

Nephrotoprotective Effect of *Vernonia amygdalina*

Extract on Benign Prostatic

Abstract

Background: Benign prostatic hyperplasia (BPH) is a noncancerous enlargement of the prostate gland. The condition is associated with symptoms like frequency in urination, hesitancy, nocturnal, weak urine stream and sexual dysfunction. The effect of *Vernonia Amygdalina* extract (VA) on kidney and liver function indices in BPH was investigated.

Methods: A total of 30 rats weighing 200-300 g were divided according to body weight into five groups (n=6). One group was used as a control and the other groups received subcutaneous injections of testosterone and estradiol for 3 weeks to induce BPH. Groups I and II were treated with different doses of VA extracts and group III received finasteride, all by gavages for thirty-five days, while group IV was left untreated, group V served as normal control. After thirty-five days of treatment with VA extract, the rats were anaesthetised by short contact with trichloromethane vapour. Blood was collected by cardiac puncture and the sera centrifuged and used for the determination of different biochemical indices. The prostates were harvested and weighed.

Results: The level of urea and creatinine were significantly ($P<0.05$) reduced when compared to the BPH control. No significant differences in serum concentrations of AST, ALT, ALP, and GGT were recorded in all treatment groups compared to the BPH control.

23 **Conclusion:** The extract of *Vernonia amygdalina* seed exhibited nephroprotective effect on
24 the kidney of BPH induced rats, while there was no observable effect on the liver as benign
25 prostate hyperplasia appeared not to have had any alteration on the liver enzymes.

26 **Keywords:** Creatinine, urea, aminotransferases, alkaline phosphatase, nephroprotective

27 1.0 Introduction

28 Benign prostatic hyperplasia (BPH) is a progressive noncancerous enlargement of the
29 epithelial cells and smooth muscle of the prostate gland accompanied by lower urinary tract
30 symptoms [1]. The enlarged prostate impinges on the urethra and therefore BPH is generally
31 associated with impairment in urinary function [2, 3, 4]. The narrowing of the urethra and
32 urinary retention—the inability to empty the bladder completely—cause many of the
33 problems associated with benign prostatic hyperplasia. The prevalence of BPH is age
34 dependent with approximately 50% of men developing BPH-related symptoms at 50 years of
35 age but the condition is not common before age 40. At the age of 85, the prevalence is as high
36 as 95% and 20-30% of men at the age of 80 years require surgical intervention to manage
37 BPH [1, 5].

38 The mechanism underlying the pathogenesis of BPH remains largely unidentified,
39 however, a number of overlapping and complementary theories have been proposed. Ageing
40 and androgens are established risk factors for the development of benign prostatic
41 enlargement, which may lead to lower urinary tract symptoms (LUTS) in elderly men [6, 7].
42 Androgens and dihydrotestosterone (DHT) play key roles in BPH development. DHT, an
43 androgen derived from testosterone through the action of 5- α -reductase and its metabolite, 3-
44 α -androstenediol, seems to be the major hormonal stimuli for stromal and glandular
45 proliferation in men with nodular hyperplasia [8]. Experimental work has also identified age-
46 related increases in estrogen levels that may increase the expression of DHT, the progenitor

47 of BPH [9]. The incrimination of DHT in the pathogenesis of BPH forms the basis for the
48 current use of 5- α -reductase inhibitors in the treatment of symptomatic nodular hyperplasia.
49 Several types of therapeutic agent, such as 5- α -reductase inhibitors, are currently available
50 for treating BPH [8, 10, 11, 12, 13, 14, 15, 16, 17].

51 Phytomedicine has been in existence for centuries ever before colonial administration
52 and it is in use today with about 80% population depending on herbal medicine for its
53 primary health values [18]. *Vernonia amygdalina* (bitter leaf) has been confirmed to have
54 some vital phytochemical constituents [19]. Phytochemicals are plant secondary metabolites
55 that plants naturally produce to protect themselves against viruses, bacteria and fungi. They
56 are non-nutritive substance with potent biological activities that help in strengthening human
57 immune system and help to lower the risk of many chronic diseases and infections [20].

