Original Research Article

Tillage and Rice Straw Management affect Soil Enzyme Activities and Chemical Properties after Three Years of Conservation Agriculture Based Rice-Wheat System in North-Western India

7 ABSTRACT

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Aims: Water, energy and labour scarcity, and increasing cost of production are major challenges faced by the farmers under intensive tillage- based conventional rice–wheat production system of Indo-Gangetic Plains in South Asia. To address these challenges, various Resource Conservation Technologies (RCTs) such as zero tillage in wheat, dry seeded rice and crop residue retention are being developed and promoted to increase the productivity and profitability of rice–wheat cropping sequence in the region. We evaluated the effects of rice establishment, tillage and rice straw management on changes in soil enzyme activities and chemical properties in soil after three cycles of continuous rice-wheat system. **Study design:** Split plot design

Place and Duration of Study: Department of Soil Science, Punjab Agricultural University, Ludhiana, Punjab-141004, India, between June 2010 and July 2013 **Methodology:** The experiment was started during kharif season of 2010. The design of an experiment was having 12 treatments with 3 replications. The main plot treatments in rice (zero till direct seeded rice, ZT-DSR; conventional till direct seeded rice, CT-DSR; zero till direct transplanted rice, ZT-DTR and puddled transplanted rice, PTR) and three sub-plot treatments in wheat (conventional till wheat without rice straw, CTW-R; ZT wheat without rice straw, ZTW-R, and ZT wheat with rice straw retained as surface mulch using Happy Seeder, ZTW+R).

Results: Zero tillage with rice straw retention (ZTW) as surface mulch (+R) increased wheat yield 9% and 15% compared with conventional tillage (CTW) and ZTW with no residue (-R). Significantly higher dehydrogenase, fluorescein diacetate, alkaline phosphatase, phytase and urease activities were recorded under ZTW+R compared with ZTW/CTW-R in 0-5 cm soil layer. Organic carbon, Olsen-P, available K and DTPA-extractable micronutrients (Zn, Fe, Mn and Cu) in the surface 0-5 cm soil layer were significantly higher in ZTW+R compared with ZTW/CTW-R. Soil enzyme activities were significantly and positively correlated with each other, soil organic carbon, Olsen-P and grain yield of wheat. **Conclusion:** We concluded that RCTs (ZTW and rice residue retention) improve soil enzyme activities and chemical properties in surface 0-5 cm soil layer and enhance productivity and sustainability of rice-wheat system.

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10 Keywords: conservation agriculture, rice straw surface mulch, tillage, enzyme activities, rice-wheat 11 system

12 **1. INTRODUCTION**

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The rice-wheat cropping system (RWS) occupies about 13.5 million hectares (M ha) in the Indo-Gangetic 14 Plains (IGP) of South Asia (Gupta and Seth, 2007) and is fundamental to employment, income and 15 16 livelihood for millions of people in the region. However, sustainability of intensive tillage-based 17 conventional RWS is threatened by water, energy and labour scarcity, increasing cost of production, increasing air pollution and deteriorating soil health. To address these issues various Resource 18 Conservation Technologies (RCTs) such as zero tillage, direct seeded rice and crop residue retention are 19 being developed and promoted for RWS (Sidhu et al., 2007; Gathala et al., 2013). Puddling in rice and 20 21 intensive tillage in wheat are known to cause sub-soil compaction, deterioration of soil structure, and 22 decrease in permeability in the subsurface layer and thereby adversely affecting productivity of RWS (Jat 23 et al., 2009).

24 The increasing constraints of labour and time under intensive agriculture have led to the adoption 25 of mechanized farming in RWS leaving large amounts of crop residues in the fields. As crop residues 26 interfere with tillage and seeding operation for the next crop, farmers in northwestern (NW) India often 27 prefer to burn surplus rice residues on-farm after grain harvest to establish the next wheat crop. Residue 28 burning impacts human and animal health both medically, and by traumatic road accidents due to 29 restricted visibility (Yadvinder-Singh et al., 2014). Establishment of wheat crop by ZTW with retention of crop residues on the soil surface potentially offers a labour-saving and cost-effective alternative to the 30 31 burning of rice residues. A new machine, known as the 'Happy Seeder' (HS), has now been developed 32 for this purpose which is capable of direct drilling wheat into heavy rice residue loads, without burning in a 33 single operation (Sidhu et al., 2007, 2015).

There is currently much interest in dry direct seeded rice (DSR) as an alternative to conventional transplanted rice in North-West India due to labour scarcity for transplanting, and because puddling and transplanting require large amounts of water to establish the rice crop. Compared with removal/ burning of crop residues, ZT with retention of rice residue (ZTW+R) has been suggested to conserve water content of soil, improve overall soil physical and chemical health through replenishing soil organic matter and to support sustainable RWS (Jat *et al.*, 2009; Gathala *et al.*, 2013). Tillage, crop establishment and residue management options may also have significant effect on soil biological health.

41 Soil enzyme activities respond much more quickly to the changes in tillage and soil management as compared with soil organic matter (Jiang et al., 2006) and therefore could be used as potential 42 43 biological indicators of soil quality (Yang et al., 2011). Dehydrogenase activity is involved in oxidative 44 phosphorylation and reflects the total oxidative potential of the soil microbial community (Dick, 1997). 45 Fluorescein diacetate (FDA) hydrolyzed by a number of different enzymes, can provide comprehensive 46 microbial activity (Bendick and Dick, 1999). The activity of alkaline phosphatase is linked to 47 transformation of organic P in soil to inorganic P compounds (Yang et al., 2008). The soil phytase 48 enzyme increases the availability of P from phytate in soils. Urease activity controls hydrolysis of urea in 49 soils, which is an important source of N.

