

Original Research Article

Spermatotoxic effects of some medicinal plants (*Carica papaya*, *Hibiscus rosa-sinensis* and *Ipomoea batatas*) on sperm quality and testicular weight in male African catfish (*Clarias gariepinus*)

ABSTRACT

This study was designed to evaluate the effects of pawpaw (*Carica papaya*) seeds, hibiscus plant (*Hibiscus rosa-sinensis*) leaves, sweet potato (*Ipomoea batatas*) leaves on sperm quality (sperm motility, sperm density, semen volume) and weight of testes of male *Clarias gariepinus*. One hundred and twenty (120) juveniles of *C. gariepinus* were collected from the University of Calabar fish farm. The 120 fish were randomly divided into 12 experimental tanks measuring 80x80x80cm (L x W x H), with three tanks for each treatment, using a completely randomized design (CRD). Three grams (3g) of each test plant were incorporated into 1kg of Coppens feed (3g/kg) and reformulated into four experimental diets; Treatment A- Control, B- Pawpaw seed meal (PSM), C-Hibiscus leaf meal (HLM) and D- sweet potato leaf meal (SPLM). The experiment was done in three replications. The fish were fed twice daily for 6 months. Data obtained were analyzed using a one way analysis of variance (ANOVA). Results showed that fish fed with HLM had significantly ($p = .05$) higher testicular weight when compared with the control and other test plants. Moreover, sperm volume and density significantly ($p = .05$) reduced in fish samples treated with PSM and SPLM when compared with the control and fish fed with HLM. The highest mean sperm volume and density were obtained in fish samples fed with HLM. No significant difference was observed in the sperm motility of the fish in all the treatment groups. Conclusively, this study reveals the pro-fertility potential of *H. rosa-sinensis* in male *C. gariepinus* while *C. papaya* and *I. batatas* possess anti-fertility properties. Therefore, HLM can be utilized as feed additive to minimize the dependence on synthetic drugs as fertility enhancing agents.

KEYWORDS: *C. gariepinus*, medicinal plants, spermatotoxicity, sperm quality, testicular weight.

Introduction

The use of medicinal plants as a fertility enhancer in aquaculture has now received much attention [1]. With the shift away from synthetic drugs, the use of plants for enhancing growth and reproductive performance in animals and fishes is becoming acceptable [2-5]. However, some plants have been shown to have deleterious impact on aquatic organisms [6-9].

39 Aquaculture is a fast growing sector in Nigeria contributing less than 5% of
40 the total fish supply but at a growth rate of about 2% per year [10]. Among the
41 culturable fishes in Nigeria, *C. gariepinus* is a major tropical aquaculture species and
42 the most popular among fish farmers and consumers [2]. Fish farming has contributed
43 greatly to the availability of food in Nigeria and the world over with products from
44 fish farming widely exported and traded to earn income. Fisheries sector employs over
45 44.5 million people and a lot of them are from developing countries. Also, industries
46 engaged in the marketing, supply and distribution of fish product create job
47 opportunities for over 150 million individuals [2].

48 The African catfish (*Clarias gariepinus*) belongs to the family Clariidae and is
49 the most cultivated fish in Nigeria, and highly demanded freshwater fish all over the
50 world due to its resistant to stress, ability to tolerate a wide range of environmental
51 conditions, high stocking densities under culture conditions and relatively fast growth
52 [11–12]. They are found throughout Africa and the Middle East, and live in freshwater
53 lakes, rivers, and swamps, as well as human-made habitats, such as oxidation ponds or
54 even urban sewage systems. Due to the high demand of quality fish and fish dietary
55 proteins, there have been an increase in various researches in different ways to
56 improve fish fertility to meet the demand and target productivity in aquaculture, with a
57 dramatic movement from synthetic drugs to medicinal plants of natural importance.

58 Sweet potato (*Ipomoea batatas*) is a member of the family convolvulaceae with
59 almost 1650 predominately tropical species. The genus *Ipomoea* comprises the largest
60 number of the convolvulaceae family. The family is characterized by climbing or
61 twinning woody or herbaceous plants that usually have heart-shaped leaves and
62 funnel-shaped flowers [13]. *I. batatas* is a tuberous-rooted perennial plant mainly
63 grown annually. The roots are adventitious, mostly located within the top 25 cm of the
64 soil. Some of the roots produce elongated starchy tubers. Tuber flesh colour can be
65 white, yellow, orange and purple while skin colour can be red, purple, brown or white.
66 Sweet potatoes and its derivatives are powerful antioxidant and may be potent in
67 boosting the immune system and treating fever, asthma, bug bites, burns, catarrh,
68 ciguatera, convalescence, diarrhoea, nausea, stomach distress, tumors and whitlows. It
69 has also been reported to affect fertility [14–15, 16–17].