58 Bitter leaf extracts may help suppress, delay or kill cancerous cell in many ways, such
59 as induction of apoptosis as determined in cell culture and animal studies, enhance
60 chemotherapy sensitivity, inhibition of the growth or growth signals of cancerous cells,
61 suppression of metastasis of cancerous cells in the body by the inhibition of an anti-apoptotic
62 transcription factors as demonstrated in animal studies and reduction of estrogen level in the
63 body by the suppression of aromatase activity [21]. *Vernonia amygdalina* (VA) has
64 demonstrated several medicinal properties enumerated above, hence the need to investigate
65 the possible ameliorative effect of *Vernonia amygdalina* extract on the kidney and liver of
66 BPH induced rats.

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71 **2.0 Materials and Methods**

72 **2.1 Plant Material**

73 Fresh leaves of *Vernonia amygdalina* was harvested from a garden in Okuku in Yala
 74 Local Government of Cross River State, South-South, Nigeria. The plant was identified at the
 75 herbarium unit of the Department of Biological Sciences, University of Calabar. Their fresh
 76 leaves were washed with clean water and dried under the shade for six days. The dried leaves
 77 were pulverized using pestle and mortar to get a powder that was used for extraction.

78 **2.1.1 Preparation of extract**

79 One hundred grams (100 g) of powered sample of *Vernonia amygdalina* was soaked
 80 into 100 mL of distilled water and filtered after 48 hours and the filtrate was concentrated in a
 81 water bath. The concentrates were diluted with corn oil, to produce a solution 100 mg /mL.

82 **2.2 Hormones**

83 Testosterone propionate Brand name: Ricostrone; a product of Greenfield pharma,
 84 Jiangsu Co Ltd., China. Estradiol valerate (by Medipharma Ltd., 108-Kotlakhpat industrial
 85 Est; Lahore, India. Testosterone propionate (T) and estradiol valerate E 2 (puregynon depot)
 86 were used for the induction of prostate enlargement at a dose of 400µg T and 80µg E2 [4, 7,
 87 22, 23, 24]. This was administered to the rats for three weeks subcutaneously in the inguinal
 88 region after which a few rats were sacrificed and inspected for gross examination of prostate
 89 enlargement. All Chemicals used in this study were of analytical grade and were obtained
 90 from reputable companies.

91 **2.3 Animals**

A total of thirty (30) Wistar rats weighing between 200-300g were obtained from the animal house of the Faculty of Basic Medical Sciences, Cross River University of Technology, Okuku Campus, Nigeria and used for the experiment. The rats were acclimatized for two weeks before the experiment commenced. The rats were exposed to approximately 12-hour light/dark cycles under humid tropical conditions, given tap water and feed *ad libitum*, and were housed in standard plastic cages (six per cage) throughout the 35-day duration of the study. The animal room was well ventilated with a temperature range of 27-29 °C. The Cross River University of Technology, Calabar, Nigeria, Animal Ethics Committee approved the study before the experiment and certified all experimental protocols.

2.3.1 Induction of BPH

BPH was induced by exogenous administration of testosterone and estradiol in staggered doses (three times a week respectively) for three weeks according to [22] with modification by [25] and [4, 7, 23, 24].

2.3.2 Animal grouping and treatment

The animals were divided into five (5) groups each which comprised of six (6) male rats. Four groups were induced with BPH which were grouped I, II, III and IV). Groups I and II received 50 and 100 mg kg⁻¹ body weight (bw) of *Vernonia amygdalina* extract while group III received finasteride (orthodox drug) at 0.1mg kg⁻¹; all by gavages for thirty-five days, group IV was left untreated and group V served as normal control. The animals were weighed prior to the commencement of the experiment and subsequently every week till the end of the experiment. The water intake was daily and lasted throughout the duration of the experiment.

2.4 Preparation and collection of samples for biochemical assay

115 After 35 days, the rats were anaesthetized by a brief exposure to trichloromethane
116 vapour and bled by cardiac puncture. The sera were carefully separated and used for the
117 determination of various biochemical analyses. Each rat's carcass was promptly dissected and
118 the prostates were carefully excised. The prostates were freed of external fascias, washed in
119 cold normal saline, blotted with filter paper and weighed on a sensitive balance. Afterward,
120 they were homogenized in ice-cold normal saline and the homogenates was used for the
121 estimation of the protein content of the prostate gland. The procedure used previously by [4,
122 7, 23, 24] was adopted.

123 **2.4.1 Determination of Aminotransferases and Alkaline Phosphatase**

124 The assay for alkaline phosphatase (ALP), aspartate amino transferase (AST), alanine
125 amino transferase (ALT) and γ -glutamyl transferase (GGT) were done using kits from
126 Randox Laboratory, Ltd, United Kingdom. Urea and creatinine concentrations were done
127 using Agape Diagnostic kits. All chemicals and reagents used in this research were of
128 analytical grade.

