

Effect of Urine Sources on Some Soil Health indicators, Maize yield and Its Heavy Metals Uptake in Abakaliki, Southeastern Nigeria

Abstract:

The need to find alternative to inorganic fertilizer which is costly and not easily affordable to local farmers necessitates research in rare areas. Thus, this experiment was carried out at the Plant and Screen house to study effect of urine sources on some soil health indicators, maize yield and its heavy metals uptake. Completely Randomized Design was used in laying the experiment. 5kg of sieved soil was treated with different urine sources replicated five times. The result indicates that soil pH, total N and organic matter were respectively significantly ($P<0.05$) higher in different urine sources than control. Human urine had significantly ($P<0.05$) higher treatment effect on soil pH, percent total N and organic matter compared to other sources of urine. Similarly, human urine was 9-10%, 15-27%, 10-47% and 6-5% higher in number of leaves, plant height, grain yield and leaf area index when compared to those of cattle and goat urine sources. Significantly ($P<0.05$) higher copper uptake by maize grains was obtained in control relative to those of urine sources. Copper and lead uptake by maize grains were respectively higher by 20, 80, 87% and 87, 47, 7% in control when compared to human, cattle and goat urine sources. Generally, heavy metals uptake by maize grains is below recommended safe limits for toxicity. Urine from adult animals is recommended as credible alternative for improvement of soil health status and sustainable productivity.

Key words: effect, maize yield, heavy metals uptake, urine sources, soil health indicators.

INTRODUCTION

Traditional agriculture relies heavily on mineral fertilizer NPK for crop production in Nigeria and other developing countries (Nwite, 2015) and incidentally, use of fertilizer is confronted with problems of unavailability, high cost and increase in soil acidity. As a result, use of fertilizer is considered to be counterproductive and there is need for its alternative source. This alternative source is urine since it easily affordable as it could be accessed from livestock and man. It has been reported (Adeoluwa and Sulaiman, 2002), that urine contains useful nutrients which if carefully harnessed could sustain soil health status and increase its

36 productivity. Well preserved urine has good quality and could have the same effect
37 as inorganic fertilizer in optimizing soil fertility status of soil (Nwite, 2015).
38 Research shows that urine contains major nutrients including nitrogen, phosphorus,
39 potassium as well as calcium and magnesium which is dependent on age and feed
40 of the animals (Marino, 2008).

41 When there is no planned disposal of urine it naturally constitutes health hazard
42 due to its pungent odour which could be curtailed through its proper treatment and
43 conversion in treating soil for higher productivity (Nwite, 2015). This offensive
44 odour is attributed to freshly accumulated urine at pH of 6.7 (Hoglung, 2001).
45 Researchers (Heinonen Tanski and Van Wijk-Sibesma, 2005; Kichman and
46 Peterson, 1995; Steineck *et al.*, 1999; Richert *et al.*,2002; Malkki and Heinonen-
47 Tanski, 1999) have shown that human urine source was successfully used as
48 fertilizer in crop production and raising flowers in Europe and other countries.
49 Confirmatory studies have been carried out using Barley and under crop and field
50 trials or even under home gardening (Richert *et al.*, 2002).

51 With the wide spread scarcity of inorganic fertilizer and its associated problems in
52 food production, there is need for alternative source. If appropriate quantity of
53 urine is applied to the soil at right time, its nitrogen contents could have the same
54 value as that of inorganic fertilizer (Adeoluwa and Sulaiman, 2012). For instance,
55 100kg N per hectare of urine improved Barley production between 90 – 110 days
56 of planting in Sweden (Richert *et al.*,2002).

57 Naturally, human being could not easily accept food crops produced with urine due
58 to suspicion of its health hazard status and safe for consumption. This, however
59 could be overcome by treating urine for quality assurance and safe from health
60 hazards (Nwite, 2015). In Nigeria food crops that grow around urinals or where
61 urine is disposed are normally eaten by human beings and animals without any
62 complaints of health problems. The objective of this experiment was to study effect

63 of urine sources on some soil health indicators, maize yield and its heavy metals
64 uptake under Abakaliki agroecological environment.