50 Significant increases in the activity of soil enzymes have been observed under ZT and residue 51 retention in non-rice based cropping systems (Eivazi *et al.*, 2003) and the magnitude of the change 52 depends on soil type and climate. Limited information, however, is available in the literature on the effects 53 of RCTs (DSR, tillage and rice straw management) on changes in soil biochemical properties under RWS 54 in South Asia. The present study was therefore, undertaken to determine the effects of rice establishment 55 methods, tillage and rice straw management on biochemical properties after three cycles of RWS in 56 northwestern India.

58 2. MATERIAL AND METHODS

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60 Description of the experimental site and climate

61 A three-year (2010-2013) field experiment on irrigated RWS was conducted on a Typic Ustochrept sandy 62 loam soil at the experimental farm of the Punjab Agricultural University, Ludhiana (30°56'N and 75°52'E) 63 in the IGP in the northwestern India. The top-soil (0-15 cm layer) at the start of experiment was nonsaline (electrical conductivity 0.36 dS m⁻¹) with pH 7.88 and contained 4.5 g kg⁻¹ Walkley-Black carbon, 64 65 8.2 mg kg⁻¹ 0.5 M NaHCO₃-extractable P (Olsen *et al.*, 1954) and 50.4 mg kg⁻¹ 1 N NH₄OAc-extractable K. The region has a sub-tropical climate, with hot, wet summers and cool dry winters. Annual mean 66 rainfall is 760 mm, about 80% of which occurs from June to September. The long-term average (30 years) 67 mean minimum and maximum temperatures in wheat (November to April) are 6.7 and 22.6^oC while in rice 68 (June to October) are 18 and 35^oC, respectively. 69

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71 Experimental layout and treatments

72 Treatments applied to RWS were arranged in a split-plot design with a total of twelve treatments 73 replicated three times. Main plot treatments applied to rice were four combinations of tillage and crop 74 establishment methods in rice (zero till direct seeded rice, ZT-DSR; conventional till direct seeded rice, 75 CT-DSR; zero till mechanically transplanted rice, ZT-DTR and puddled transplanted rice, PTR). The three 76 sub-plot treatments in wheat were combinations of tillage and residue management options (conventional 77 till wheat without residues, CTW-R; ZT wheat without residues, ZTW-R, and ZT wheat with residues 78 retained as surface mulch using Happy Seeder, ZTW+R). The treatments were assigned to the same 79 experimental plots in the 3 years of the study. The treatment details are summarized in Table 1.

81 Soil and crop management

82 Wheat straw was removed at harvest in April each year and the plots were fallowed until pre-83 irrigation for rice in early June. ZT-DSR was sown in a single operation using zero-till-fertilizer cum seed 84 drill at 20 cm row spacing. In CT-DSR, plots were prepared by 2 harrowings + 1 cultivator + 1 planking 85 followed by dry seeding of rice using a seed rate of 20 kg ha⁻¹. Light irrigation (50 mm) was applied at 1 day after seeding to ensure satisfactory germination, and then at 4-5 day intervals until physiological 86 87 maturity depending on the rainfall events during the growing season. In ZT-DTR, rice seedlings were 88 transplanted in ZT plots with standing water using mechanical transplanter. In PTR, tillage included 89 disking twice in early June followed by two cultivator operations in standing water to puddle the soil and 90 planking. Rice (variety PR 114) was transplanted manually using 30-day-old seedlings spaced at 15 cm x 20 cm in the second week of June. All treatments received a uniform dose of 150 kg N as urea, 26 kg P 91 92 as diammonimum phosphate and 25 kg K as muriate of potash. Whole of P and K was applied at rice planting on all the plots. Fertilizer N in PTR and ZT-DTR was applied in three equal split doses at 93 94 transplanting and at 3 and 6 weeks after transplanting. While in CT-DSR and ZT-DSR fertilizer N was 95 applied in three equal spilt doses at 3, 5 and 9 weeks after sowing. Rice was harvested manually in the 96 second week of October in -R to remove the rice straw from the field plots and the combine harvester 97 was used in +R plots to retain the rice straw in the field.

98 All plots received flood irrigation (75-80 mm) prior to planting of wheat. In conventional plots, seed 99 bed was prepared by 2 dry harrowings followed by 2 cultivators and one planking. Wheat (variety PBW 100 621) was sown in the second week of November using a seed rate of 100 kg ha⁻¹. Sowing of wheat was done on the same day in all the treatments at row spacing of 20 cm using Happy Seeder machine. 101 102 Fertilizer N (120 kg ha⁻¹) as urea was applied in three equal split doses at sowing, and three weeks and 8 weeks after planting. A basal dose of 26 kg P ha⁻¹ as single super phosphate and 25 kg K ha⁻¹ as muriate of potash were applied on all plots at planting. Wheat was irrigated (each of 75 mm) at crown root 103 104 initiation (CRI), maximum tillering (MT), panicle initiation and dough stages recommended for wheat crop 105 106 in the region. Grain yield of wheat was recorded from 1.4 m x 9 m area in the center of each plot and was 107 reported on dry weight basis.

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109 Soil sampling and analysis

Soil samples were collected from 0-5 cm, 5-10 cm and 10-15 cm soil layers, after the harvest of wheat crop (after 3 cycles of rice-wheat rotation). The soil samples were collected with the help of tube auger (25 mm internal diameter) from randomly selected 4 places within each treatment plot. After removing visible root debris, the soil samples were mixed, sieved (2 mm) and stored at 4° C for subsequent analysis for biochemical soil properties. The methods used for assaying biochemical soil properties are listed in Table 2.

117 Statistical analysis

All the dataset was analyzed using analysis of variance (ANOVA) and differences among treatments were compared at p=0.05 level of significance using the IRRISTAT data analysis package (IRRI, 2000). Correlations between the variables were assessed by determining Pearson correlation coefficients (r) and probabilities. In all the analyses, significance was accepted at a level of probability (p) of <0.05.

