Original Research Article 1 PHYTOCHEMICAL SCREENING AND 2 ANTIMICROBIAL ACTIVITY OF LEAVES 3 AND STEM-BARK AQUEOUS EXTRACTS OF 4 DIOSPYROS MESPILIFORMIS 5 6 7 ABSTRACT The leaves and stem-bark extracts of Diospyros mespiliformis from Ebanaceae family, which 8 is used as herbal remedies for the cure of many ailments by natives in northern part of 9 10 Nigeria, were collected from Mubi in Mubi North Local Government area of Adamawa State, air dried, pulvurised, extracted by simple overnight maceration techniques and analyzed. 11 Aqueous extracts of the aforementioned parts of the plant were screened phytochemically for 12

its chemical constituents and subjected to antimicrobial activity against Escherichia coli, 13 14 Pseudomonas aeruginosa, Shigella spp, Staphylococcus aureus, Streptococcus pneumonia, Streptococcus pyogenes and Salmonella typhi. The results show that alkaloids, 15 16 anthraquinones, carbohydrates, flavonoids, phlobatannins, saponins, steroids, tannins and terpenoids are present in both the leaves and the stem-bark extracts of the plant while 17 18 glycosides is present in the leaves extract but absent in the stem-bark extract. The antimicrobial activity reveals that, both the leaves and the stem-bark extracts of the plant 19 showed high sensitivity to Escherichia coli, Pseudomonas aeruginosa, Shigella spp, 20 21 Staphylococcus aureus, Streptococcus pneumonia, Streptococcus pyogenes and Salmonella 22 typhi. The phytochemical constituents and the antimicrobial potential of the plant part may account for varied enthnobotanical uses of the plants in traditional medicine in Nigeria, 23 24 which if further purified can be used to source novel antibiotics.

25 Keywords: *Diospyros mespiliformis*, Phytochemical Screening, Antimicrobial,
26 Ethnobotanical

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28 INTRODUCTION

29 Diospyros mespiliformis is a tall, evergreen tree 15-20 m high, with dense, rounded and buttressed stem belonging to the family Ebenaceae. It is known in Hausa as "Kanya" and 30 31 in Yoruba as "Igidudu". It is commonly called Jackal-berry or African ebony. It is found in 32 Savannah and Northern low land forest [1]. The leaves are simple, alternate dark green with 33 small hairs on the underside of older leaves. D. mespiliformis is dioecious, pollinated by bees and flowers in the rainy season while fruit ripening, which coincides with the dry season 34 takes place 6-8 months after flower fertilization [2]. The edible it of this plant is a 35 traditional food of high nutritive value in Africa that is either used fresh in fermented drink or 36 37 dried and stored for later use [3]. D. mespiliformis is reportedly one of the most important genera of *Ebenaceae* which species have been used over the millennia in traditional 38

UNDER PEER REVIEW

39 medicinal systems [4]. The leaves, roots, stem-bark and fruits contain antibiotic qualities and 40 posses many medicinal uses in West Africa. Roots and stem-bark is used to facilitate 41 childbirth, and are used for infections such as pneumonia, syphilis, leprosy, dermatomycoses, 42 as an anthelmintic [5] while the leaves decoction is used as a remedy for fever, otitis and 43 wound dressing agent [2]. Different parts of the plant are used against diarrhea, headache, 44 toothache and as a psycho-pharmacological drug. The leaves are eaten by elephants, giraffe, 45 black rhino, baboons while the wood, which is hard, strong, fungal and termite resistant, is 46 used for construction purposes, furniture, carvings, walking sticks, stamping blocks, pestles 47 and makes good firewood and charcoal [6, 7]. Plant parts, which have one or more of its organs containing substances that can be 48 49 used for therapeutic purpose, are called medicinal plants [8]. Ethnobotanical studies carried 50 out throughout Africa confirm that indigenous plants are the main constituents of traditional 51 African medicines [9, 10]. 52 Phytochemical is a non - nutritive plant chemical compounds produced by plants, generally to help them thrive or thwart competitors, predators or pathogens [11] and protect 53 54 them against environmental stresses like cold and heat [1]. Phytochemical compounds have 55 curative and prophylactic properties against a wide range of human and animal diseases [12]. 56 There are many phytochemical in fruits, leaves, stem-bark and roots of plants, each of which 57 may cure or prevent a disease singly or in synergism with some others [1]. Phytochemical 58 screening is of great importance in providing information about chemicals found in a plant in 59 term of their nature and range of occurrence [13]. The plant *D. mespiliformis* possess a number of medicinal uses, hence the need to 60

phytochemically screen the plant extracts to establish its phytochemical profile for the better
prediction and easier confirmation of the medicinal value of this plant and to test the plant
extract against organisms known to be the causes of these diseases, so as to see if their
growth can be inhibited in order to prove their folk uses and claims by traditional healers in
Nigeria.

