

Original Research Article

Spermatotoxic effects of some medicinal plants (*Carica papaya*, *Hibiscus rosa-sinensis* and *Ipomoea batatas*) on sperm quality and testicular weight in male African catfish (*Clarias gariepinus*)

ABSTRACT

This study was designed to evaluate the effects of pawpaw (*Carica papaya*) seeds, hibiscus plant (*Hibiscus rosa-sinensis*) leaves, sweet potato (*Ipomoea batatas*) leaves on sperm quality (sperm motility, sperm density, semen volume) and weight of testes of male *Clarias gariepinus*. One hundred and twenty (120) juveniles of *C. gariepinus* were collected from the University of Calabar fish farm. The 120 fish were randomly divided into 12 experimental tanks measuring 80x80x80cm (L x W x H), with three tanks for each treatment, using a completely randomized design (CRD). Three grams (3g) of each test plant were incorporated into 1kg of Coppens feed (3g/kg) and reformulated into four experimental diets; Treatment A- Control, B- Pawpaw seed meal (PSM), C-Hibiscus leaf meal (HLM) and D- sweet potato leaf meal (SPLM). The experiment was done in three replications. The fish were fed twice daily for 6 months. Data obtained were analyzed using a one way analysis of variance (ANOVA). Results showed that fish fed with HLM had significantly ($p = .05$) higher testicular weight when compared with the control and other test plants. Moreover, sperm volume and density significantly ($p = .05$) reduced in fish samples treated with PSM and SPLM when compared with the control and fish fed with HLM. The highest mean sperm volume and density were obtained in fish samples fed with HLM. No significant difference was observed in the sperm motility of the fish in all the treatment groups. Conclusively, this study reveals the pro-fertility potential of *H. rosa-sinensis* in male *C. gariepinus* while *C. papaya* and *I. batatas* possess anti-fertility properties. Therefore, HLM can be utilized as feed additive to minimize the dependence on synthetic drugs as fertility enhancing agents.

KEYWORDS: *C. gariepinus*, medicinal plants, spermatotoxicity, sperm quality, testicular weight.

Introduction

The use of medicinal plants as a fertility enhancer in aquaculture has now received much attention [1]. With the shift away from synthetic drugs, the use of plants for enhancing growth and reproductive performance in animals and fishes is becoming acceptable [2-5]. However, some plants have been shown to have deleterious impact on aquatic organisms [6-9].

39 Aquaculture is a fast growing sector in Nigeria contributing less than 5% of
40 the total fish supply but at a growth rate of about 2% per year [10]. Among the
41 culturable fishes in Nigeria, *C. gariepinus* is a major tropical aquaculture species and
42 the most popular among fish farmers and consumers [2]. Fish farming has contributed
43 greatly to the availability of food in Nigeria and the world over with products from
44 fish farming widely exported and traded to earn income. Fisheries sector employs over
45 44.5 million people and a lot of them are from developing countries. Also, industries
46 engaged in the marketing, supply and distribution of fish product create job
47 opportunities for over 150 million individuals [2].

48 The African catfish (*Clarias gariepinus*) belongs to the family Clariidae and is
49 the most cultivated fish in Nigeria, and highly demanded freshwater fish all over the
50 world due to its resistant to stress, ability to tolerate a wide range of environmental
51 conditions, high stocking densities under culture conditions and relatively fast growth
52 [11–12]. They are found throughout Africa and the Middle East, and live in freshwater
53 lakes, rivers, and swamps, as well as human-made habitats, such as oxidation ponds or
54 even urban sewage systems. Due to the high demand of quality fish and fish dietary
55 proteins, there have been an increase in various researches in different ways to
56 improve fish fertility to meet the demand and target productivity in aquaculture, with a
57 dramatic movement from synthetic drugs to medicinal plants of natural importance.

