- 1 2
- 3

4

# **Phosphorus Influence on Plant Tissue Nitrogen Contents and Yield Attributes of Finger Millet varieties** in Semi-Arid Region of Kenya

5 6

#### 7 ABSTRACT

8

The experiment was conducted with the aim of evaluating the effect of differential levels of P on finger millet accumulation of N and yield components. On-station experiments were conducted at the KALRO-Kiboko crops research station in Makueni County during the short and long rains of 2014 and 2015 respectively. There were 4 levels of P (0, 12.5, 25 and 37.5 kg ha<sup>-1</sup>  $P_2O_5$ ) and three varieties (U-15, P-224 and Ekalakala). Ekalakala is the local check while 0 kg/ha  $P_2O_5$  was the control. The trial was laid out in a randomized complete block design and fitted in factorial arrangement with three replicates given a total of 36 plots. Soil sampling was at a depth of 0-30 cm on all the plots and analytical results showed moderately available P but very low N, organic carbon and zinc. Significant differences (P=.05) were observed between the phosphate levels on the nitrogen contents in plant parts with the control showing the lowest N accumulation of 4.95% and 4.90% for the short and long rains respectively while the 25 kg  $ha^{-1} P_2 O_5$  rate had the highest with 5.66% in the short rains and 5.14% in the long rains. The stem contained the highest nitrogen content while the roots had the lowest. Phosphate rates did not have significant influence on the yield components except the finger width while the varieties varied significantly (P=.05) on the productive tillers, panicle number, grains per spikelet and the harvest index. Variety U-15 had the highest yield for both seasons with a maximum of 3410 kg ha<sup>-1</sup> for the short rains. The newly released variety (U-15) responded well to moderate P supplemented at 25 kg ha<sup>-1</sup> P<sub>2</sub>O<sub>5</sub> thus can efficiently utilize N in soils with low N like in Makueni and is highly recommended. The optimal P for the yield and N accumulation was 25 kg ha<sup>-1</sup>  $P_2O_5$  and beyond this point the P would not be translated to profitable yield but a loss to the farmer in the short run.

- 9
- 10

Keywords: Finger millet; available P; N accumulation; grains per spikelet.

11

#### **1. INTRODUCTION** 12

13 Finger millet (Eleusine coracana) is the most important small millet grown in eastern and southern Africa 14 and it serves as a subsistence and food security crop that is especially important for its nutrition and 15 resilience to harsh weather conditions. Of all major cereals, this crop is one of the most nutritious [1] 16 especially in calcium. It is a food crop in traditionally low input cereal-based farming systems in Africa, 17 and is of particular importance in upland areas of Eastern Africa. While developing countries in Asia still produce the majority of the world's millets, Africa is becoming the hub of production [2]. In Kenya, the 18 crop is mainly produced in the part of the country west of the Rift Valley and it is cultivated on around 65 19 20 000 ha yr<sup>-1</sup> [1]. However, the yields of finger millet on farmers' fields are generally low, just about 15-16 % 21 of their potential maximum in Kenya [3]. Soil infertility is one of the major constraints to finger millet 22 production throughout much of the Sub-Saharan Africa.

23 An understanding of the internal and external P efficiencies of modern finger millet varieties is very 24 important in selection of varieties adaptable to P deficient and moderate P conditions. Phosphorus deficiency has been identified as one of the most limiting soil nutrient after nitrogen due to soil erosion, 25 26 continuous cultivation and fast reversion of soluble P where P fertilization is of fundamental importance in 27 replenishing, enhancing and maintaining soil fertility [4]. Achievement of higher efficacy and efficiency of 28 P mineral fertilization is possible through searching for and improving the methods of assessment of plant

nutritional status as well as aiming at optimization of fertilizer use. Knowledge on factual needs of plants 29

concerning balancing of mineral nutrients and utilization efficiency is an important aspect of reducing
 agricultural negative effects on environment through improper P fertilization.

32 Nutrient limitation of ecosystems is typically determined by fertilization experiments, with increased 33 biomass or growth rates taken as evidence of limitation [5, 6]. A less direct index of nutrient limitation is 34 foliar nutrient concentration, which is predicted to increase in response to addition of the limiting nutrient, 35 although the positive relationship between biomass and foliar nutrients is not necessarily a linear one. This index is reasonable given that foliar nutrient concentrations (expressed either as N concentration, P 36 37 concentration, or a ratio of N-to-P) reflect soil nutrient concentrations [7, 8]). Foliar N and P 38 concentrations also relate to the functioning of plants, as comparisons across agro-ecologies have shown that they are correlated with physiological traits such as photosynthesis and dark respiration, and leaf 39 40 properties that affect resource capture such as specific leaf area and leaf lifespan [9].

