

Review Paper

ANATOMICAL FEATURES OF SUGARCANE TREATED WITH THIAMINE

ABSTRACT. This worked aimed to know the anatomical features of sugarcane treated with thiamine. A completely randomized experiment was installed and designed in a double factorial scheme at 3x5 levels, in which the first factor consists of a variety of sugarcane: RB86-7515; RB96-6928 and CTC-4; the second factor was thiamine doses in five levels: zero mgL^{-1} ; 100 mgL^{-1} ; 200 mgL^{-1} ; 400 mgL^{-1} and 800 mgL^{-1} ; fifteen treatments were made with five replications, 75 plots in total. Tissues from the leaves and roots were influenced by exogenous action of thiamine as used at planting, displaying a positive response, doses above 400 mg L^{-1} can be a limiting factor to the development of these tissues. Factor regarding the sugarcane variety did not influenced on the anatomy of leaves and roots. Concentrations till 400 mg L^{-1} of thiamine, at exogenous administration, promoted a better development on morph-anatomic features of leaves and roots in planting of sugarcane seedlings.

Keywords: *Saccharum* sp., vitamin, phloem, xylem, B1

1. INTRODUCTION

Sugarcane belongs to family Poaceae and has a fasciculate root system that do not deeply reach its substrate, which may harm the development as it suffers water stress, once it does not have the capacity of water absorption in deeper layers of soil [1]. The use of synthetic or natural molecules at exogenous administration in sugarcane seedlings can promote better roots at initial stage, leading to a greater development of the stems. Among these molecules, vitamin B1 highlights [2].

The active form of Vitamin B1, thiamine pyrophosphate (TPP), works as a cofactor to the reaction of enzymes that acts on carbohydrate synthesis as well as some amino acids [2]. Its synthesis occurs on formation of independent compounds, mainly pyrimidine and thiazole. In prokaryotes the way of vitamin B1 synthesis has been explained, however, regarding eukaryotes there is a lack of studies [3]. In

Arabidopsis thaliana, Thi1 protein is likely responsible by thiazole synthesis, once a compound related to TPP has been found in its structure [4].

These modifications in organs of plants as submitted to use of vitamins are barely studied, once leaves present high plasticity of adaptations as exposed to environmental stimulus and even to these biomolecules [5]. That way, thiamine function on vegetables' metabolism can be related to process of formation of Acetyl-CoA in the Krebs cycle, favoring the development of roots, which presents a higher cellular respiration [6], and, consequently, promotes a greater exploration of deeper layers of soil, guarantying a higher absorption of nutrients and water, which will be direct to aerial part of the plant, to the photosynthetic process [7].

Use of thiamine can cause alterations on anatomy of plants, in its structures and arrangement of fundamental and vessels tissues [8], being necessary a deep knowledge of this features regarding the response to the use of this biomolecules. The transformations caused by the changes in the ambient where the vegetable was inserted [9] makes the symptomatology an important tool in understanding the damage caused by the mechanisms that cause morphological changes [10].

In this way, this work aimed to know the anatomical features of sugarcane treated with thiamine.

2. MATERIAL AND METHODS

The experiment was carried out at College of Agricultural and Technological Sciences – São Paulo State University, in Dracena, São Paulo State, Brazil, geographical coordinates: 21° 29' 10.24" S and 51° 31' 41.29" W, with a 411m average above the sea level, on April, 2016.

The local weather, according to Köppen classification, is the Cwa type: hot weather in summer and dry terms on winter, with the biggest rain rates between November and March. The annual average of temperature varies between 30.4°C and 19.2°C, average precipitation of 1311.6 mm and air humidity of 78%.

The soil was collected in the depth of 30-50 cm and classified as Argissolo red yellow [11], with the following chemical parameters, as shown in Table 1 [12]:

Table 1: Soil chemical parameters

pH	MO	P	K	Ca	Mg	H	+	Al	SB	CTC	V%	m%	S	B	Cu	Fe	Mn	Zn
								Al										
CaCl ₂	g dm ⁻³	mg dm ⁻³						mmol _c dm ⁻³							mg dm ⁻³			
4.7	9.0	2.0	1.2	12	5.0	16.0	2.0	20.2	36.2	56.0	9.0	6.0	0.22	0.6	4.0	13.6	0.2	

SB: Sum of bases; V%: Base Saturation; m%: Saturation Al.