58 Sweet potato (*Ipomoea batatas*) is a member of the family convolvulaceae with
59 almost 1650 predominately tropical species. The genus *Ipomoea* comprises the largest
60 number of the convolvulaceae family. The family is characterized by climbing or
61 twinning woody or herbaceous plants that usually have heart-shaped leaves and
62 funnel-shaped flowers [13]. *I. batatas* is a tuberous-rooted perennial plant mainly
63 grown annually. The roots are adventitious, mostly located within the top 25 cm of the
64 soil. Some of the roots produce elongated starchy tubers. Tuber flesh colour can be
65 white, yellow, orange and purple while skin colour can be red, purple, brown or white.
66 Sweet potatoes and its derivatives are powerful antioxidant and may be potent in
67 boosting the immune system and treating fever, asthma, bug bites, burns, catarrh,
68 ciguatera, convalescence, diarrhoea, nausea, stomach distress, tumors and whitlows. It
69 has also been reported to affect fertility [14–15, 16–17].

Hibiscus (Malvaceae) is a genus of herbs, shrubs and trees. Its 250 species are widely distributed in tropical and subtropical regions of the world and are reported to possess various medicinal properties viz; antitumor, antihypertensive, antioxidant, anti-ammonemic [18–22]. The flowers have been reported to possess anti-implantation and anti-spermatogenic activities [23–24]. The petroleum ether extracts of the leaves and flowers have been shown to potentiate hair growth in vivo and in vitro [25]. Leaves and flowers also possess hypoglycemic activity [26–27]. The mucilage of the leaf has anti-complementary activity [28].

Carica papaya is a soft-wooded perennial plant that has a life span of 5–10 years although commercial plantations are usually replanted [29]. It normally grows a single-stemmed tree with a crown of large palmate leaves emerging from the apex of the trunk but plant strands may become multi-stemmed when damaged. The fruit, seeds, leaves contained novel biologically active compounds which are potent as therapeutics [30]. *C. papaya* seeds have been reported to contain glycosides and polyphenols in excess among other compounds such as alkaloids, saponins, flavonoids and quinones [30].

Therefore, this study was aimed at investigating the effect of pawpaw (*Carica papaya*) seeds, hibiscus plant (*Hibiscus rosa-sinensis*) leaves, sweet potato (*Ipomoea batatas*) leaves on sperm quality and testicular weight in male African catfish (*Clarias gariepinus*).

Materials and methods

Duration and location of the study

The study was conducted for six months at the University of Calabar fish farm.

Collection and preparation of plant samples

The plant samples (seeds of *Carica papaya*, leaves of *Hibiscus rosa-sinensis* and *Ipomoea batatas*) were collected within the University of Calabar campus and authenticated in the herbarium unit of the Department of Botany, University of Calabar. The samples were washed with clean water, air dried for three weeks, ground using electric blender (Qlink-Q15L40) to get the powdery form and extracted using Soxhlet method with 70 percent ethanol as solvent. The filtrate was obtained using

rotary evaporator at 45⁰C, while the extract was reduced into pastes with hot-air oven at 60⁰c. The pastes obtained were stored in plastic screw capped bottles, labeled and stored in refrigerator for use.

Collection of fish samples

One hundred and twenty (120) juveniles of *C. gariepinus* were purchased from the University of Calabar fish farm. An average initial body weight of 46.3g, and 17.7cm length were obtained using weighing balance (Scout-pro; 3000g), and a measurement meter, respectively at the time of stocking. The fish were acclimated for 7 days in tanks and the water parameters tested to be ideal, before feeding with commercial feed (Coppens) twice daily (morning and evening) throughout the period of the experiment.

Experimental design and procedure

Experimental design and procedure

Twelve experimental tanks measuring; 80cm x 80cm x 80cm (L x W x H) were constructed with an outlet and inlet pipe in the University of Calabar fish farm hatchery complex and each tank was filled with clean water. The 120 fish were randomly divided into four experimental groups using a completely randomized design (CRD) in three replicates with each treatment containing 10 male fish. Each group had three experimental tanks containing 10 male fish giving rise to the 12 experimental tanks used and 120 animals. Sex determination was done through visual examination of the gonad. Three grams (3g) of each plant extract were incorporated into 1 kilogram of commercial feed (3g/kg; Coppens). The plants extract made up 75% of each experimental diet. The extracts were dissolved in 5ml dimethylsulphoxide (DMSO) and made into solution with water, and mixed with fish feed homogeneously using a spreader and air dried for 48 hours. This procedure was repeated for each plant and the prepared diets stored in airtight containers, labeled as follows; Treatment A-Control, B-Pawpaw seed meal (PSM), C-Hibiscus leaf meal (HLM) and D-Sweet potato leaf meal (SPLM). The physico-chemical parameters of the water were measured using the APHA [31] method of water quality assessment.