129 **2.4.2 Determination of Protein Content of the Prostate**

130 Cupric ions, in an alkaline medium, interact with protein peptide bonds resulting in
131 the formation of coloured complex. The protein content of the prostate was determined using
132 the modified Biuret method [26] and [4]. Briefly, 3.9ml of deionized water and 4.0 ml of
133 Biuret reagent were added to 0.1ml of the aliquot and allowed for 30 minutes at room
134 temperature to develop. A standard and blank were also prepared by adding 4.0ml of Biuret
135 reagent and 3.9ml of deionized water to 0.1ml of standard albumin and water respectively.
136 Subsequently, the absorbance of the test and standard were read against the blank at 540nm
137 using a UV/VIS spectrophotometer.

138 **2.5 Statistical Analysis**

139 The data obtained from the experiment was presented as mean \pm SD after calculation
140 using Microsoft Office Excel 2007. The data was also subjected to a one-way analysis of
141 variance (ANOVA) and post hoc (LSD) for levels of significance using SPSS version 16.0.
142 The level of significance was accepted at $P < 0.05$

143 **3.0 RESULTS**

144 **3.1 Body weight**

145 Reduction in body weight was observed in the BPH-control group when compared
146 with normal control (Table 1). The extract and standard drug treated groups showed
147 significant ($P < 0.05$) increase in body weight when compared with the BPH control group.
148 Administration of extract and finasteride enhanced the body weight when compared with
149 normal control.

150 **3.2 Prostate gland and Prostate/body weight (P/PW)**

151 The average weight of the prostate gland and prostate/body weight ratio were
152 significantly increased in the BPH control group compared with normal control group (Table
153 1). The extract and finasteride treated groups showed a decrease in prostate gland and
154 prostate/body weight ratio when compared with the BPH-control group.

155 **3.3 Kidney indices of BPH-induced rats**

156 There were significant ($P < 0.05$) increase in level of serum urea concentration and
157 creatinine in BPH control group when compared with normal control and test groups. The
158 value of the doses of VA and finasteride were similar to the normal control. The results

showed that all the treated groups exhibited reduction in the level of urea and creatinine concentration (Table 2).

3.4 Liver function enzymes activities of BPH-induced rats

Serum ALT, AST, ALP and GGT concentrations are given in (Table 3). The result of the investigation showed no significant difference ($P>0.05$) in all the test groups compared with both the BPH control and normal control. There was also no significant difference ($P>0.05$) among the test groups.

Table 1: Effect of extract of VA and finasteride prostate weight and protein content of prostate

GROUP	BW (g)	PW (g)	PW (mg)	P/BW ratio (mg/g)	PC (g/dl)
BPH + 50mg VA	275.40±5.68 ^b	0.39±0.05 ^a	388.00±45.50 ^a	1.41±0.14 ^a	5.30±0.20 ^a
BPH + 100mg VA	271.60±5.68 ^b	0.36±0.06 ^a	360.00±57.01 ^a	1.33±0.21 ^a	5.09±0.21 ^a
BPH + FINASTERIDE	271.80±2.77 ^b	0.35±0.05 ^a	352.00±50.70 ^a	1.30±0.18 ^a	5.27±0.89 ^a
BPH CONTROL	220.40±8.9b ^a	0.96±0.03 ^b	962.00±32.71 ^b	4.37±0.20 ^b	7.41±0.96 ^b
NORMAL CONTROL	279.20±4.97 ^b	0.36±0.03 ^a	356.00±33.62 ^a	1.28±0.12 ^a	5.08±0.73 ^a

Values are expressed as mean ± SD.
^{a, b} Values with different superscripts are significantly different at $P<0.05$
 BPH (Benign prostate hyperplasia) and VA (*Vernonia amygdalina*).

Table 2: Effect of extract VA and finasteride on kidney function parameters

GROUP	UREA (mg/dl)	CREATININE (mg/dl)
BPH + 50mg VA	19.49±1.07 ^a	0.92±0.21 ^a

BPH + 100mg VA	18.46±1.46 ^a	0.87±0.16 ^a
BPH + FINASTERIDE	18.97±1.07 ^a	0.83±0.15 ^a
BPH CONTROL	26.41±2.81 ^b	1.96±0.33 ^b
NORMAL CONTROL	17.69±1.07 ^a	0.67±0.35 ^a

Values are expressed as mean ± SD.