66 MATERIALS AND METHODS

67 Sampling and sample preparation

Fresh leaves and stems-bark of *D. mespiliformis* were collected from Mubi in
Mubi North Local Government area of Adamawa State and identified by Prof. Mohammed S.
of Department of Biological Sciences, Adamawa State University Mubi, Adamawa State,
Nigeria. The identified plant parts were cleaned under a running tap water before being algorithm.

dried at room temperature and pulverized, using a clean, sterile mortar and pestle into fine
particles suitable for extraction. The pulverized leaves and stem-bark samples were each
collected into sterile labeled cellophane bags to prevent mix up.

75 **Preparation of the plant extract**

76 Extraction was carried out for the leaves and stem-bark of D. Mespiliformis by using overnight maceration techniques according to the method reported in [14]. About 50 g each 77 78 of leaves and stem-bark material were macerated in 250 ml of water in a flask. Each of the 79 soaked samples was stirred, sealed with aluminium foil and allowed to stand for 72 hours at 80 room temperature and the supernatant decanted. Thereafter, the supernatant was filtered using 81 Whatman No. 1 filter paper followed by evaporation at 45°C using a rotary evaporator at a 82 minimum pressure. The extracts were kept and stored in a refrigerator at 4°C until further 83 analysis.

84 Phytochemical screening

Qualitative phytochemical analysis of the leaves and stem-bark extract of *D. mespiliformis* was carried out using standard qualitative methods as described by [14,15]. Alkaloids, anthraquinones, carbohydrates, flavonoids, glycosides, phlobatannins, saponins, steroids, tannins and terpenoids tests were conducted in all the extracts and the results are shown in Table 1.

90 Test Organism

The bacterial and fungal strains used for the investigation are:- *Escherichia coli*(EC114ADSUC), *Pseudomonas aeruginosa* (PA214ADSUC), *Shigella spp* (SS314ADSUC), *Staphylococcus aureus* (SA414ADSUC), *Streptococcus pneumonia* (SP514ADSUC), *Streptococcus pyogenes* (SP614ADSUC) and *Salmonella typhi* (ST714ADSUC).

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Antimicrobial investigation

The microorganisms were clinically isolated from Adamawa State University Clinic, Mubi. The stocks bacterial cultures were maintained at 4°C on nutrient agar slant and subculture in nutrient broth for incubation at 37°C for 24 hours prior to each antimicrobial testing. Inoculation of the test organisms on nutrient agar prepared plates was achieved by flaming a wire loop on a spirit lamp, cooling the wire loop (air cooling) and fetching the test organisms. The discs were prepared using a Whatman No. 3 filter paper and putting in vialsbottles and sterilizing in an oven at 150°C for 15 minutes. Prepared discs containing the various extracts were carefully placed on the inoculated plates using a sterilized forceps in each case. The plates were then turned upside-down and inoculate at 37° C for 24 hours in an incubator [16]. After incubation, the inoculated plates were observed for zones of inhibition (in mm diameter). Ampiclox (10μ g/mL) was used as the control. The result was taken by considering the zone of growth and inhibition of the organisms by the test fractions [17]. Activity and inactivity were observed in accordance with the standard and accepted method. Results are shown in Table 2.

110 **RESULTS AND DISCUSSION**

111 Phytochemical screening conducted on leaves and stem-bark extracts of D. 112 *mespiliformis* revealed that alkaloids, anthraquinones, carbohydrates, flavonoids, 113 phlobatannins, saponins, steroids, tannins and terpenoids were present in both the leaves and 114 the stem-bark extract of the plant while glycosides is present in the leaves extract but absent 115 in the stem-bark extract. The plant extracts contain different phytochemical, with various biological activities that can be of value in both medical and veterinary practic \mathcal{O} hus, the 116 117 component of these chemicals suggests that these plants could posses some pharmacological 118 behavior.

119 Alkaloids like morphine and codeine have been reported to have analgesic properties. 120 However, some of them like the morphine have addictive tendencies [18]. Anthraquinones 121 have been associated with anticancer, laxative and anti-arthritic properties [19]. 122 Phytochemical constituents such as flavonoids, alkaloids and saponins present in the leaves 123 and stem-bark extracts were reported to possess biological activity against microbes [14, 20].

124 Tannin a compound that is able to react with protein to appear as stable water 125 insoluble compound, precipitate proteins of the wound, forming a protective layer on the 126 wound, thus assisting in the arrest of bleeding and therefore promoting wound healing. Both the leaves and the stem-bark extracts of *D. mespiliformis* in this study contain tannin and 127 128 could all be investigated for wound healing activity. Tannins have also been reported to be used in the treatment of diarrhea as an effective astringent medicine that does not only stop 129 130 the flow of the disturbing substance in the stomach rather controls the irritation in the small 131 intestine [21]. In addition, phlobatannin a subtype of tannin is also confirmed to be present in 132 all the extracts.