Evaluation of sperm quality

At the end of the feeding trial, 3 male fish at table size, randomly selected per dietary treatment were sacrificed under chloroform anesthesia and the testes were removed to determine the following sperm quality indices: sperm volume, motility duration, percentage motility and spermatozoa count.

Sperm volume

Small incision was made into the lobes of the testes, the sperm squeezed out into a Petri dish. This was measured with plastic syringe in mL.

Percentage motility

Each sample was estimated using light microscope at 400x magnification immediately after addition of 20 µL distilled water as an activating solution. During spermatozoa activation, Immotile Sperm Cell (ISC) was counted and when the activation stopped, Whole Sperm Cells (WSC) were counted [32]. The Motile Sperm Cells (MC) were calculated as:

$$MC = WSC - ISC$$

$$\% MC = \frac{MC}{WSC} \times 100$$

Spermatozoa count

Sperm count was determined by counting the number of spermatozoa in sample dilute with distilled water (100x) in a Burkner haemocytometer, under 400x magnification [33]

Statistical analysis

All data collected on the weight of testes, sperm volume; sperm count and sperm motility were subjected to analysis of variance (ANOVA) using predictive analysis software (PASW), version 18.0. Significant means were separated using the Least Significant Difference (LSD) at 5% probability level.

Results

Effect of *Carica papaya*, *Hibiscus rosa-sinensis* and *Ipomea batatas* on some reproductive parameters in male catfish

Testes weight

Results obtained on the effects of the different treatments are presented in Table 1. Results revealed that fish treated with extract of *H. rosa-sinensis* had significantly higher ($p = .05$) testicular weight (11.22g) when compared with the control (7.22g) and other treatment groups (6.11 and 7.56g for *C. papaya* and *I. batatas*, respectively).

Semen volume

Results presented in Table 1 also showed that sperm volume was significantly lower ($p = .05$) in animals treated with extracts of *C. papaya* and *I. batatas* (1.79 and 1.60mL, respectively) when compared with the control and *H. rosa-sinensis* groups. The highest volume of sperm was obtained in group of fish treated with *H. rosa-sinensis* (2.18mL) while the control had mean sperm volume of 1.9mL.

Semen density

There was significant reduction ($p = .05$) in the density of semen obtained from the male catfish treated with extracts of *C. papaya* and *I. batatas* (2.22×10^9 sperm/ml, respectively) when compared with the control and *H. rosa-sinensis* treated fish samples (Table 1). Sperm density was highest in the catfish treated with extracts of *H. rosa-sinensis* ($2.78^a \times 10^9$ sperm/ml) and closely followed by the control ($2.66^a \times 10^9$ sperm/ml). Sperm density of the control and *H. rosa-sinensis* groups were statistically similar.

Semen motility

No significant difference ($p = .05$) was observed in the sperm motility of fish in all the experimental groups. However, numerically, *H. rosa-sinensis* had the highest percentage of motile sperm cells (1.20%) followed by the control animals (1.08%). *C. papaya* and *I. batatas* treated animals had 1.02 and 0.98%, respectively (Table 1).