70 Hibiscus (Malvaceae) is a genus of herbs, shrubs and trees. Its 250 species are
71 widely distributed in tropical and subtropical regions of the world and are reported to
72 possess various medicinal properties viz; antitumor, antihypertensive, antioxidant,
73 anti- amnonemic [18–22]. The flowers have been reported to possess anti-
74 implantation and anti – spermatogenic activities [23–24]. The petroleum ether extracts
75 of the leaves and flowers have been shown to potentiate hair growth in vivo and in
76 vitro [25]. Leaves and flowers also possess hypoglycemic activity [26-27]. The
77 mucilage of the leaf has anti – complementary activity [28].

78 *Carica papaya* is a soft-wooded perennial plant that has a life span of 5–10
79 years although commercial plantations are usually replanted [29]. It normally grows a
80 single – stemmed tree with a crown of large palmate leaves emerging from the apex of
81 the trunk but plant strands may become multi-stemmed when damaged. The fruit,
82 seeds, leaves contained novel biologically active compounds which are potent as
83 therapeutics [30]. *C. papaya* seeds have been reported to contain glycosides and
84 polyphenols in excess among other compounds such as alkaloids, saponins, flavonoids
85 and quinones [30].

86 Therefore, this study was aimed at investigating the effect of pawpaw (*Carica*
87 *papaya*) seeds, hibiscus plant (*Hibiscus rosa-sinensis*) leaves, sweet potato (*Ipomoea*
88 *batatas*) leaves on sperm quality and testicular weight in male African catfish (*Clarias*
89 *gariiepinus*).

90

91 **Materials and methods**

92 **Duration and location of the study**

93 The study was conducted for six months at the University of Calabar fish farm.

94 **Collection and preparation of plant samples**

95 The plant samples (seeds of *Carica papaya*, leaves of *Hibiscus rosa-sinensis*
96 and *Ipomoea batatas*) were collected within the University of Calabar campus and
97 authenticated in the herbarium unit of the Department of Botany, University of
98 Calabar. The samples were washed with clean water, air dried for three weeks, ground
99 using electric blender (Qlink-Q15L40) to get the powdery form and extracted using
100 Soxhlet method with 70 percent ethanol as solvent. The filtrate was obtained using

101 rotary evaporator at 45⁰C, while the extract was reduced into pastes with hot-air oven
102 at 60⁰c. The pastes obtained were stored in plastic screw capped bottles, labeled and
103 stored in refrigerator for use.

104 **Collection of fish samples**

105 One hundred and twenty (120) juveniles of *C. gariepinus* were purchased from
106 the University of Calabar fish farm. An average initial body weight of 46.3g, and
107 17.7cm length were obtained using weighing balance (Scout-pro; 3000g), and a
108 measurement meter, respectively at the time of stocking. The fish were acclimated for
109 7 days in tanks and the water parameters tested to be ideal, before feeding with
110 commercial feed (Coppens) twice daily (morning and evening) throughout the period
111 of the experiment.

112 **Experimental design and procedure**

113 **Experimental design and procedure**

114 Twelve experimental tanks measuring; 80cm x 80cm x 80cm (L x W x H) were
115 constructed with an outlet and inlet pipe in the University of Calabar fish farm
116 hatchery complex and each tank was filled with clean water. The 120 fish were
117 randomly divided into four experimental groups using a completely randomized
118 design (CRD) in three replicates with each treatment containing 10 male fish. Each
119 group had three experimental tanks containing 10 male fish giving rise to the 12
120 experimental tanks used and 120 animals. Sex determination was done through visual
121 examination of the gonad. Three grams (3g) of each plant extract were incorporated
122 into 1 kilogram of commercial feed (3g/kg; Coppens). The plants extract made up
123 75% of each experimental diet. The extracts were dissolved in 5ml
124 dimethylsulphoxide (DMSO) and made into solution with water, and mixed with fish
125 feed homogeneously using a spreader and air dried for 48 hours. This procedure was
126 repeated for each plant and the prepared diets stored in airtight containers, labeled as
127 follows; Treatment A-Control, B-Pawpaw seed meal (PSM), C-Hibiscus leaf meal
128 (HLM) and D-Sweet potato leaf meal (SPLM). The physico-chemical parameters of
129 the water were measured using the APHA [31] method of water quality assessment.