Besides tannin and phlobatannin, all of the investigated plant extracts also contained flavonoids and terpenoids. Flavonoids have been known to have antioxidant, antibacterial, antifungal and antiviral activity [22]. Other studies confirmed that flavonoids and terpenoids
produce antidiabetic activity, possibly by their antioxidant effects [12, 22].

137 Saponins have been associated with decreased cholesteraemia. They are also thought 138 to aid in the absorption of calcium [23]. The two extracts analyzed in this study contain saponin as one of their phytochemical. The leaves and the stem-bark extracts of D. 139 *mespiliformis* are therefore candidate extracts for the prevention of artherosclerosis, which is 140 141 a condition precipitated by cholesterol. Glycosides were only observed in leaves extracts of the plant. A recent study [22] revealed that glycoside is a phytochemical which contributes to 142 one of the hypoglycaemic effects. These phytochemical particularly tannins and flavon 143 are proven to induce an important antimicrobial activity due to their possession of ability to 144 145 inactivate microbial adhesions, enzymes, cell envelope transport proteins, and so forth [24].

Table 1: Phytochemical analysis of aqueous extracts of leaves and stem-bark of *D*. *mespiliformis*

Bioactive compounds	Leaves extract	Stem- bark extract
Alkaloids	+	+
Anthraquinones	+	+
Carbohydrates	+	+
Flavonoids	+	+
Glycosides	+	-
Phlobatannains	+	+
Saponins	+	+
Steroids	+	+
Tannins	+	+
Terpenoids	+	+

148 Key: + = Presence of Phytochemical substance, - = Absence of Phytochemical
 149 substance

150 Antimicrobial activities of the leaves and stem-bark extract of *D. mespiliformis* were 151 studied by measuring the zone of inhibition formed around the disc and the results are given 152 in table 2 and display in figure 1. The result revealed that the stem-bark extract of the plant 153 showed high sensitivity to Escherichia coli, Pseudomonas aeruginosa, Shigella spp, 154 Staphylococcus aureus, Streptococcus pneumonia, Streptococcus pyogenes and Salmonella typhi than the leaves extracts. D. mespiliformis shows significant antimicrobial activity 155 156 against the tested microorganism as shown in figure 1. This study has provided some biochemical basis of the medical use of the extracts from D. mespiliformis in the treatment of 157

UNDER PEER REVIEW

infection, as a potential source of useful drugs and will help to develop new drugcompositions.

160	Table 2:	Antimicrobial	activity	of	aqueous	extracts	of	leaves	and	stem-bark	of	D.
161	mespilifor	mis										

	Zone of inhibition (mm)				
	Leaves	Stem-bark	Ampiclox		
Strains of Microorganisms	50 mg/mL	50 mg/mL	(Control)		
Escherichia coli	12	17	30		
Pseudomonas aeruginosa	12	20	25		
Shigella spp	15	19	22		
Staphylococcus aureus	14	18	26		
Streptococcus pneumonia	5	8	20		
Streptococcus pyogenes	28	29	35		
Salmonella typhi	24	26	32		





166 CONCLUSION

167 The result of study reveals that, the aqueous extracts of the leaves and stem-bark part 168 of *D. mespiliformis* plant contains some major bioactive compounds that inhibit the growth of

169 the test microorganisms, thereby proving to have significant antimicrobial potential. The 170 studies provided some biochemical basis of the medical use of the extracts from D. 171 *mespiliformis* in the treatment of infectious diseases, particularly those causes by the test organism susceptible to the extracts. Further work on the identification of specific 172 173 phytochemical from class of bioactive components identified in the leave and stem-bark 174 extract of the plant is required in order to explore more enormous benefits of *D. mespiliformis* 175 to mankind, especially with regard to the growing résistance of most pathogens to the 176 activities of the common antibiotics. Other related pharmacological studies such as in vivo 177 investigation, drug formulation and clinical trials are highly recommended.