65
66

MATERIALS AND METHODS

67 **Experimental site**

68 The research was conducted in 2014 at Plant and Screen House of Teaching
69 and Research Farm, Faculty of Agriculture and Natural Resources
70 Management, Ebonyi State University, Abakaliki. The area is located by latitude
71 $06^{\circ} 4'N$ and longitude $08^{\circ} 65'E$ in the derived savanna of southeast agro-
72 ecological zone of Nigeria. The area experiences bimodal pattern of rainfall
73 which is spread from April-July and September-November of each year. There is
74 a break in August normally referred by residents as “August break”. At the
75 beginning of rainfall, it is torrential and violent and is characterized by
76 thunderstorm and lightning. The minimum and maximum rainfalls are 1700
77 and 2000 mm with a mean of 1800 mm (ODNRI, 1989). The temperature during
78 rainy season is usually low ($27^{\circ}C$) but increases to $31^{\circ}C$ in dry season. Relative
79 humidity is 80% in rainy season which declines to 60% during the cold
80 Harmattan periods and dry season of the year (ODNRI, 1989) being
81 characteristics of tropical climate.

82 The soil is derived from sedimentary deposits from cretaceous and tertiary
83 periods. According to Federal Department of Agricultural Land Resources
84 (FDALR, 1985), Abakaliki agricultural zone lies within “Asu River” and is
85 associated with Olive brown shale, fine grained sandstones and mudstone. It is
86 unconsolidated within 1 m depth (Shale residuum) and belongs to the order
87 ultisol classified as *typic haplustult*. The area was grown of short vegetation
88 and medium to tall trees. There is also growth of native grasses, herbs and shrubs
89 with patches of ground.

90 **Experimental Design and Treatment Application**

91 The experimental design used in this study was Completely
92 Randomized Design (CRD). Human urine of male adult was collected from
93 researchers immediate family while cattle and goat urine was sourced from
94 Animal Farm of Faculty of Agriculture and Natural Resources
95 Management, Ebonyi State University, Abakaliki. The animals used were of
96 matured age. The choice of these animals was based on ease of accessibility since
97 every farming family in the locality can afford to keep them. These animals too
98 are omnivorous and have common feeding habit at adult age. The urine was
99 stored in air-tight plastic containers for 6 months before application to ensure
100 sanitation process. Urine treatments used as fertilizer was based on a mean
101 of 4.55 gN/liter content of urine (Table 1) as follows:

102 Human urine = 100 LNha⁻¹ (0.05 litre) equivalent to 100 kg Nha⁻¹

103 Cattle urine = 100 LNha⁻¹ (0.05 litre) equivalent to 100 kg Nha⁻¹

104 Goat urine = 100 LNha⁻¹ (0.05 litre) equivalent to 100 kg Nha⁻¹

105 Control = 0 LNha⁻¹ (0 litre) equivalent to 0 kg Nha⁻¹

106 The urine rates were applied to 5kg of soil weighed into pots two weeks
107 after germination of maize seeds. These treatments were replicated six times to
108 give a total of twenty four experimental pots in the experiment. The pots were
109 watered to field capacity as often as moisture is required.

110 The pots were separated by 0.5m spaces while replicates were set 1m apart.

111 **Planting of maize**

112 Maize variety (Oba super II hybrid) (*Zea mays* L.) collected from
113 Ebonyi State Agricultural Development Programme (EBADEP), Onu Ebonyi
114 Izzi, Abakaliki was used as a test crop. The maize seeds were planted at two
115 seeds per hole and at 5 cm depth in each pot. Two weeks after germination

116 (WAG), thinning was carried out to allow one plant per stand. Weeds were
117 removed by handpicking at regular intervals till harvest.

118 **Agronomic parameters**

119 A total of twelve tagged maize plants were used for study. When the
120 husks were dried, the cobs were harvested, dehusked, shelled and grain yield
121 adjusted to 14% moisture content. Plant height was measured with metric
122 ruler from the base of plant to tallest plant leaf at tasseling. Leaf area index
123 (LAI) was determined by the **formula** according to Nwite *et al.* (2014).

$$124 \quad \text{LAI} = \frac{\text{Leaf area (m}^2\text{)}}{\text{Ground cover (m}^2\text{)}} \dots\dots\dots (1)$$

125

126 **Soil Sampling**

127 Auger sampler was used to collect soil samples at 0-20 cm depth from
128 site where soil used for experiment was collected. The samples were bulked and
129 used for routine laboratory analysis. Samples were further collected from each
130 pot for post-harvest chemical properties determination.