124 **3. RESULTS AND DISCUSSION**

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There was no significant interaction effects of main (rice establishment methods) and sub-treatments (tillage and rice residue management in wheat) on wheat yield, enzymes activities and chemical properties in all the three soil layers (0-5, 5-10 and 10-15 cm). Therefore, main effects of the treatments are reported and discussed in the following sections. Rice establishment systems also showed no significant effects on the grain yield of subsequent wheat and any of the soil property measured in the study.

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134 Enzyme activities in soil

Tillage and rice straw management in wheat significantly (P≤0.05) influenced all the enzyme activities in 135 0-5 cm soil layer but showed no effect at 5-10 cm and 10-15 cm soil layers (Tables 3-7). In general, 136 137 enzyme activities decreased with depth. In 0-5 cm soil laver, DHA under ZTW+R were 6% and 14 % 138 higher as compared to ZTW-R and CTW-R, respectively (Table 3). DHA was 9 % higher under ZTW-R 139 than CTW-R. FDA and APA at 0-5 cm depth were 9 and 13 % higher in ZTW+R as compared to ZTW-R 140 and CTW-R, respectively (Table 4 and 5). PA was increased by 9% and 24 % under ZTW+R in 0-5 cm 141 layer compared to ZTW-R and CTW-R, respectively (Table 6). Three enzyme activities (FDA, APA and 142 PA) were significantly higher under ZTW-R than CTW-R. The increase in UA under ZTW+R was 8 and 13 143 % higher as compared to ZTW-R and CTW-R in 0-5 cm soil layer, respectively (Table 7).

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145 Chemical properties of soil

146 Tillage and rice straw management practices in wheat significantly affected the soil chemical properties, 147 except soil pH and EC in the 0-5 cm layer only (data not shown) (Tables 8-11). Soil organic carbon 148 content in 0-5 cm layer in ZTW+R treatment was increased by 10.3% and 23.1% compared to ZTW-R 149 and CTW-R, respectively (Table 8). ZTW-R plots had significantly higher organic carbon content than 150 CTW-R. Similarly, Olsen-P, NH₄OAc-extratable - K and DTPA-extractable micronutrients (Zn, Fe, Mn and 151 Cu) were significantly higher in ZTW+R compared to ZTW-R and CTW-R treatments (Table 9-11). The 152 increase in micronutrient availability was greater for Fe and Cu (12-14%) compared with the increase in 153 the Zn and Mn (3-6%) (Table11).

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156 Grain yield and yield attributes of wheat

157 Grain yield of subsequent wheat was not significantly affected by the rice establishment methods, irrespective 158 of tillage and straw management in wheat (Table 12). However, tillage and rice straw management practices 159 in wheat significantly affected grain yield of third wheat crop (Table 12). ZTW+R produced 9% and 15% higher grain yield than ZTW-R and CTW-R, respectively. CTW-R produced significantly higher (6 %) grain 160 yield than ZTW-R. As the grain yield of subsequent wheat was not significantly affected by the rice 161 162 establishment methods so the yield attributes did not differ significantly (Table 12). However, tillage and straw 163 management practices in wheat significantly affected wheat grain yield attributes. All the yield attributes 164 namely Spike length, grains per spike, grain weight per spike and 1000 grain weight were highest under 165 ZTW+R followed by CTW-R and ZTW-R.

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167 Correlation between soil enzyme activities, grain yield and soil chemical properties

168 Activities of all of the enzymes assayed in the study were significantly and positively correlated 169 with each other as well as with organic carbon and Olsen-P in surface 0-5 cm soil layer (Table 13). The 170 values of correlation coefficient (r) for soil enzyme activities ranged from 0.62 to 0.96. While the lowest 171 value of 'r' was between APA and PA, the highest value was recorded for FDA and PPA. Except for APA, 172 all the other enzymes showed significant and positive relationship (r = 0.61 - 0.72) with organic carbon in 173 the 0-5cm soil layer. A strong and positive relationship (r = 0.65 - 0.83) was observed between all the 174 enzyme activities and Olsen P in 0-0.5 m layer. Grain yield of wheat showed significant positive 175 relationship (r = 0.69 - 0.86) with FDA, PA, PPA and UA in the surface 0-5 cm soil layer (Table 13). The 176 activities of all the enzymes were significantly and positively correlated (r = 0.63 - 0.86) with DTPA-177 extractable Fe and Zn in 0-5 cm layer (Table 14). The relationship with DTPA-Mn was significant only for 178 FDA and PA.

179 180 **DISCUSSIO**

180 **DISCUSSION**

181 In our study, ZTW+R improved all soil enzyme activities in the 0-5 cm soil layer of than ZTW-R and CTW-182 R. Tillage and crop residues management practices have been reported to increase enzyme activities in 183 soils (Klose et al., 1999). The stimulating effect of organic materials on enzyme activities in soil is 184 consistent with results earlier reported (Elfstrand et al., 2007; Garcia et al., 1998). The rice straw provided 185 carbon for enhancing microbial activity and thereby increased activity of several enzymes (Caravaca and 186 Roldan, 2003). The increases in microbiological activities under ZT compared to CT have been linked to 187 the increases in organic matter content of surface soils (Roldan et al., 2007; Melero et al., 2009). The 188 addition of organic matter on CT fields can rapidly increase the soil enzyme activities (Kandeler et al., 189 2006).

DHA activity occurs intracellular in all living microbial cells (Yuan and Yue, 2012; Zhao *et al.*, 2010) and can be used as an indicator of overall soil microbial activity (Salazar *et al.*, 2011). DHA responded to tillage and residue management treatments in a similar manner as SOC. DHA activity in soil decreased with depth possibly due to the decrease in SOC, soil aeration and root biomass. Consistent with our study, reduced tillage or ZT increased DHA compared to CT and the values decreased with soil depth (Madejon *et al.*, 2007; Tao *et al.*, 2009).

