# Anti-*Alternaria solani* activity of onion (*Allium cepa*), Ginger (*Zingiber officinale*) and Garlic (*Allium sativum*) *in vitro*


## ABSTRACT

Plant pathogens cause serious losses in quantity and quality of agricultural products. Use of fungicides is gradually becoming unpopular due to their negative effects on ecosystems, human and animal health, and due to resistance by pathogens to the fungicides. *In vitro* studies were carried out in order to determine the effects of three plant extracts; onion (*Allium cepa*), ginger (*Zingiber officinale*) and garlic (*Allium sativum*) on the control of *Alternaria solani.* The experiment was laid in a Completely Randomized Design (CRD) with a 3x3 factorial arrangement plus one control. The first factor was plant extract, with three levels (garlic, onion and ginger) the second was plant extract concentration, with three levels (50%, 75% and 100%). The experiment was carried out in the laboratory at Midlands State University, Zimbabwe, in October 2014. Data on mycelia growth diameter, mycelia inhibition percent and spore germination percent was collected. Results showed that the plant extracts had strong anti-*A. solani* activity and their effect increased with increase in their concentration. Ginger and garlic had significantly stronger effect on reducing mycelia growth, reducing spore germination and causing high inhibition percentage of *A. solani.* Ginger was the most effective in controlling *A. solani* across all concentrations. It can be concluded that the plant extracts (onion, ginger and garlic) can be used as natural fungicides to control pathogenic fungi. It is recommended that further research be done on the plant extracts so as to identify the active compounds which are in the extracts as these are responsible for this fungicidal activity and to carry out more studies to test antifungal activity of these studied plant extracts on other different fungi, at different concentration levels. Further experiments may also be done in the field to determine effects of these plant extracts in controlling diseases caused by *A. solani*.



**Key words**: Antifungal activity, plant extracts, *Alternaria solani*


## 1. INTRODUCTION

In agriculture, the crop loss due to plant pathogens has become a major concern and one of such pathogens is *A. solani*. *A. solani* is a soil inhabiting air borne pathogen [1] responsible for early blight, an important chronic foliar disease of mainly the Solanacea family including tomatoes (*Lycopersicon esculentum*) and potato (*Solanum tuberosum*) [2]. Basal girdling and death of seedlings may occur, a symptom known as collar rot. Despite the name "early," foliar symptoms usually occur on older leaves, [3]. The disease causes yield losses through defoliation of plants and this may result in a reduction in yields by as much as 20 to 30% for example in potatoes [4].

Chemical control is the most effective and applied method in controlling *A. solani* and there are numerous fungicides on the market for controlling early blight. The disease is commonly managed using succinate dehydrogenase inhibitor (SDHI) fungicides. Unfortunately, recent studies have shown that SDHI

resistance has increased dramatically over the years in *A. solani* populations [5]. In addition, conventional pesticides; over the past five decades have led to a range of problems in agriculture, the environment, and human health [6]. There are numerous costs derived from pesticide use and these include monitoring and sanitation for contamination of soils, drinking water, or food, poisoning of pesticide users and farm workers, and the deleterious effects on non-target organisms such as bees and other beneficial insects, fish, and birds [7]. To overcome these problems, some alternative control methods must be employed.

Natural plant products (botanicals) are becoming a new source of agricultural chemicals to manage plant diseases [8]. Plant extracts have been known for their medicinal and antimicrobial properties since ancient times [9]. Many higher plants produce economically important organic compounds, pharmaceuticals and pesticides. Plant based secondary metabolites, which have defensive role may be exploited for the management of foliar diseases [10]. The antifungal action of plant extracts has gained much attention. Nowadays, plants are being used against many plant pathogenic fungi. The plants serve as eco-friendly and economic bio-control agents [11]. Natural chemicals from plants are cheap, readily available and cost-effective in developing countries where synthetic fungicides are scarce and expensive for resource-poor farmers [12]. A number of researches have been documented which demonstrate the antimicrobial efficacy of various plant extracts which have been seen to contain some antifungal properties against *A. solani*. These botanicals include onions, (*Allium cepa*), ginger (*Zingiber officinale*) and garlic (*Allium sativum*) [11,13,14]. These three botanicals have antifungal properties, which enable them to distort the life cycle of *A. solani* [15]. The present study was designed to evaluate the efficacy of three plant extracts, onion, ginger and garlic on *A. solani* development *in vitro*.

