travelling into distant tissues and not merely stop seepage of individual 110 cells from the primary tumors [20]. Therefore, designing of new cancer therapeutics should consider this fact as many anticancer therapeutic fail to achieve this. Lactaptin should therefore be optimized to improve it therapeutic efficacy and serum stability. As noted by Gilbert and Hemann [20, 21], it is likely that vicinity of metastasis site formed resistance against anticancer therapeutics, thus become chemoprotective. It is also stated by Aguirre-Ghiso [20, 22] that agents that are toxic to cell inhibiting division cycle and active growth may be resisted by slow growing micro metastasis, in this regard, improved therapeutics are urgently needed.

Many of the biochemical event leading to the metastatic cascade can be controlled with number of intervention such as small molecules, monoclonal antibodies and miRNA [23]. Due it inhibition of metastases, lactaptin can play critical role in regulating the spread of these cancer cells by targeting this cascade (Fig. 3). The major event in metastasis cascade include primary tumor formation, local invasion, survival of the tumor in circulation, distant organ arrest, extravasation, micro metastasis formation, metastasis colonization and finally clinical detected metastasis [20] (fig 3). Lactaptin may emerge as one of the promising therapeutics that can target the whole event of metastasis cascades. In this way, our capacity to successfully treat malignancy is to a great extent subject to our ability to comprehend—and maybe even turn around—the metastasis cascades.



145

146 5. CYTOTOXIC ACTIVITY OF LACTAPTIN

147

MTT has been exploited to test the cytotoxic activity of lactaptin. The cytotoxic activity RL2 was analyzed using human endometrial cancer cells in which cultures of both normal and malignant human endometrial [6] were carried out by digestion of endometrial tissue obtained from biopsy material. Results indicated that RL2 employ it cytotoxic activities to the human endometrial cancer with apoptosis – like features [6]

153

154 6. ENHANCING OF LACTAPTIN WITH TUMOR SPECIFIC PEPTIDE

155 156

157 Enhancement of lactaptin with tumor specific peptide may play important role in helping lactaptin 158 exerting it therapeutic effects, poor penetration of cancer peptides might be the limitation of some 159 anticancer agent for exerting their activity. Systemic administration with iRGD, a tumor-targeting and 160 -penetrating peptide, enhanced the therapeutic efficacy of drugs of various compositions, including a 161 monoclonal antibody (trastuzumab), a small molecule (doxorubicin) and nanoparticles (nab-paclitaxel 162 and doxorubicin liposomes). Thus, co-administration of iRGD may be an important way to improve the 163 usefulness of anticancer drugs while decreasing their side effects, a main target of cancer treatment 164 investigations [25,26]. An in vitro study of recombinant lactaptin with a number of fusion proteins has 165 been carried out against two different cancer cell lines MCF-7 and MDA-MB-231 [25,26]. The result 166 indicates that fusion proteins enhanced with lactaptin exert inhibition of the cell lines proliferation and 167 antitumor activity of the enhanced peptide is greater than the lactaptin alone [25,26]. In vivo studies of 168 one of the fusion proteins with lactaptin, T3 - RL3 in mouse xenograft using cancer cell lines indicates 169 that T3 – RL3 inhibit tumor higher than that of the RL2 in comparison [25,26]. Accordingly, the 170 enhancement of lactaptin with tumor specific proteins will greatly increase it specificity and therefore increase it overall therapeutic effects. A number of possible modifications of peptides has been 171 172 provided [27]. This include adjustment of C- and N-terminal of peptide, hybridization, cyclization and 173 substitution of amino acids in the existing peptide [27]. Accordingly, enhancing lactaptin activity will 174 greatly enhance it therapeutic efficacy and will provide insight into designing of new therapeutics.

175 176

177 **7.** Discussion178

179 Research into the development of new anti-cancer molecules is urgently needed considering the fact 180 that resistance rises on almost all the chemotherapeutics agents used at the moments. Peptides has 181 shown promising role in resolving a number of ailments apart from cancers and Their role in cancer 182 therapeutics and beyond has been well acknowledged [26-29]. Multiple biochemical changes result 183 due to progression of cancer and recurrent abnormal signals leading to the cell proliferation. There 184 has been extensive research on milk peptides often relating these peptides to many functions 185 including anti- cancer, anti-metastasis, anti-microbial, antioxidant effects and immunomodulatory 186 properties [27]. Peptide from milk are generally tolerated with well bioavailability, a property that will 187 enhance successful treatment of cancer ailments [27]. 188

189 190 As at the moment, research and development of cancer therapeutics based on peptides obtained 191 from milk capable of causing apoptosis, cytotoxicity and anti-metastasis has become very popular and 192 many more are in progress [27, 28]. Lactaptin, a pepetide with a potent property to induced program 193 cell death of many different cancers activate proteases- the caspases that execute cancer cell to 194 apoptosis [6,7]. An analog of this particular peptides possesses cytotoxic activity to human and mice 195 cancer cell both in vivo and invitro. The pre-clinical trial studies have been well established following 196 the evaluation of its anti-tumor activities in breast cancer cell mouse xenograft [8,9]. The safety of this 197 paper has been therefore demonstrated as concluded by the toxicity and pharmacokinetics studies 198 [8].

