QUANTIFICATION OF PHOTOSYNTHETIC PIGMENTS OF PLANTS, WATER AND SEDIMENT SAMPLES IN CHIRACKAL AND KATTIPARAMBU OF ERNAKULAM DISTRICT, KERALA

4

5 ABSTRACT

Aims : The present study intended to investigate the pigment composition of four selected mangrove
 plants viz., Avicennia officinalis, Excoecaria agallocha, Rhizophora mucranata and Sonnaratia alba,
 water and sediment samples. And to quantify the concentration of various pigments found in the above
 samples.

Place and Duration of Study: The samples were collected from the mangrove creeks of Chirackal and
 Kattiparambu of Ernakulam district, Kerala. Duration of the study was from 2013 December to 2015
 December.

Methodology: The estimation of the total pigments, chlorophyll a, chlorophyll b and carotenoid concentration of the biotic samples, water and sediments were done using standard methods in Spectrophotometer.

16 **Results:** Plants showed high pigment concentration compared to water and sediments. High 17 <mark>c</mark>hlorophyll 'a'(2%), <mark>c</mark>hlorophyll 'b'(0.8%) and <mark>t</mark>otal <mark>c</mark>hlorophyll(2.74%) were observed in *Excoecaria* 18 agallocha of Kattiparambu and carotenoids (0.72%) observed in Rhizophora mucranata, Chirackal. In 19 sediment samples, high chlorophyll 'a'(0.85%), total chlorophylls(1.31%) and carotenoids (0.725%) were 20 reported in Chirackal area and chlorophyll 'b'(0.595%) reported in Kattiparambu. Chlorophyll 'b'(0.6%) 21 and carotenoids(0.86%) were reported high in the water samples of Kattiparambu region and chlorophyll 22 'a'(0.61%) and total chlorophylls(0.86%) in Chirackal. In Pearson's correlation coefficient studies, the 23 content of KEA-chlorophyll was found to have a strong positive correlation(1,0.999,0.998 and 0.997) 24 among themselves, with water and with sediment samples between the two media implying common 25 source of plants and sediments.

26 Conclusion: Seasonal changes and local geological conditions are the major factors for variations in 27 pigment concentrations in plants, water and sediment samples. Sediment pigments proved to be good 28 indicators of lake-ecosystem response to climate change and long-term variability in the photo trophic 29 community.

- 30 Keywords: Chlorophyll, Carotenoids, Pigments, Sediments, Mangroves, Correlation.
- 31

32 1. INTRODUCTION

33

34 Total leaf pigment includes chlorophyll-a(chl.a), chlorophyll-b(chl.b) and carotenoids that are necessary 35 for photosynthesis process. Variation in leaf pigments (chlorophylls and carotenoids) and its relation can 36 be due to internal factors and environmental conditions. Chlorophyll and carotenoids content varied with 37 microclimatic conditions in species (1). The ratio of chlorophyll-a and chlorophyll-b in terrestrial plants has 38 been used as an indicator of response to light shade conditions (2). The small proportion of chlorophyll 39 a/b is considered as sensitive biomarker of pollution and environmental stress (3). Acetone gives very 40 sharp chlorophyll absorption peaks and has great merit as the solvent for assay of chlorophylls (4). 41 Chlorophyll is a pigment that has a clear impact on the spectral responses of plants, mainly in the visible 42 spectrum portion. N is a key element in chlorophyll, therefore is usually a high correlation between them 43 (5).

43 (· 44

45 Previous studies indicated that chlorophyll pigments have antioxidant, anti inflammatory and wound 46 healing properties. It has been observed that chlorophyll pigments contain chlorophyllin which is 47 responsible for increasing the number and activity of dominant immune cells like Bcells, T- cells and 48 macrophages essential to human health (6,7). Photoactive pigments such as chl-a cause distinct changes 49 in the color of water by absorbing and scattering the light incident on water. In deep ocean waters, phytoplankton is usually the predominant constituent and the concentrations of other constituents covary 50 51 with chl-a concentration. Thus, the optical properties of these waters are dominated by phytoplankton and 52 the observed spectral features in the reflected light can be directly related to chl-a concentration (8,9). In 53 most island, estuarine, and coastal waters, constituents such as suspended solids and dissolved organic 54 matter occur in abundance and their concentrations do not co-vary with chl-a concentration (10,11).