## 2. MATERIALS AND METHODS

### 2.1 Site description and experimental design

The experiment was carried out in the laboratory at Midlands State University which is located in Gweru, Zimbabwe. The area is found in Agro-ecological Region III [16] on the following coordinates 29°45'E, 19°45'S and the altitude is 1420m above sea level.

The experiment was laid in a Completely Randomized Design (CRD) with a 3x3 factorial arrangement plus one control. The first factor was plant extract type, with three levels; garlic, onion and ginger, while the second factor was plant extract concentration, with three levels; 50%, 75% and 100%.The control used was 70% ethanol. The experiment was replicated three times.

## 2.2 Experimental Procedure

### 2.2.1 Isolation of *A. solani*.

The infected tissues along with adjacent small unaffected tissue are cut into small pieces (2–5 mm squares) and by using flame-sterilized forceps, they are transferred to sterile Petri dishes containing 5% sodium hypochloride for 30-60 s for surface sterilization of plant tissues. The sterilized pieces are aseptically transferred to Petri dishes containing solidified Potato Dextrose Agar and were incubated at 27$^{o}$C for 72 hours as according to Abou-Zeid *et al*, 2004.

### 2.2.2 Preparation of plant extracts and inoculation of *A.solani*

The research material ginger (*Z. officinale*) rhizomes, onion (*A. cepa*) bulbs, and garlic (*A. sativum)* bulbs, was obtained from a local vegetable market. Fifty grams of the plant material of each plant species was washed with water and surface sterilized with sodium hypochloride for 30-60seconds and crushed in a mortar with pestle by adding sterile distilled water at the rate of 10 ml/10g of plant tissue and the homogenates were centrifuged at 10 000 rpm for 15 min at 4$^{o}$C and the supernatant solutions were collected [18]. The supernatant was filtered through Whatman No. 1 filter paper and sterilized at 120$^{o}$C for 30 min. The obtained extracts served as the crude extract which is the 100% concentration as according to Mohana and Raveesha, (2007). The obtained concentrates were stored at 4$^{o}$C. Out of the 100% crude extract from the different plant materials, the respective dilutions of 50% and 75% were then prepared.

## 2.3 Determination of mycelia growth diameter

Five ml of 50%, 75 % and 100% of natural concentrate of onion (*A. cepa*), garlic (*A. sativum*) and ginger (*Z. officinale),* was then administered separately into Petri dishes and blended with cooled liquid PDA. One ml of 70% ethanol (positive control) was poured per Petri dish using an inoculating needle. Fifteen ml PDA was separately poured into Petri dishes, allowed to cool and solidify. After complete solidification of the medium, five mm disc of 72 hour old culture of the *A. solani* was inoculated into PDA at the centre of the Petri dishes. The plates were incubated at 28$^{o}$C. The Petri dishes containing media devoid of the extract but with same amount of distilled water served as control. *A. solani* mycelia growth diameter was measured using a string diagonally and the string was put on a 30 cm measuring ruler. This was done daily for four consecutive days. Mean diameter was calculated respectively to plant type and concentration level.

## 2.4 Determination of mycelial Inhibition percentage by Poisoned food technique

After incubation the colony diameter was measured in mm as described by Singh and Tripathi (1999). Each treatment was repeated three times. The toxicity of the extracts in terms of percentage inhibition of mycelia growth was calculated using the formula: Gc - Gt/Gc x 100, where Gc =diameter in control and Gt= diameter in plant extract.

### 2.5 Spore germination

The counting of conidia was done by means of haemocytometer for this purpose one disc (one cm) from each Petri dish was taken from seven days old culture of *A. solani*. The disc (one cm) was washed using two ml of distilled water for the collection of spores. One drop of solution was put on haemocytometer and spores were counted under microscope. The percentage was found using the formula:

Number of spore germinated/number of examined spores x100

### 2.6 Data Analysis

Analysis of variance (ANOVA) was done on data collected using Genstat 14$^{th}$ edition. Separation of means was done using Duncan Multiple Range Test at 5% level of significance.