196 The FDA hydrolysis is a sensitive and nonspecific test to depict the hydrolytic activity of soil 197 microorganisms and has been commonly accepted as a simple measure of total microbial activity (Adam 198 and Duncan, 2001). The large increase in FDA hydrolysis due to combined effect of ZTW and rice residue could be attributed to organic matter enrichment in 0-5 cm soil layer. Earlier studies have showed a 199 200 similar increase in FDA in ZT systems under different climatic conditions (Perez-Brandan et al., 2012; Gajda et al., 2013). The activity of APA is linked to the transformation of organic to inorganic P (Yang et 201 al., 2008). Consistent with our results, several studies have reported higher APA activities under ZT than 202 CT (Tao et al., 2009; Qin et al., 2010). Higher PA in ZT with retaining rice straw may be due to organic 203 204 matter build up after 3 years of continuous ZT than under CT (Yadav and Tarafdar, 2004). The increase 205 in PA under ZTW+R is likely to increase the release of P from phytate present in soil.

Urease (UA) was significantly increased under ZTW+R in 0-5 cm soil layer demonstrating an improvement of soil quality. Mikanova *et al.*, (2009) observed that UA was significantly higher under reduced tillage compared to the CT at soil depth of 0–10 cm. Similarly, Qin *et al.*, (2010) also observed UA was significantly higher in ZT with maize residue as compared to CT without maize residue in 0-10 cm soil layer. An increase in UA is generally accompanied by increase in SOC (Kheyrodin *et al.*, 2012). Activity of UA decreased with depth in all treatments which might be owing to a decrease in SOC.

212 Retention of rice residue in ZT caused significant increase in the contents of OC, Olsen-P, NH₄OAc-K and DTPA-micronutrients compared to CTW-R (Table 3). The increases in OC under ZT+R 213 214 have been observed in a range of soil and climatic conditions (Melero et al., 2009; Yadvinder-Singh et al., 215 2010b). The significant increases in the contents of Olsen-P, NH₄OAc-K and DTPA-micronutrients in 0-5 216 cm soil layers under ZTW+R compared to CTW-R were probably positive effects of rice residue retention 217 and lower rate of soil organic decomposition under ZT conditions (Salinas-Garcia et al., 2002; Yadvinder-218 Singh et al., 2014). In addition, the increase in the availability of P in surface layer under ZTW+R compared to CTW-R could also be due to lower P adsorption by soil and the release of organic P during 219 220 decomposition of rice residues (Qin et al., 2010; Wang et al., 2008). Being a rich source of K, rice straw 221 increased the availability K under ZT+R than the other treatments (Yadvinder Singh et al., 2010a).

In our study, ZTW+R significantly increased the grain yield over ZTW-R and CTW-R. The increase in grain yield of wheat under ZTW+R may be ascribed to the positive effects of soil temperature modifications, soil moisture supply and improved soil fertility due to rice residue mulch (Yadvinder-Singh and Sidhu, 2014). Consistent with our results, earlier studies have also reported higher yields of wheat under ZT+R compared to CT-R (Sidhu *et al.*, 2007). When the entire rice residue was removed (-R), yield of ZTW was significantly lower than CTW. In an earlier study, Arora *et al.*, (2010) reported that poor root growth and lower N use efficiency was responsible for lower wheat yields in ZT-R compared to CT-R.

229 Significant correlations between different enzyme activities observed in our study (Table 4) 230 suggest a general relationship between soil microbiological properties. Retention of crop residues 231 improved the organic carbon status of soils, which in turn was reflected in the higher soil enzymatic activities (Omidi et al., 2008; Gajda et al., 2013). Qin et al., (2010) found that UA and APA activities were 232 233 significantly correlated with available P content in soil. The relationship between all assayed soil enzyme 234 activities and chemical properties indicate that these enzymes play an important role in the cycling of 235 elements and initial phases of the decomposition of crop residues. This implies that continuous addition of 236 diversified carbon sources through rice residues as organic manuring enhanced the microbial biomass 237 carbon and abundance of different microbial communities of soil and perhaps the enzyme activities.

238239 4. CONCLUSION

Our study showed that zero tillage with rice residues as surface mulch (ZTW+R) markedly improved grain yield of wheat over zero tillage without residue (ZTW-R) after three years of RW cropping sequence. ZTW+R was more effective than ZTW/CTW for increasing the soil organic carbon, Olsen-P, NH₄OAc-K and DTPA-extractable micronutrients in soil surface 0-5 cm layer. A significant change in soil enzyme activities occurred after 3 years of ZTW+R. All soil enzymes were positively correlated with each other and with soil chemical properties and grain yield. The increase in enzyme activities in soil may contribute to a long-term sustainability of RWS under semi-arid climate conditions of South Asia. The beneficial
 effect of RCTs on soil quality was mainly confined to the soil surface layer.