130 **Evaluation of sperm quality**

131 At the end of the feeding trial, 3 male fish at table size, randomly selected
132 per dietary treatment were sacrificed under chloroform anesthesia and the testes
133 were removed to determine the following sperm quality indices: sperm volume,
134 motility duration, percentage motility and spermatozoa count.

135 **Sperm volume**

136 Small incision was made into the lobes of the testes, the sperm squeezed
137 out into a Petri dish. This was measured with plastic syringe in mL.

138 **Percentage motility**

139 Each sample was estimated using light microscope at 400x magnification
140 immediately after addition of 20 µL distilled water as an activating solution.
141 During spermatozoa activation, Immotile Sperm Cell (ISC) was counted and
142 when the activation stopped, Whole Sperm Cells (WSC) were counted [32]. The
143 Motile Sperm Cells (MC) were calculated as:

144

$$MC = WSC - ISC$$

$$\% MC = \frac{MC}{WSC} \times 100$$

145 **Spermatozoa count**

146 Sperm count was determined by counting the number of spermatozoa in
147 sample dilute with distilled water (100x) in a Burker haemocytometer, under
148 400x magnification [33]

149

150 **Statistical analysis**

151 All data collected on the weight of testes, sperm volume; sperm count and
152 sperm motility were subjected to analysis of variance (ANOVA) using predictive
153 analysis software (PASW), version 18.0. Significant means were separated using the
154 Least Significant Difference (LSD) at 5% probability level.

155

156 **Results**

157 **Effect of *Carica papaya*, *Hibiscus rosa-sinensis* and *Ipomea batatas* on some**
158 **reproductive parameters in male catfish**

159 **Testes weight**

160 Results obtained on the effects of the different treatments are presented in
161 Table 1. Results revealed that fish treated with extract of *H. rosa-sinensis* had
162 significantly higher ($p= .05$) testicular weight (11.22g) when compared with the
163 control (7.22g) and other treatment groups (6.11 and 7.56g for *C. papaya* and *I.*
164 *batatas*, respectively).

165 **Semen volume**

166 Results presented in Table 1 also showed that sperm volume was significantly
167 lower ($p=.05$) in animals treated with extracts of *C. papaya* and *I. batatas* (1.79 and
168 1.60mL, respectively) when compared with the control and *H. rosa-sinensis* groups.
169 The highest volume of sperm was obtained in group of fish treated with *H. rosa-*
170 *sinensis* (2.18mL) while the control had mean sperm volume of 1.9mL.

171 **Semen density**

172 There was significant reduction ($p= .05$) in the density of semen obtained from
173 the male catfish treated with extracts of *C. papaya* and *I. batatas* (2.22 and 2.22
174 $\times 10^9$ sperm/ml, respectively) when compared with the control and *H. rosa-sinensis*
175 treated fish samples (Table 1). Sperm density was highest in the catfish treated with
176 extracts of *H. rosa-sinensis* ($2.78^a \times 10^9$ sperm/ml) and closely followed by the control
177 ($2.66^a \times 10^9$ sperm/ml). Sperm density of the control and *H. rosa-sinensis* groups were
178 statistically similar.

179 **Semen motility**

180 No significant difference ($p= .05$) was observed in the sperm motility of fish in
181 all the experimental groups. However, numerically, *H. rosa-sinensis* had the highest
182 percentage of motile sperm cells (1.20%) followed by the control animals (1.08%). *C.*
183 *papaya* and *I. batatas* treated animals had 1.02 and 0.98%, respectively (Table 1).

184 **TABLE 1**

185 Effects of *C. papaya*, *H. rosa-sinensis* and *I. batatas* on weight of testes and some
186 sperm parameters in male catfish (*Clarias gariepinus*)

Sperm parameters	Control	<i>C. papaya</i>	<i>H. rosa-sinensis</i>	<i>I. batatas</i>
Weight of testes (g)	$7.22^b \pm 0.67$	$6.11^b \pm 0.59$	$11.22^a \pm 0.22$	$7.56^b \pm 0.48$

Sperm volume (mL)	1.91 ^a ± 0.15	1.79 ^b ± 0.87	2.18 ^a ± 0.19	1.60 ^b ± 0.07
sperm density (x10 ⁹ sperm/ml)	2.66 ^a ± 0.11	2.22 ^b ± 0.11	2.78 ^a ± 0.20	2.22 ^b ± 0.11
Semen motility (%)	1.08. ^a ± 0.00	1.02 ^a ± 0.00	1.20 ^a ± 0.00	0.98 ^a ± 0.00

187 *Means with different superscript letters along each horizontal array differ
 188 significantly (p= .05)

189

190 Discussion

191 Viable sperm is an essential component of any successful animal production
 192 operation and the success of reproduction process is dependent on a supply of high
 193 quality gametes [34].

