

Original Research Article

Mango (*Mangifera indica*) Seed Kernel: Proximate Properties and Effect on Normal and Monosodium Glutamate-Hepato-Compromised Wistar Rats

ABSTRACT

Monosodium glutamate, MSG, a widely used flavour enhancer could induce hepatic injury in humans while mango (*Mangifera indica*) seed kernel, MSK, a common fruit waste could be nutritive and therapeutic. Thus, this study investigated the proximate properties of MSK and the effect of MSK on normal and monosodium glutamate-hepato-compromised Wistar rats using standard protocols involving five groups (n=4) viz: control (distilled water, 2ml/kg), MSG (8g/kg), MSK ethanolic extract, MSKE (0.3g/kg), MSG (8g/kg) + MSKE (0.2g/kg), MSG (8g/kg) + MSKE (0.3g/kg) and MSG (8g/kg) + MSKE (0.4g/kg) which were respectively fed *via* gavage daily for 14 days. Proximate content of MSK were: total carbohydrate (71.99%), moisture (10.53%), crude protein (5.68%), ash (4.31%), fat (4.29%), and crude fiber (3.20%). The determined serum bio-indicators of hepatic function (alanine aminotransferase, aspartate aminotransferase and alkaline phosphatase activity and total bilirubin concentration) in the rats exposed to MSG alone which were significantly ($p < 0.05$) higher than those in the control were reduced ($p < 0.05$) dose dependently following concomitant exposure of the rats to MSG and increasing concentration of MSKE. Multifocal necrosis with periportal infiltration of mononuclear inflammatory leucocytes in the liver histology of MSG-treated rats contrasted the normal hepatic histo-architecture in that of the control and MSKE groups. The study highlighted the dietary potential of MSK and confirmed a definite MSG-induced adverse effect on the liver histology and serum functional indices of rats, probably ameliorated on concomitant exposure with MSKE. Further studies to exploit MSK in diets and drugs could reduce its waste status hence warranted.

Keywords: Total bilirubin, Alanine aminotransferase, Alkaline phosphatase, Aspartate aminotransferase

1. INTRODUCTION

Monosodium glutamate (MSG) improves food flavour but with adverse effects including on the liver of animals [1-5]. The liver plays a major role in protein and amino acids metabolism among other metabolic, synthestic and regulatory processes [6]. Plants wastes could contribute to environmental pollution [7]. However, potentials exist to utilize plant wastes for new foods sources and drugs development thereby reducing the possible adverse environmental impact [8-10].

Mango by-products, such as peels and seeds, contain high levels of various health-enhancing substances and exhibited clear cytoprotective effect on the liver injury [11,12]. In particular, *Mangifera indica* L. seed kernel which currently not utilized for any commercial purpose but discarded as waste showed promising biochemical effects in recent studies [13-15]. These warranted the present study aimed at determining the proximate property of *Mangifera indica* L. seed kernel and the effect of the ethanolic extract on normal and monosodium glutamate-hepato-compromised Wistar rats. The liver is a major organ and central to xenobiotic detoxification hence unique in detecting possible adverse effects of novel diets and drugs.

2. MATERIAL AND METHODS / EXPERIMENTAL DETAILS

2.1. Sample procurement and identification

A commercially available brand of MSG (99 % purity) was used in this study as procured from *Uban* market, a daily food condiments market in Umuahia, south east Nigeria. Chemicals and solvents used in this study were products of reputable companies procured from reputable chemical dealers and were used without further purification. This study was conducted between June and August, 2016.

Fresh mango fruits collected from a particular mango tree were purchased in June, 2016 at *Orie ugba*, a fruit and foodstuff market in Umuahia, Abia state, Nigeria. The mango fruits were identified and authenticated as *Mangifera indica* (German variety) by a taxonomist in the department of Plant Science and Biotechnology, Michael Okpara University of Agriculture Umudike, Nigeria.

2.2. Sample preparation and extraction

The mango fruits were thoroughly washed with tap water. The fleshy part of each of the fruits was removed to obtain the seed stones which were sun-dried for three days. The sun-dried seed stones were carefully cut with clean table knife to remove the stony seed coat and obtain the seed kernels. The kernels thus obtained were chopped with home choice knife into bits and sun-dried for one week (seven days). The dried mango seed kernels were pulverized using Arthur Thomas Laboratory Mill, Crypto Model, USA. The pulverized mango seed kernel was extracted with ethanol (98 %) as described earlier [16].