131

132 **Laboratory methods**

133 The samples were dried, ground and passed through 2 mm sieve and used
134 to determine soil properties. Particle size distribution of the experimental soil
135 was determined using the Bouyoucous method as outlined in Gee and Or
136 (2002) procedure. Soil pH determination was carried out in soil/water solution
137 ratio of 1:2.5. The pH values were read off using pH meter with glass
138 electrode (Peech, 1965). Total nitrogen was determined using Micro-kjeldahl
139 procedure (Bremner, 1996). Available phosphorus determination was done
140 using Bray-2 method as outlined in Page *et al.* (1982). Organic matter was
141 determined by Walkley and Black (1934) digestion method. Exchangeable
142 bases of calcium (Ca), Magnesium (Mg), Potassium (K), and Sodium (Na) were

143 extracted using ammonium acetate (NH_4OAC) extraction method. Potassium
144 and sodium were determined using flame photometer. The compositions of
145 urine were determined by Atomic Absorption spectrophotometer as well as crop
146 uptake copper (Cu) lead (Pb) using Dewis and Freitas (1976) procedure.

147

148 **Data analysis**

149 Data collected from the experiment were subjected to Analysis of
150 Variance (ANOVA). Means were separated using Fishers' Least Significant
151 Difference (FLSD) as outlined in Steel and Torrie (1980). Significance was
152 reported at 5% probability level.

153

154

RESULTS AND DISCUSSION

Composition of Urine

Table 1 shows some major nutrients and heavy metals composition of urine source. There were variations in values of nutrients and heavy metals in urine source. Nevertheless, human urine have highest values of nutrients when compared to livestock sources although, comparable. Cattle and goat urine contained 0.10 (g/litre) each of copper (Cu) and lead (Pb) but was not found in human urine. The comparable composition of elemental concentrations in animal urine could be attributed to their adult age, omnivorous nature as well as similarity in their dietary needs.

Table 1. Compositions of some major nutrients and heavy metals in urine sources

Parameter	Human urine	Cattle urine	Goat urine
pH	9.1	9.0	8.9
Ammonia (g/litre)	0.01	0.01	0.01
Nitrogen (g/litre)	4.54	4.52	4.51
Phosphorus (g/litre)	0.04	0.03	0.02
Potassium (g/litre)	0.05	0.03	0.03
Sodium (g/litre)	0.29	0.28	0.28
Copper (g/litre)	-	0.10	-
Lead (g/litre)	-	-	0.10

178 **Properties of Soil before initiation of study**

179 Table 2 shows physicochemical properties of soil before initiation of study.
180 Sand fraction was dominant in the particle size distribution. The textural
181 class was sandy loam. The pH was 5.0 which indicates strongly acidic soil
182 (Landon, 1991). Nitrogen was 0.13% and according to Enwezor *et al.* (1981)
183 is very low and organic matter which had 2.2% value was moderate using
184 Benchmark of FMAWRD (2002) for Tropical soils. Phosphorus (20.40
185 mgkg⁻¹) was high (Enwezor *et al.*, 1989). Exchangeable calcium was of
186 medium value but magnesium, potassium and sodium were very low (Asadu
187 and Nweke (1999). Cation exchange capacity recorded very low values
188 (Asadu and Nweke, 1999). It implies that the soil was of low fertility status
189 as obtained in Abakaliki areas for soils used for maize production as well as
190 other crops.

191 **Table 2.** Properties of soil before initiation of study

192 Soil properties	193 Values
193 Sand (gkg ⁻¹)	750
194 Silt (gkg ⁻¹)	140
195 Clay (gkg ⁻¹)	110
196 Texture class	Sandy Loam
197 pH (H ₂ O)	5.0
198 Total Nitrogen (%)	0.13
199 Organic matter (%)	2.2
200 Available phosphorus (mgkg ⁻¹)	20.40
201 Calcium (cmol kg ⁻¹)	3.10
202 Magnesium (cmol kg ⁻¹)	0.92
203 Potassium (cmol kg ⁻¹)	0.17
204 Sodium (cmol kg ⁻¹)	0.10
205 Cation exchange capacity (cmol kg ⁻¹)	7.50

