

# **Tillage and Rice Straw Management affect Soil Enzyme Activities and Chemical Properties after Three Years of Conservation Agriculture Based Rice-Wheat System in North-Western India**

**Sushil Kumar Kharia<sup>1\*</sup>, H.S. Thind<sup>1</sup>, Sandeep Sharma<sup>1</sup>, H.S. Sidhu<sup>2</sup>, M.L. Jat<sup>3</sup>, Yadvinder-singh<sup>1</sup>**

<sup>1</sup>Department of Soil Science, Punjab Agricultural University, Ludhiana-141004, India

<sup>2</sup>Borlaug Institute for South Asia, Ludhiana 141008, India

<sup>3</sup>International Maize and Wheat Improvement Centre (CIMMYT), New Delhi 110012, India

## **ABSTRACT**

**Aims:** To evaluate the effects of rice establishment, tillage and rice straw management on changes in soil enzyme activities and chemical properties in soil after three cycles of continuous rice-wheat system.

**Study design:** The experiment was laid in split plot design with three replications.

**Place and Duration of Study:** PAU, Ludhiana, 2010-2013

**Methodology:** The experiment was started during kharif season of 2010. The design of an experiment was having 12 treatments with 3 replications. The main plot treatments in rice (zero till direct seeded rice, ZT-DSR; conventional till direct seeded rice, CT-DSR; zero till direct transplanted rice, ZT-DTR and puddled transplanted rice, PTR) and three sub-plot treatments in wheat (conventional till wheat without rice straw, CTW-R; ZT wheat without rice straw, ZTW-R, and ZT wheat with rice straw retained as surface mulch using Happy Seeder, ZTW+R).

**Results:** Zero tillage with rice straw retention (ZTW) as surface mulch (+R) increased wheat yield by 9% and 15% compared with conventional tillage (CTW) and ZTW with no residue (-R). Significantly higher dehydrogenase, fluorescein diacetate, alkaline phosphatase, phytase and urease activities were recorded under ZTW+R compared with ZTW/CTW-R in 0-5 cm soil layer. Organic carbon, Olsen-P, available K and DTPA-extractable micronutrients (Zn, Fe, Mn and Cu) in the surface 0-5 cm soil layer were significantly higher in ZTW+R compared with ZTW/CTW-R. Soil enzyme activities were significantly and positively correlated with each other, soil organic carbon, Olsen-P and grain yield of wheat. **Conclusion:** We concluded that RCTs (ZTW and rice residue retention) improve soil enzyme activities and chemical properties in surface 0-5 cm soil layer and enhance productivity and sustainability of rice-wheat system.

**Keywords:** conservation agriculture, rice straw surface mulch, tillage, enzyme activities, rice-wheat system

## **1. INTRODUCTION**

The rice-wheat cropping system (RWS) occupies about 13.5 million hectares (M ha) in the Indo-Gangetic Plains (IGP) of South Asia (Gupta and Seth, 2007) and is fundamental to employment, income and livelihood for millions of people in the region. However, sustainability of intensive tillage-based

conventional RWS is constrained or limited by water, energy and labour scarcity, increasing cost of production, increasing air pollution and deteriorating soil health. To reverse this situation various Resource Conservation Technologies (RCTs) such as zero tillage, direct seeded rice and crop residue retention are being developed and promoted for RWS (Sidhu *et al.*, 2007; Gathala *et al.*, 2013). Puddling in rice and intensive tillage in wheat are known to cause sub-soil compaction, deterioration of soil structure, and decrease in permeability in the subsurface layer and thereby adversely affecting productivity of RWS (Jat *et al.*, 2009).

The increasing constraints of labour and time under intensive agriculture have led to the adoption of mechanized farming in RWS leaving large amounts of crop residues in the fields. As crop residues interfere with tillage and seeding operation for the next crop, farmers in northwestern (NW) India often prefer to burn surplus rice residues on-farm after grain harvest to establish the next wheat crop. Residue burning impacts human and animal health both medically, and by traumatic road accidents due to restricted visibility (Yadvinder-Singh *et al.*, 2014). Establishment of wheat crop by ZTW with retention of crop residues on the soil surface potentially offers a labour-saving and cost-effective alternative to the burning of rice residues. A new machine, known as the 'Happy Seeder' (HS), has now been developed for this purpose which is capable of direct drilling wheat into heavy rice residue loads, without burning in a single operation (Sidhu *et al.*, 2007, 2015).

There is currently much interest in dry direct seeded rice (DSR) as an alternative to conventional transplanted rice in North-West India due to labour scarcity for transplanting, and because puddling and transplanting require large quantities of water to establish the rice crop. Zero tillage with rice residue retained on soil surface significantly improve water content in soil, improve overall soil physical and chemical health through replenishing soil organic matter and to support sustainable RWS (Jat *et al.*, 2009; Gathala *et al.*, 2013). Tillage, crop establishment and residue management options may also have significant effect on soil biological health.

Soil enzyme activities respond much more quickly to the changes in tillage and soil management as compared with soil organic matter (Jiang *et al.*, 2006) and therefore could be used as potential biological indicators of soil quality (Yang *et al.*, 2011). Dehydrogenase activity is involved in oxidative phosphorylation and reflects the total oxidative potential of the soil microbial community (Dick, 1997). Fluorescein diacetate (FDA) hydrolyzed by a number of different enzymes, can provide adequate microbial activity (Bendick and Dick, 1999). The activity of alkaline phosphatase is linked to transformation of organic P in soil to inorganic P compounds (Yang *et al.*, 2008). The soil phytase enzyme increases the availability of P from phytate in soils. Urease activity controls hydrolysis of urea in soils, which is an important source of N.