## 3. RESULTS

### 3.1 Effects of plant extracts on *A. solani* mycelia growth diameter

There was an interaction between plant type and concentration level of the plant extracts on mycelia growth diameter of *A. solani.* The mycelia colony diameter decreased with an increase in concentration rate of the different plant extracts. Of the three plant extracts, the highest mycelial growth diameter (3.7cm) was recorded for garlic at 50% concentration level while the lowest was recorded for ginger at 100% and this was not significantly different ($P<0.05$) from that of the control (ethanol). Generally ginger resulted in the highest decrease in *A. solani* colony diameter across all respective concentrations (50%, 75% and 100%) though its effect at 50% and 75% were not significantly different from that of onion at these respective concentrations (Fig. 1).

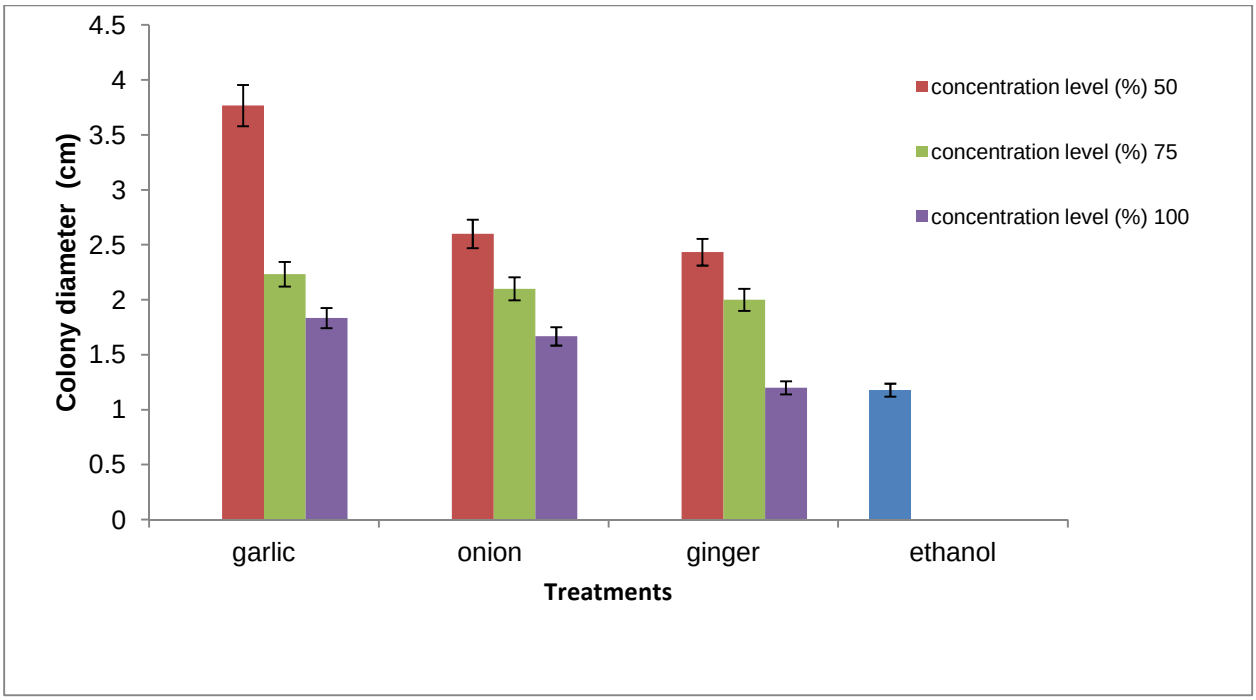


**Fig. 1.** Effects of plant extracts and different concentrations on mycelial diameter growth of *A.solani*

### 3.2 Effects of plant extracts on inhibition percentage

There was an interaction between plant type and concentration level on their effects on inhibition percentage. As the concentration of the plant extracts increased; the *A. solani* inhibition percentage also increased (Fig. 1). Of the three plant extracts, garlic applied at 100% concentration resulted in the highest inhibition percentage followed by 100% onion although this was not significantly different ($P$<0.05) from that of 100% ginger. Ethanol (70%) recorded the highest *A. solani* inhibition percentage (100%).