2.3. Animal study

2.3.1. Animal procurement and exposure groups

Twenty adult male albino rats (weight range, 104-170 g) used in this study were procured from the animal house of the Faculty of Biological Sciences, University of Nigeria, Nsukka. The animals were acclimatized for 2 weeks and then randomized (based on weight) to five experimentation groups with sample size of four rats. Rats in the control group were sham-dosed with distilled water (without either the extract or MSG) while rats in the MSG group were fed intoxicating dose (8000 mg/kg body weight) of MSG according to Mariyamma *et al.* [17] and as in several recent studies [13-15,18-20]. Rats in the extract group were fed mango seed kernel extract at 300 mg/kg body weight while rats in the MSG + low extract group were concomitantly fed the mango seed kernel extract (200 mg/kg body weight) and intoxicating dose of MSG (8000 mg/kg body weight) whereas rats in the MSG + high extract group were co-administered 400 mg/kg body weight of the mango seed kernel extract and intoxicating dose of MSG (8000 mg/kg body weight). The exposure was *per oral* and daily for 14 days.

2.3.2. Sacrifice, blood sample collection and preparation

After 2 weeks (14 days) exposure, the rats were sacrificed the next day after overnight fast by cardiac puncture technique and the blood sample of the respective rats was collected individually into clean polystyrene tubes. The blood samples thus collected were respectively centrifuged at 3000 rpm for 10 minutes. The resultant serum was respectively collected into polystyrene tubes and stored in deep freezer for the determination of serum alanine aminotransferase (ALT), aspartate aminotransferase (AST) and alkaline phosphatase (ALP) activity as well as serum total bilirubin (TBIL) concentration.

2.3.3. Ethical consideration

This study which is a continuation of our line of studies on the evaluation of biochemical effects of mango seed kernel extract on normal and monosodium glutamate-challenged experimental models considered and adhered to the standard ethical use of experimental animals. Throughout out the experimentation (acclimatization and exposure periods), all rats were housed at 25 °C in stainless steel cages under normal daylight/dark cycle and humid tropical conditions. The rats were allowed free access to rat feed (Vital feed, Jos Nigeria) and tap water, and generally received humane care in accordance with the guidelines of the National institute of Health, USA for ethical treatment of laboratory animals as approved by the various (departmental and college) ethical committees of Michael Okpara University of Agriculture Umudike, Nigeria.

2.4. Determination of proximate content

Proximate content viz: moisture, ash, fibre, fat and protein was determined as described previously [21-23]. The total carbohydrate content of MSK was estimated as the Nitrogen free extract (NFE) by arithmetic difference using the relation: Total CHO % = 100 – (fibre +protein +Moisture + ash + fats).

2.5. Determination of seric chemistry parameters

Determination of serum bilirubin was according to instruction provided in Randox™ assay kit based on the method of Jendrassik and Grof [24]. The method was based on the principle that bilirubin reacts with diazotized sulphanilic acid in an alkaline medium to form a blue coloured complex which is measured with a spectrophotometer and calculated to give the concentration of bilirubin in the serum sample.

The serum activity for AST and ALT was determined using a Randox™ commercial kits according to the method of Reitman and Frankel [25]. The method was based on the colorimetric

estimation of oxaloacetate (for AST) or pyruvate (for ALT) produced through transamination of aspartate or alanine respectively on reacting with 2-4-dinitrophenyl hydrazine (DNPH). The intensity of the resultant brown-colored hydrazone was measured with a colorimeter after 5 minutes at 540 nm and the activity (AST or ALT) was read up from standard curve.

The serum ALP activity was determined using a Randox™ commercial assay kit according to the method of King and Armstrong [26]. This was based on the colorimetric estimation, at 520 nm, of red colored complex formed following the hydrolysis of phenyl phosphate by alkaline phosphatase to release hydroxybenzene (aside phenol and phosphate) which combines with 4-aminophenazone to form a red complex in a reaction catalyzed by potassium ferricyanide (K₂Fe(CN)₂). The alkaline phosphatase activity (IU/L) was calculated from the relation:

$$\text{ALP (IU/L)} = \frac{\text{Abs of sample}}{\text{Abs of standard}} \times \text{conc of standard}$$

2.6. Histopathological examination

Collected sample (Liver) was fixed in 10% phosphate buffered formalin for a minimum of 48 hours. The tissue was subsequently trimmed, dehydrated in 4 grades of alcohol (70%, 80%, 90% and absolute alcohol), cleared in 3 grades of xylene and embedded in molten wax. On solidifying, the blocks were sectioned, 5µm thick with a rotary microtome, floated in water bathe and incubated at 60°C for 30 minutes. The 5µm thick sectioned tissues were subsequently cleared in 3 grades of xylene and rehydrated in 3 grades of alcohol (90%, 80% and 70%). The sections were stained with Hematoxylin for 15 minutes. Blueing was done with ammonium chloride. Differentiation was done with 1% acid alcohol before counterstaining with Eosin. Permanent mounts were made on degreased glass slides using a mountant; DPX. Prepared slides were examined with a Motic™ compound light microscope using x4, x10 and x40 objective lenses. Photomicrographs were taken using a Motic™ 9.0 megapixels microscope camera at x100 and x400 magnifications.