55 56

57 2. MATERIALS AND METHODS

58 Collection of Samples : Fresh leaf samples were washed thoroughly first in tap water followed by 59 distilled water in the laboratory, kept to dry in room temperature and ground in an electric mixer (12). 60 Then analyzed for the determination of chlorophylls (Chl-a and Chl-b) and carotenoids content. Water 61 sample collected from three locations of Kattiparambu and Chirackal area in clean sampling bottles. For 62 chlorophyll estimation, sample was collected from the sub surface water in sampling bottle and add 1 ml 63 saturated MgCl₂ per litre of sample and keep in chill condition, then used for analysis. Sediment also 64 collected in polythene bags from three locations of these two areas, dried, powdered and used for the 65 analysis.

66

67 Estimation of pigments

68 The amount of chlorophyll present in the leaves was estimated by the standard method. 500 mg of leaf 69 tissue was kept in a pestle and mortar with 10 ml of 80% acetone and it was ground well and the 70 homogenate was centrifuged at 3000 rpm for 15 minutes and the supernatant was stored for analysis. In 71 water and finely powdered sediment sample, 90% acetone was added, mixed and kept for overnight at 72 low temperature under dark for extraction. The supernatant extract was centrifuged at 2000 to 3000 rpm 73 to get clear solution and the solution was used for analysis. Absorbance of the samples was measured at 74 645nm, 663nm and 480nm in a spectrophotometer. The chlorophyll content was determined by using the 75 following formula,

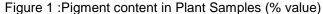
-	
77	Chlorophyll a (%.fr.wt) = $\frac{12.7 \times A663 - 2.69 \times A645}{X V \times 100}$
	a×1000×W
78	
79	Chlorophyll b (%.fr.wt) = $\frac{22.9 \times A645 - 4.68 \times A663}{22.9 \times A645 - 4.68 \times A663} \times V \times 100$
19	a×1000×W
80	
	20.2×A645+8.02×A663
81	Total Chlorophylls (%.fr.wt) = $\frac{20.2 \times 10049 + 0.02 \times 1000}{a \times 1000 \times W} \times V \times 100$
82	
83	Carotenoids (%.fr.wt) = A480+ (0.114×A663)-0.638×A645×100 (13)
84	Where,
85	A - Absorbance at respective wave length
86	a - Length of path in the cell
87	W - Fresh weight of sample (g)
88	V - Volume of extract (ml)
89	

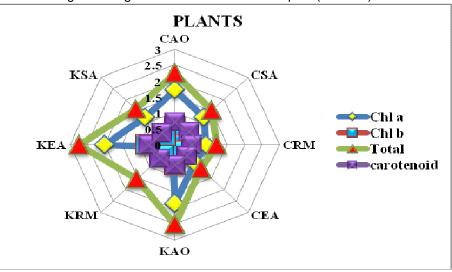
90 3. RESULTS AND DISCUSSION

91 Chloropyll a (Chl a) is a ubiquitous pigment and can be used as a global biomass indicator. In 92 Angiosperms (most land plants) there are typically two types of Chlorophyll (Chl) molecules, namely,