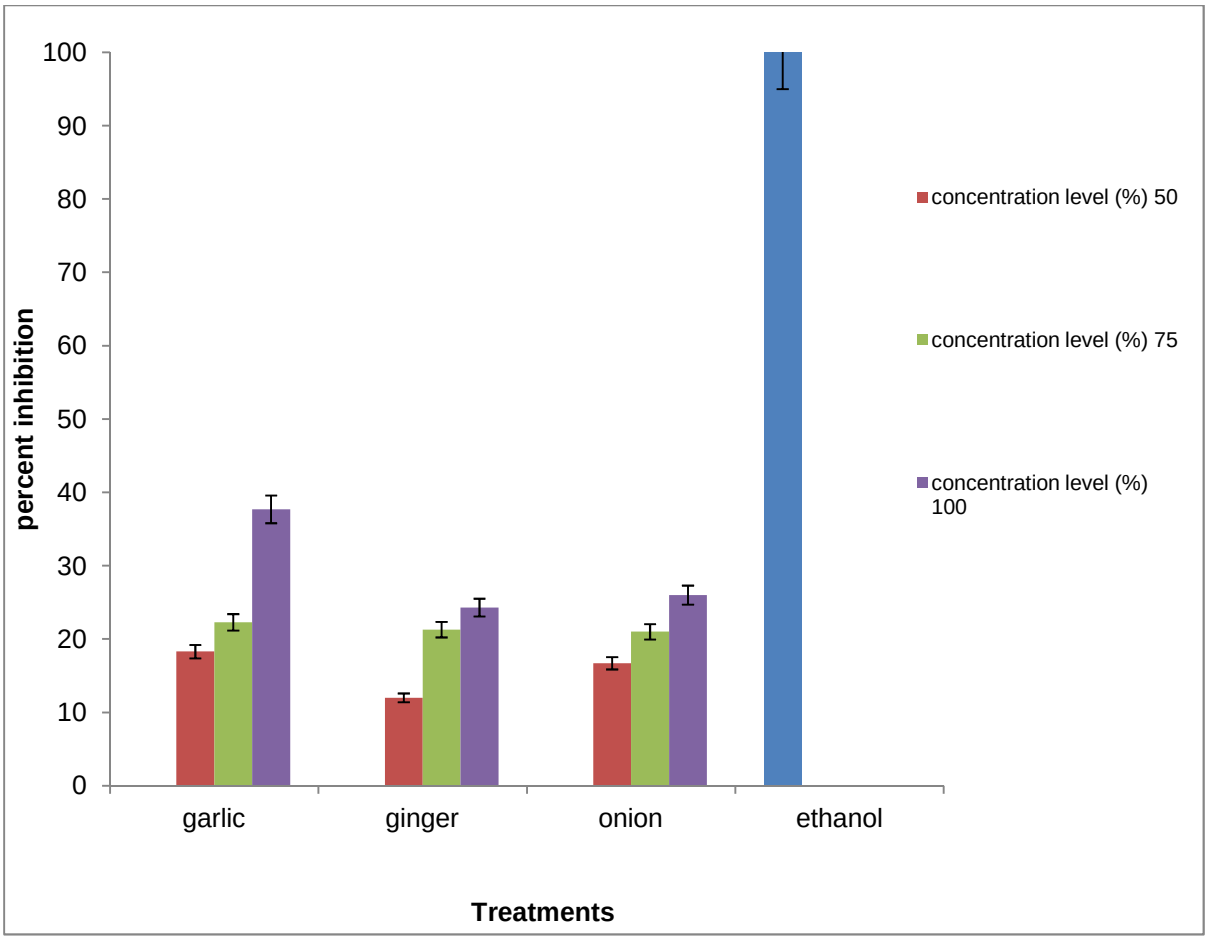


**Fig. 2.** Effects of plant extracts and concentrations on inhibition percentage of *A.solani*

### 3.3 Effects of plant extracts on spore germination

There was an interaction between plant extract type and concentration level on *A. solani* spore germination percentage. There was a reduction in spore germination percentage as concentration of the respective plant extracts increased. Results showed that ginger resulted in a significantly ($P$<0.05) greatest reduction in spore germination percentage while onion resulted in the highest spore germination percentage under the three concentration levels (Fig. 3). Where 70% ethanol (control) was used, no spores germinated at all.