TABLE 1

Effects of *C. papaya*, *H. rosa-sinensis* and *I. batatas* on weight of testes and some sperm parameters in male catfish (*Clarias gariepinus*)

Sperm parameters	Control	<i>C. papaya</i>	<i>H. rosa-sinensis</i>	<i>I. batatas</i>
Weight of testes (g)	$7.22^b \pm 0.67$	$6.11^b \pm 0.59$	$11.22^a \pm 0.22$	$7.56^b \pm 0.48$

Sperm volume (mL)	1.91 ^a ± 0.15	1.79 ^b ± 0.87	2.18 ^a ± 0.19	1.60 ^b ± 0.07
sperm density (x10 ⁹ sperm/ml)	2.66 ^a ± 0.11	2.22 ^b ± 0.11	2.78 ^a ± 0.20	2.22 ^b ± 0.11
Semen motility (%)	1.08. ^a ± 0.00	1.02 ^a ± 0.00	1.20 ^a ± 0.00	0.98 ^a ± 0.00

187 *Means with different superscript letters along each horizontal array differ
188 significantly (p= .05)
189

190 Discussion

191 Viable sperm is an essential component of any successful animal production
192 operation and the success of reproduction process is dependent on a supply of high
193 quality gametes [34].

194 Result of the present study showed a significant reduction in the sperm
195 parameters of fish samples treated with *C. papaya* and *I. batatas* when compared with
196 the control and *Hibiscus rosa-sinensis* treated fish which agree with Ekpo et al. [8],
197 Ikpeme et al. [35] and Jegede [36]. Sperm volume and density significantly decreased
198 (p= .05) in animals treated with *C. papaya* and *I. batatas* suggesting the presence
199 phytochemicals such as saponins, alkaloids, terpenoids, flavonoids, etc. [35], that
200 might have altered the spermatogenic processes and pathways. This assertion is
201 corroborated by Elham et al. [37], Ayotunde et al. [38], Udoh and Kehinde [39], Uno
202 et al. [40] and Ekpo et al. [8,41] who reported anti-fertility properties of *C. papaya*
203 and *I. batatas*, respectively. In human, mammals and fish, the length of time and
204 intensity of spermatozoa motility, the percentage motile sperm and sperm density are
205 all parameters that have been measured in an attempt to assess sperm quality [42].

206 On the other hand, sperm count and density significantly increased in animals
207 treated with extract of *H. rosa-sinensis* which indicate the pro-fertility potential of the
208 plant. This disagrees with the findings of Jegede [36] who reported reproduction
209 inhibitory potential of *H. rosa-sinensis*. This suggests that the plant contains inherent
210 phytochemicals that might have enhanced the hormonal milieu and/or spermatogenic
211 processes in the fish. Hormonal balance and bioavailability have been shown to play
212 very important role in spermatogenesis [43-44].

The Results obtained also showed that testicular weight significantly reduced (p= .05) in *C. papaya* and *I. batatas* treated animals which suggest distortions in the testicular integrity of the affected fish samples. The decrease in the testicular weight of *C. papaya* and *I. batatas* treated animals support the concomitant decrease observed in the sperm volume and density of the same fish.

Moreover, there was no significant difference in the sperm count of the fish in all the treatment groups. This suggests that the plant did not significantly alter the vigor of the sperm cells when compared with the control. However, numerically, animals treated with *H. rosa-sinensis* had the highest sperm count while those treated *C. papaya* and *I. batatas* had the lowest which is also in line with result of other parameters studied. The fertility enhancing property of *H. rosa-sinensis* may be attributed to the antioxidant properties of the of the plant extract against oxidative stress, which has been implicated in altering the production of hormones necessary for spermatogenesis with its concomitant effect on fertility[4,45]. Turner [46] and Adeparusi *et al.* [2] reported that spermatozoa motility varies in rigor and duration not only among male but also within an individual male depending on the ripeness, age and time of sampling.

Conclusion

In conclusion, this study reveals the pro-fertility potential of Hibiscus leaf meal in male *C. gariepinus* while *C. papaya* and *I. batatas* possess anti-fertility properties. This suggests that *H. rosa-sinensis* has the potential to enhance fertility in male *C. gariepinus* while *C. papaya* and *I. batatas* are toxic to fertility in male *C. gariepinus*. Therefore, future studies should focus on the enhancement of seedling production strategies for different fishes using *H. rosa-sinensis* since the main objective of fish farming is to improve fish production and this plant has a promising pro-fertility property which can be harnessed in aquaculture.