Significant increases in the activity of soil enzymes have been observed under ZT and residue retention in non-rice based cropping systems (Eivazi *et al.*, 2003) and the magnitude of the change depends on soil type and climate. However, little information is available in the literature on the effects of RCTs (DSR, tillage and rice straw management) on changes in soil biochemical properties under RWS in South Asia. The present study was therefore, carried out to determine the effects of rice establishment methods, tillage and rice straw management on biochemical properties after three cycles of RWS in northwestern India.

## 2. MATERIALS AND METHODS

### Description of the experimental site and climate

A three-year (2010-2013) field experiment on irrigated RWS was conducted on a Typic Ustochrept sandy loam soil at the experimental farm of the Punjab Agricultural University, Ludhiana (30°56'N and 75°52'E) in the IGP in the northwestern India. The top-soil (0–15 cm layer) at the start of experiment was non-saline (electrical conductivity 0.36 dS m<sup>-1</sup>) with pH 7.88 and contained 4.5 g kg<sup>-1</sup> Walkley-Black carbon, 8.2 mg kg<sup>-1</sup> 0.5 M NaHCO<sub>3</sub>-extractable P (Olsen *et al.*, 1954) and 50.4 mg kg<sup>-1</sup> 1 N NH<sub>4</sub>OAc-extractable K. The region has a sub-tropical climate, with hot, wet summers and cool dry winters. Annual mean rainfall is 760 mm, about 80% of which occurs from June to September. The long-term average (30 years) mean minimum and maximum temperatures in wheat (November to April) are 6.7 and 22.6°C while in rice (June to October) are 18 and 35°C, respectively.

### Experimental layout and treatments

Treatments applied to RWS were arranged in a split-plot design with a total of twelve treatments replicated three times. Main plot treatments applied to rice were four combinations of tillage and crop establishment methods in rice (zero till direct seeded rice, ZT-DSR; conventional till direct seeded rice, CT-DSR; zero till mechanically transplanted rice, ZT-DTR and puddled transplanted rice, PTR). The three sub-plot treatments in wheat were combinations of tillage and residue management options (conventional till wheat without residues, CTW-R; ZT wheat without residues, ZTW-R, and ZT wheat with residues retained as surface mulch using Happy Seeder, ZTW+R). The treatments were assigned to the same experimental plots in the 3 years of the study. The treatment details are summarized in Table 1.

### Soil and crop management

Wheat straw was removed at harvest in April each year and the plots were fallowed until pre-irrigation for rice in early June. ZT-DSR was sown in a single operation using zero-till-fertilizer cum seed drill at 20 cm row spacing. In CT-DSR, plots were prepared by 2 harrowings + 1 cultivator + 1 planking followed by dry seeding of rice using a seed rate of 20 kg ha<sup>-1</sup>. Light irrigation (50 mm) was applied at 1 day after seeding to ensure satisfactory germination, and then at 4–5 day intervals until physiological maturity depending on the rainfall events during the growing season. In ZT-DTR, rice seedlings were transplanted in ZT plots with standing water using mechanical transplanter. In PTR, tillage included disking twice in early June followed by two cultivator operations in standing water to puddle the soil and planking. Rice (variety PR 114) was transplanted manually using 30-day-old seedlings spaced at 15 cm x 20 cm in the second week of June. All treatments received a uniform dose of 150 kg N as urea, 26 kg P as diammonium phosphate and 25 kg K as muriate of potash. Whole of P and K was applied at rice planting on all the plots. Fertilizer N in PTR and ZT-DTR was applied in three equal split doses at transplanting and at 3 and 6 weeks after transplanting. While in CT-DSR and ZT-DSR fertilizer N was applied in three equal split doses at 3, 5 and 9 weeks after sowing. Rice was harvested manually in the second week of October in –R to remove the rice straw from the field plots and the combine harvester was used in +R plots to retain the rice straw in the field.

All plots received flood irrigation (75–80 mm) prior to planting of wheat. In conventional plots, seed bed was prepared by 2 dry harrowings followed by 2 cultivators and one planking. Wheat (variety PBW 621) was sown in the second week of November using a seed rate of 100 kg ha<sup>-1</sup>. Sowing of wheat was done on the same day in all the treatments at row spacing of 20 cm using Happy Seeder machine. Fertilizer N (120 kg ha<sup>-1</sup>) as urea was applied in three equal split doses at sowing, and three weeks and 8 weeks after planting. A basal dose of 26 kg P ha<sup>-1</sup> as single super phosphate and 25 kg K ha<sup>-1</sup> as muriate of potash were applied on all plots at planting. Wheat was irrigated (each of 75 mm) at crown root initiation (CRI), maximum tillering (MT), panicle initiation and dough stages recommended for wheat crop in the region. Grain yield of wheat was recorded from 1.4 m x 9 m area in the center of each plot and was reported on dry weight basis.

### Soil sampling and analysis

Soil samples were collected from 0–5 cm, 5–10 cm and 10–15 cm soil layers, after the harvest of wheat crop (after 3 cycles of rice-wheat rotation). The soil samples were collected with the help of tube auger (25 mm internal diameter) from randomly selected 4 places within each treatment plot. After removing visible root debris, the soil samples were mixed, sieved (2 mm) and stored at 4°C for subsequent analysis for biochemical soil properties. The methods used for assaying biochemical soil properties are listed in Table 2.

### Statistical analysis

All the dataset was analyzed using analysis of variance (ANOVA) and differences among treatments were compared at p=0.05 level of significance using the IRRISTAT data analysis package (IRRI, 2000). Statistical software used for this study was SPSS version 20.0. Correlations between the variables were determined using Pearson correlation coefficients (r) and probabilities. In all the analyses, significance was accepted at a level of probability (p) of <0.05.

## 3. RESULTS AND DISCUSSION

There was no significant interaction effects of rice establishment methods and tillage and rice residue management in wheat) on enzymes activities and chemical properties in all the three soil layers (0–5, 5–10

and 10-15 cm). Rice establishment systems also showed no significant effects on the grain yield of subsequent wheat and any of the soil property measured in the study.