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68 The experimental design was completely randomized with three varieties of
 69 sugarcane: RB86-7515; RB96-6928 and CTC-4, the second factor refers to doses of
 70 thiamine, in different five levels: null L⁻¹; 100 mg L⁻¹; 200 mg L⁻¹; 400 mg L⁻¹ e 800 mg
 71 L⁻¹; within 15 treatments and 5 repetitions, in total 75 plots.

72 The plots were planted in plastic pots with a capacity of 9.0 dm³ of sieved soil
 73 and corrected according to the nutritional requirements of the crop, the urea, super
 74 simple and potassium chloride fertilizers were used [12]. The experiment was
 75 ~~installed and~~ conducted in an unprotected environment and irrigated according to the
 76 soil moisture factor.

77 Forty five days after planting the total leaf area (TLA cm²) was determined
 78 using the Easy leaf area image program [13]. At the same time, ultra structural
 79 characteristics of sugarcane leaf were also evaluated, from fragment five with five-
 80 centimeter was taken from the median region of leaves from the central middle third
 81 of the stem and a five-centimeter fragment from the median root region. After 24
 82 hours, the fragments were washed and stored in 70% ethanol until the date of
 83 analyzes. All fragments of plant tissues received the pertinent procedures for
 84 dehydration, diaphanization, inclusion and embedding. By using a microtome that
 85 contains steel razors, eight- μm transversal sections were **done** in each embedded
 86 fragment.

87 The first transversal sections without damage caused by cut of plants tissues
 88 were chosen for preparation of the histological slides. These sections were fixed with
 89 patches (albumin), were tinted with safranin with a 1% ratio, and were set in
 90 microscope and glass slides with Entellan®. All slides were observed with an
 91 Olympus optical microscope model BX 43, with an attached camera in order to
 92 ~~perform~~ the photographs of the cuts. Pictures were used to measure anatomic
 93 parameters through the software cellSens Standart that was calibrated with a
 94 microscopic ruler in the same gains [14].

By using transversal sections, the following morph-anatomic characteristics were measured: adaxial epidermal thickness (ADET); abaxial epidermal thickness (ABET); adaxial cuticle thickness (ADCT); abaxial cuticle thickness (ABCT); diameter of the beam buliform cells (DBBC); root phloem diameter (RPD); root xylem diameter (RXD) and thickness of the endoderm (TE). Five measurements were done for all characteristics in each microscope slide. Plots were represented by average value obtained on each characteristic.

All variables were submitted to the F test ($p < 0.05$) and the regression analysis was applied to the Thiamine doses, in which their models were tested: linear, quadratic and cubic [15]. For the sugarcane varieties, the Tukey test was applied at a 5% probability. Assistat 7.7 static software was used [16].

3. RESULTS

Variety RB96-6928 highlights among the studied varieties by presenting greater averages to the variable total leaf area (TLA), as Table 2 shows.

Table 2: Mean values of total leaf area (TLA); adaxial epidermal thickness (ADET); abaxial epidermal thickness (ABET); adaxial cuticle thickness (ADCT); abaxial cuticle thickness (ABCT); diameter of the beam buliform cells (DBBC) of sugarcane cultivated with thiamine.

	TLA cm ²	ADET	ABET	ADCT	ABCT	DBBC
		μm				
RB86-7515	221.93b	9.89a	9.64a	4.93a	5.13a	4.93a
RB96-6928	453.16a	9.81a	9.57a	4.83a	4.75a	4.83a
CTC-4	260.81b	9.46a	8.56a	4.73a	4.32a	4.73a
MSD	121.64	1.19	1.38	0.71	0.91	0.71
CV%	57.25	18.03	21.96	21.71	28.25	21.71
MG	311.96	9.72	9.26	4.83	4.73	4.83
f	12.01**	0.43ns	2.19ns	0.24ns	2.30ns	0.24ns

S: Stomata. MSD: Minimum significant difference. CV: Coefficient of variation. MG: Overall mean. f: value of F calculated in the analysis of variance; ** significant at the 1% probability level ($p < 0.01$); * significant at the 5% probability level ($0.01 \leq p < 0.05$); ns—not significant ($p \geq 0.05$). The averages in the column followed by the same letter do not differ statistically from each other. The Tukey test was applied at a 5% probability level. Source: August, 2018.

To others variables significant difference was not observed among the sugarcane varieties (Table1). However, as thiamine doses are considered, leaf area were not changed, which displays only the effect of sugarcane varieties, as Table 3 shows.

Table 3: The analysis of variance of the regressions of the thiamine doses applied, where the models were tested: linear, quadratic and cubic of variety sugarcane.