Finger millet has high genetic diversity [10]. All finger millet varieties do not respond to nutrients in the 41 same manner. Genotypic variability among different finger millet cultivars has been reported for 42 responsiveness to N and P [11]. Gupta et al. [12] evaluated the N use efficiency (ratio of grain yield to N 43 44 supply) and N utilization efficiency (ratio of grain yield to total N uptake) of three finger millet genotypes 45 under different N inputs and found that there was genotypic variability among the finger millet genotypes' 46 responses to different N inputs, wherein some varieties were highly responsive to N. Therefore, the 47 understanding of the existence in genetic variability in finger millet genotypic response to nutrients 48 prompted the need to study yield and N accumulation responses to P among different varieties in 49 marginal Kiboko, of lower area in Eastern Kenya.

# 50 2. MATERIALS AND METHODS

51

# 52 **2.1.** Site description

53 The experiment was conducted during the 2014 short and 2015 long rains season at the International Crops Research Institute for the Semi-Arid Tropics (ICRISAT) and Kenya Agricultural and Livestock 54 Research Organization (KALRO)-Kiboko Crops Research Station where the previous crop planted was 55 56 pigeon pea. The station is in Kiboko Location, Makueni County. It lies within longitudes 3738" East, latitudes 2°16" South and at an elevation of 975 m above sea level. The soils are classified as Acri-57 58 Rhodic Ferrassols [13] composed of sand (43.08%), silt (33.91%) and clay (23.01%) with an average pH 59 unit of 8.9. Mean available P was found to be moderate while nitrogen, organic carbon and zinc were 60 extremely low as shown in Table 1. The station received a total of 441 and 286.3 mm rainfall during the short and long rains seasons respectively. The mean maximum and minimum temperature for the short 61 62 and long rains were 31.8°C and 17.9°C, and 29.7°C a nd 15.7°C respectively with mean precipitation of 63 83.3% and 85% potential evaporation during the short and long rains respectively.

# 64 **Table 1: Physiochemical properties of experimental site in Kiboko**

Physical properties					
Sand %	43.08				
Silt %	33.91				
Clay %	23.01				
Chemical properties	Short Rains	Long Rains			
Soil pH	8.55	9.25			
Available N %	0.03	0.03			
Organic Carbon %	0.24	0.26			
Extractable P ppm	13.25	15.5			
Potassium me%	0.49	1.53			
Calcium me%	4.37	10.7			
Magnesium me%	2.95	3.26			
Manganese me%	0.26	0.14			
Copper ppm	2.38	2.51			
Iron ppm	25.2	44.2			

Zinc ppm	1.45	2.86
Sodium me%	0.47	1.05
Electrical Conductivity mS/cm	0.47	0.27

### 65 **2.2. Experimental design, treatments and data collection**

66 The experiment was laid out in a Randomized Complete Block Design in a factorial arrangement and replicated three times. That is, 4 P rates (0, 12.5, 25 and 37.5 kg ha<sup>-1</sup> P<sub>2</sub>O<sub>5</sub>) x 3 varieties (U-15, P-224 67 68 and Ekalakala) × 3 replicates (total of 36 treatments). Where Ekalakala was the local check and 0 kg ha 69 P<sub>2</sub>O<sub>5</sub> was the absolute control. Phosphate fertilizer was measured according to the treatment and divided per row and hand applied as whole during planting as triple superphosphate. Seeds were drilled in the 70 rows and thinned four weeks after planting to a spacing of 10 cm between plants and 50 cm between 71 72 rows giving a total of 205 plants per plot. The experimental units measured 4 m long and 2 m wide with a 73 2 m pathway which also separated the plots. Recommended cultural practices were applied throughout 74 the crop growth as per crop demands. The crop was harvested at physiological maturity when 90% 75 browning of the heads was observed. The three middle rows were harvested on a net area of 3m<sup>2</sup>. Data 76 was collected on the number of productive tillers, finger width, finger length, grains per spikelet, number 77 of panicles harvested and the grain yield according to the IBPGR [14]. The harvest index was calculated 78 as ratio of the grain to biological yield.