- 199
- 200

201 One of the major downsides was that laptaptin was distributed evenly through the body and therefore 202 reduces the amount available to cancer cells. Improvement of the anti-cancer efficacy of RL2 through 203 conjugation with tumor specific peptides is expected to promote delivery of RL2 to target tissues [9]. 204 Tumor-specific peptides proved as a better target for delivering therapeutic substances and are used 205 for a number of purposes including delivering vehicles to pro-apoptotic peptides, genes, therapeutic 206 drugs and cytokines [9].

8. Conclusions

Review of lactaptin as a new candidate molecule for anticancer therapy has been carried out. The peptide has been found to play important role in program cell death, cytotoxic to cancer cells, activation of caspases and suppression of metastasis. In general, the peptide shows promising activity against cancer cells. Accordingly, enhancing it activity will greatly enhance it therapeutic efficacy and will provide insight into designing of new therapeutics.

References

- 1. Lozza C, Navarro-Teulon I, Pelegrin A, Pouget J-P, Vives E. Peptides in receptor-mediated radiotherapy: from design to the clinical application in cancers. Front Oncol. 2013;3(247):1-13.
- Nekipelaya VV, Semenov DV, Potapenko MO, Kuligina EV, Kit Yu, Romanova IV and Richter 2. VA. Lactaptin is a human milk protein inducing apoptosis of MCF-7 adenocarcinoma cells. Dokl Biochem Biophys. 2008; 419: 58-61.
- Vlassov VV, Richter VA, Semenov DV, Nekipelaya VV, Kuligina EV and Potapenko MO. 3. Peptide inducing apoptotic death of human cancer cells. 2008; Patent RF N 2317304.
- Tikunova NV, Semenov DV, Babkina IN, Kuligina EV, Koval O A., Fomin A S, Matveeva V A, 4. Matveev L E, Matveev AL, Richter VA. Recombinant plasmid DNA pFK2, providing synthesis of the recombinant peptide which is the analog of human kappa-casein, and recombinant peptide - the analog of human kappa-casein fragment, with the apoptotic activity against human tumor cells. 2010; Patent RF N.2401307.
- 5. Semenov DV, Fomin AS, Kuligina EV, Koval OA, Matveeva VA, Babkina IN, Tikunova NV and Richter VA. Recombinant analogs of a novel milk pro-apoptotic peptide, lactaptin, and their effect on cultured human cells. Protein J. 2010; 29: 174-180.
- 6. Koval OA, Fomin AS, Kaledin VI, Semenov DV, Potapenko MO, Kuligina EV, Nikolin VP, Nikitenko EV and Richter VA. A novel pro-apoptotic effector lactaptin inhibits tumor growth in mice models. Biochimie. 2012; 94: 2467-2474.
- 7. Koval OA, Tkachenko AV, Fomin AS, Semenov DV, Nushtaeva AA, Kuligina EV, Zavjalov EL and Richter VA. Lactaptin induces p53-independent cell death associated with features of apoptosis and autophagy and delays growth of breast cancer cells in mouse xenografts. PLoS One. 2014; 9(4): e93921
- 8 Bondarenko DA, Richter VA, Kuligina EV, Koval OA, Fomin AS, et al: Toxicity studies and pharmacokinetics of Lactaptin. Russian Journal of Biopharmaceuticals 2015; 7: 40-47.