249 **REFERENCES**

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- Adam G, Duncan H. Development of a sensitive and rapid method for the measurement of total microbial activity using fluorescein diacetate (FDA) in a range of soils. Soil Biology and Biochemistry. 2001;33:943–951.
- 2. Ames BN. Assay of inorganic phosphate, total phosphate and phosphatases. Methods Enzymology. 1966;8:115–118.
- 3. Arora VK, Sidhu S, Sandhu KS, Thind S S. Effects of tillage intensity, planting time and nitrogen rate on wheat yield following rice. Experimental Agriculture. 2010;46:267-275.
- 4. Bendick AK, Dick RP. Field management effects on soil enzyme activities. Soil Biology and Biochemistry. 1999;31:1471–1479.
- Caravaca F, Roldan A. Effect of Eisenia fetida earthworms on mineralization kinetics, microbial biomass, enzyme activities, respiration and labile C fractions of three soils treated with a composted organic residue. Biology and Fertility of Soils. 2003;38: 45–51.
- Dick RP. Soil enzyme activities as integrative indicators of soil health. In: Pankhurst C, Doube B, Gupta V (eds.) Biological Indicators of Soil Health, CAB International, Wallingford, UK, 1997;pp. 121–156.
- 266
 7. Douglas LA, Bremner JM. Extraction and colorimetric determination of urea in soils. Soil Science
 267 Society of America Journal.1970;34:859–862.
 - 8. Eivazi F, Bayan MR, Schmidt K. Select soil enzyme activities in the historic Sanborn field as affected by long-term cropping systems. Communications in Soil Science and Plant Analysis. 2003;3:2259–2275.
 - Elfstrand S, Bath B, Martersson A. Influence of various forms of green manure amendment on soil microbial community composition, enzyme activity and nutrient levels in leek. Applied Soil Ecology. 2007;36:70–82.
 - 10. Gajda AM, Przewoka B, Gawryjoek K. Changes in soil quality associated with tillage system applied. International Agrophysics. 2013;27:133–141.
 - 11. Garcia C, Hernandez MT, Albaladejo J, Castillo V, Roldan A. Revegetation in semiarid zones: influence of terracing and organic refuse on microbial activity. Soil Science Society of America Journal. 1998;62:670–676.
 - 12. Gathala MK, Kumar V, Sharma PC, Saharawat YS, Jat HS, Singh M, Kumar A, Jat ML, Humphreys E, Sharma DK, Sharma S, Ladha JK. Optimizing intensive cereal-based cropping systems addressing current and future drivers of agricultural change in the Northwestern Indo-Gangetic Plains of India. Agriculture, Ecosystems & Environment. 2013;177:85–97.
 - 13. Gupta RK, Seth A. A review of resource conserving technologies for sustainable management of the rice-wheat cropping system of the Indo-Gangetic plains. Crop Protection. 2007;26:436–447.
 - 14. International Rice Research Institute (IRRI) 2000 IRRISTAT for window (CD-ROM) version 4.02b. Los Banos, Philippines:IRRI.
 - 15. Jackson ML. Soil chemical analysis. Prentice Hall of India, Private Limited, New Delhi. 1973;pp. 38-56.
- 16. Jat ML, Gathala MK, Ladha JK, Saharawat YS, Jat AS, Kumar V, Sharma SK, Gupta R.
 Evaluation of precision land leveling and double zero-till systems in the rice-wheat rotation:
 Water use, productivity, profitability and soil physical properties. Soil and Tillage Research. 2009;
 105:112–121.
 17. Jiang HM, Jiang JP, Jia Y, Li FM, Xu JZ. Soil carbon pool and effects of soil fertility in seeded
 - Jiang HM, Jiang JP, Jia Y, Li FM, Xu JZ. Soil carbon pool and effects of soil fertility in seeded alfalfa fields on the semi-arid Loess Plateau China. Soil Biology and Biochemistry. 2006;38:2350– 2358.
- 18. Kandeler E, Mosier AR, Morgan JA, Milchunas DG, King JA, Rudolph S, Tscherko D. Response
 of soil microbial biomass and enzyme activities to the transient elevation of carbon dioxide in
 semi-arid grassland. Soil Biology and Biochemistry. 2006;38:2448–2460.
- 29919. Kharia Sushil Kumar, Thind HS, Goyal Avinash, Sharma Sandeep and Dhaliwal SS. Yield and300Nutrient Uptakes in Wheat under Conservation Agriculture Based Rice-Wheat Cropping System

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in Punjab, India. International Journal of Current Microbiology and Applied Sciences. 2017;6(2): 302 1698-1708 303