# 79 **2.3.** Soil samples and plant tissue analysis

80 Soil samples (500 g) were collected from every experimental plot at a depth of 0-30 cm using a soil auger then air-dried in a well-ventilated room for 3 days. The samples were then ground and passed through a 81 2-mm sieve to obtain 50 g uniform samples from each plot for analysis and storage. The soil pH was 82 determined electrometrically in water as outlined by Okalebo et al. [15]. Plant samples from five plants 83 84 from each experimental unit were collected and separated into roots, stem, leaves and grain from five 85 plants collected from each experimental unit. The samples were weighed and oven-dried to a constant 86 weight at 70°C for 48 hours. The dried plant material was ground using a Crompton Willey mill and passed through a 2-mm sieve to a uniform mass of 8 g for analysis. Soil and plant tissue nitrogen was 87 88 determined the by Kjeldahl method while organic carbon content was determined using modified Walkley and Black wet oxidation [16]. The analysis of N and organic carbon were done as described by Ryan et 89 90 al. [16]. Extractable potassium was determined by flame photometer [8] whereby a neutral salt solution 91 replaced cations present on the soil exchange complex. Extractable soil nutrients (Ca, Mg, Mn, Cu, Fe, 92 Zn, and Na) were determined by the Diethylenetriamine Pentaacetic Acid (DTPA) method then measured 93 with an AAS [17]. Determination of P involved digestion of soil sample with a strong acid and the dissolution of all insoluble inorganic minerals and organic P forms which followed the procedure 94 95 described by Olsen and Sommers [18]. For the electrical conductivity, a 5 g sample of soil was placed in 96 a 100 ml disposable plastic cup; 50 ml of deionized water was added. The slurry was shaken on a 97 reciprocating shaker for 45 minutes, and then filtered. Electrical conductivity of the filtrate was then read 98 with a conductivity bridge.

### 99 **2.4. Data analysis**

Data collected was compiled, cleaned and tabulated for statistical analyses. Analysis of variance
 (ANOVA) was performed using GenStat statistical software version 15.1. Where a significant F-test was
 observed, the means were separated using Fischer's protected LSD test at 5% probability level.

103

### 104 **3. RESULTS AND DISCUSSION**

### 105 **3.1. Nitrogen accumulation**

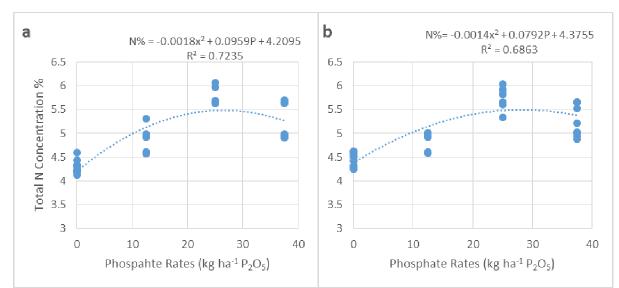
106 Phosphorus application had significant (P=.05) influence on the nitrogen contents of finger millet. The 25 107 kg ha<sup>-1</sup> P<sub>2</sub>O<sub>5</sub> rate showed the highest total accumulation of nitrogen of 5.66 % during the short rains and 108 5.49 % during the long rains. The control had the lowest nitrogen content in the plant for both seasons 109 (Table 2). The stem contained the highest nitrogen where all the treatments with phosphorus had higher N contents compared to the control during the short rains season. There was no clear trend in 110 accumulation of N in stems in response to P treatments. The 25 kg ha<sup>-1</sup> P<sub>2</sub>O<sub>5</sub> rate had the highest 111 112 nitrogen accumulation in the leaves and grains of the crop for both seasons with the control having the 113 lowest except during the short rains on the leaves where the highest rate had the least. Nitrogen contents 114 were enhanced due to the added P which resulted in profuse root development and shoot growth that in 115 turn activated greater absorption of nitrogen from the soil. These results are in conformity with earlier 116 reports [19] in cluster beans. It has been reported that fragile lands such as those in Makueni usually support cropping systems with lower nitrogen contents that also use water less efficiently leading to poor 117 118 crop yields. It has been reported that as phosphorus rates increase towards the optimum, crop productivity increases but at a decreasing rate, the nitrogen contents typically declines [20]. On the other 119 120 hand, the roots showed the lowest nitrogen accumulation with the control having the highest during the 121 long rains while no significant differences at P=.05 were observed between the P treatments in the short 122 rains. This low accumulation of N in the roots might be due to the switching mechanism of the crop to partition N to the reproductive parts at the expense of vegetative ones especially under low soil N 123 124 conditions. Varietal differences were observed on the nitrogen accumulation in both seasons where U-15 125 had the highest in all the plant parts.