### Enzyme activities in soil

Tillage and rice straw management in wheat significantly ( $P \leq 0.05$ ) influenced all the enzyme activities in 0-5 cm soil layer but showed no effect at 5-10 cm and 10-15 cm soil layers (Tables 3-7). In general, enzyme activities decreased with depth. In 0-5 cm soil layer, DHA under ZTW+R were 6% and 14 % higher as compared to ZTW-R and CTW-R, respectively (Table 3). DHA was 9 % higher under ZTW-R than CTW-R. FDA and APA at 0-5 cm depth were 9 and 13 % higher in ZTW+R as compared to ZTW-R and CTW-R, respectively (Table 4 and 5). PA was increased by 9% and 24 % under ZTW+R in 0-5 cm layer compared to ZTW-R and CTW-R, respectively (Table 6). Three enzyme activities (FDA, APA and PA) were significantly higher under ZTW-R than CTW-R. The increase in UA under ZTW+R was 8 and 13 % higher as compared to ZTW-R and CTW-R in 0-5 cm soil layer, respectively (Table 7).

### Chemical properties of soil

Tillage and rice straw management practices in wheat significantly affected the soil chemical properties, except soil pH and EC in the 0-5 cm layer only (data not shown) (Tables 8-11). Soil organic carbon content in 0-5 cm layer in ZTW+R treatment was increased by 10.3% and 23.1% compared to ZTW-R and CTW-R, respectively (Table 8). ZTW-R plots had significantly higher organic carbon content than CTW-R. Similarly, Olsen-P,  $\text{NH}_4\text{OAc}$ -extractable - K and DTPA-extractable micronutrients (Zn, Fe, Mn and Cu) were significantly higher in ZTW+R compared to ZTW-R and CTW-R treatments (Table 9-11). The increase in micronutrient availability was greater for Fe and Cu (12-14%) compared with the increase in the Zn and Mn (3-6%) (Table 11).

### Grain yield and yield attributes of wheat

Grain yield of subsequent wheat was not significantly affected by the rice establishment methods, irrespective of tillage and straw management in wheat (Table 12). However, tillage and rice straw management practices in wheat significantly affected grain yield of third wheat crop (Table 12). ZTW+R produced 9% and 15% higher grain yield than ZTW-R and CTW-R, respectively. CTW-R produced significantly higher (6 %) grain yield than ZTW-R. As the grain yield of subsequent wheat was not significantly affected by the rice establishment methods so the yield attributes did not differ significantly (Table 12). However, tillage and straw management practices in wheat significantly affected wheat grain yield attributes. All the yield attributes namely Spike length, grains per spike, grain weight per spike and 1000 grain weight were highest under ZTW+R followed by CTW-R and ZTW-R.

### Correlation between soil enzyme activities, grain yield and soil chemical properties

Activities of all of the enzymes in the study were significantly and positively correlated with each other as well as with organic carbon and Olsen-P in surface 0-5 cm soil layer (Table 13). The values of correlation coefficient ( $r$ ) for soil enzyme activities ranged from 0.62 to 0.96. While the lowest value of ' $r$ ' was between APA and PA, the highest value was recorded for FDA and PPA. Except for APA, all the other enzymes showed significant and positive relationship ( $r = 0.61 - 0.72$ ) with organic carbon in the 0-5cm soil layer. A strong and positive relationship ( $r = 0.65 - 0.83$ ) was observed between all the enzyme activities and Olsen P in 0-0.5 m layer. Grain yield of wheat showed significant positive relationship ( $r = 0.69 - 0.86$ ) with FDA, PA, PPA and UA in the surface 0-5 cm soil layer (Table 13). The activities of all the enzymes were significantly and positively correlated ( $r = 0.63 - 0.86$ ) with DTPA-extractable Fe and Zn in 0-5 cm layer (Table 14). The relationship with DTPA-Mn was significant only for FDA and PA.

### DISCUSSION

In our study, ZTW+R improved all soil enzyme activities in the 0-5 cm soil layer of than ZTW-R and CTW-R. Tillage and crop residues management practices have been reported to increase enzyme activities in soils (Klose *et al.*, 1999). The stimulating effect of organic materials on enzyme activities in soil is consistent with results earlier reported (Elfstrand *et al.*, 2007; Garcia *et al.*, 1998). The rice straw provided carbon for enhancing microbial activity and thereby increased activity of several enzymes (Caravaca and Roldan, 2003). The increases in microbiological activities under ZT compared to CT have been linked to

the increases in organic matter content of surface soils (Roldan *et al.*, 2007; Melero *et al.*, 2009). The addition of organic matter on CT fields can rapidly increase the soil enzyme activities (Kandeler *et al.*, 2006).

DHA activity occurs intracellular in all living microbial cells (Yuan and Yue, 2012; Zhao *et al.*, 2010) and can be used as an indicator of overall soil microbial activity (Salazar *et al.*, 2011). DHA responded to tillage and residue management treatments in a similar manner as SOC. DHA activity in soil decreased with depth possibly due to the decrease in SOC, soil aeration and root biomass. Consistent with our study, reduced tillage or ZT increased DHA compared to CT and the values decreased with soil depth (Madejon *et al.*, 2007; Tao *et al.*, 2009).

The FDA hydrolysis is a sensitive and nonspecific test to depict the hydrolytic activity of soil microorganisms and has been commonly accepted as a simple measure of total microbial activity (Adam and Duncan, 2001). The large increase in FDA hydrolysis due to combined effect of ZTW and rice residue could be attributed to organic matter enrichment in 0-5 cm soil layer. Earlier studies have showed a similar increase in FDA in ZT systems under different climatic conditions (Perez-Brandan *et al.*, 2012; Gajda *et al.*, 2013). The activity of APA is linked to the transformation of organic to inorganic P (Yang *et al.*, 2008). Consistent with our results, several studies have reported higher APA activities under ZT than CT (Tao *et al.*, 2009; Qin *et al.*, 2010). Higher PA in ZT with retaining rice straw may be due to organic matter build up after 3 years of continuous ZT than under CT (Yadav and Tarafdar, 2004). The increase in PA under ZTW+R is likely to increase the release of P from phytate present in soil.