chlorophyll a (Chl a) and chlorophyll b (Chl b). Both of these pigments absorb photons of light in the blue 93 94 and red spectral regions, but the specific wavelengths of light they absorb are different. These natural 95 pigments exhibit various beneficial biological activities such as antioxidant, anticancer, anti-inflammatory, 96 anti-obesity, anti-angiogenic and neuroprotective activities(14). Therefore, various natural pigments 97 isolated from plants have attracted much attention in the fields of food, cosmetic and pharmacology(15).In 98 the present study, pigment level of plants gave good results when compared to water and sediments. E. 99 agallocha in Kattiparambu showed high range of Chl.a and Chl.b (2.01% and 0.804%). Total chlorophylls 100 were found to be higher in E. agallocha (2.74%) of Kattiparambu, and lower in E. agallocha (1.09) of 101 Chirackal. Similarly, carotenoids were measured to be higher in leaves of A. officinalis (0.72%) and E. 102 agallocha (0.76%) of Chirackal and Kattiparambu respectively, minimum levels of caretenoid was present 103 in R. mucranata (0.48%) of Chirackal compared to other plants (Figure-1). Acetone is known to have a 104 lower extractability of chlorophylls from the protein matrix (16). The change in the carotenoids and 105 tocopherols during seed maturation of Cassia species is studied (17). Water and sediment samples of 106 Chirackal showed high chl.a (0.61% and 0.83%) and total chlorophyll (1.074% and 1.31%) contents. High 107 range of chl.b in water (0.61%) and sediment (0.85) was reported from Kattiparambu. High range of 108 carotenoids (0.86%) reported in Kattiparambu water and sediment carotenoids (0.73%) from Chirackal 109 (Figures 2 &3). Chlorophyll capture sunlight and make it available to plant system for its cultivation on photosynthesis (18). Chlorophyll a/b ratio is an index for determining the photosynthetic efficiency of the 110 111 mangrove plant system (19). But, in this study, we claim that ratio between the bound and free forms of 112 chlorophylls can be used as an index for determining the photosynthetic efficiency of the mangrove species. Similar reports have been made earlier in mangrove species, such as R. apiculata, R. mucronata 113 and Avicennia marina (20,21,22) and pine species (23). Higher content of chlorophyll in reaction centre 114 might enhance the light - induced photosynthetic activity of the chloroplast, thereby high energy transfer 115 116 (24) and energy production could be assumed.

- 117
- 118
- 119
- 120
- 121





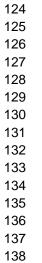
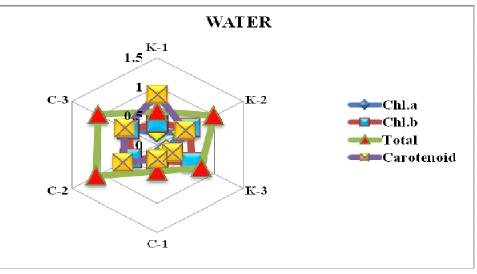
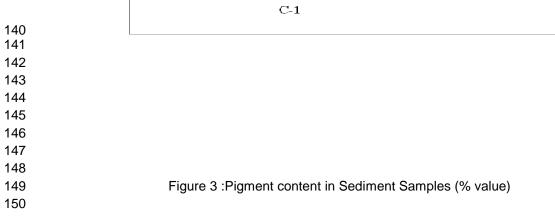
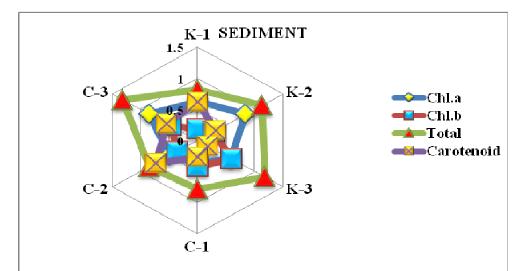




Figure 2 :Pigment content in Water Samples (% value)