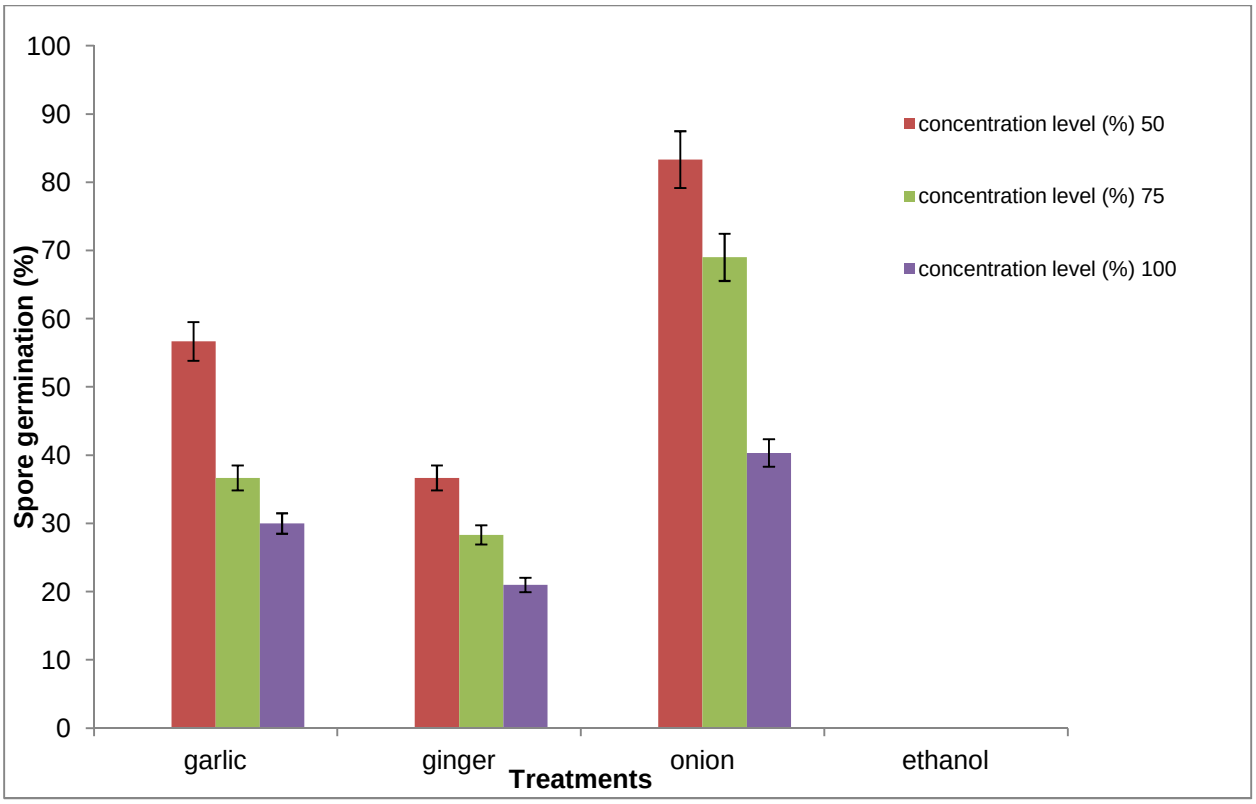


**Fig. 3.** Effects of plant extracts and concentrations on spore germination percent of *A.solani.*

## 4. DISCUSSION

The results from our study showed that the plant extracts tested (ginger, garlic and onion) have some antifungal property and have the capacity to suppress development of *A. solani*. The reduction in mycelia growth increased with increase in concentration of the extracts. This is in concurrence with some *in-vitro* action tests conveyed on some plant extracts on seed borne pathogens of wheat, for example, *Aspergillus* spp. [21]. Similar findings were reported by Swame and Alane, 2013 who found that at higher concentrations tested, plant extracts were effective in controlling seed borne fungi of mungbean seed. Tagoe *et al*, 2009 also noted the antifungal properties of garlic in inhibiting the growth of *Aspergillus* species. Results of this study are also in line with those of other researchers who showed that plant extracts result in inhibition of mycelial growth and these extracts include *Allium cepa* and *Allium sativum* [23], *Azadirachta indica* [13], *Zinger officinale* [14].

Ginger had the highest antifungal activity on *A. solani* with mycelial diameter mean of (2.4cm) at 50%, (2.1cm) at 75% and (1.2cm) at 100%. The strong inhibition potential of ginger is attributed to the fact that it contains over 400 different compounds, a mixture of both volatile and non-volatile chemical constituents

such as zingerone, shogaols and gingerols, sesquiterpenoids (β-sesquiphellandrene, bisabolene and farnesene) and a small monoterpenoid fraction (β-phelladrene, cineol, and citral [24].The main constituents of the garlic essential oils are diallyl monosulfide, diallyl disulfide (DADS), diallyl trisulfide, and diallyl tetrasulfide [25]. G*ingerols* and *shogals*, found in ginger are less volatile as compared to *alliin* in garlic and onion which could have been lost through diffusion during plant extracts preparation process.

There was an interaction between plant extract type and concentration level on spore germination percentage. As plant extract concentration level increased, this resulted in a corresponding decrease in spore germination percentage. Ginger at 100 % was most effective with the lowest spore germination percentage of 22%. Results on the effectiveness of ginger as a bio control is in line with findings by Fawzi *et al*., 2009, who showed that plant extracts including cinnamon (*Cinnamomum zeylanicum*), laurel (*Laurus nobilis*) and ginger (*Zinger officinale*) had strong antifungal activity with high inhibition on growth of *Alternaria alternata* and *Fusarium oxysporum*. According to this study by Fawzi *et al*., 2009 ginger proved to be the most effective in inhibiting