Urease (UA) was significantly increased under ZTW+R in 0-5 cm soil layer demonstrating an improvement of soil quality. Mikanova *et al.*, (2009) observed that UA was significantly higher under reduced tillage compared to the CT at soil depth of 0–10 cm. Similarly, Qin *et al.*, (2010) also observed UA was significantly higher in ZT with maize residue as compared to CT without maize residue in 0-10 cm soil layer. An increase in UA is generally accompanied by increase in SOC (Kheyrodin *et al.*, 2012). Activity of UA decreased with depth in all treatments which might be owing to a decrease in SOC.

Retention of rice residue in ZT caused significant increase in the contents of OC, Olsen-P,  $\text{NH}_4\text{OAc-K}$  and DTPA-micronutrients compared to CTW-R (Table 3). The increases in OC under ZT+R have been observed in a range of soil and climatic conditions (Melero *et al.*, 2009; Yadvinder-Singh *et al.*, 2010b). The significant increases in the contents of Olsen-P,  $\text{NH}_4\text{OAc-K}$  and DTPA-micronutrients in 0-5 cm soil layers under ZTW+R compared to CTW-R were probably positive effects of rice residue retention and lower rate of soil organic decomposition under ZT conditions (Salinas-Garcia *et al.*, 2002; Yadvinder-Singh *et al.*, 2014). In addition, the increase in the availability of P in surface layer under ZTW+R compared to CTW-R could also be due to lower P adsorption by soil and the release of organic P during decomposition of rice residues (Qin *et al.*, 2010; Wang *et al.*, 2008). Being a rich source of K, rice straw increased the availability K under ZT+R than the other treatments (Yadvinder Singh *et al.*, 2010a).

In our study, ZTW+R significantly increased the grain yield over ZTW-R and CTW-R. The increase in grain yield of wheat under ZTW+R may be ascribed to the positive effects of soil temperature modifications, soil moisture supply and improved soil fertility due to rice residue mulch (Yadvinder-Singh and Sidhu, 2014). Consistent with our results, earlier studies have also reported higher yields of wheat under ZT+R compared to CT-R (Sidhu *et al.*, 2007). When the entire rice residue was removed (-R), yield of ZTW was significantly lower than CTW. In an earlier study, Arora *et al.*, (2010) reported that poor root growth and lower N use efficiency was responsible for lower wheat yields in ZT-R compared to CT-R.

Significant correlations between different enzyme activities observed in our study (Table 4) suggest a general relationship between soil microbiological properties. Retention of crop residues improved the organic carbon status of soils, which in turn was reflected in the higher soil enzymatic activities (Omidi *et al.*, 2008; Gajda *et al.*, 2013). Qin *et al.*, (2010) found that UA and APA activities were significantly correlated with available P content in soil. The relationship between all assayed soil enzyme activities and chemical properties indicate that these enzymes play an important role in the cycling of elements and initial phases of the decomposition of crop residues. This implies that continuous addition of diversified carbon sources through rice residues as organic manuring enhanced the microbial biomass carbon and abundance of different microbial communities of soil and perhaps the enzyme activities.

#### 4. CONCLUSION

The study showed that zero tillage with rice residues as surface mulch (ZTW+R) markedly improved grain yield of wheat over zero tillage without residue (ZTW-R) after three years of RW cropping sequence. ZTW+R was more effective than ZTW/CTW for increasing the soil organic carbon, Olsen-P,  $\text{NH}_4\text{OAc-K}$



and DTPA-extractable micronutrients in soil surface 0-5 cm layer. A significant change in soil enzyme activities occurred after 3 years of ZTW+R. All soil enzymes were positively correlated with each other and with soil chemical properties and grain yield. The increase in enzyme activities in soil may contribute to a long-term sustainability of RWS under semi-arid climate conditions of South Asia. The beneficial effect of RCTs on soil quality was mainly confined to the soil surface layer.

## ACKNOWLEDGEMENTS

The land and infrastructure support of Punjab Agricultural University, Ludhiana for the field study and financial as well technical support of International Maize and Wheat Improvement Center (CIMMYT) through USAID and BMGF funded Cereal System Initiative for South Asia (CSISA) project and CGIAR Research Program on wheat (CRP 3.1) are gratefully acknowledged.

## COMPETING INTERESTS

Authors have declared that no competing interests exist.

## AUTHORS' CONTRIBUTIONS

*This work was carried out in collaboration between all authors. Author HST, HSS, MLJ and YS designed the study, started the experiment in 2010. Author SKK collected the data, performed the statistical analysis and wrote the first draft of the manuscript. All co-authors were give technical support in data collection, determination of different parameters and statistical analyses of the study. All authors read and approved the final manuscript.*