	151		L																	
	152																			
	153																			
	154																			
	155																			
	156																			
	157																			
	158																			
	159																			
	160																			
	161																			
	162																			
	162 163																			
	163	Table	ə 1: Co	rrelatio	on Analy	sis of p	plants,	water a	and se	dimen	ts									
	163 164	Table	е 1: Со	rrelatio	on Analy	/sis of p	plants,	water a	and se	dimen	ts sкз	SC1	SC2	SC3	WK1	WK2	WK3	WC1	WC2	WC3
САО	163 164 165											SC1	SC2	SC3	WK1	WK2	WK3	WC1	WC2	WC3
CAO CSA	163 164 165 _{CAO}											SC1	SC2	SC3	WK1	WK2	WK3	WC1	WC2	WC3
	163 164 165 CAO	CSA										SC1	SC2	SC3	WK1	WK2	WK3	WC1	WC2	WC3
CSA	163 164 165 CAO 1 0.994**	CSA 1	CRM									SC1	SC2	SC3	WK1	WK2	WK3	WC1	WC2	WC3
CSA CRM	163 164 165 CAO 1 0.994** 0.994**	CSA 1 0.998**	CRM 1	CEA								SC1	SC2	SC3	WK1	WK2	WK3	WC1	WC2	WC3
CSA CRM CEA	163 164 165 CAO 1 0.994** 0.994**	CSA 1 0.998** 0.994**	CRM 1 0.997**	CEA 1	KAO							SC1	SC2	SC3	WK1	WK2	WK3	WC1	WC2	WC3
CSA CRM CEA KAO	163 164 165 CAO 1 0.994** 0.994** 0.984* 0.985**	CSA 1 0.998** 0.994** 0.978*	CRM 1 0.997** 0.981*	CEA 1 0.964*	KAO 1	KRM						SC1	SC2	SC3	WK1	WK2	WK3	WC1	WC2	WC3
CSA CRM CEA KAO KRM	163 164 165 CAO 1 0.994** 0.994** 0.995** 0.536	CSA 1 0.998** 0.994** 0.978* 0.518	CRM 1 0.997** 0.981* 0.575	CEA 1 0.964* 0.588	KAO 1 0.545	KRM	KEA					SC1	SC2	SC3	WK1	WK2	WK3	WC1	WC2	WC3

SK1	0.793	0.851	0.851	0.886	0.731	0.522	0.726	0.795	1									
SK2	0.999**	0.987*	0.989*	0.975*	0.999**	0.545	0.998**	0.998**	0.763	1								
SK3	0.787	0.720	0.751	0.716	0.838	0.713	0.850	0.785	0.357	0.815	1							
SC1	0.745	0.689	0.732	0.710	0.786	0.861	0.801	0.743	0.425	0.769	0.970*	1						
SC2	0.563	0.613	0.644	0.692	0.510	0.792	0.514	0.565	0.865	0.539	0.342	0.513	1					
SC3	0.970*	0.958*	0.975*	0.969*	0.970*	0.724	0.974	0.970*	0.787	0.972*	0.855	0.861	0.677	1				
WK1	-0.224	0.159	-0.123	0.058	0.282	0.458	0.275	-0.221	0.316	0.251	0.295	0.056	0.680	0.063	1			
WK2	0.612	0.569	0.624	0.618	0.643	0.966*	0.660	0.611	0.431	0.631	0.862	0.959*	0.645	0.780	0.217	1		
WK3	0.247	0.169	0.230	0.205	0.314	0.789	0.336	0.244	0.061	0.284	0.758	0.830	0.252	0.437	0.078	0.874	1	
WC1	0.589	0.552	0.609	0.607	0.615	0.981*	0.633	0.589	0.450	0.606	0.827	0.939*	0.684	0.765	0.283	0.998**	0.862	1

WC2	0.846	0.845	0.879	0.890	0.837	0.892	0.846	0.847	0.802	0.845	0.783	0.869	0.848	0.945*	0.249	0.882	0.543	0.886	1	
WC3	0.763	0.745	0.790	0.795	0.769	0.955*	0.781	0.763	0.672	0.769	0.829	0.926*	0.798	0.897	0.264	0.958*	0.698	0.961*	0.980*	1