COMPETING INTEREST

Authors have declared that no competing interests exist.

Ethical Approval:

All authors hereby declare that "Principles of laboratory animal care" (NIH publication No. 85-23, revised 1985) were followed, as well as specific national laws where applicable. All experiments have been examined and approved by the appropriate ethics committee.

Consent: NA

References

1. Dada AA and Ajilore VO. Use of ethanol extracts of *Garcinia Kola* as fertility enhancer in female catfish *Clarias gariepinus* broodstock. *Interenational Journal of Fisheries and Aquaculture*, 2009; 1(1):1-5.
2. Adeparusi EO, Dada AA and Alale OV,. Effects of medicinal plant (*Kigelia Africana*) on sperm quality of African catfish (*Clarias gariepinus*) broodstock. *Journal of Agricultural. Science*, 2010; 2: 193 – 199
3. Ekaluo UB, Uno UU, Edu NE, Ekpo PB, Etta SE and Volunteer BO. Protective role of onion (*Allium cepa*) on caffeine induced spermatotoxicity in albino rats. *Journal of Applied Life Science International*, 2016; 4(4): 1-7.
4. Ekaluo UB, Ikpeme EV, Uno UU, Umeh SO and Erem FA. Protective effect of aqueous guava leaf extract on caffeine induced spermatotoxicity in albino rats. *Research Journal of Medicinal Plant*, 2016; 10(1): 98-105.
5. Uno UU, Ogbe HO, Okolo CM, Ekaluo, UB. and Akwa BU. Effect of soursop (*Annona muricata* L.) leaf extract on sperm toxicity induced by caffeine in albino rats. *The Pharmaceutical and Chemical Journal*, 2017; 4(1): 82 – 87.
6. Ekpo PB, Uno UU, Okolo CM, Agu RB and Onwudike CF. Acute toxicity of *Adenia cissampeloides* in Farmed African Catfish (*Clarias gariepinus*). *Annual Research & Review in Biology*, 2017; 5: 1-5
7. Ekpo PB, Uno UU, Ogbe HO and Ekaluo UB. Effect of Sweet Potato (*Ipomoea batatas*) Tuber on Sperm Profile and Testicular Integrity of Male Albino Rats. *Archives of Current Research International*, 2017; 9(2): 1-7.
8. Ekpo PB, Uno UU, Adilieje CM, Umoyen AJ, Okey FO. Reproductive performance of *Carica papaya*, *Hibiscus rosa-sinensis* and *Ipomoea batatas* on female African catfish (*Clarias gariepinus*). *Biotechnology Journal International*, 2018; 21(2): 1-8.
9. Uno UU, Ekpo PB, Onwudiwe CF and Agu RC. Comparative Acute Toxicity of Ichthyotoxic Plants (*Tephrosia vogelii*, *Adenia cissampeloides* and *Asystasia vogeliana*) on Farmed African Catfish (*Clarias gariepinus*). *Asian Journal of Biology*, 2018; 5(4): 1-7
10. Moses BS. Fisheries and Ecotourism: A Tool for National Development. *Proceedings of the fisheries society of Nigeria (FISON)* (E.I. Chukwu; P.O. Ajah; D.A. Aina- Abasi and F.M. Nwogu ed). p412, 2006.