## REFERENCES

1. Adam G, Duncan H. Development of a sensitive and rapid method for the measurement of total microbial activity using fluorescein diacetate (FDA) in a range of soils. *Soil Biology and Biochemistry*. 2001;33:943–951.
2. Ames BN. Assay of inorganic phosphate, total phosphate and phosphatases. *Methods Enzymology*. 1966;8:115–118.
3. Arora VK, Sidhu S, Sandhu KS, Thind S S. Effects of tillage intensity, planting time and nitrogen rate on wheat yield following rice. *Experimental Agriculture*. 2010;46:267–275.
4. Bendick AK, Dick RP. Field management effects on soil enzyme activities. *Soil Biology and Biochemistry*. 1999;31:1471–1479.
5. Caravaca F, Roldan A. Effect of *Eisenia fetida* earthworms on mineralization kinetics, microbial biomass, enzyme activities, respiration and labile C fractions of three soils treated with a composted organic residue. *Biology and Fertility of Soils*. 2003;38: 45–51.
6. Dick RP. Soil enzyme activities as integrative indicators of soil health. In: Pankhurst C, Doube B, Gupta V (eds.) *Biological Indicators of Soil Health*, CAB International, Wallingford, UK, 1997;pp. 121–156.
7. Douglas LA, Bremner JM. Extraction and colorimetric determination of urea in soils. *Soil Science Society of America Journal*. 1970;34:859–862.
8. Eivazi F, Bayan MR, Schmidt K. Select soil enzyme activities in the historic Sanborn field as affected by long-term cropping systems. *Communications in Soil Science and Plant Analysis*. 2003;3:2259–2275.
9. Elfstrand S, Bath B, Martersson A. Influence of various forms of green manure amendment on soil microbial community composition, enzyme activity and nutrient levels in leek. *Applied Soil Ecology*. 2007;36:70–82.
10. Gajda AM, Przewoka B, Gawryjock K. Changes in soil quality associated with tillage system applied. *International Agrophysics*. 2013;27:133–141.
11. Garcia C, Hernandez MT, Albaladejo J, Castillo V, Roldan A. Revegetation in semiarid zones: influence of terracing and organic refuse on microbial activity. *Soil Science Society of America Journal*. 1998;62:670–676.

12. Gathala MK, Kumar V, Sharma PC, Saharawat YS, Jat HS, Singh M, Kumar A, Jat ML, Humphreys E, Sharma DK, Sharma S, Ladha JK. Optimizing intensive cereal-based cropping systems addressing current and future drivers of agricultural change in the Northwestern Indo-Gangetic Plains of India. *Agriculture, Ecosystems & Environment*. 2013;177:85–97.
13. Gupta RK, Seth A. A review of resource conserving technologies for sustainable management of the rice-wheat cropping system of the Indo-Gangetic plains. *Crop Protection*. 2007;26:436–447.
14. International Rice Research Institute (IRRI) 2000 IRRISTAT for window (CD-ROM) version 4.02b. Los Banos, Philippines:IRRI.
15. Jackson ML. Soil chemical analysis. Prentice Hall of India, Private Limited, New Delhi. 1973;pp. 38–56.
16. Jat ML, Gathala MK, Ladha JK, Saharawat YS, Jat AS, Kumar V, Sharma SK, Gupta R. Evaluation of precision land leveling and double zero-till systems in the rice–wheat rotation: Water use, productivity, profitability and soil physical properties. *Soil and Tillage Research*. 2009; 105:112–121.
17. Jiang HM, Jiang JP, Jia Y, Li FM, Xu JZ. Soil carbon pool and effects of soil fertility in seeded alfalfa fields on the semi-arid Loess Plateau China. *Soil Biology and Biochemistry*. 2006;38:2350–2358.
18. Kandeler E, Mosier AR, Morgan JA, Milchunas DG, King JA, Rudolph S, Tschierko D. Response of soil microbial biomass and enzyme activities to the transient elevation of carbon dioxide in semi-arid grassland. *Soil Biology and Biochemistry*. 2006;38:2448–2460.
19. Kharia Sushil Kumar, Thind HS, Goyal Avinash, Sharma Sandeep and Dhaliwal SS. Yield and Nutrient Uptakes in Wheat under Conservation Agriculture Based Rice-Wheat Cropping System in Punjab, India. *International Journal of Current Microbiology and Applied Sciences*. 2017;6(2): 1698-1708
20. Kheyrodin H, Ghazvinian K, Taherian M. Tillage and manure effect on soil microbial biomass and respiration, and on enzyme activities. *African Journal of Biotechnology*. 2012;11:14652–14659.
21. Klose S, Moore JM, Tabatabai MA. Arylsulfatase activity of microbial biomass in soils as affected by cropping systems. *Biology and Fertility of Soils*. 1999;29:46–54.
22. Lindsay WL, Norwell WA. Development of a DTPA test for zinc, iron, manganese and copper. *Soil Science Society of America Journal*. 1978;42:421-428.
23. Madejon E, Moreno F, Murillo JM, Pelegrin F. Soil biochemical response to long-term conservation tillage under semi-arid Mediterranean conditions. *Soil and Tillage Research* 2007;94:346–352.
24. Melero S, Lopez-Garrido R, Madejon E, Murillo JM, Vanderlinden K, Ordonez R, Moreno F. Long-term effects of conservation tillage on organic fractions in two soils in South-West of Spain. *Agriculture, Ecosystems & Environment*. 2009;133: 68–74.
25. Mikanova O, Javurek M, Simon T, Friedlova M, Vach M. The effect of tillage systems on some microbial characteristics. *Soil and Tillage Research*. 2009;105:72–76.
26. Olsen S R, Cole C V, Waternabe F S, Dean L A. 1954. Estimation of available phosphorous in soil by extraction with sodium bicarbonate. USDA Cir 939:19.
27. Omid H, Tahmasebi Z, Torabi H, Miransari M. Soil enzymatic activities and available P and Zn as affected by tillage practices, canola (*Brassica napus* L.) cultivars and planting dates. *European Journal of Soil Biology*. 2008;44:443–450.
28. Perez-Brandan C, Arzeno JL, Huidobro J, Grumberg B, Conforto C, Hilton S, Bending GD, Meriles JM, Vargas-Gil S. Long-term effect of tillage systems on soil microbiological, chemical and physical parameters and the incidence of charcoal rot by *Macrophomina phaseolina* (Tassi) Goid in soybean. *Crop Protection*. 2012;40:73–82.
29. Qin S, He X, Hu C, Zhang Y, Dong W. Responses of soil chemical and microbial indicators to conservational tillage versus traditional tillage in the North China Plain. *European Journal of Soil Biology*. 2010;46:243–247.
30. Roldan A, Salinas-Garcia JR, Alguacil MM, Caravaca F. Soil sustainability indicators following conservation tillage practices under subtropical maize and bean crops. *Soil and Tillage Research*. 2007;93: 273–282.
31. Salazar S, Sanchez L, Alvarez J, Valverde A, Galindo P, Igual J, Peix A, Santa-Regina I. Correlation among soil enzyme activities under different forest system management practices. *Ecological Engineering*. 2011;37:1123–1131.