Middle Square								
System	FV	GL	TLA	ADET	ABET	ADCT	ABCT	DBBC
RB86-7515	Concentration	4	16073.88	0.84	42.15	0.35	0.79	0.35
	Residue	21	27519.00	4.76	3.86	0.70	0.52	0.70
	Regression	1	Ns	Ns	Q**	Ns	Ns	Ns
RB96-6928	Concentration	4	43391.33	18.36	18.31	2.63	2.79	2.63
	Residue	21	57441.27	4.08	3.39	1.75	0.82	1.75
	Regression	1	Ns	L*	Q*	Ns	Ns	Ns
CTC-4	Concentration	4	3804.79	1.52	11.85	0.21	0.55	0.21
	Residue	21	20092.79	0.98	5.60	1.07	3.85	1.07
	Regression	1	Ns	Ns	Ns	Ns	Ns	Ns

Ns— $p \geq 0.05$; * $0.01 \leq p < 0.05$; ** $p < 0.01$. L: polynomial of 1st degree. Q: polynomial of 2nd degree. NL—number leaf; PH—plant height; DMAP—Dry mass of the air part. Source: August, 2018.

Only variety RB-96-6928 presented a linear response to the use of thiamine doses in the variable adaxial epidermis thickness (ADET) (Table 2), as Figure 1 shows.

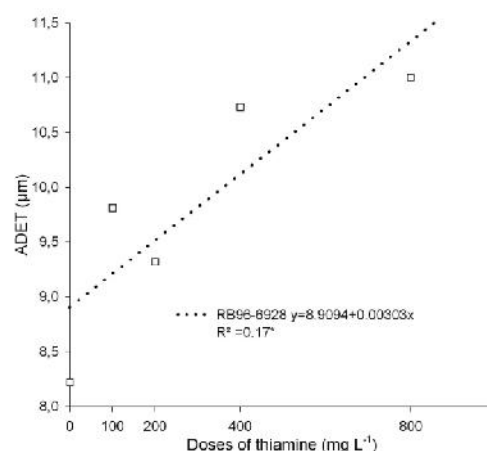


Fig. 1: Variable adaxial epidermis thickness (ADET) of variety RB96-6928 thirty days after use of thiamine following the planting.

To the variable abaxial epidermis thickness (ABET) varieties of sugarcane RB86-7515 and RB96-6928 presented a quadratic growth curve as submitted to

126 thiamine, in which they reach their peak at 441.17 and 410.44 mg L⁻¹ doses. Doses
127 above 400 mg L⁻¹ presented a harmful effect, as Figure 2 shows.
128

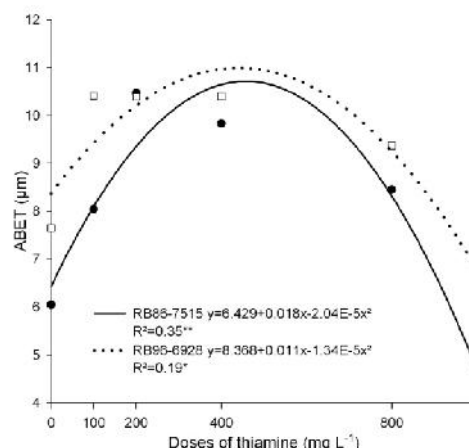


Fig. 2: Variable abaxial epidermis thickness (ABET) of varieties RB86-7515 and RB96-6928 thirty days after use of thiamine following the planting.

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130 No statistical difference was observed among the sugarcane varieties for the
131 variables: root phloem diameter (RPD); root xylem diameter (RXD) and thickness of
132 endoderm (TE) as Table 2a demonstrates.
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Table 2a: Mean values of root phloem diameter (RPD); root xylem diameter (RXD) and thickness of the endoderm (TE) of sugarcane cultivated with thiamine.

	RPD (μm)	RXD (μm)	TE (μm)
	----- μm -----		
RB86-7515	9.99a	96.91a	22.47a
RB96-6928	9.14a	95.21a	20.83a
CTC-4	9.02a	88.73a	20.37a
MSD	2.15	9.31	2.13
CV%	33.70	14.60	14.73
MG	9.38	93.62	21.22
f	0.70ns	2.49ns	3.13ns

S: Stomata. MSD: Minimum significant difference. CV: Coefficient of variation. MG: Overall mean. f: value of F calculated in the analysis of variance; **significant at the 1% probability level ($p < 0.01$); *significant at the 5% probability level ($0.01 \leq p < 0.05$); ns—not significant ($p \geq 0.05$). The averages in the column followed by the same letter do not differ statistically from each other. The Tukey test was applied at a 5% probability level. Source: August, 2018.