11. James R, Sampath K. Effect of meal frequency on growth and reproduction in the ornamental red swordtail, *Xiphophorus helleri*. *Israel Journal of Aquaculture*, 2003; 55:197 – 202.
12. Eyo VO, and Ekanem, A. Effect of feeding frequency on the growth, food utilization and survival of African catfish (*Clarias gariepinus*) using locally formulated diets. *African Journal Environmental Pollution Health*, 2011; 9(2): 11-17
13. Uno UU, Ekpo PB, Ogbe HO, Okolo CM, Ekaluo UB. Effect of soursop (*Annona muricata* L.) leaf extract on oxidative stress caused by caffeine in albino rat model. *Asian Journal of Biology*, 2011; 1(2): 1 – 7.
14. Austin DF, Huaman ZA. Synopsis of *Ipomoea* (Convolvulaceae) in the Americas. *Taxon*, 1996; 45: 3 – 38
15. Duke J, Wain K. Medicinal plant of the World, Vol. 3 Computer index with more than 85,000 entries. Plant genetics and germplasm Institute. Agriculture Research service, Beltsville, Maryland, 2008; 231-239.
16. Uno UU, Ekpo PB, Okolo CM and Ekaluo UB. Comparative Effects of Sweet Potato (*Ipomoea batatas*) Leaf and Tuber on Male Albino Rats. *Asian Journal of Research in Medical and Pharmaceutical Sciences*, 2017; 1(2): 1-7.
17. Hou DX, Tong X, Terahara N, Lou D, Fujii M. Delphinidin-3-sambubioside, a Hibiscus anthocyanin, induces apoptosis in human leukemia cells through reactive oxygen species-mediated mitochondrial pathway. *Arch. Biochem. Biophys.*, 2005; 440:101–9.
18. Hirunpanich V, Utaipat A, Morales NP, Bunyapraphatasara N, Sato H, Herunsalee A, et al. Hypocholesteremic and antioxidant effect of the aqueous extracts of *Hibiscus sabdariffa* Linn. In hypercholesteremic rats. *J. Ethanopharmacol.*, 2006; 103:252–60.
19. Chang YC, Haung KX, Haung AC, HO YC, Wang CJ. Hibiscus anthocyanins-rich extract inhibited LDL. Oxidation and oxLDL-mediated macrophages apoptosis. *Food Chem Toxicol.*, 2006; 44:1015–23.
20. Herrera AA, Flores RS, Chavez-Soto MA, Tortoriello J. Effectiveness and tolerability of a standardized extract from *Hibiscus sabdariffa* in patients with mild to moderate hypertension: a controlled and randomized clinical trial. *Phytomedicine.*, 2004; 11:375–82.
21. Mohamed Essa M, Subramanian P. *Hibiscus sabdariffa* affects ammonium chloride-induced hyperammonemic rats. *Evid. Based Complement. Altern. Med.*, 2007; 4:321–326.
22. Mudgal VN. Botanical description of *Hibiscus rosa-sinensis* (China rose or shoe flower or japakusum). *J. Res. Indian Med.*, 1974; 9:105.
23. Murthy DRK, Reddy CM, Patil SB. Effect of benzene extract of *Hibiscus rosa-sinensis* on the oestrous cycle and ovarian activity in albino mice. *Biol. Pharm. Bull.*, 1997; 20:756–8
24. Adhirajan N, Ravi kumar T, Shanmugasundram N, Babu M. in vivo and in vitro evaluation of hair growth potential of *Hibiscus rosa-sinensis* Linn. *J. Ethanopharmacol.*, 2003; 88: 235–9.