32. Salinas-Garcia JR, Velazquez-Garcia JJ, Gallardo-Valdez M, Diaz-Mederos P, Caballero-Hernandez F, Tapia-Vargas L M, Rosales-Robles E. Tillage effects on microbial biomass and nutrient distribution in soils under rain-fed corn production in central-western Mexico. *Soil and Tillage Research*. 2002;66:143–152.
33. Sidhu HS, Singh M, Humphreys E, Yadvinder Singh, Bijay Singh, Dhillon SS, Blackwell J, Bector V, Singh M, Singh S. The Happy Seeder enables direct drilling of wheat into rice stubble. *Australian Journal of Experimental Agriculture*. 2007;47:844–854.
34. Sidhu HS, Singh M, Yadvinder Singh, Blackwell J, Lohan SK, Humphreys E, Jat ML, Singh V, Singh S. Development and evaluation of the Turbo Happy Seeder for sowing wheat into heavy rice residues in NW India. *Field Crops Research*. 2015;184:201–212.
35. Tabatabai M A. 1982. Soil Enzymes. In: A. L. Page, R. H. Miller, D. R. Keeney (eds.) *Methods of Soil Analysis. Part 2. Agronomy 9*, American Society of Agronomy. Madison, USA, pp. 903–947.
36. Tabatabai MA, Bremner JM. Use of p-nitrophenyl phosphate for assay of soil phosphatase activity. *Soil Biology and Biochemistry*. 1969;1:301–307.
37. Tao J, Griffiths B, Zhang S, Chen X, Liu M, Hu F, Li H. Effects of earthworms on soil enzyme activity in an organic residue amended rice–wheat rotation agro-ecosystem. *Applied Soil Ecology*. 2009;42:221–226.
38. Walkey A, Black CA. An examination of the Degtjareff method for determination of soil organic matter and a proposed modification of the chromic acid titration method. *Soil Science*. 1934;37:29–39.
39. Wang Q, Bai Y, Gao H, He J, Chen H, Chesney RC, Kuhn NJ, Li H. Soil chemical properties and microbial biomass after 16 years of no-tillage farming on the Loess Plateau, China. *Geoderma*. 2008;144:502–508.
40. Yadav B K, Tarafdar J C. Phytase activity in the rhizosphere of crops, trees and grasses under arid environment. *Journal of Arid Environments*. 2004;58:285–293.
41. Yadvinder Singh, Gupta R K, Jagmohan S, Gurpreet S, Gobinder S, Ladha J K. Placement effects on rice residue decomposition and nutrient dynamics on two soil types during wheat cropping in rice–wheat system in northwestern India. *Nutrient Cycling in Agroecosystems*. 2010a;88: 471–480.
42. Yadvinder Singh, Singh M, Sidhu HS, Khanna PK, Kapoor S, Jain AK, Singh AK, Sidhu GK, Avtar S, Chaudhary DP, Minhas PS. 2010b. Options for effective utilization of crop residues. Directorate of Research, Punjab Agricultural University, Ludhiana, India. 32pp.
43. Yadvinder Singh, Kukal SS, Jat ML, Sidhu HS. Improving water productivity of wheat-based cropping systems in South Asia for sustained productivity. *Advances in Agronomy*. 2014;127: 157–258.
44. Yadvinder Singh, Sidhu HS. Management of cereal crop residues for sustainable rice–wheat production system in the Indo-Gangetic plains of India. *Proceedings of the Indian National Science Academy*. 2014;80: 95–114.
45. Yang LJ, Li TL, Li FS, Lemcoff J H, Cohen S. Fertilization regulates soil enzyme activity and fertility dynamics in a cucumber field. *Scientia Horticulturae*. 2008;116:21–26.
46. Yang X, Chen C, Luo Q, Li L, Yu Q. Climate change effects on wheat yield and water use in oasis cropland. *International Journal of Plant Production*. 2011;5:83–94.
47. Yuan B, Yue D. Soil microbial and enzymatic activities across a chronosequence of chinese pine plantation development on the loess plateau of China. *Pedosphere*. 2012;22:1–12.
48. Zhao B, Chen J, Zhang J, Qin S. Soil microbial biomass and activity response to repeated drying–rewetting cycles along a soil fertility gradient modified by long-term fertilization management practices. *Geoderma*. 2010;160:218–222.

**Table 1. Description of experimental treatments (Kharia et al 2017)**



Rice (main plots)	Wheat (sub-plots)
T <sub>1</sub> . Zero till direct seeded rice (ZT-DSR). Residues of previous wheat crop were removed. DSR was sown using same drill as in treatment.	1. Conventional till wheat after removal of rice residue (CTW-R). Residues of previous rice removed. Tillage operations for CTW included two passes of harrows and two passes of Tyne plough <sup>y</sup> followed by planking.
T <sub>2</sub> . Conventional direct seeded rice (CT-DSR). Residues of previous wheat crop were removed. Tillage for DSR included two passes of harrows and two passes of Tyne plough <sup>y</sup> followed by planking, The DSR was sown using seed cum fertilizer drill in 20 cm apart.	
T <sub>3</sub> . ZT transplanted rice using mechanical trans planter (ZT-DTR). Residues of previous wheat crop were removed.	2. Zero till wheat after removal of rice residue (ZTW-R). Wheat was direct seeded in the no till plots using zero till drill.
T <sub>4</sub> . Conventional till puddled transplanted rice (PTR): Residues of preceding wheat crop removed. Puddling (wet tillage) was done twice in 6-8 cm of standing water using a tractor-mounted puddler followed by planking.	3. Zero till wheat + residues (ZTW+R) Residues of previous rice crop retained. Wheat was directly seeded into residues using Turbo Happy Seeder (Sidhu <i>et al.</i> , 2015).*