- 20. Khevrodin H. Ghazvinian K. Taherian M. Tillage and manure effect on soil microbial biomass and respiration, and on enzyme activities. African Journal of Biotechnology. 2012;11:14652-14659.
- 21. Klose S, Moore JM, Tabatabai MA. Arylsulfatase activity of microbial biomass in soils as affected by cropping systems. Biology and Fertility of Soils. 1999;29:46-54.
- 22. Lindsay WL, Norwell WA. Development of a DTPA test for zinc, iron, manganese and copper. Soil Science Society of America Journal. 1978;42:421-428.
- 23. Madejon E, Moreno F, Murillo JM, Pelegrin F. Soil biochemical response to long-term conservation tillage under semi-arid Mediterranean conditions. Soil and Tillage Research 2007;94:346-352.
- 24. Melero S, Lopez-Garrido R, Madejon E, Murillo JM, Vanderlinden K, Ordonez R, Moreno F. Longterm effects of conservation tillage on organic fractions in two soils in South-West of Spain. Agriculture, Ecosystems & Environment. 2009;133: 68-74.
- 25. Mikanova O, Javurek M, Simon T, Friedlova M, Vach M. The effect of tillage systems on some microbial characteristics. Soil and Tillage Research. 2009;105:72-76.
- 26. Olsen S R, Cole C V, Waternabe F S, Dean L A. 1954. Estimation of available phosphorous in soil by extraction with sodium bicarbonate. USDA Cir 939:19.
 - 27. Omidi H, Tahmasebi Z, Torabi H, Miransari M. Soil enzymatic activities and available P and Zn as affected by tillage practices, canola (Brassica napus L) cultivars and planting dates. European Journal of Soil Biology. 2008;44:443-450.
 - 28. Perez-Brandan C, Arzeno JL, Huidobro J, Grumberg B, Conforto C, Hilton S, Bending GD, Meriles JM, Vargas-Gil S. Long-term effect of tillage systems on soil microbiological, chemical and physical parameters and the incidence of charcoal rot by Macrophomina phaseolin (Tassi) Goid in soybean. Crop Protection. 2012;40:73-82.
 - 29. Qin S, He X, Hu C, Zhang Y, Dong W. Responses of soil chemical and microbial indicators to conservational tillage versus traditional tillage in the North China Plain. European Journal of Soil Biology. 2010;46:243-247.
 - 30. Roldan A, Salinas-Garcia JR, Alguacil MM, Caravaca F. Soil sustainability indicators following conservation tillage practices under subtropical maize and bean crops. Soil and Tillage Research. 2007;93: 273-282.
 - 31. Salazar S, Sanchez L, Alvarez J, Valverde A, Galindo P, Igual J, Peix A, Santa-Regina I, Correlation among soil enzyme activities under different forest system management practices. Ecological Engineering. 2011;37:1123–1131.
 - 32. Salinas-Garcia JR, Velazquez-Garcia JJ, Gallardo-Valdez M, Diaz-Mederos P, Caballero-Hernandez F. Tapia-Vargas L M. Rosales-Robles E. Tillage effects on microbial biomass and nutrient distribution in soils under rain-fed corn production in central-western Mexico. Soil and Tillage Research. 2002;66:143-152.
 - 33. Sidhu HS, Singh M, Humphreys E, Yadvinder Singh, Bijay Singh, Dhillon SS, Blackwell J, Bector V, Singh M, Singh S. The Happy Seeder enables direct drilling of wheat into rice stubble. Australian Journal of Experimental Agriculture. 2007;47:844-854.
- 34. Sidhu HS, Singh M, Yadvinder Singh, Blackwell J, Lohan SK, Humphreys E, Jat ML, Singh V, Singh S. Development and evaluation of the Turbo Happy Seeder for sowing wheat into heavy rice residues in NW India. Field Crops Research. 2015;184:201-212.
 - 35. Tabatabai M A. 1982. Soil Enzymes. In: A. L. Page, R. H. Miller, D. R. Keeney (eds.) Methods of Soil Analysis. Part 2. Agronomy 9, American Society of Agronomy. Madison, USA, pp. 903–947.
- 36. Tabatabai MA, Bremner JM. Use of p-nitrophenyl phosphate for assay of soil phosphatase activity. Soil Biololgy and Biochemistry. 1969;1:301-307.
 - 37. Tao J, Griffiths B, Zhang S, Chen X, Liu M, Hu F, Li H. Effects of earthworms on soil enzyme activity in an organic residue amended rice-wheat rotation agro-ecosystem. Applied Soil Ecology. 2009;42:221-226.
- 352 38. Walkey A, Black CA. An examination of the Degtjareff method for determination of soil organic 353 matter and a proposed modification of the chromic acid titration method. Soil Science. 354 1934;37:29-39.

- 39. Wang Q, Bai Y, Gao H, He J, Chen H, Chesney RC, Kuhn NJ, Li H. Soil chemical properties and 355 microbial biomass after 16 years of no-tillage farming on the Loess Plateau, China. Geoderma. 356 357 2008:144:502-508. 358
 - 40. Yadav B K, Tarafdar J C. Phytase activity in the rhizosphere of crops, trees and grasses under arid environment. Journal of Arid Environments. 2004;58:285-293.
 - 41. Yadvinder Singh, Gupta R K, Jagmohan S, Gurpreet S, Gobinder S, Ladha J K. Placement effects on rice residue decomposition and nutrient dynamics on two soil types during wheat cropping in rice-wheat system in northwestern India. Nutrient Cycling in Agroecosystems. 2010a;88: 471-480.
 - 42. Yadvinder Singh, Singh M, Sidhu HS, Khanna PK, Kapoor S, Jain AK, Singh AK, Sidhu GK, Avtar S, Chaudhary DP, Minhas PS. 2010b. Options for effective utilization of crop residues. Directorate of Research, Punjab Agricultural University, Ludhiana, India. 32pp.
 - 43. Yadvinder Singh, Kukal SS, Jat ML, Sidhu HS. Improving water productivity of wheat-based cropping systems in South Asia for sustained productivity. Advances in Agronomy. 2014;127: 157-258.
 - 44. Yadvinder Singh, Sidhu HS. Management of cereal crop residues for sustainable ricewheat production system in the Indo-Gangetic plains of India. Proceedings of the Indian National Science Academy. 2014;80: 95-114.
 - 45. Yang LJ, Li TL, Li FS, Lemcoff J H, Cohen S, Fertilization regulates soil enzyme activity and fertility dynamics in a cucumber field. Scientia Horticulturae. 2008;116:21-26.
 - 46. Yang X, Chen C, Luo Q, Li L, Yu Q. Climate change effects on wheat yield and water use in oasis cropland. International Journal of Plant Production. 2011;5:83-94.
 - 47. Yuan B, Yue D. Soil microbial and enzymatic activities across a chronosequence of chinese pine plantation development on the loess plateau of China. Pedosphere. 2012;22:1-12.
 - 48. Zhao B, Chen J, Zhang J, Qin S. Soil microbial biomass and activity response to repeated drying-rewetting cycles along a soil fertility gradient modified by long-term fertilization management practices. Geoderma. 2010:160:218-222.
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Table 1. Description of experimental treatments (Kharia et al 2017)