Table 2. Effect of phosphate rates and varieties on N % concentrations in finger millet plant parts
 in Kiboko

P <sub>2</sub> O <sub>5</sub>		S	hort Rai	ns		Long Rains				
Rates	Root	Stem	Leaf	Grain	Total	Root	Stem	Leaf	Grain	Total
<mark>(kg ha⁻¹</mark> )	N%	N%	N%	%	N %	N%	N%	N%	%	N %
Control	0.71 <sup>a</sup>	1.53 <sup>b</sup>	1.41 <sup>b</sup>	1.30 <sup>d</sup>	4.95 <sup>b</sup>	1.17 <sup>a</sup>	1.28 <sup>c</sup>	1.17 <sup>c</sup>	1.17 <sup>c</sup>	4.79 <sup>c</sup>
12.5	0.71 <sup>a</sup>	1.65 <sup>a</sup>	1.41 <sup>b</sup>	1.41 <sup>°</sup>	5.18 <sup>b</sup>	1.05 <sup>b</sup>	1.17 <sup>d</sup>	1.40 <sup>a</sup>	1.28 <sup>b</sup>	4.90 <sup>b</sup>
<mark>25.0</mark>	0.72 <sup>a</sup>	1.65 <sup>a</sup>	1.65 <sup>a</sup>	1.64 <sup>a</sup>	5.66 <sup>a</sup>	1.05 <sup>b</sup>	1.52 <sup>a</sup>	1.40 <sup>a</sup>	1.52 <sup>a</sup>	5.49 <sup>a</sup>
37.5	0.71 <sup>a</sup>	1.65 <sup>a</sup>	1.29 <sup>c</sup>	1.50 <sup>b</sup>	5.15 <sup>b</sup>	1.05 <sup>b</sup>	1.40 <sup>b</sup>	1.28 <sup>b</sup>	1.28 <sup>b</sup>	5.01 <sup>b</sup>
LSD	0.03	0.02	0.01	0.01	0.36	0.04	0.07	0.08	0.08	0.07
Variety										
U-15	0.72 <sup>a</sup>	1.68 <sup>a</sup>	1.77 <sup>a</sup>	1.58 <sup>a</sup>	5.75 <sup>a</sup>	1.14 <sup>a</sup>	<mark>1.40<sup>a</sup></mark>	<mark>1.50<sup>a</sup></mark>	<mark>1.50<sup>a</sup></mark>	<mark>4.40<sup>a</sup></mark>
P-224	0.72 <sup>a</sup>	1.59 <sup>b</sup>	1.33 <sup>b</sup>	1.32 <sup>°</sup>	4.96 <sup>c</sup>	1.14 <sup>a</sup>	<mark>1.30<sup>b</sup></mark>	<mark>1.20<sup>b</sup></mark>	<mark>1.40<sup>b</sup></mark>	<mark>3.90<sup>b</sup></mark>
Ekalakala	0.7 <sup>b</sup>	1.59 <sup>b</sup>	1.24 <sup>c</sup>	1.50 <sup>b</sup>	5.03 <sup>b</sup>	0.96 <sup>b</sup>	<mark>1.30<sup>b</sup></mark>	<mark>1.20<sup>b</sup></mark>	1.00 <sup>c</sup>	<mark>3.50<sup>°</sup></mark>
LSD	0.01	0.05	0.08	0.06	0.30	0.04	0.06	0.07	0.07	0.27
CV%	1.60	<mark>1.00</mark>	<mark>1.00</mark>	<mark>21.10</mark>	5.40	<mark>4.10</mark>	<mark>5.40</mark>	<mark>6.60</mark>	<mark>6.20</mark>	<mark>5.70</mark>

128 Values followed by same letters within the column are not significantly different

129 These genotypic variations could be due to the efficiency in acquisition of P and N from the rhizosphere, nitrogen use efficiency and phosphorus use efficiency as well as the internal mobilization of N in all parts 130 of plant. Such variations could be explained mainly by the diversity of the finger millet genotypes which is 131 highly influenced by the environmental conditions [21]. Also, organic compounds secreted by the different 132 133 varietal roots stimulated microbial activity in the rhizosphere, which might also influenced the P availability 134 [22] which is related to N uptake since it is the source of ATP. Phosphorus and Nitrogen acquisition plays 135 an important role for crop adaptation to low P and N soils [22] and a higher internal P and N use efficiency 136 could help to limit soil nutrient mining [23], especially in low-input farming conditions and this has been 137 exhibited by varieties like U-15.



138 139

Fig. 1: The influence of phosphate rates on the total nitrogen concentration in finger millet plant 140 for the short rains (a) and long rains (b).