^{a, b} Values with different superscripts are significantly different at $P < 0.05$

BPH (Benign prostate hyperplasia) and VA (*Vernonia amygdalina*).

Table 3: Effect of extract of VA and finasteride on serum enzyme activities

GROUP	ALT (U/L)	AST (U/L)	ALP (U/L)	GGT (U/L)
BPH + 50mg VA	23.49±0.58 ^a	33.35±0.51 ^a	241.15±3.01 ^a	17.88±1.40 ^a
BPH + 100mg VA	22.99±1.33 ^a	33.31±0.46 ^a	241.20±2.36 ^a	18.17±1.21 ^a
BPH + FINASTERIDE	23.07±1.14 ^a	32.55±3.18 ^a	241.14±2.62 ^a	18.17±1.71 ^a
BPH CONTROL	23.56±1.50 ^a	33.82±1.27 ^a	241.58±2.40 ^a	18.15±0.60 ^a
NORMAL CONTROL	23.32±1.66 ^a	33.01±0.99 ^a	241.12±2.97 ^a	18.15±0.97 ^a

Values are expressed as mean ± SD.

Values with identical superscript (a) are not significantly different at $P > 0.05$.

BPH (Benign prostate hyperplasia) and VA (*Vernonia amygdalina*)

4. Discussion

Given the many side effects of surgery and pharmacological therapy and the long latency of BPH, phytotherapy based on products derived naturally from plants has emerged as an alternative treatment for BPH because it is thought to be less toxic [27]. Despite the

many possible causes of obstructive uropathy, in studies of elderly patients with acute renal failure, the most common cause among all patients was BPH [28]. Previous studies showed that acute renal failure in patients with obstructive uropathy were due to BPH [29, 30]. This necessitated the evaluation of the effect of *Vernonia Amygdalina* on the kidney and liver integrity of rats induced with BPH.

The prostate weight is used as one important marker of BPH development [12, 31, 32]. In previous studies, animals with BPH have shown an increased prostate weight indicating increase in cell number [13, 14]. Finasteride or other agents used to treat BPH decrease the prostate weight [11, 16, 23]. In the present study, the animals with BPH showed an increased prostate weight compare to the control group. In contrast, the animals treated with *Vernonia amygdalina* showed a reduction in prostate weight compared to BPH group. These results indicate that *Vernonia amygdalina* attenuated the prostatic enlargement induced by testosterone. Increase in cell number (hyperplasia) of the prostate would come with a collateral increase in its weight (especially its relative weight) [8, 15]. Also increase in cell number in a tissue also goes with a collateral increase in the protein content of the tissue [4, 33]. The protein content of the prostate was significantly high in BPH untreated group compared with the treated groups.

The liver enzymes found within organs and tissues are released into the bloodstream following cellular necrosis and cell membrane permeability and are used as diagnostic measure of liver damage [34, 35]. Tissue cells contain characteristic enzymes which enter the blood only when the cells to which they are confined are damaged or destroyed [36]. The tissue activities of the transaminase (ALT and AST) enzyme are markers for the functions and integrity of the liver and heart [37, 38]. The present study was therefore conducted to

provide scientific data on the effect of aqueous extract of *Vernonia Amygdalina* on alanine transaminase (ALT), aspartate transaminase (AST), alkaline phosphatase (ALP), γ -glutamyltransferase (GGT), creatinine and urea levels in male Wistar rats induced with BPH.

The extract did not affect the activities of ALT, AST, ALP and GGT indicating normal liver function. This implied that benign prostatic hyperplasia may not have exhibited adverse effect on the liver function and that the extract had no toxic effect on this organ [24, 39]. Earlier studies showed that oral administration of the aqueous extract of some plant could accelerate the reversion of liver damage through reduction of liver marker enzymes, including aspartate aminotransferase (AST), alanine transaminase (ALT) and alkaline phosphatase (ALP), glutamate-oxaloacetate transaminase, glutamate- pyruvate transaminase, lactate dehydrogenase and bilirubin indices in liver biochemical tests [24, 40].

5. Conclusion

The extract of *Vernonia Amygdalina* leaf exhibited nephroprotective effect on the kidney of BPH induced rats, while there was no observable effect on the liver as benign prostate hyperplasia appeared not to have had any alteration on the liver enzymes.

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