Rice (main plots)	Wheat (sub-plots)
T ₁ . Zero till direct seeded rice (ZT-DSR). Residues of previous wheat crop were removed. DSR was sown using same drill as in treatment. T ₂ . Conventional direct seeded rice (CT-DSR). Residues of previous wheat crop were removed. Tillage for DSR included two passes of harrows and two passes of Tyne plough ^{\vee} followed by planking, The DSR was sown using seed cum fertilizer drill in 20 cm apart.	1. Conventional till wheat after removal of rice residue (CTW-R). Residues of previous rice removed. Tillage operations for CTW included two passes of harrows and two passes of Tyne plough ^Y followed by planking.
T_3 . ZT transplanted rice using mechanical trans planter (ZT-DTR). Residues of previous wheat crop were removed.	2. Zero till wheat after removal of rice residue (ZTW-R). Wheat was direct seeded in the no till plots using zero till drill.
T ₄ .Conventional till puddled transplanted rice (PTR): Residues of preceding wheat crop removed. Puddling (wet tillage) was done twice in 6-8 cm of standing water using a tractor-mounted puddler followed by planking.	3. Zero till wheat + residues (ZTW+R) Residues of previous rice crop retained. Wheat was directly seeded into residues using Turbo Happy Seeder (Sidhu <i>et al.</i> , 2015).*
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Soil properties	Method used	Reference
Organic carbon	Wet digestion method	Walkley and Black (1934)
Olsen P	0.5 M NaHCO ₃ -extractable	Ölsen <i>et al</i> ., (1954)
Available K	1M NH₄OAc-extractable	Jackson (1973)
DTPA- Fe, Mn, Cu and Zn	DTPA extractable	Lindsay and Norwell (1971)
Dehydrogenase activity (DHA)	Triphenylformazan (TPF) produced by the reduction of 2, 3, 5-triphenyltetrazolium chloride (TTC).	Tabatabai (1982)
Fluorescein di-acetate activity (FDA)	Fluorescein released by the hydrolysis of fluorescein di-acetate.	Adam and Duncan (2001)
Alkaline phosphatase activity (APA)	p-nitrophenyl method.	Tabatabai and Bremner (1969)
Phytase activity (PA)	Inorganic phosphate released by hydrolysis using sodium phytate	Àmes [´] (1966)
Urease activity (UA)	2M KCI- Phenyl mercuric acetate	Douglas and Bremner (1970)

388 Table 2. Methods used for analysis of soil samples for different soil parameters

Table 3. Effect of rice establishment, tillage and rice straw management on dehydrogenase (μ g TPF g⁻¹ 24 hr⁻¹) activities at various soil layers. 393

Treatments		Soil layers (cm)	
	0-5	5-10	10-15
Rice establishment s	ystems		
DSR-ZT-DSR	303	177	77.4
DSR-CT-DSR	275	168	76.2
ZT-DTR	290	163	74.7
PTR	286	159	72.4
LSD(0.05)	NS	NS	NS
Tillage and rice straw	management practi	ces	
CTW-R	265	158	73.4
ZTW-R	291	167	73.8
ZTW+R	309	175	78.4
LSD (0.05)	25.8	NS	NS

Table 4. Effect of rice establishment, tillage and rice straw management on flourescein diacetate (μ g fluorescein g⁻¹ dry soil) activities at various soil layers

Treatments		Soil layers (cm)	
	0-5	5-10	10-15
Rice establishment	systems		
DSR-ZT-DSR	3.32	2.68	2.45
DSR-CT-DSR	3.24	2.74	2.45
ZT-DTR	3.16	2.73	2.50
PTR	3.12	2.74	2.42
LSD(0.05)	NS	NS	NS
Tillage and rice stra	w management praction	ces	
CTW-R	3.02	2.69	2.42
ZTW-R	3.15	2.72	2.41
ZTW+R	3.47	2.76	2.52
LSD (0.05)	0.120	NS	NS

Table 5. Effect of rice establishment, tillage and rice straw management on alkaline phosphatase (μ g p-nitrophenol g⁻¹ dry soil hr⁻¹) activities at various soil layers

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Treatments		Soil layers (cm)	
	0-5	5-10	10-15
Rice establishment	systems		
DSR-ZT-DSR	49.4	42.2	33.9
DSR-CT-DSR	42.5	39.0	34.3
ZT-DTR	43.8	39.1	30.5
PTR	41.4	34.7	28.7
LSD(0.05)	NS	NS	NS
Tillage and rice stra	w management practi	ces	
CTW-R	41.4	37.6	31.2
ZTW-R	43.6	37.6	31.7
ZTW+R	47.8	41.0	32.7
LSD (0.05)	3.72	NS	NS

Table 6. Effect of rice establishment, tillage and rice straw management on Phytase (µg inorganic

P released g⁻¹ dry soil hr⁻¹) activities at various soil layers

Treatments		Soil layers (cm)	
	0-5	5-10	10-15
Rice establishment	systems		
DSR-ZT-DSR	0.341	0.273	0.238
DSR-CT-DSR	0.340	0.265	0.234
ZT-DTR	0.328	0.268	0.233
PTR	0.340	0.262	0.237
LSD(0.05)	NS	NS	NS
Tillage and rice stray	w management practi	ces	
CTW-R	0.303	0.254	0.232
ZTW-R	0.310	0.269	0.232
ZTW+R	0.398	0.277	0.241
LSD (0.05)	0.031	NS	NS

Table 7. Effect of rice establishment, tillage and rice straw management on urease (μ g urea g⁻¹ dry soil min⁻¹) activities at various soil layers 429 430

Treatments		Soil layers (cm)	
	0-5	5-10	10-15
Rice establishment	systems		
DSR-ZT-DSR	2.60	2.13	1.68
DSR-CT-DSR	2.55	2.14	1.70
ZT-DTR	2.58	2.17	1.60
PTR	2.51	2.18	1.66
LSD(0.05)	NS	NS	NS
Tillage and rice stra	w management praction	ces	
CTW-R	2.40	2.12	1.63
ZTW-R	2.54	2.16	1.64
ZTW+R	2.75	2.20	1.72
LSD (0.05)	0.070	NS	NS