166

167 Correlation studies

168 The result of Pearson's correlation coefficient studies conducted between the pigment contents in Plants, 169 Water and Sediments in Table 1. Plants showed strong positive correlation among themselves, with 170 water and sediment samples and also very strong correlation (0.998 and 0.997) with sediment between 171 the two media implying common source of plants and sediments. The content of KEA-chl was found to 172 have a high positive correlation with the photosynthetic efficiency of mangrove species. The correlation 173 coefficient between the KEA-chl and assimilation rate was 1,0.999,0.998 and 0.997. There was a strong 174 negative correlation between water and plants (-0.224,-0.221 and -0.123). This suggests that in plants 175 there is less production of pigments in the presence of certain sediments and water or vice versa in a 176 particular condition.

177

178 Abbreviations

- 179 CAO- Chirackal Avicennia officinalis
- 180 CRM-Chirackal Rhizophora mucranata
- 181 CEA Chirackal Excoecaria agallocha
- 182 CSA Chirackal Sonneratia alba
- 183 KAO Kattipapambu Avicennia officinalis
- 184 KRM Kattiparambu Rhozophora mucranata
- 185 KEA Kattiparambu Excoecaria agallocha
- 186 KSA Kattiparambu Sonneratia alba
- 187 SK-1 Sediment of Kattiparambu-1
- 188 SK-2 Sediment of Kattiparambu-2
- 189 SK-3 Sediment of Kattiparambu-3
- 190 SC-1 Sediment of Chirackal-1
- 191 SC-2- Sediment of Chirackal -2
- 192 SC-3 Sediment of Chirackal -3
- 193 WK-1 Water of Kattiparambu-1
- 194 WK-2 Water of Kattiparambu-2
- 195 WK-3 Water of Kattiparambu-3
- 196 WC-1 Water of Chirackal -1
- 197 WC-2 Water of Chirackal -2
- 198 WC-3 Water of Chirackal -3
- 199 K-1 Kattiparambu 1
- 200 K-2 Kattiparambu 2
- 201 K-3 Kattiparambu 3
- 202 C-1 Chirackal -1
- 203 C-2 Chirackal -2
- 204 C-3 Chirackal -3
- 205

206 4. CONCLUSION

Results from the above analysis clearly indicate that extraction of photosynthetic pigments depend on chemical nature of bio-molecules (cholorophyll-a, chlorophyll-b and carotenoids). The pigment content were influenced by environmental parameters. Temporal and seasonal changes and local geological conditions are the reasons for variations in pigment concentrations in plants, water and sediment samples. Sediment pigments proved to be good indicators of lake-ecosystem response to climate change and long-term variability in the photo trophic community, which is needed for predicting possible effects of future climate change. It was also recognized that the quality of the pigment record is highly dependent on the preservation regime in the sediment and water. Therefore further study in this context is recommended.