141 A quadratic trendline (Fig. 1) revealed that the yield reached the peak (optimal) point with 25 kg ha<sup>-1</sup>  $P_2O_5$ . Beyond this point, there is likelihood of luxury consumption, which may have economic implication to a 142 farmer. Accumulation of nitrogen by the plants was not linear, but rather polynomial and after about 25 kg 143 of P per hectare for both seasons (Fig. 1) the nitrogen contents started to descent. Previous work by 144 Mohidin et al. [24] reported peak yield and accumulation on nutrients at 90 mg L<sup>-1</sup> as opposed to 120 mg 145 L<sup>1</sup> with vield reducing at 120 mL<sup>1</sup>. Depending the type of soil, P beyond optimal level can have a 146 negative interaction with micronutrients such as Mn and Zn as previously reported [25]. Other findings 147 [26] working with different onion varieties and different rates of N showed concurring results to current 148 work. The results of the authors showed  $R^2$  values of between 0.82-0.91 amongst varieties. 149 150 Understanding optimal nutrient requirements is important particularly in regards to maintaining safe environment. 151

152 The control showed the lowest N because phosphorus deficiency restricts activity of meristematic sink of

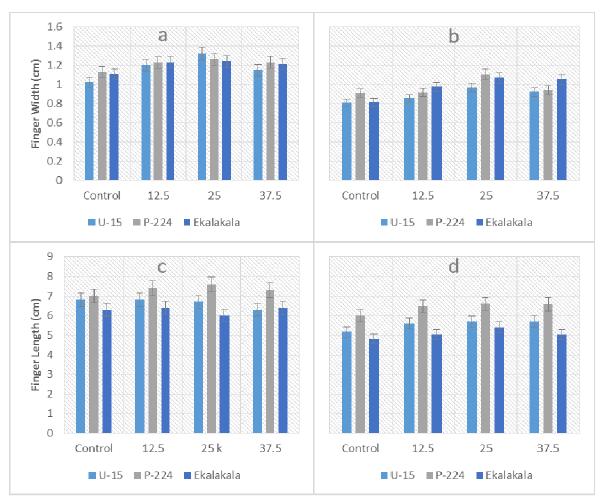
153 plants and leads to a demand for assimilates in growth that are responsible for reduction of source activity

154 and partitioning of photo-assimilates [27].

#### 155 3.2. Yield attributes

Phosphate rates showed significant differences (P=.05) on the finger widths with the 25 kg ha<sup>-1</sup> P<sub>2</sub>O<sub>5</sub> rate 156 157 having the widest spikes for both seasons as shown in Fig. 2. The finger length, harvest indices, threshability, number of harvested panicles and productive tillers were not responsive to applied P and 158 were similar to the control. This was probably due to the availability of P above the critical value of 10 159 160 ppm in the study soil and therefore the additional P through phosphate fertilizer did not lead to significant differences. 161

162



163

Fig. 2. Influence of phosphate rates on the finger width during the short (a) and long (b) rains season and finger length during the short (c) and long (d) rains seasons

166 Previous work based on multi location field experiments conducted in Eastern Uganda, Tenvwa et al. [28] found that application of P fertilizer (20–40 kg  $P_2O_5$  ha<sup>-1</sup>) increased the growth and yield of finger millet 167 168 compared to the no fertilizer control under row planting conditions. However, Hedge and Gowda [29] 169 reported a reduction in finger millet grain yields from 16.3 to 14.7 kg ha<sup>-1</sup>  $P_2O_5$  when the P application 170 rate was increased from 30 to 60 kg ha<sup>-1</sup>  $P_2O_5$ . This could be due to negative interactions with 171 micronutrients when applied beyond certain level (depending on soil characteristics). Similar to inorganic 172 N, this result suggests that application of excess P does not improve yield, but rather that application of 173 balanced fertilizer is crucial.

The pattern in yields and yield components in response to P supply is in line with findings reported by Sankar [30] in semi-arid Alfisols. There could be other limiting factors that could limit P uptake and hence these yield components. Various other scientists [31, 32] also reported similar trends which are in agreement with our findings. They concluded that phosphorous had a significant impact on yield attributes of various cereals crops and the yield reduction after peak value a paradox in regards to some yield enhancing factors.

Notably, all the above traits were significantly influenced by varieties for both seasons as shown on Table 3. U-15 had the highest harvest index for both seasons but with the lowest number of grains per spikelet. Significant genotypic variation for traits related to P acquisition and P use efficiency has been observed in various crops. Cereal improvement in the recent decade has been mainly attributed to the increased harvest index. So far this trait has not been fully exploited in finger millet. Among the various factors influencing the harvest index, mineral nutrition is of utmost significance [33]. Reddy [34] on the effect of P on stability of harvest index with two contrasting genotypes of finger millet contradicted this finding, he found that as P level increased from 20 to 80 kg ha<sup>-1</sup> there was a significant increase in biomass and grain yield but the harvest index and partitioning percentage decreased significantly with increased P levels. This contradiction was probably due to the difference in the available phosphorus values in the study soil and because of the rate where the highest in the study was more than double to that used in the current study.