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REFERENCES

179 [1] Bala SA. (2006). Common Ethnomedicinal Plants of the Semiarid Regions of West

- Africa: Their Description and Phytochemicals, Vol 1. Triumph Publishing Company. Page 1-266
- [2] National Research Council (2008). "Ebony". Lost Crops of Africa: Volume III: Fruits.
 Lost Crops of Africa 3. National Academies Press. ISBN 978-0-309-10596-5.
- 184 [3] Breslin A. (2017). "The Chemical Composition of Green Plants". Sciencing, Leaf Group185 Ltd.
- [4] Mallavadhni UV, Panda AK, and Rao YR. (1998). Pharmacology and chemotaxonomy of
 Diospyros mespiliformis. Phytochemistry, 49:901-51.
- [5] Mohamed IE, El Nur EE, Choudhary MI, and Khan SN. (2009). Bioactive natural
 products from two Sudanese medicinal plants *Diospyros mespiliformis* and *Croton zambesicus*. Rec. Nat. Prod.; 3:198-203.
- [6] Caston, J.A. (2008), "Infections Bursal Disease virus (IBDV)." (HTTP: //www. horizon
 press. Com/Ma). Segmented Double Stranded RNA viruses: structure and Molecular
 Biology. Caster Academic Press. ISBN 978-1-90 4455-21-9
- [7] Cazarolli LH, Zanatta L, Alberton EH, Figueiredo MS, Folador P, Damazio RG,
 Pizzolatti MG, and Silva FR (2008). "Flavonoids: Prospective Drug Candidates". Mini
 Reviews in Medicinal Chemistry. 8 (13): 1429–1440. doi:10.2174/138955708786369564.
- 197 [8] Sofowora A (1982). Med. Plants and Traditional Med. in Africa: John Willy. New York,198 pp. 289.
- 199 [9] Adesina SK, and Sofowora EA (1992). Sweet Fruits from Niger. J. Afr. Med. Plants, 2,17.
- 200 [10] Mann A, Yahaya Y, Banso A, and John F(2008). Phytochem. and antimicrobial 201 activity of Terminalia avicennioides extracts against some bacteria pathogens associated with

patients suffering from complicated respiratory tract diseases. J. Med. Plants Res., 2(5): 9497.

[11] Hostettmann, K. A. and Marston (1995). Saponins. Cambridge: Cambridge University
 Press. p. 3ff. ISBN 0-521-32970-1. OCLC 29670810.

[12] Ebbo AA, Mammam M, Suleiman MM, Ahmed A, Bello A (2014) Preliminary
Phytochemical Screening of *Diospyros Mespiliformis*. Anat Physiol 4: 156.
doi:10.4172/2161-0940.1000156

[13] Sundaramoorthy, S., A. Rastogi and S. Arunachalam, (2014). Phytochemical testing,
antioxidant activity, HPTLC and FTIR analysis of antidiabetic plants *Nigella sativa*, *Eugenia jambolana*, *Andrographis paniculata* and *Gymnema sylvestre*. *Res. J. Biotechnol.*, 9: 65-72.

[14] Harborne JB (1993). Phytochemical method. Chapman and Hall, London.Vol (3):135203.

[15] Sofowora A (1984). Medicinal plants and traditional medicine in Africa published in
association with spectrum Books Ltd. Ibadan by John Wiley and Sons.NY pp. 142-143.

[16] Fatope MO, and Adoum OA (1993). "Bioactivity of some savannah plants in the brine
shrimp lethality test and in-vitro anti-microbial Assays". *Int. J. pharmacognosy.* 35(5): 334336.

- [17] Mackie R, McCartney C(1989). Practical Med. Microbiol. 3rd edition, Vol. 2 Churchill
 Livingstone (publishers), London and New York Pp. 100 106,121, 141, 163-167, 303, 432491
- [18] Pilbeam, J.D., and Lyon-Joyce, A.J. (1983). Occurrence of the pyrrolizidine alkaloids
 monocrotaline in Crotalaria seeds. *J. Natl. Prod.* 46(5): 601-605.
- [19] Hemen D., Lalita L. (2012). A review of anthraquinones isolated from Cassia spp. and
 their applications. *Indian J. Nat. Prod. Resour.* 3(3): 291-319
- [20] Shagal M. H, Kubmarawa D and Alim H (2012). Preliminary phytochemical
 investigation and antimicrobial evaluation of roots, stem-bark and leaves extracts of *Diospyros mespiliformis. International Research Journal of Biochemistry and Bioinformatics*
- 229 (ISSN-2250-9941) Vol. **2(1)** pp.011-015
- [21] Chen, H.Y., Lin, C.C., and Lin, T.C. (2002). Antiherpes simplex virus type 2 activity of
 casuarinin from the bark of Terminalia arjuna Linn. Antiviral Res. 55: 447-455.
- [22] Dangoggo, S.M., L.G. Hassan, I.S. Sadiq and S.B. Manga, 2012. Phytochemical analysis
- and antibacterial screening of leaves of *Diospyros mespiliformis* and *Ziziphusspina christi* .
- 234 J. Chem. Eng., 1: 31-37.
- [23] Güçlü-Üstünda, Ö and Mazza, G. (2007). Saponins: Properties, applications and
 processing. Crit. Rev. Food Sci. Nutr., 47: 231-258.

- 237 [24] Cowan MM (1999). Plant products as antimicrobial agents. Clinical Microbiology
- 238 Reviews.12 (4):564–582.