25. Sachdewa A, Nigam R, Khemani LD. Hypoglycemic effects of *Hibiscus rosa-sinensis* L. leaf extract in glucose and streptozotocin induced hyperglycemic rats. Indian J Exp Biol 2001; 39:284–6.
26. Sachdewa A, Khemani LD. Effect of *Hibiscus rosa-sinensis* Linn. ethanol flower extract on blood glucose and lipid profile in streptozotocin induced diabetes in rats. J. Ethnopharmacol., 2003; 89:61–6.
27. Hiremath SP, Rao SH, Jain PK, Jaya Y, Sembulingam K. Antifertility activity of *Striga lutea* – part I. Indian J. Physiol. Pharmacol., 1990; 34: 23–5.
28. Clay – Prove P, Ross P, O’Hare N, Macleod N and Kernot I. Agrilink series: your growing guide to better farming. Queensland Horticulture Institute, Nambour.
29. Adebiyi A, Adaikan PG, and Prasad RNV. Papaya (*Carica papaya*) consumption and usage in some parts of Asia using a rat model. Br. J. Nutr., 2002; 88: 99 – 203.
30. American Public Health Association (APHA). Standard method for the examination of water and waste. New York: APHA press; 1988.
31. Musa YM, Haruna AK, Ilyas AH, Yaro AA, Ahmadu AA and Usman H. Phytochemical, analgesic and anti-inflammatory effects of the ethylacetate extract of the leaves of *Pseudocedrella kotschyii*. Afr. J. Trad. Complemetary Altern. Med., 2008; 5: 92 – 96.
32. Canyurt MA. and Akhan S. Effect of Ascorbic Acid Supplementation on Sperm Quality of Rainbow Trout (*Oncorhynchus mykiss*). Turkish Journal and Aquatic Sciences, 2008; 8:171-175.
33. Rainis S, Mylonas CC, Kyriakou Y, and Divanach P. Enhancement of spermiation in European sea bass (*Dicentrarchus labrax*) at the end of the reproductive season using GnRHa implants. Aquaculture, 2003; 19: 873-890.
34. Cruz-Casallas PE, Lombo-Rodriguez DA and Velasco- Santamaria YM. Milt quality and spermatozoa morphology of captive *Brycon siebenthalee* (Eigenmann) broodstock. Aquaculture Research, 2005; 36: 628-688.
35. Ikpeme EV, Udensi O, Ekaluo UB, Uyoh EA, Asuquo BO, Udoh FV and Udoh PB. Effect of crude extract of *Carica papaya* seeds on the reproductive efficiency of male albino rats. Global J. Pure Applied Sci., 2007; 13: 365-368.
36. Jegede T. Control of reproduction in *Oreochromis niloticus* (Linnaeus 1758) using *Hibiscus rosa-sinensis* (Linn.) leaf meal as reproduction inhibitor. Journal of Agricultural Science 2010; 2(4):149-154.
37. Elham MA, Fedekar FM and Ashraf MI. Effect of pawpaw (*Carica papaya*) seeds on the reproductive performance and histological characteristic of gonads in Nile tilapia (*Oreochromis niloticus*). Indian Journal of Applied Research, 2013; 3:12
38. Ayotunde EO and Offem BO. Acute and chronic toxicity of pawpaw to adult Nile tilapia (*Oreochromis niloticus* (Linn 1757). African Journal of Biotechnology, 2008; 7(13): 2265-74.
39. Udo P and Kehinde A. Studies of antifertility effect of pawpaw seeds (*Carica papaya*) on the gonada of male albino rats. Phytother. Res., 1999; 13: 226 – 228.

- 375 40. Uno UU, Okolo CM, Ogbe HO, Ekaluo UB. and Benjamin ME. Evaluation of
376 Spermatotoxic Effect of Sweet Potato (*Ipomoea batatas*) Leaf Extract on Male
377 Albino Rats. Asian Journal of Biotechnology and Bioresource Technology,
378 2017; 1(3): 1-5.
- 379 41. Ekpo PB, Uno UU, Ogbe HO and Ekaluo UB. Effect of Sweet Potato (*Ipomoea*
380 *batatas*) Tuber on Sperm Profile and Testicular Integrity of Male Albino Rats.
381 Archives of Current Research International, 2017; 9(2): 1-7.
- 382 42. Billard R and Cosson MP. Some problems related to the assessment of sperm
383 motility in freshwater fish. J. Exp. Zool., 1992; 261: 122-131.
- 384 43. Uno UU, Ekaluo UB, Okoi PE, Ogbe HO. and Peter N. Mitigating role of
385 Trevo dietary supplement on hormonal toxicity induced by caffeine in albino
386 rats. International Journal of Advance Research, 2015; 3(11): 586 – 590.
- 387 44. Ono Y, Suzuki K, Kashiwagi B, Shibata Y, Ito K and Fukabori Y. Role of
388 androgen on blood flow and capillary structure in rat seminal vesicles. Tohoku
389 Journal of Experimental Medicine, 2004; 202:193-201.
- 390 45. Makker K, Agarwal A, Sharma R. Oxidative stress and male infertility. Indian
391 Journal of Medical Research, 2009; 129: 357-367.
- 392 46. Turner C. Evaluation of salmonid sperm motility for cryopreservation. Prog.
393 Fish Cult., 1986; 48: 230-232.