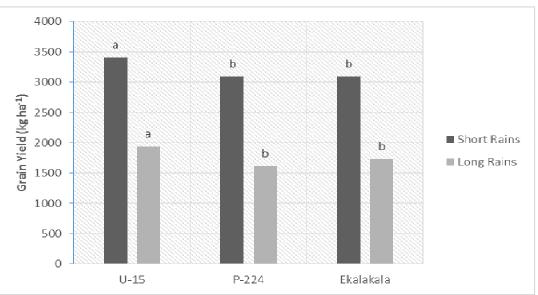
### 192 Table 3. The effect of varieties on the harvest index, threshability, grains per spikelet, panicles and 193 productive tillers during the long and short rains season

				Grains/					Productive tillers (3 m <sup>2</sup> )	
	Harvest Index		Threshing %		Spikelet		Panicles (3 m <sup>2</sup> )			
	Short	Long	Short	Long	Short	Long	Short	Long	Short	Long
Variety	rains	rains	rains	rains	rains	rains	rains	rains	rains	rains
U-15	0.36 <sup>a</sup>	0.41 <sup>a</sup>	<mark>65.90<sup>a</sup></mark>	<mark>81.70<sup>a</sup></mark>	<mark>6.20<sup>b</sup></mark>	<mark>6.10<sup>b</sup></mark>	<mark>270.20<sup>a</sup></mark>	<mark>141.00<sup>ab</sup></mark>	<mark>125.00<sup>a</sup></mark>	<mark>38.00<sup>a</sup></mark>
P-224	0.31 <sup>c</sup>	0.35 <sup>c</sup>	<mark>63.00<sup>b</sup></mark>	<mark>72.60<sup>b</sup></mark>	<mark>7.20<sup>a</sup></mark>	<mark>6.80<sup>a</sup></mark>	<mark>238.90<sup>b</sup></mark>	<mark>123.00<sup>b</sup></mark>	107.00 <sup>b</sup>	<mark>19.00<sup>b</sup></mark>
Ekalakala	0.33 <sup>b</sup>	0.38 <sup>b</sup>	<mark>66.50<sup>a</sup></mark>	<mark>79.30<sup>a</sup></mark>	<mark>7.00<sup>a</sup></mark>	<mark>6.80<sup>a</sup></mark>	<mark>246.40<sup>ab</sup></mark>	<mark>149.00<sup>a</sup></mark>	<mark>130.00<sup>a</sup></mark>	<mark>37.00<sup>a</sup></mark>
L.S.D	0.02	0.02	2.09	3.11	0.45	0.58	25.37	<mark>19.70</mark>	13.09	6.60
CV (%)	<mark>11.50</mark>	<mark>7.80</mark>	<mark>7.40</mark>	<mark>4.70</mark>	<mark>7.90</mark>	<mark>10.20</mark>	<mark>11.90</mark>	<mark>25.50</mark>	<mark>19.80</mark>	<mark>7.40</mark>

194 Values followed by same letters within the column are not significantly different

U-15 and Ekalakala had the greatest threshing ability with the highest of 81.7% observed during the long 195 196 rains season on U-15. The highest number of grains per spikelet was shown on P-224 and Ekalakala. U-197 15 and Ekalakala had the highest number of panicles harvested and number of productive tillers on the 198 net plot. The productive tillers accounted for almost 50% of the harvested panicles on U-15 and Ekalakala 199 which directly impacted on the final grain yield. The superiority of U-15 on most of the yield components 200 translated to the highest yield among the varieties for both seasons as shown on Fig. 3. The short rains had a higher mean grain yield compared to the long rains where U-15 revealed 3.41 and 1.94 tonnes per 201 hectare respectively. The performance of U-15 probably indicates the predominance of additive gene 202 203 effects in controlling these traits in the variety hence the high potential in yielding.

204



205 206

Fig. 3. Varietal effect on the grain yield during the short and long rains seasons

Matsuo *et al.* [35] also found significant differences in grain yield of rice varieties with variable P levels. Path coefficient analysis by Manyasa [36] revealed that productive tillers per plant, grains per spikelet and threshing percent had positive direct genetic effects on grain yield. The same results were also reported by Bezawelataw *et al.* [37].