194 Result of the present study showed a significant reduction in the sperm
 195 parameters of fish samples treated with *C. papaya* and *I. batatas* when compared with
 196 the control and *Hibiscus rosa-sinensis* treated fish which agree with Ekpo et al. [8],
 197 Ikpeme et al. [35] and Jegede [36]. Sperm volume and density significantly decreased
 198 (p= .05) in animals treated with *C. papaya* and *I. batatas* suggesting the presence
 199 phytochemicals such as saponins, alkaloids, terpenoids, flavonoids, etc. [35], that
 200 might have altered the spermatogenic processes and pathways. This assertion is
 201 corroborated by Elham et al. [37], Ayotunde et al. [38], Udoh and Kehinde [39], Uno
 202 et al. [40] and Ekpo et al. [8,41] who reported anti-fertility properties of *C. papaya*
 203 and *I. batatas*, respectively. In human, mammals and fish, the length of time and
 204 intensity of spermatozoa motility, the percentage motile sperm and sperm density are
 205 all parameters that have been measured in an attempt to assess sperm quality [42].

206 On the other hand, sperm count and density significantly increased in animals
 207 treated with extract of *H. rosa-sinensis* which indicate the pro-fertility potential of the
 208 plant. This disagrees with the findings of Jegede [36] who reported reproduction
 209 inhibitory potential of *H. rosa-sinensis*. This suggests that the plant contains inherent
 210 phytochemicals that might have enhanced the hormonal milieu and/or spermatogenic
 211 processes in the fish. Hormonal balance and bioavailability have been shown to play
 212 very important role in spermatogenesis [43-44].

213 The Results obtained also showed that testicular weight significantly reduced
214 ($p = .05$) in *C. papaya* and *I. batatas* treated animals which suggest distortions in the
215 testicular integrity of the affected fish samples. The decrease in the testicular weight
216 of *C. papaya* and *I. batatas* treated animals support the concomitant decrease observed
217 in the sperm volume and density of the same fish.

218 Moreover, there was no significant difference in the sperm count of the fish in
219 all the treatment groups. This suggests that the plant did not significantly alter the
220 vigor of the sperm cells when compared with the control. However, numerically,
221 animals treated with *H. rosa-sinensis* had the highest sperm count while those treated
222 *C. papaya* and *I. batatas* had the lowest which is also in line with result of other
223 parameters studied. The fertility enhancing property of *H. rosa-sinensis* may be
224 attributed to the antioxidant properties of the of the plant extract against oxidative
225 stress, which has been implicated in altering the production of hormones necessary for
226 spermatogenesis with its concomitant effect on fertility[4,45]. Turner [46] and
227 Adeparusi *et al.* [2] reported that spermatozoa motility varies in rigor and duration not
228 only among male but also within an individual male depending on the ripeness, age
229 and time of sampling.

230 **Conclusion**

231 In conclusion, this study reveals the pro-fertility potential of Hibiscus leaf
232 meal in male *C. gariepinus* while *C. papaya* and *I. batatas* possess anti-fertility
233 properties. This suggests that *H. rosa-sinensis* has the potential to enhance fertility in
234 male *C. gariepinus* while *C. papaya* and *I. batatas* are toxic to fertility in male *C.*
235 *gariepinus*. Therefore, future studies should focus on the enhancement of
236 seedling production strategies for different fishes using *H. rosa-sinensis* since the
237 main objective of fish farming is to improve fish production and this plant has a
238 promising pro-fertility property which can be harnessed in aquaculture.

239

240 **COMPETING INTEREST**

241 Authors have declared that no competing interests exist.

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245 **Ethical Approval:**

246 All authors hereby declare that "Principles of laboratory animal care" (NIH
247 publication No. 85-23, revised 1985) were followed, as well as specific national laws
248 where applicable. All experiments have been examined and approved by the
249 appropriate ethics committee.

250

251 **Consent: NA**

252

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