fungal growth, similar to our findings. Of the three extracts used garlic and ginger were comparatively most effective in controlling *A. solani*. This is in line with studies by Islam and Faruq, 2013, who also showed that garlic clove and ginger rhizome were effective in controlling *F. oxysporum* and *Scleretonium rolfsii*; fungi which cause damping off disease. However on spore germination garlic across all concentrations turned to be more effective as compared to onion. This is likely because garlic is known to have some added phytochemicals which inhibit spore germination [22]. These findings are in agreement with those of many researches [27,28, 29] which indicate positive antifungal spore germination effect of the plant extracts *A. cepa* and *A. sativum.* Garlic has also been shown to effectively reduce mycelia growth of *Pythium aphanidermatum*, a causal organism of damping of chilli [30].

Experiment by Mohana and Raveesha 2007, confirmed the antimicrobial activity of six plant extracts including sweat Basil, neem, eucalyptus, Jimson weed, oleander and garlic, against *A. solani in vitro*. In this study, neem and garlic were shown to be the most effective in causing highest reduction of mycelia growth of *A. solani* (43.3% and 42.2% respectively). The inhibitory effects of plant extracts may be due to their direct toxic effects on the pathogen or the plant extracts may induce systemic resistance in host plants resulting in a reduction of the disease development [31].

## 5. CONCLUSION AND RECOMMENDATIONS

From our findings it can be concluded that plant extracts onion (*Allium cepa*), ginger (*Zingiber officinale*) and garlic (*Allium sativum*) can be used for biocontrol of *A. solani* since they have antifungal properties. It has been demonstrated that these plant extracts can effectively reduce *A. solani* mycelia growth, and cause significant inhibition of fungal growth. Of the plant extracts used; ginger proved to be most effective followed by garlic, and lastly onion. It can also be concluded that plant extracts may be more effective in

fungal growth control at high concentrations. Use of plant extracts as control method of *A. solani* can contribute to minimizing risks and hazards of toxic fungicides. We recommend for further research to be done on the plant extracts so as to identify the active compounds which are in the extracts as these are responsible for this fungicidal activity. In addition, it is recommended that more studies be done to test antifungal activity of the studied plant extracts on other different fungi, at different concentration levels. Further experiments may also be done in the field to determine effects of these plant extracts in controlling diseases caused by *A. solani* for example early blight.