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When is considered the effect of the application of thiamine on sugarcane, variety RB86-7515 presented a quadratic response in the variable root phloem diameter, as Table 3a shows.

Table 3a: The analysis of variance of the regressions of the thiamine doses applied, where the models were tested: linear, quadratic and cubic of variety sugarcane.

Middle Square					
System	FV	GL	RPD	RXD	TE
RB86-7515	Concentration	4	112.61	2623.75	261.40
	Residue	21	8.87	157.85	16.13
	Regression	1	Q**	Q**	Q**
RB96-6928	Concentration	4	12.61	29.50	64.92
	Residue	21	10.05	364.07	5.17
	Regression	1	Ns	Ns	L**
CTC-4	Concentration	4	26.81	24.78	363.47
	Residue	21	10.46	125.37	11.34
	Regression	1	Ns	Ns	Q**

Ns– $p \geq 0.05$; * $0.01 \leq p < 0.05$; ** $p < 0.01$. L: polynomial of 1st degree. Q: polynomial of 2nd degree. NL–number leaf; PH–plant height; DMAP–Dry mass of the air part. Source: August, 2018.

There was an increase in the phloem diameter of the variety RB86-7515 till, approximately, 434.13 mg L⁻¹ thiamine doses, in which concentrations above 400 mg L⁻¹ of thiamine may is a limiting factor for the development of sugarcane root phloem, as Figure 3 shows.

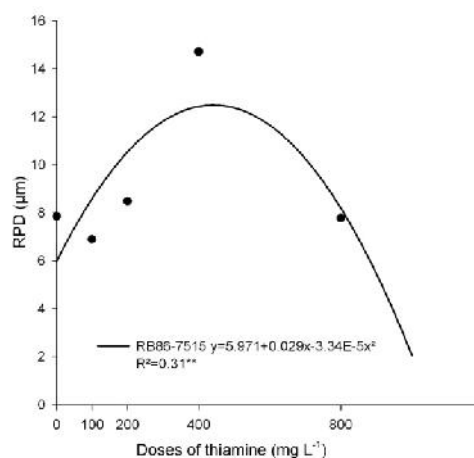


Fig. 3: Root Phloem Diameter (RPD) of the variety RB86-7515 thirty days after use of thiamine at the planting.

It similarly occurs with Root Xylem Diameter (RXD), since only variety RB86-7515 presents a quadratic response to the use of thiamine at the sugarcane planting, reach its peak at 381.98 mg L^{-1} thiamine doses, as Figure 4 shows.

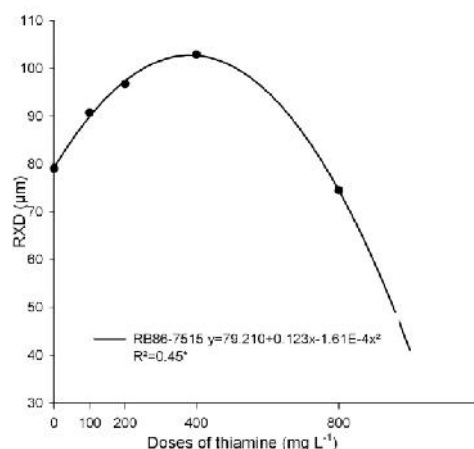


Fig. 4: Root Xylem Diameter (RXD) of variety RB86-7515 thirty days after use of thiamine at the planting.

To the variable thickness of endoderm (TE) all studied varieties significantly responded to the presence of thiamine at the planting. RB86-7515 presented a quadratic response till 491.15 mg L^{-1} thiamine doses, while CTC-4 displayed a negative quadratic response to the increasing of thiamine till 416.55 mg L^{-1} doses, however, RB96-6928 present a linear positive response, as Figure 5 shows.

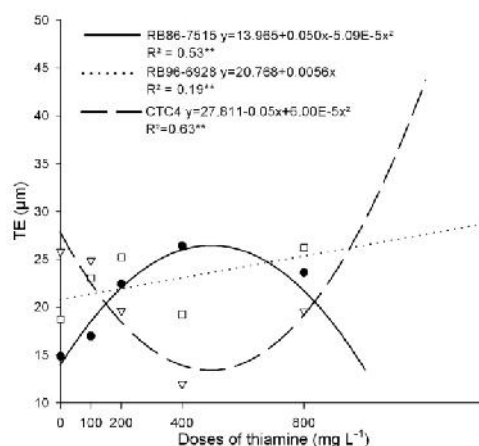


Fig. 5: Thickness of endoderm of varieties RB86-7515; RB96-6928 and CTC-04 thirty days after use of thiamine at the planting.