216 217

218

219

220

221

222 223 224

229

230

231 232

233

234

235 236

237

238 239

240

241

242

243

244 245

246

247

248 249

250

251

252 253

254

255

256

257 258 259

260

261		
262	9	Nemudraya AA, Kuligina EV, Ilyichev AA, Fomin AS, Stepanov GA, Savelyeva AV, Koval OA,
263	0.	Richter VA. Selection of antitumor displayed peptides for the specific delivery of the
264		anticancer drug lactaptin. Oncol Lett. 2016; 12(6):4547–4555.
265		
	10	WUM Ding H. Fisher DE. Anontonia: Malagular Machaniama, Engualanadia of Life Sajanaga
266	10.	Wu M, Ding H, Fisher DE. Apoptosis: Molecular Mechanisms. Encyclopedia of Life Sciences.
267		Nature Publishing Group; 2001.
268		
269	11.	Richter VA, Vaskova AA, Koval OA, Kuligina EV. Antitumor Potential of Lactaptin. Biol Med
270		(Aligarh). 2015; S2:004.
271		
272		
273	12.	Samejima K, Earnshaw WC. Trashing the genome: the role of nucleases during apoptosis.
274		Nat Rev Mol Cell Biol. 2005; 6:677–688.
275		
276	13	Larsen BD, Sørensen CS. The caspase-activated DNase: apoptosis and beyond. The FEBS
277	10.	Journal. 2017; 284 :1160–1170
		Journal. 2017, 204 .1100-1170
278		
279	14.	Elliott MR, Chekeni FB, Trampont PC, Lazarowski ER, Kadl A, Walk SF, Park D, Woodson
280		RI, Ostankovich M, Sharma P Et al. Nucleotides released by apoptotic cells act as a find-me
281		signal to promote phagocytic clearance. Nature. 2009; 461:282–286.
282		
283	15.	Radic M, Marion T, Monestier M. Nucleosomes are exposed at the cell surface in apoptosis. J
284		Immunol. 2004; 172: 6692–6700.
285		
286	16.	Yan B, Wang H, Wang H, Zhuo D, Li F, Kon T, Dewhirst M, Li CY. Apoptotic DNA
287		fragmentation factor maintains chromosome stability in a P53-independent manner.
288		Oncogene. 2006; 25:5370–5376.
289		Cheogene. 2000, 23.5070-5070.
209	17	Liu X, Zou H, Slaughter C, Wang X. DFF, a heterodimeric protein that functions downstream
	17.	
291		of caspase-3 to trigger DNA fragmentation during apoptosis. Cell.1997; 89:175–184.
292		
293	18.	Enari M, Sakahira H, Yokoy ama H, Okawa K, Iwamatsu A, Nagata S. A caspase-activated
294		DNase that degrades DNA during apoptosis, and its inhibitor ICAD. Nature.1998; 391: 43–50.
295		
296	19.	Halenbeck R, MacDonald H, Roulston A, Chen TT, Conroy L, Williams LT. CPAN, a human
297		nuclease regulated by the caspase-sensitive inhibitor DFF45. Curr. Biol. 1998; 8:537–540.
298		
299	20.	Valastyan S, Weinberg RA. Tumor metastasis: molecular insights and evolving paradigms.
300		Cell. 2011; 147:275–292.
301		
302		
303	21	21 Gilbert LA, Hemann MT. DNA damage-mediated induction of a chemo resistant niche.
304	۷١.	Cell. 2010; 143:355–366.
		Cell. 2010, 143.335–306.
305		
306		
307	22.	Aguirre-Ghiso, J A. (2007). Models, mechanisms and clinical evidence for cancer dormancy.
308		Nat. Rev. Cancer. 2007; 7: 834–846.
309		
310	23.	Guan X. Cancer metastases: challenges and opportunities. Acta Pharmaceutica Sinica B.
311		2015; 5: 402–418.
312		
313		
314	24	Sugahara KN, Teesalu T, Karmali PP, Kotamraju VR, Agemy L, Greenwald DR, et al.
315		Coadministration of a tumor-penetrating peptide enhances the efficacy of cancer drugs.
316		Science. 2010;328(5981): 1031–5.
317		
318		
319		

Nemudraya AA, Makartsova AA, Fomin AS, Nushtaeva AA, Koval OA, Richter VA, et al. Tumor-Specific Peptide, selected from a Phage Peptide Library, Enhances Antitumor Activity of Lactaptin. PLoS ONE. 2016; 11(8): e0160980.
Hu C, Chen X, Zhao W, Chen Y, Huang Y. Design and Modification of Anticancer Peptides. Drug Des.2016; 5:138.
Chen HY, Mollstedt O, Tsai MH, Kreider RB. Potential clinical applications of multi-functional milk proteins and peptides in cancer management. Curr Med Chem. 2014; 21: 2424-2437.
Millimouno FM, Dong J, Yang L, Li J and Li X: Targeting apoptosis pathways in cancer and perspectives with natural compounds from mother nature. Cancer Prev Res (Phila) 2014; 7: 1081-1107.
Thundimadathil J. Cancer treatment using peptides: current therapies and future prospects. J. Amino Acids 2012; 2012: 1–13