206 **Effect of Urine Sources on Some Soil Health Indicators**

207 Effect of urine sources on some soil health indicators is shown in Table 3. **Soil**
208 **health indicators are used in this study to indicate parameters that are**
209 **determinant of soil fertility status.** Urine sources had significantly ($P < 0.05$)
210 higher treatment effect on pH when compared with the control. Human source
211 of urine had significantly ($P < 0.05$) higher pH than those of cattle and goat urine
212 sources, respectively. On the other hand, human urine was 5 and 6% higher in
213 pH than the urine from cattle and goat. Similarly, significantly ($P < 0.05$) higher
214 treatment effect was obtained in percent total nitrogen in human and cattle
215 sources of urine relative to control. Furthermore, human urine showed
216 significantly ($P < 0.05$) higher treatment effect on percent total nitrogen
217 compared to those of cattle and goat sources of urine. Available phosphorus
218 obtained in different urine sources slightly varied from that of the control. The
219 available phosphorus of human source of urine was 14% higher than control and
220 generally marginally higher than those of cattle and goat sources of urine. There
221 was significantly ($P < 0.05$) higher treatment effect of urine sources on percent
222 organic matter relative to control. Urine obtained from human and goat was
223 significantly ($P < 0.05$) higher in percent organic matter than the one from cattle.
224 This represents 21 and 14% increments in percent organic matter in human and
225 goat sources of urine compared to that of cattle source.

226 The significant increments of pH, percent nitrogen, organic matter and
227 improvement of available phosphorus show that these soil health indicators
228 were released into the soil by urine sources. This finding indicates that urine
229 could substitute mineral inorganic fertilizer as it could be used as fertilizer to
230 supply essential and major nutrients to soil on one land and on the other
231 improve soil health status. These findings are in line with the report of
232 Adeoluwa and Sulaiman (2012) that urine used as fertilizer improved soil health

233 status. Several researchers (Gutser *et al.*, 2005; Schonning, 2001; Adeoluwa and
 234 Cofie, 2012) reported positive impact of urine on nitrogen which increased and
 235 sustained soil fertility. Higher significant positive effect of human urine source
 236 on soil health indicators suggests that it could be more superior than other urine
 237 sources in improving soil health indicators (Nwite, 2015). This finding had
 238 earlier been reported by Benge, (2006) and Adeoluwa and Sulaiman (2012).
 239 This by extension suggests that indeed, urine sources and particularly human
 240 urine could serve as useful alternative fertilizer for crop production.
 241 The positive impacts of the urine fertilizer on soil health indicators indicate an
 242 improvement on the soil health status. The human urine source improved the
 243 soil pH keeping it within a safe range of 5.6-6.0, nitrogen and organic matter at
 244 significant levels than other sources. Peverly and Gates (1973) stated that
 245 organic fertilizers perform better with some crops. This is further supported by
 246 Adeoluwa and Cofie (2012) findings that urine fertilizer improved some health
 247 indicators.

248

249 **Table 3.** Effect of urine sources on some soil health indicators

250 Treatment	pH H₂O	Total N%	Pmgkg⁻¹	OM%
251 Control	5.1d	0.10b	25.60	1.05d
252 Human urine	6.0a	0.14a	29.65	1.76a
253 Cattle urine	5.7b	0.12b	28.24	1.40a
254 Goat urine	5.6c	0.11b	28.30	1.62b
255 FLSD(0.05)	0.1	0.02	NS	0.05

256 P – Available phosphorus, OM(%) – Percent organic matter, N(%) -Percent
 257 Total nitrogen. Treatment means with different letters indicate significant
 258 differences.

259

260

261 **Effect of Urine Sources on Agronomic Yield of Maize**

262 Table 4 shows effect of urine sources on agronomic yield of maize. Urine
263 sources had higher number of leaves, plant height, taller maize plants and larger
264 leaf area when compared with the control, respectively. Human urine source
265 was 9-10%, 15%-27%, 10-47% and 6-5% higher in these maize agronomic yield
266 than those of cattle and goat urine sources. The effect of urine sources not
267 having any significant effect on agronomic parameters of maize could be linked
268 to inherent capability of the maize plant (Benge, 2006). Results in Table 1 did
269 not show much variations on nutrients compositions and this could have
270 influenced maize agronomic parameters. The generally higher agronomic yield
271 of maize in urine sources relative to control could be attributed to improved soil
272 health status by urine treatment. This by implication underscores the usefulness
273 of urine as credible of alternative to inorganic fertilizer for sustenance of soil
274 fertility status and in increased maize production that is safe for human
275 consumption. Human urine increased agronomic yield of maize due to its high
276 nutrients (Table 1) and its ability to release same into the soil. Benge (2006) and
277 Adeoluwa and Sulaiman (2012) pointed out that human urine increased soil
278 fertility and Jathropha production.