# 211 **4. CONCLUSION**

Phosphorus had a positive influence on the nitrogen accumulation in the plant parts and total nitrogen uptake of finger millet as well as the finger width for both seasons with the peak observed at 25 kg ha<sup>-1</sup>  $P_2O_5$ . The newly released variety U-15 responded well to the low N in Makueni and yielded the highest (3,410 kg ha<sup>-1</sup>) which is five times the national average production and is ideal to achieve food security, poverty eradication and economic growth which are topmost set targets for sustainable development goals.

# 218 **COMPETING INTERESTS**

219 Authors have declared that no competing interests exist.

# 220 **REFERENCES**

229

230

231

237

238

239

240

241

245

246 247

248

- Oduori CO. Small Millets Production and Research in Kenya. In: Riley, K.W., Gupta, S.C.,
   Seetharam, A. and Mushonga, J.N. (eds). Advances in small millets. New Delhi: Oxford and IBH;
   1993.
- 224 2. Consultative Group on International Agricultural Research. CGIAR Research: Areas of 225 Research–Millet; 2001.
- Takan JP, Muthumeenakshi S, Sreenivasaprasad B, Akello R. Bandyopadhyay R, Talbot NJ.
   Characterization of finger millet blast pathogen populations in East Africa and strategies for disease management; 2002.
  - Opala AP, Omami NO, Christopher J, Opile NR. Response of the African nightshade to phosphate fertilizer application in Western Kenya. Archive of Applied Science Research. 2013; 5 (1): 195-201.
- LeBauer DS, Treseder KK. Nitrogen limitation of net primary productivity in terrestrial ecosystems is globally distributed. Ecology. 2008; 89:371–379.
- Elser JJ, Bracken MES, Cleland EE, Gruner DS, Harpole WS, Hillebrand H, Ngai JT, Seabloom
   EW, Shurin JB, Smith JE. Global analysis of nitrogen and phosphorus limitation of primary
   producers in freshwater, marine and terrestrial ecosystems. Ecol Lett. 2007; 10:1135–1142.
  - 7. Han W, Fang J, Guo D, Zhang Y. Leaf nitrogen and phosphorus stoichiometry across 753 terrestrial plant species in China. New Phytol. 2005;168:377–385
  - Ordoñez JC, PMv B, Witte J-PM, Wright IJ, Reich PB, Aerts R. A global study of relationships between leaf traits, climate and soil measures of nutrient fertility. Glob Ecol Biogeogr. 2005; 18:137–149.
- Wright IJ, Reich PB, Cornelissen JHC, Falster DS, Garnier E, Hikosaka K, Lamont BB, Lee W,
   Oleksyn J, Osada N, Poorter H, Villar R, Warton DI, Westoby M. Assessing the generality of
   global leaf trait relationships. New Phytol. 2005; 166:485–496
  - 10. Goron TL, Raizada MN. Genetic diversity and genomic resources available for the small millet crops to accelerate a new green revolution. Front. Plant Sci. 2015; 6, 157.
    - 11. Bhoite SV, Nimbalkar VS. Response of finger millet cultivars to nitrogen and phosphorus under rain-fed condition. J. Maharashtra Agric. Univ. 1996; 20:189–190.
- 249
   12. Gupta N, Gupta AK, Gaur VS, Kumar A. Relationship of nitrogen use efficiency with the activities of enzymes involved in nitrogen uptake and assimilation of finger millet genotypes grown under different nitrogen inputs. Sci. World J. 2012; 1–10.
- 252 13. CIMMYT. Kiboko Crops Research Station: A brief and visitors' guide: CIMMYT; 2013.
- 14. International Board for Plant Genetic Resources. Descriptors for Finger Millet. IBPGR Secretariat,
   Rome; 1985.
- 255 15. Okalebo JR, Gathua KW, Woomer PL. Laboratory methods of soil and plant analysis: A Working
   256 Manual. UNESCO, Nairobi, Kenya; 1993.

- Ryan J, George E, Rashid A. Soil and Plant Analysis Laboratory Manual. Second edition. Jointly
   published by international Center for Agricultural Research in the dry areas (ICARDA) and the
   National Agricultural Research Centre (NARC), 2001; 46-48.
- 17. Olsen DN, Sommers LE. Methods of soil analysis, part 2, chemical and microbiological properties. In: Page A.L., Miller, R.H. and Keeney, D.R. (Eds.). Agronomy monograph No. (2nd edition). ASA, Madison, Wisconsin; 1982.
- 18. Yadav BK, Niwas R, Yadav RS, Tarafdar JC. "Effect of Chaetomium globosum inoculation and organic matter on phosphorus mobilization in soil and yield of clusterbean,"Annals of Arid Zone.
   2009; 48(1): 41–44.
  - 19. Barbieri P, Echeverría HE, Saínz Rozas HR, Andrade FH. Nitrogen Use Efficiency in Maize as Affected by Nitrogen Availability and Row Spacing. Agron. J. 2008; 100:1094-1100.