217 REFERENCES

216

234

235

236

244

- Shaikh S. D and Dongare M. Analysis of photosynthesis pigments in *Adiantum lunulatum*, Burm.
 At different localities of Sindhu durg District (Maharastra). *Indian Fern J.* 2008; 25: 83–86.
- Vicas S. I, Laslo V, Pantea S. and Bandict G. E. Chlorophyll and carotenoids pigments from Mistletoe (*Viscum album*) leaves using different solvents. *Fascicula Biol.* 2010; (2): 213–218.
- Tripathi A. K and Gautam M. Biochemical parameters of plants as indicators of air Pollution. *J. Environ. Biol.* 2007; 28: 127–132.
- Ritchie R. J. Consistent sets of spectrophotometric chlorophyll equations for acetone, methanol and ethanol solvents. *Photosynth. Res.* 2006; 89: 27–41.
- Schlemmer, M. R, Francis, D. D, Shanahan, J. F & Schepers, J. S. Remotely measuring
 chlorophyll content in corn leaves with differing nitrogen levels and relative water content.
 Agronomy Journal. 2005; 97(1): 106–112.
- Rajalakshmi K and Banu N. Antioxidant capacity of chlorophyll in from *Mimosa pudica* by
 formation of a phosphomolybdenum complex. *International Journal of Frontiers in Science and Technology*. 2014; 2: 1-14.
- Durgadevi, M and Banu, N. Study of antioxidant activity of chlorophyll from some medicinal
 plants. Paripex Indian Journal of research. 2015; 4(2): 6-8.
 - 8. Morel A and Prieur L . Analysis of variations in ocean color Limnol. Oceanogr. 1977; 22 709–22
 - 9. O'Reilly J E. SeaWiFS Postlaunch Calibration and Validation Analyses (Part 3. NASA Tech. Memo. 2000-206892) vol 11 (MD: NASA Goddard Space Flight Center). 2000; 49.
- 237 10. Carder K L, Chen F R, Cannizzaro J P, Campbell J W and Mitchell B G. Performance of the
 238 MODIS semi-analytical ocean color algorithm for chlorophyll-*a Adv. Space Res.* 2004; 33 1152–9
- 239 11. Dall'Olmo G and Gitelson A A. Effect of bio-optical parameter variability on the remote estimation
 240 of chlorophyll-*a* concentration in turbid productive waters: experimental results *Appl. Opt.* 2005;
 241 4412–22
- Kupper, H, Kupper, F and Spiller, M. *In situ* detection of heavy metal substituted chlorophylls in water plants. *Photosynthesis Res*.1998; 58: 123–133.
 - 13. Arnon D. I. Copper enzymes in isolated chloroplasts. Polyphenol oxidase in Beta vulgaris. Plant. Physiol.1949; ,24 : 1-5.
- Rehman, A.M, Mohamed, M.I. Effect of cement dust deposition on physiological behaviors of
 some halophytes in the salt marshes of red sea. Egyptian Academic Journal of Biological
 Sciences. 2012; 3(1): 1-11.
- Pangestuti, R., and S-K. Kim, (2011). Biological activities and health benefit effects of natural pigments derived from marine algae. J. Func. Food., 3(4): 255-266.
- 16. Nakamura, A., T. Watanabe, (2001). Separation and determination of minor photosynthetic
 pigments by reversed-phase HPLC with minimal alteration of chlorophylls. Anal. Sci. 17: 503–
 508.
- 254 17. Zako S. M, W. Akht A. R, Khan S.A and Bhathy M.K. Characterization of *Cassia* seed oil. Proc.
 255 Pakistan Acad. Sci. 1986; 23 : 167-172.
- 18. Rao, A.V. and Rao, L.G. Carotenoids and human health. *Pharmacological Research*.2007; 55:
 207-216.
- 19. Kathiresan, K. & L. Kannan.. Photosynthetic productivity in species of Rhizophora. In: The
 Mangroves. Proc. Natl. Symp. Biol. Util. Cons. Mangroves Shivaji university, Kolhapur, India.
 1985; 262-265.

- 20. Kathiresan, K. & P. Moorthy. Influence of different irradiance on growth and photosynthetic characteristics in seedlings of Rhizophora species. Photosynthetica. 1993; (29): 143-146.
- 263 21. Kathiresan, K. & P. Moorthy. Photosynthetic responses of *Rhizophora apiculata* Blume seedlings
 264 to a long chain aliphatic alcohol. Aquat. Bot. 1994; 47: 191-193.
- 265 22. Kathiresan, K. & P. Moorthy.. Hormone-induced physiological responses of a tropical mangrove
 266 species. Bot Mar. 1994a ;37: 139-141.
- 267 23. Krivosheeva, A., S.A. Shavnin, V.A. Kalinin & P.S. Venedikov. Effect of industrial pollutants on
 268 seasonal changes of chlorophyll content in scotch pine seedlings. Fiziol. Rastenii. 1991; 38: 162269 168.
 - 24. Moorthy, P.& K. Kathiresan. Physiological responses of a mangrove seedling. Biol. Plant. 1993; 35: 577-581.