279 **Table 4.** Effect of urine sources on agronomic yield of maize

280 Treatment	No of leaf	Plant height (cm)	Grain yield (g/pot)	LAI
281 Control	11.2	58.12	2.0	0.60
282 Human urine	13.4	85.08	4.2	0.66
283 Cattle urine	12.2	72.14	3.8	0.62
284 Goat urine	12.0	61.98	2.2	0.63
285 FLSD (0.05)	NS	NS	NS	NS

286 LAI – Leaf area index.

287
288
289

290 **Effect of Urine Sources on Heavy Metals Uptake by Maize Grains**

291 Effect of urine sources on heavy metals uptake by maize grains is shown
292 in Table 5. The result showed significantly ($P < 0.05$) higher effect of copper
293 uptake by maize grains in control compared to urine sources of human, cattle
294 and goat. This accounted for 20, 80 and 37% increments of copper uptake in
295 maize grains in control respectively when compared to human, cattle and goat
296 sources of urine. Lead uptake by maize grains was 87, 47 and 7% higher in
297 control relative to human, cattle and goat sources of urine.

298 The significantly higher copper uptake by maize grains grown in control plot
299 compared to those grown in urine sources treated plots could be attributed to
300 inputs from soil rather than urine fertilizer. Analysis of urine sources indicated
301 very low presence of copper and lead (Table 1). The same trend of higher Cu
302 uptake by maize grains was shown by control in Pb uptake by maize grains
303 compared to those obtained under urine sources treatment. These findings show
304 that urine could be used as fertilizer for crop production without placing man at
305 a risk of ecotoxicity of heavy metals. This finding could be attributed to
306 improved health status of soil (Table 3) and low presence of heavy metals in
307 urine sources. The likelihood of heavy metals to build up in soil amended with
308 urine fertilizer appears to be higher in soils treated with goat and cattle urine
309 than human urine. Adewole *et al.* (2008) reported heavy metal uptake by crops
310 in their work and noted that these heavy metals were stored in crop parts.
311 Anikwe and Nwobodo (2002) and corroborated by Asadu *et al.* (2008) in their
312 findings observed that human beings were at risk of heavy metals toxicity if
313 they could utilize crops grown around areas contaminated with heavy metals
314 due to eco-toxicity. This could be possible through recycling of heavy metals
315 through food chain. Heavy metal of lead has the capacity to cause brain, renal or
316 reproductive disorders in human beings. The heavy metals of copper and lead

317 are below 0.0-2.0 and 0.01 rated as medium to low (LASEPA, 2005) values and
 318 far below 0.0-5.0 (WHO, 1996) or 2-1500 and 2-300 recommended as normal
 319 by Alloway (1990). However, the interesting result is that heavy metal uptake
 320 by maize grains could not be linked to urine treatment of soil.

321
 322

323 **Table 5.** Effect of urine sources on heavy metals uptake by maize grains

Treatment	$\xrightarrow{\hspace{1cm}}$ mgkg⁻¹ $\xleftarrow{\hspace{1cm}}$
	Cu Pb
Control	0.30a 0.30
Human urine	0.04a 0.04
Cattle urine	0.24b 0.16
Goat urine	0.06c 0.28
FLSD (0.05)	0.05 NS

331 Cu – Copper, Pb – Lead, Treatment means with different letters indicate
 332 significant differences from each other.

333 **Conclusion**

334 This study has shown that urine sources could improve soil health status
 335 and serve as useful alternative fertilizer for maize crop production. Urine
 336 sources significantly improved soil health indicators. Agronomic parameters
 337 responded positively to improved soil health status and performed better in
 338 urine sources than control. Perhaps, the greatest beneficial aspect of use of urine
 339 as fertilizer is low input of heavy metals which keep them below safe limits and
 340 without any danger of eco-toxicity. In view of its superior performance over
 341 other urine sources, human urine could be harvested for treatment of soil for
 342 higher productivity rather than be allowed to be wasted through improper
 343 disposal.

344

345

References

346 Adeluwa, O. O. and Cofie, O. (2012). Urine as an alternative fertilizer in
347 Agriculture: **In:Effects on** Amaranths (*Amaranthus caudatus*) Production.
348 Renewable Agriculture and Food Systems 8:1. doi:
349 10.1017/51742170511000512.