- Kamboj Machinery Manufacturers, Ramdas, Amritsar, Punjab

**Table 2. Methods used for analysis of soil samples for different soil parameters**

Soil properties	Method used	Reference
Organic carbon	Wet digestion method	Walkley and Black (1934)
Olsen P	0.5 M NaHCO <sub>3</sub> -extractable	Olsen <i>et al.</i> , (1954)
Available K	1M NH <sub>4</sub> OAc-extractable	Jackson (1973)
DTPA- Fe, Mn, Cu and Zn	DTPA extractable	Lindsay and Norwell (1978)
Dehydrogenase activity (DHA)	Triphenylformazan (TPF) produced by the reduction of 2, 3, 5-triphenyltetrazolium chloride (TTC).	Tabatabai (1982)
Fluorescein di-acetate activity (FDA)	Fluorescein released by the hydrolysis of fluorescein di-acetate.	Adam and Duncan (2001)
Alkaline phosphatase activity (APA)	p-nitrophenyl method.	Tabatabai and Bremner (1969)
Phytase activity (PA)	Inorganic phosphate released by hydrolysis using sodium phytate	Ames (1966)
Urease activity (UA)	2M KCl- Phenyl mercuric acetate	Douglas and Bremner (1970)

**Table 3. Effect of rice establishment, tillage and rice straw management on dehydrogenase ( $\mu\text{g TPF g}^{-1} 24 \text{ hr}^{-1}$ ) activities at various soil layers.**

Treatments	Soil layers (cm)		
	0-5	5-10	10-15
<b>Rice establishment systems</b>			
DSR-ZT-DSR	303	177	77.4
DSR-CT-DSR	275	168	76.2
ZT-DTR	290	163	74.7
PTR	286	159	72.4
LSD(0.05)	NS	NS	NS
<b>Tillage and rice straw management practices</b>			
CTW-R	265	158	73.4
ZTW-R	291	167	73.8
ZTW+R	309	175	78.4
LSD (0.05)	25.8	NS	NS

**Table 4. Effect of rice establishment, tillage and rice straw management on flourescein diacetate ( $\mu\text{g fluorescein g}^{-1} \text{ dry soil}$ ) activities at various soil layers**

Treatments	Soil layers (cm)		
	0-5	5-10	10-15
<b>Rice establishment systems</b>			
DSR-ZT-DSR	3.32	2.68	2.45
DSR-CT-DSR	3.24	2.74	2.45
ZT-DTR	3.16	2.73	2.50
PTR	3.12	2.74	2.42
LSD(0.05)	NS	NS	NS
<b>Tillage and rice straw management practices</b>			
CTW-R	3.02	2.69	2.42
ZTW-R	3.15	2.72	2.41
ZTW+R	3.47	2.76	2.52
LSD (0.05)	0.120	NS	NS

**Table 5. Effect of rice establishment, tillage and rice straw management on alkaline phosphatase ( $\mu\text{g p-nitrophenol g}^{-1} \text{ dry soil hr}^{-1}$ ) activities at various soil layers**

Treatments	Soil layers (cm)		
	0-5	5-10	10-15
<b>Rice establishment systems</b>			
DSR-ZT-DSR	49.4	42.2	33.9
DSR-CT-DSR	42.5	39.0	34.3
ZT-DTR	43.8	39.1	30.5
PTR	41.4	34.7	28.7
LSD(0.05)	NS	NS	NS
<b>Tillage and rice straw management practices</b>			
CTW-R	41.4	37.6	31.2
ZTW-R	43.6	37.6	31.7
ZTW+R	47.8	41.0	32.7
LSD (0.05)	3.72	NS	NS

**Table 6. Effect of rice establishment, tillage and rice straw management on Phytase ( $\mu\text{g inorganic P released g}^{-1} \text{ dry soil hr}^{-1}$ ) activities at various soil layers**

Treatments	Soil layers (cm)		
	0-5	5-10	10-15
<b>Rice establishment systems</b>			
DSR-ZT-DSR	0.341	0.273	0.238
DSR-CT-DSR	0.340	0.265	0.234
ZT-DTR	0.328	0.268	0.233
PTR	0.340	0.262	0.237
LSD(0.05)	NS	NS	NS
<b>Tillage and rice straw management practices</b>			
CTW-R	0.303	0.254	0.232
ZTW-R	0.310	0.269	0.232
ZTW+R	0.398	0.277	0.241
LSD (0.05)	0.031	NS	NS

**Table 7. Effect of rice establishment, tillage and rice straw management on urease ( $\mu\text{g urea g}^{-1}$  dry soil  $\text{min}^{-1}$ ) activities at various soil layers**

Treatments	Soil layers (cm)		
	0-5	5-10	10-15
<b>Rice establishment systems</b>			
DSR-ZT-DSR	2.60	2.13	1.68
DSR-CT-DSR	2.55	2.14	1.70
ZT-DTR	2.58	2.17	1.60
PTR	2.51	2.18	1.66
LSD(0.05)	NS	NS	NS
<b>Tillage and rice straw management practices</b>			
CTW-R	2.40	2.12	1.63
ZTW-R	2.54	2.16	1.64
ZTW+R	2.75	2.20	1.72
LSD (0.05)	0.070	NS	NS

**Table 8. Effect of rice establishment, tillage and rice straw management on OC ( $\text{g kg}^{-1}$ ) content at various soil layers**