Table 8. Effect of rice establishment, tillage and rice straw management on OC (g kg⁻¹) content at

various soil layers

Treatments		Soil layers (cm)	
	0-5	5-10	10-15
Rice establishment s	systems		
DSR-ZT-DSR	5.88	4.65	3.37
DSR-CT-DSR	5.49	4.93	3.17
ZT-DTR	5.90	4.65	3.43
PTR	5.82	5.03	3.58
LSD(0.05)	NS	NS	NS
Tillage and rice strav	v management praction	ces	
CTW-R	5.18	4.56	3.32
ZTW-R	5.79	4.86	3.43
ZTW+R	6.35	5.02	3.41
LSD (0.05)	0.30	0.36	NS

Treatments	-	Soil layers (cm)	
	0-5	5-10	10-15
Rice establishment	systems		
DSR-ZT-DSR	9.98	8.35	6.24
DSR-CT-DSR	9.69	7.14	4.72
ZT-DTR	10.1	8.32	5.66
PTR	9.13	7.02	5.72
LSD(0.05)	NS	NS	NS
Tillage and rice stra	w management practi	ces	
CTW-R	8.90	6.82	5.23
ZTW-R	9.64	7.81	6.03
ZTW+R	10.7	8.49	5.50
LSD (0.05)	0.799	0.687	NS

Table 9. Effect of rice establishment, tillage and rice straw management on Olsen P (kg ha⁻¹)
 content at various soil layers

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Table 10. Effect of rice establishment, tillage and rice straw management on available K (kg ha⁻¹)

442 content at various soil layers

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Treatments		Soil layers (cm)	
	0-5	5-10	10-15
Rice establishment	systems		
DSR-ZT-DSR	44.8	38.1	34.8
DSR-CT-DSR	45.4	37.3	37.8
ZT-DTR	47.0	39.1	38.3
PTR	42.7	34.4	36.9
LSD(0.05)	NS	NS	NS
Tillage and rice stray	w management praction	ces	
CTW-R	42.6	35.1	36.1
ZTW-R	40.7	35.3	34.9
ZTW+R	51.6	41.3	39.8
LSD (0.05)	6.34	4.01	NS

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Table 11. Effect of rice establishment, tillage and rice straw management on DTPA micronutrients

in 0-5 cm soil layer after three cycles of rice-wheat system.

Treatments	I	DTPA- extractable micronutrients (mg kg ⁻¹)			
	Zn	Mn	Fe	Cu	
Rice establishment systems					
ZT-DSR	5.17	8.32	30.0	1.34	
CT-DSR	4.95	8.20	28.4	1.29	
ZT-DTR	4.87	8.12	29.9	1.23	
PTR	4.77	8.30	29.4	1.38	
LSD (p=0.05)	NS	NS	NS	NS	
Tillage and rice straw management	t practices				
CTW-R	4.85	8.19	28.0	1.25	
ZTW-R	4.85	8.04	28.5	1.26	
ZTW+R	5.14	8.47	31.8	1.43	
LSD (p=0.05)	0.24	0.30	2.40	0.09	

Table 12. Yield and yield attributing parameters of wheat as influenced by rice establishment systems, tillage and rice straw management practices under rice-wheat system (Kharia et al 2017).

Treatment	Grain yield (Mg ha ⁻¹)	Spike length (cm)	Grains spike ⁻¹	Grain weight spike ⁻¹ (gram)	1000 grain weight (gram)
Rice establish	nment systems				
ZT-DSR	4.84	10.5	53.5	2.08	38.0
CT-DSR	4.84	10.8	52.7	1.97	38.0
ZT-DTR	4.88	10.7	53.7	2.04	35.7
PTR	4.77	11.0	52.9	2.01	37.8
LSD (0.05)	NS	NS	NS	NS	NS
Tillage and st	raw manageme	nt practices			
CTW-R	4.77	10.5	53.1	2.01	36.8
ZTW-R	4.47	10.9	52.1	1.90	35.8
ZTW+R	5.26	10.8	54.3	2.16	39.6
LSD (0.05)	0.19	0.30	1.10	0.08	2.20

456	Table 13. Correlation between soil enzyme activities, grain yield of wheat, pH, organic carbon and Olsen-P in 0-5 cm soil layer after
457	three cycles of rice-wheat cropping system

Properties	DHA	FDA	ΑΡΑ	ΡΑ	UA	GY	рН	ОС	Olsen-P
0-5 cm layer									
DHA	1	0.826**	0.843**	0.742**	0.896**	0.488	-0.198	0.674*	0.739**
FDA		1	0.787**	0.886**	0.910**	0.690**	-0.232	0.608*	0.824**
APA			1	0.624*	0.775**	0.500	-0.006	0.497	0.652*
PA				1	0.892**	0.857**	-0.283	0.694*	0.759**
UA					1	0.710**	-0.381	0.717**	0.831**

DHA-Dehydrogenase activity, FDA-Fluorescein di-acetate, APA-Alkaline phosphatase activity, PA-Phytase activity, UA-Urease activity, GY-Grain yield, OC-Organic carbon.

** Significant at p<0.01, * Significant at p<0.05

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Table 14. Correlation between soil enzyme activities and DTPA-extractable micronutrients (mg kg⁻¹) in 0-5 cm soil layer 465 466 467

Parameter	Fe	Mn	Cu	Zn
Dehydrogenase (DHA)	0.775**	0.453	0.650*	0.627*
Fluorescein di-acetate (FDA)	0.760**	0.624*	0.717**	0.782**
Alkaline phosphatase activity (APA)	0.685*	0.428	0.519	0.771**
Phytase activity (PA)	0.820**	0.747**	0.847**	0.643*
Urease (UA)	0.857**	0.505	0.681*	0.657*

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** Significant at p<0.01, * Significant at p<0.05