350 Adeluwa, O. O. and Suleiman, O. N. (2012). Effect of human urine on the **growth**
351 **performances** of *Jathropha curcas* seedlings and some soil health indices.
352 Nigerian Journal of Soil Science 22 (2):186-193.

353 Adewole, M.B. Adeoye, G.O. and Sridhar, M.K.C. (2008). Effect of inorganic
354 and organo mineral fertilizers on the uptake of selected heavy metals by
355 *Helianthus annuus L* and *Tithornia diversifolia* (Hems h) under green house
356 condition. **Journal of Toxicological and Environmental**. Chemistry 91(5):
357 970-980.

358 Alloway, B. J. (1990). Heavy metals on soils. New York: John Wiley and Sons,
359 Inc 280p.

360 Anikwe, M.A.N. and Nwobodo, K. C. A. (2002). Long-term effect of municipal
361 wastes disposal on soil properties and productivity in sites used for urban
362 agriculture in Abakaliki, Nigeria. Bio-resources Technology 83:24-50.

363 Asadu, C. L. A. and Nweke, F. I. (1999). Soils of arable crop fields in sub-Sahara
364 Africa: Focus on cassava growing areas, collaborative study of cassava in
365 Africa. Working Paper No. 18. Resources and Crop -Management Division,
366 IITA, Ibadan, Nigeria. 1782p.

367 Benge, M. (2006). Assessment of the potential of *Jathropha curcas*, (*biodiesel*) for
368 the energy production and other uses in developing countries. Posted on EC
369 Ho's website with permission of the author. July 2006 and updated August
370 2006. 2: 22. [http://www.ascension-](http://www.ascension-publishing.com/B/Z/Jathropha)
371 [publishing.com/B/Z/Jathropha](http://www.ascension-publishing.com/B/Z/Jathropha). Pdf
download on January 9, 2012.

372 Bremner, J. M. (1996). Nitrogen-Total. *In: Sparks, D. L. (ed). Methods of Soil*
373 *Analysis. Chemical Methods. American Society of Agronomy* 5(3):1085-
374 1121.

375 Dewis, J. and Freitas, F. (1976). Physical and Chemical Methods of Soil and Water
376 Analysis. Soil Bulletin 18, FAO, UN, Rome.

- 377 Enwezor, W. O., Udo, E. J., Usoroh, N. J. Ayoade, K. A., Adepetu, J. A., Chude,
378 V. O. and Udegbe, C. J. (1989). Fertilizer Use and Management Practices for
379 Crops in Nigeria, Series No 2. Fertilizer Procurement and Distribution
380 Division, Federal Ministry of Agriculture, Water Resources and Rural
381 Development, Lagos, Nigeria.
- 382 Enwezor, W.O., Udo, F. J. and Sobulo, R.A. (1981). Fertility status and
383 productivity of acid sands: *In: Acid sands of Southeastern Nigeria*. Soil
384 Science Society of Nigeria. 1: 56-73.
- 385 Federal Department of Agricultural Land Resources (FDALR) (1985).
386 Reconnaissance Soil Survey of Anambra State, Nigeria. Soil Report, Kaduna.
387 3p.
- 388 Federal Ministry of Agriculture and Rural Development (FMARD) (2002).
389 Fertilizer Use and Management Practice for Crops in Nigeria. Produced by the
390 Federal Fertilizer Department. *In: Aduayi, E. A., Chude, V. O. Adebusuyi, B.*
391 *A. and Olayiwola, S. O. (eds), Abuja 2002. 188p.*
- 392 Gee, G. W. and Or, D. (2002). Particle Size Analysis. *In: Dane. J. H. and Topp, G.*
393 *C. (eds). Methods of Soil Analysis. Physical Methods. Soil Science Society*
394 *America. 5(4):255-293.*
- 395 Gutser, R., Ebertseder, T., Weber, A., Schrami, M. and Schmmidhlter, U. (2005).
396 Short-term and residual availability of nitrogen after long-time application of
397 organic fertilizers on arable land. *Journal of Plant Nutrition and Soil Science*
398 *168:439-446.*
- 399 Heinnonen-Tanski, and Van Wijk-Sibesma, C. (2005). Human excreta for plant
400 production. *Bioresource Technology* 96:403-411.
- 401 Hoglung, C. (2001). Evaluation of Microbial Health Risks Associated with the
402 Reuse of Source-Separated Human urine. Ph.D Thesis, Department of
403 Biotechnology, Applied Microbiology, Royal Institute of Technology (KTH)
404 Stolkolm Sweden. 62-64.
- 405 Kirchmann, H. and Peterson, S. (1995). Human urine-chemical composition
406 and fertilizer use efficiency. *Fertilizer Resources* 40:149-154.
- 407 Lagos State Environmental Protection Agency (LASEPA, 2005). Lagos State
408 Environmental Reports 3: 28-33.