4. DISCUSSION

Due to agronomic differences between varieties of sugar cane and the capacity of adaptation to different environmental conditions, these variations may reflect the productivity of culture, especially with the variation in leaf area, since, by reducing it, the photosynthetic rate is impaired, harming the carbon fixation in the culture' dry mass [9].

Non-variation of leaves' and roots' anatomical features among the studied varieties affirms the importance of the exogenous use of thiamine, once the used doses of the vitamin entails a better development of leaf endoderm, due to the some carbohydrates' promoting action [3; 17; 2]. It was expected sharp changes in other areas of tissues, as in cuticle of the leaves, once it presents a high accumulation of oils and waxes, also, with the possible action of thiamine as cofactor in the synthesis of Acetyl-CoA, it could have entailed a bigger deposition of theses biomolecules [5; 7].

Sheath cells of the vascular bundle could also have shown a greater development, since they present high biochemical reactions, mainly in the action of RuBPco molecule in the Calvin cycle [6; 18; 19] which could have potentialized in its reactions, leading to a greater development of the leaf area, and even enhancing the opening of the stomatal fissure, in which recent researches display that thiamine

may act as an important factor in the opening and closure process, influencing the photosynthetic rate [5].

Due to the presence of thiamine, some polyamines may turn into synthesized and were carried by the vases of the vegetal organs, the presence of these biomolecules is an indicator of the reactions triggered by the presence of this vitamin [20; 21; 22], that way, conductors vases of the roots was influenced by the exogenous application at the planting, showing a positive response till 400 mg L⁻¹, doses above can be a limiting factor to the development of the tissues.

5. CONCLUSION

Factors regarding the sugarcane variety did not influenced on the anatomy of leaves and roots.

Concentrations till 400 mg L⁻¹ of thiamine, at exogenous administration, promoted a better development on morph-anatomic features of leaves and roots in planting of sugarcane seedlings.

REFERENCES

1. Clemente PRA, Bezerra BKL, Silva VSG, Santos JCM, Endres L. 2017. Root growth and yield of sugarcane as a function of increasing gypsum doses. *Pesquisa Agropecuária Tropical*, 47(1): 110-117. <http://dx.doi.org/10.1590/1983-40632016v4742563>
2. Kamarudin AN, Seman IA, Yusof ZNB. 2017. Thiamine biosynthesis gene expression analysis in *Elaeis guineensis* during interactions with *Hendersonia toruloidea*. *Journal of Oil Palm Research*, 29(2): 218-226. <http://dx.doi.org/10.21894/jopr.2017.2902.06>
3. Park J, Dorrestein PC, Zhai H, Kinsland C, McLafferty FW, Begley TP. 2003. Biosynthesis of the thiazole moiety of thiamin pyrophosphate (Vitamin B1). *Biochemistry*, 42: 12430-12438. <http://dx.doi.org/10.1021/bi034902z>
4. Garcia AF, Dyszy F, Munte CE, Demarco R, Beltramini LM, Oliva G, Costa-Filho AJ, Araujo APU. 2014. THI1, a protein involved in the biosynthesis of thiamin in *Arabidopsis thaliana*: Structural analysis of THI1(A140V) mutant.