## REFERENCES


1. Datar VV, Mayee CD. Conidial dispersal of *Alternaria solani* in tomato. Indian Phytopathology 1982; 35:68 -70.
2. Gudmestad NC, Arabiat S, Miller JS, Pasche JS. Prevalence and impact of SDHI fungicide resistance in *Alternaria solani.* Plant Dis 2013; 97:952-960.
3. Kemmitt G. Early blight of potato and tomato. The Plant Health Instructor. 2002. *Updated 2013.* http://www.apsnet.org/edcenter/intropp/lessons/fungi/ascomycetes/Pages/PotatoTomato.aspx
4. Shahbazi H, Aminian H, Sahebani N, Halterman DA. Biochemical evaluation of resistance responses of potato to different isolates of *Alternaria solani*. Phytopathology 2010;100:454 - 459.
5. Miles TD, Fairchild KL, Merlington A, Kirk WW, Rosenzweig N, Wharton PS. First Report of Boscalid and Penthiopyrad-Resistant Isolates of *Alternaria solani* Causing Early Blight of Potato in Michigan 2013; 97 (12):1655.
6. Geiger F, Bengtsson J, Berendse F, Weisserc WW, Emmerson M, Morales MB, Ceryngier P, Liir J, Tscharntke T, Winqvist C, Eggers S, Bommarco R, Pa¨rt T, Bretagnolle V, Plantegenest M, Clement LW, Dennis C, Palmer C, On˜ate JJ, Guerrero I, Hawro V, Aavik T, Thies C, Flohre A, Ha¨nke S, Fischer C, Goedhart PW, Inchausti P. Persistent negative effects of pesticides on biodiversity and biological control potentialon European farmland. Basic and Applied Ecology 2010;11:97-105.
7. Lamichhane JR, Dachbrodt-Saaydey S, Kudsk P, Messean A. Toward a reduced reliance on conventional pesticides in European Agriculture. 2015; The American Phytopathological Society.
8. Hubert J, Mabagala RB, Mamiro DP. Efficacy of Selected Plant Extracts against *Pyricularia grisea*, Causal Agent of Rice Blast Disease. American Journal of Plant Sciences 2015; 6: 602-611. http://dx.doi.org/10.4236/ajps.2015.65065
9. Lalitha P, Sripathi SK, Jayanthi P. UV Protecting Ability of Sunscreen Lotions Prepared with Extracts of Pisonia Grandis R.Br World Journal of Pharmacy and Pharmaceutical Sciences. 2015; 4: 324-329.
10. Saurabh S, Seweta S, Jyotiranjan M, Richa R, Asha S. Extract Against Predominant Seed Mycoflora of Mungbean *Vigna Radiata* (L.) Wilczek Seed. Life Sciences Leaflets. 2013; 51:83-89.
11. Swami, CS, Alane, SK, Efficacy of Some Botanicals against Seed - Borne Fungi of Green Gram (*Phaseolus Aureus* Roxb.) Bioscience Discovery, 2013; 4(1):107-110.

12. Mossini SAG, Carla C, Kemmelmeier C. Effect of neem leaf extract and neem oil on Penicillium growth, sporulation, morphology and ochratoxin A production. Toxins 1, 3-13. 2009

13. Abd El-Ghany TM, Roushdy MM, Mohamed A. A. Efficacy of certain plant extracts as safe fungicides against phytopathogenic and mycogenic fungi. Agricultural and Biological Sciences Journal. 2015; 1 (3); 71-75

14. Fawzi EM, Khalil AA, Afifi AF. Antifungal effect of some plant extracts on *Alternaria alternata* and *Fusarium oxysporum* African Journal of Biotechnology. 2009; 8 (11), 2590-2597.