266 267

268

269

279

280

281

288

289

290

291

292

296 297

- 20. Li YF, Luo AC, Wei XH, Yao XG. Changes in phosphorus fractions, pH and phosphatase activity in rhizosphere of two rice genotypes. Pedosphere. 2008; 18:785–794.
- 270 21. Bowen GD, Rovira AD. The rhizosphere and its management to improve plant growth. Adv.
   271 Agron. 1999; 66:1–102.
- 272 22. Wang X, Shen J, Liao H. Acquisition or utilization, which is more critical for enhancing 273 phosphorus efficiency in modern crops? Plant Sci. 2010; 179:302–306.
- 274 23. Rose TJ, Wissuwa M. Rethinking internal phosphorus utilization efficiency. Advances in agronomy. Elsevier. 2012; 185–217.
- 276 24. Mohidin H, Hanafi MM, Rafii YM, Abdullah SNA, Idris AB, Man S, Idris J, Sahebi M.
   277 Determination of optimum levels of nitrogen, phosphorus and potassium of oil palm seedlings in 278 solution culture. Bragantia, Campinas. 2015; 74 (3):247-254.
  - Ogembo JO. Effects of phosphorus deficiency on secondary metabolites and distribution of African Nightshade in Siaya and Kisii Counties, Kenya. MSc Dissertation, Kenyatta University; 2015
- 282 26. Boyhan GE, Torrance RL, Hill CR. Effects of Nitrogen, Phosphorus, and Potassium Rates and
   283 Fertilizer Sources on Yield and Leaf Nutrient Status of Short-day Onions. HORTSCIENCE.
   284 2007;42(3): 653-660
- 285 27. Sankar GR, Sharma KL. Dhanapal GN, Shankar MA, Mishra PK, Venkateswarlu B, Grace JK.
  286 Influence of soil and fertilizer nutrients on sustainability of rainfed finger millet yield and soil
  287 fertility in semi-arid Alfisols. Commun. Soil Sci. Plant Anal. 2011; 42:462–1483.
  - Tenywa JS, Nyende P, Kidoido M, Kasenge V, Oryokot J, Mbowa S. Prospects and constraints of finger millet production in Eastern Uganda. Afr. Crop Sci. J. 1999; 7:569–583
  - 29. Hegde BR, Gowda L. Cropping systems and production technology for small millets in India. In Proceedings of the First International Small Millets Workshop, Bangalore, India, 29 October–2 November; 1986: 209–236.
- 30. Bertrand I, Holloway RE, Armstrong RD, McLaughlin MJ. Chemical characteristics of phosphorus
   in alkaline soils from southern Australia. Australian Journal of Agricultural Research. 2003; 41:61 76.
  - 31. Glassop D, Smith SE, Smith FW. Cereal phosphate transporters associated with the mycorrhizal pathway of phosphate uptake into roots. Planta. 2005; 222:688-698.
- 298 32. Rose TJ, Pariasca-Tanaka J, Rose MT. Genotypic variation in grain phosphorus concentration,
   and opportunities to improve P-use efficiency in rice. Field Crop Res. 2010; 119:154–160.
- 300 33. Shankar AG, Udayakumar M, Prasad TG. Genotypic variability for net photosynthesis in finger
   301 millet (*Eleusine coracana* G.) genotypes: An approach to identity high CER types. Journal of
   302 Agronomy and Crop Science. 1990; 165:240-252.
- 30334. Reddy DVV, Udayakumar M, Prasad TG, Seethram A, Nanjareddy YA. Influence of NPK on304Relative Stability of Harvest Index in Finger Millet. Karnataka J. Agric. Sci. 2004; 17(4):691-695.
- 305 35. Matsuo T, Kumazawa K, Ishii R, Ishihara K, Hirata H. Science of the rice plant Physiology. Tokyo:
   306 Food and Agricultural Policy Research Center. ISBN; 1996.

- 307 36. Manyasa EO. A study of the diversity, adaptation and gene effects for blast resistance and yield
   308 traits in East African finger millet (*Eleusine coracana* (L.) Gaertn) landraces. PhD Dissertation,
   309 University of KwaZulu-Natal; 2013.
- 37. Bezawelataw K, Sripichitt P, Wongyai W, Hongtrakul V. Genetic variation, heritability and pathanalysis in Ethiopian finger millet (*Eleusine coracana* (L.) Gaertn) landraces. Kasetsart Journal (National Science). 2006; 40:322-334.
- 313