Treatments	Soil layers (cm)		
	0-5	5-10	10-15
<b>Rice establishment systems</b>			
DSR-ZT-DSR	5.88	4.65	3.37
DSR-CT-DSR	5.49	4.93	3.17
ZT-DTR	5.90	4.65	3.43
PTR	5.82	5.03	3.58
LSD(0.05)	NS	NS	NS
<b>Tillage and rice straw management practices</b>			
CTW-R	5.18	4.56	3.32
ZTW-R	5.79	4.86	3.43
ZTW+R	6.35	5.02	3.41
LSD (0.05)	0.30	0.36	NS

**Table 9. Effect of rice establishment, tillage and rice straw management on Olsen P (kg ha<sup>-1</sup>) content at various soil layers**

Treatments	Soil layers (cm)		
	0-5	5-10	10-15
<b>Rice establishment systems</b>			
DSR-ZT-DSR	9.98	8.35	6.24
DSR-CT-DSR	9.69	7.14	4.72
ZT-DTR	10.1	8.32	5.66
PTR	9.13	7.02	5.72
LSD(0.05)	NS	NS	NS
<b>Tillage and rice straw management practices</b>			
CTW-R	8.90	6.82	5.23
ZTW-R	9.64	7.81	6.03
ZTW+R	10.7	8.49	5.50
LSD (0.05)	0.799	0.687	NS

**Table 10. Effect of rice establishment, tillage and rice straw management on available K (kg ha<sup>-1</sup>) content at various soil layers**

Treatments	Soil layers (cm)		
	0-5	5-10	10-15
<b>Rice establishment systems</b>			
DSR-ZT-DSR	44.8	38.1	34.8
DSR-CT-DSR	45.4	37.3	37.8
ZT-DTR	47.0	39.1	38.3
PTR	42.7	34.4	36.9
LSD(0.05)	NS	NS	NS
<b>Tillage and rice straw management practices</b>			
CTW-R	42.6	35.1	36.1
ZTW-R	40.7	35.3	34.9
ZTW+R	51.6	41.3	39.8
LSD (0.05)	6.34	4.01	NS

**Table 11. Effect of rice establishment, tillage and rice straw management on DTPA micronutrients in 0-5 cm soil layer after three cycles of rice-wheat system.**

Treatments	DTPA- extractable micronutrients (mg kg <sup>-1</sup> )			
	Zn	Mn	Fe	Cu
<b>Rice establishment systems</b>				
ZT-DSR	5.17	8.32	30.0	1.34
CT-DSR	4.95	8.20	28.4	1.29
ZT-DTR	4.87	8.12	29.9	1.23
PTR	4.77	8.30	29.4	1.38
LSD (p=0.05)	NS	NS	NS	NS
<b>Tillage and rice straw management practices</b>				
CTW-R	4.85	8.19	28.0	1.25
ZTW-R	4.85	8.04	28.5	1.26
ZTW+R	5.14	8.47	31.8	1.43
LSD (p=0.05)	0.24	0.30	2.40	0.09

**Table 12. Yield and yield attributing parameters of wheat as influenced by rice establishment systems, tillage and rice straw management practices under rice-wheat system (Kharia *et al* 2017).**

Treatment	Grain yield (Mg ha <sup>-1</sup> )	Spike length (cm)	Grains spike <sup>-1</sup>	Grain weight spike <sup>-1</sup> (gram)	1000 grain weight (gram)
<b>Rice establishment systems</b>					
ZT-DSR	4.84	10.5	53.5	2.08	38.0
CT-DSR	4.84	10.8	52.7	1.97	38.0
ZT-DTR	4.88	10.7	53.7	2.04	35.7
PTR	4.77	11.0	52.9	2.01	37.8
LSD (0.05)	NS	NS	NS	NS	NS
<b>Tillage and straw management practices</b>					
CTW-R	4.77	10.5	53.1	2.01	36.8
ZTW-R	4.47	10.9	52.1	1.90	35.8
ZTW+R	5.26	10.8	54.3	2.16	39.6
LSD (0.05)	0.19	0.30	1.10	0.08	2.20





**Table 13. Correlation between soil enzyme activities, grain yield of wheat, pH, organic carbon and Olsen-P in 0-5 cm soil layer after three cycles of rice-wheat cropping system**

Properties	DHA	FDA	APA	PA	UA	GY	pH	OC	Olsen-P
<b>0-5 cm layer</b>									
<b>DHA</b>	1	0.826**	0.843**	0.742**	0.896**	0.488	-0.198	0.674*	0.739**
<b>FDA</b>		1	0.787**	0.886**	0.910**	0.690**	-0.232	0.608*	0.824**
<b>APA</b>			1	0.624*	0.775**	0.500	-0.006	0.497	0.652*
<b>PA</b>				1	0.892**	0.857**	-0.283	0.694*	0.759**
<b>UA</b>					1	0.710**	-0.381	0.717**	0.831**

DHA-Dehydrogenase activity, FDA-Fluorescein di-acetate, APA-Alkaline phosphatase activity, PA-Phytase activity, UA-Urease activity, GY-Grain yield, OC-Organic carbon.

\*\* Significant at  $p < 0.01$ , \* Significant at  $p < 0.05$

515 **Table 14. Correlation between soil enzyme activities and DTPA-extractable**  
516 **micronutrients (mg kg<sup>-1</sup>) in 0-5 cm soil layer**  
517

Parameter	Fe	Mn	Cu	Zn
<b>Dehydrogenase (DHA)</b>	0.775**	0.453	0.650*	0.627
<b>Fluorescein di-acetate (FDA)</b>	0.760**	0.624*	0.717**	0.782*
<b>Alkaline phosphatase activity (APA)</b>	0.685*	0.428	0.519	0.774*
<b>Phytase activity (PA)</b>	0.820**	0.747**	0.847**	0.649*
<b>Urease (UA)</b>	0.857**	0.505	0.681*	0.657

529 \*\* Significant at p<0.01, \* Significant at p<0.05