- 409 Landon, J. R. (eds) (1991). Booker, Tropical Soil Manual. A hand book for
410 Soil Survey and Agricultural Land Evaluation in Tropics and Subtropics
411 New York, USA, John Wiley and Sons: Inc. Third Avenue.
- 412 Malkki, S. and Heinonen-Tanski, (1999). Composition of toilets in permanent
413 houses. *In: 1 Use of municipal organic wastes. Proceedings of Nigerian*
414 *Journal of Forestry Seminar 292 DIAS-report. 13: 147-154.*
- 415 Marino, C. (2008). Urine composition depends on certain factors. *Journal of*
416 *Soil Science Society of America. 73:159-219.*
- 417 Nwite, J.N. (2015). Effect of urine source on soil properties and maize yield in
418 Abakaliki, South-Eastern Nigeria. *International Journal of Advance*
419 *Agricultural Research 3:31-36.*
- 420 Nwite, J. N. (2013). Evaluation of the productivity of a spent Automobile oil
421 contaminated soil amended with organic wastes in Abakaliki, Southeastern
422 Nigeria. Ph.D Thesis, University of Nigeria, Nsukka 130p.
- 423 Nwite, J. N., Okolo, C. C., Ezeaku, P. I. and Enyioko, C. (2014). Effect of
424 Integrated Nutrient Management on Soil Chemical properties and maize
425 yield on a sandy loam in Abakaliki, Nigeria. *International Journal of*
426 *Agriculture and Biosciences 3(6):278-282.*
- 427 Overseas Development of Natural Resources Institute (ODNR) (1989). Nigeria
428 Profile of Agricultural Potential ODA, United Kingdom. 3p.
- 429 Page, A. L., Miller, R. H. and Keeney, D. R. (1982). Methods of Soil
430 Analysis. *American Society of Agronomy 9:539-579.*
- 431 Peech, M. (1965). Hydrogen activity. *Methods of Soil Analysis. In: Black CA*
432 *(ed). American Society of Agronomy 9(1):914-926.*
- 433 Perverly, J.H. and Gates, P.B. (1993). Utilization of municipal solid waste and
434 sludge compost in crop production systems. *In: Sewage sludge, Land*
435 *utilization and the Environment. Proceedings of a conference by American*
436 *Society of Agronomy.*
- 437 Richert Stintizing, A., Rodhe, L., Akerhieln, H. and Stenieck, S. (2002).
438 Human urine as a fertilizer and plant nutrients application technique and
439 environmental effects. *In: Venglosk, J., Greserova, G. (eds). Proceedings*

- 440 of 10 International Conference Ramiran 2002 Network. FAO European
441 System of Cooperative Research Network 161-162 Pp.
- 442 Schonning, C. (2001). Urine diversion-hygienic risks and microbial guidelines
443 for reuse. Department of Parasitology, Mycology and Environmental
444 Microbiology. Swedish Institute for Infections.
- 445 Steel, G. D. and Torrie J. H. (1980). Procedures of statistics. A biometrical
446 approach, 2nd ed. New York, McGraw Hill, Book Company 63p.
- 447 Steineck, S., Richert Stintzing, A., Rodhe, L., Elinquist, H. and Jakobsson,
448 M. C. (1999). Plant nutrients in human and food refuse. *In: Proceedings*
449 *of Nigerian Journal of Forestry Seminar 292. DIAS report. 13:125-130.*
- 450 Walkey, A. and Black, A. (1934). An examination of the Degtjareff method for
451 determining soil organic matter and a proposed modification of the chronic
452 acid titration method. *Soil Science 37:29-38.*
- 453 World Health Organization (WHO, 1996). Guideline for drinking quality water.
454 Switzerland 1(2&3):50-57.