15. Benkeblia N. Antimicrobial activity of essential oil extracts of various onions (Allium cepa) and garlic (Allium sativum). 2004; 37:263-268.

16. Mugandani R, Wuta M, Makarau A, Chipindu B, Re-Classification Of Agro-Ecological Regions of Zimbabwe In Conformity with Climate Variability and Change. African Crop Science Journal 2012; 20, Issue Supplement S2: 361 - 369

17. Abou-Zeid AM, Mahmoud YAG, Talhi AD. Effect of Gaucho insectide on the efficacy of fungicides used to control root-rot and damping-off diseases in cotton seedlings in Egypt. Microbiol 2004; 9: 1-10.

18. Nashwa SMA, Abo-Elyousr KAM. Evaluation of various plant extracts against the early blight disease of tomato plants under greenhouse and field conditions. Plant Protect. Sci 2012; 48: 74–79.

19. Mohana DC, Raveesha KA. Anti-fungal evaluation of some plant extracts against some plant pathogenic field and storage fungi. Journal of Agricultural Technology 2007; 4(1): 119-137.

20. Singh J, Tripathi NN. Inhibition of storage fungi of blackgram (*Vigna mungo* L.) by some essential oils. Flavour and Fragrance Journal 1999; 14: 1-4.

21. Hasan MM, Chowdoury SP, Shahidul A, Hossain B, Alam MS. Antifungal effects of plant extracts on seedborne fungi of wheat seed regarding seed germination, seedling health and vigor index. Pakistan Journal of Biological Sciences 2005; 8, 1284-1289.

22. Tagoe D, Baidoo S, Dadzie I, Kangah V, Nyarko H. A Comparison of the Antimicrobial (Antifungal) Properties of Garlic, Ginger and Lime on Aspergillus flavus, Aspergillus niger and Cladosporium herbarum Using Organic and Water Base Extraction Methods. The Internet Journal of Tropical Medicine. 2009; 7(1).

23. Gohil VP and Vala GD. Effect of extracts of some medicinal plants on the growth of Fusarium moniliforme, J. Mycol. Pl. Pathol. 1996; 26 (1) 110- 111.

24. Chrubasik S, Pittler MH, Roufogalis BD. Zingiberis rhizoma: A comprehensive review on the ginger effect and efficacy profiles. Phytomedicine. 2005; 12(9):684-701.

25. Casella S, Leonardi M, Melai B, Fratini F and Pistelli L. The Role of Diallyl Sulfides and Dipropyl Sulfides in the In Vitro Antimicrobial Activity of the Essential Oil of Garlic, Allium sativum L., and Leek, Allium porrum L. Phytotherapy Research 2013; 27(3). DOI: 10.1002/ptr.4725

26. Islam MT, Faruq AN. Effect of some medicinal plant extracts on damping-off disease of winter vegetable. World Applied Sciences Journal.2012; 17 (11): 1498-1503.

27. Bashir S. Evaluation of some medicinal plant extracts against Fusarium oxysporum f. sp. and Alternaria sp. M.Sc (Ag) Thesis, Allahabad Agriculture Institute (Deemed University), Allahabad, U.P, India, 2001; 65 P.

28. Bhat ZA, Comparative efficacy of bio-control agents, Botanical extracts and fungicide in the management of chickpea wilt caused by Fusarium oxysporum. M. Sc. (Ag.) thesis, Allahabad Agriculture Institute (Deemed University). Allahabad-211007, (U.P) India. 2002; 65pp.

29. William Q. Least toxic controls of plant diseases. Brooklyn Botanic garden. Natural Disease Control. 2008; 11,225.

30. Kurucheve V, Padmavathi R. Effect of seed treatment with plant products on seed germination, growth and vigour of chilli seedlings (K-1). Indian Pathology 1997; 50(4): 529-530.

31. Kagale S, Marimuthu T, Thayumanavan B, Nandakumar R, Samiyappan R. Antimicrobial activity and induction of systemic resistance in rice by leaf extract of *Datura metel* against *Rhizoctonia solani* and *Xanthomonas oryzae* pv. *oryzae*. Physiological and Molecular Plant Pathology. 2004; 65: 91–100.