- 215 *Biochimica et Biophysica Acta*, 1844: 1094–1103.
216 <http://dx.doi.org/10.1016/j.bbapap.2014.03.005>
- 217 5. Li C, Wang M, Wu X, Chen D, Lv H, Shen J, Qiao Z, Zhang W. 2016. THI1, a
218 thiamine thiazole synthase, interacts with Ca²⁺-Dependent protein kinase
219 CPK33 and modulates the S-Type anion channels and stomatal closure in
220 Arabidopsis. *Plant Physiology*, 170: 1090-1104.
221 <https://doi.org/10.1104/pp.15.01649>
- 222 6. Idris ZHC, Abidin AAZ, Subki A, Norhana Z, Yusof B. 2018. The effect of
223 oxidative stress towards the expression of thiamine biosynthesis genes (THIC
224 and THI1/THI4) in oil palm (*Elaeis guineensis*). *Tropical Life Sciences*
225 *Research*, 29(1): 71–85. <http://dx.doi.org/10.21315/tlsr2018.29.1.5>
- 226 7. Pourcel L, Moulin M, Fitzpatrick TB. 2013. Examining strategies to facilitate
227 vitamin B1 biofortification of plants by genetic engineering. *Frontiers in Plant*
228 *Science*, 4: 1-8. <http://dx.doi.org/10.3389/fpls.2013.00160>
- 229 8. Woodward JB, Abeydeera ND, Paul D, Phillips K, Rapala-Kozik M, Freeling
230 M, Begley TP, Ealick SE, Mcsteen P, Scanlon MJ. 2010. A maize thiamine
231 auxotroph is defective in shoot meristem maintenance. *The Plant Cell*, 22:
232 3305–3317. <http://dx.doi.org/10.1105/tpc.110.077776>
- 233 9. Raven PH, Eichhorn SE, Evert RF. 2014. *Biologia Vegetal*. 8.ed. Guanabara
234 Koogan. 850p.
- 235 10. Reis AR, Barcelos JPQ, Osório CRWS, Santos EF, Lisboa LAML, Santini
236 JMK, Santos MJD, Junior EF, Figueiredo PAM, Lavres J, Gratão PL. 2017. A
237 glimpse into the physiological, biochemical and nutritional status of soybean
238 plants under Ni-stress conditions. *Environmental and Experimental Botany*,
239 144: 76–87. <http://dx.doi.org/10.1016/j.envexpbot.2017.10.006>
- 240 11. Empresa Brasileira de Pesquisa Agropecuária - Embrapa. 2013. *Sistema*
241 *brasileiro de classificação de solos*. 3.ed. Brasília, 353p.
- 242 12. Raij B, Cantarella H, Quaggio JÁ, Furlani AMC. 1996. *Recomendações de*
243 *adubação e calagem para o Estado de São Paulo*. 2.ed. Campinas: IAC.
244 285p.
- 245 13. Easlson HM, Bloom AJ. 2014. Easy leaf área: Automated digital image
246 analysis for rapid and accurate measurement of leaf area. *Applications in*
247 *Plant Sciences*, 2(7): 1-4. <https://doi.org/10.3732/apps.1400033>

14. Lisboa LAM, Lapaz AM, Bottan AP, Yoshida CHP, Santos LFM, Figueiredo PAM, Viana, RS. 2018. Ethephon Action on Germination and Leaf Ultrastructure of *Urochloa brizantha* cv. Marandu. *Australian Journal of Basic and Applied Sciences*, 12(4): 22-28. <http://dx.doi.org/10.22587/ajbas.2018.12.4.5>
15. Banzatto D, Kronka, SN. 2013. *Experimentação Agrícola*. 4.ed. Funep, 237p.
16. Silva FAZ, Azevedo CAV. 2016. The Assistat Software Version 7.7 and its use in the analysis of experimental data. *African Journal Agriculture Research*, 11(39): 3733-3740. <http://dx.doi.org/10.5897/AJAR2016.11522>
17. Rebeille F, Douce R. 2011. *Biosynthesis of Vitamins in Plants Part A, Volume 58: Vitamins A, B1, B2, B3, B5 (Advances in Botanical Research)*. Academic Press; 1 ed.. 322p.
18. Begley TP, Ealick SE, McLafferty FW. 2012. Thiamin Biosynthesis - still yielding fascinating biological chemistry. *Biochemical Society Transactions*, 40(3): 555–560. <http://dx.doi.org/10.1042/BST20120084>
19. Minhas AP, Tuli R, Puri S. 2018. Pathway Editing Targets for Thiamine Biofortification in Rice Grains. *Frontiers in Plant Science*, 9(975): 1-8. <https://doi.org/10.3389/fpls.2018.00975>
20. Friedman R, Levin N, Altman A. 1986. Presence and Identification of Polyamines in Xylem and Phloem Exudates of Plants. *Plant Physiology*, 82(4): 1154-1157. <https://doi.org/10.1104/pp.82.4.1154>
21. Martinis J, Gas-Pascual E, Szydlowski N, Crèvecoeur M, Gisler A, Bürkle L Fitzpatrick, T. B. 2016. Long distance transport of thiamine (vitamin B1) is concomitant with that of polyamines in Arabidopsis. *Plant Physiology Preview*, 171(1): 542-553. <https://doi.org/10.1104/pp.16.00009>
- 22.** Zimmermann MH, Milburn JA. 1975. Transport in plants I: Phloem transport. New Series, 1.ed. 525p. <http://dx.doi.org/10.1007/978-3-642-66161-7>