2.7. Method of Statistical Analysis

Results of replicate experiments were expressed as mean ± standard deviation (SD). Analysis of variance (ANOVA) and least significance difference (LSD) were carried out on the result data at 95% confidence level using SPSS statistical software package, version 20.

3. RESULTS AND DISCUSSION

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xylene and rehydrated in 3 grades of alcohol (90%, 80% and 70%). The sections were stained with Hematoxylin for 15 minutes. Blueing was done with ammonium chloride. Differentiation was done with 1% acid alcohol before counterstaining with Eosin. Permanent mounts were made on degreased glass slides using a mountant; DPX. Prepared slides were examined with a Motic™ compound light microscope using x4, x10 and x40 objective lenses. Photomicrographs were taken using a Motic™ 9.0 megapixels microscope camera at x100 and x400 magnifications.

Method of Statistical Analysis

Results of replicate experiments were expressed as mean $\pm$ standard deviation (SD). Analysis of variance (ANOVA) and least significance difference (LSD) were carried out on the result data at 95% confidence level using SPSS statistical software package, version 20.

RESULTS

Table 1: Proximate composition (%) of pulverized mango (*M. indica*) seed kernel, MSK

Proximate parameter	Composition in MSK (%)
Total carbohydrate	71.99 $\pm$ 0.06
Moisture content	10.53 $\pm$ 0.02
Crude protein	5.68 $\pm$ 0.02
Ash content	4.31 $\pm$ 0.01
Fat	4.29 $\pm$ 0.01
Crude fibre	3.20 $\pm$ 0.01

Value presented as mean $\pm$ SEM of triplicate determinations

As shown on Table 1, the proximate content of MSK in decreasing order revealed that total carbohydrate (71.99%) was highest followed by moisture (10.53%), crude protein (5.68%), fat (4.29%) and ash (4.31%) while crude fiber (3.20%) was the least.

Table 2: Effect of pulverized mango (*M. indica*) seed kernel extract, MSKE, on the activity (IU/L) of alanine aminotransferase, ALT, aspartate aminotransferase, AST, and alkaline phosphatase, ALP, and concentration of total bilirubin of normal and monosodium glutamate-challenged rats' serum

Groups	ALT (IU/L)	AST (IU/L)	ALP (IU/L)	TBIL (mg/dl)
Control (distilled water 2 ml/kg b.w)	74.48 $\pm$ 5.13	116.68 $\pm$ 1.31	34.85 $\pm$ 0.92	0.33 $\pm$ 0.03
MSG 8000 mg/kg b.w)	129.55 $\pm$ 9.83	152.50 $\pm$ 0.87	238.67 $\pm$ 1.31	1.76 $\pm$ 0.03
Extract 300 mg/kg b.w)	34.18 $\pm$ 0.68	145.75 $\pm$ 0.77	137.25 $\pm$ 1.30	0.85 $\pm$ 0.05
MSG 8000 mg/kg b.w) + low Extract 200 mg/kg b.w)	39.50 $\pm$ 1.05	151.00 $\pm$ 1.01	173.25 $\pm$ 0.86	1.24 $\pm$ 0.01
MSG 8000 mg/kg b.w) + high Extract 400 mg/kg b.w)	36.62 $\pm$ 1.23	149.71 $\pm$ 1.09	169.67 $\pm$ 1.04	1.17 $\pm$ 0.22

Value presented as mean $\pm$ SEM of sample size, n = 4 rats. + denotes higher by; - denotes lower by. Significant difference at $P = .05$.

The determined serum alanine aminotransferase, aspartate aminotransferase and alkaline phosphatase activity (IU.l⁻¹), respectively in the rats exposed to MSG alone (129.55, 152.50, 238.67) which was significantly ($p < 0.05$) higher than that in the control (74.48, 116.68, 34.85) and Extract-exposed groups (34.18, 145.75, 137.25) reduced ($p < 0.05$) dose dependently following concomitant exposure of the rats to 200 mg/kg (39.50, 151.00, 173.25) and 400 mg/kg (36.62, 149.71, 169.67) of MSKE (Table 2).

Similarly, the determined serum total albumin concentration (mg.dl⁻¹) in the rats exposed to MSG alone (1.76) which was significantly ($p < 0.05$) higher than that in the control (0.33) and Extract-

exposed groups (0.85) reduced ($p<0.05$) dose dependently following concomitant exposure of the rats to 200 mg/kg (1.24) and 400 mg/kg (1.17) of MSKE (Table 2).

Histopathological Observations

Sections from the MSG group showed a mild periportal infiltration of mononuclear inflammatory leucocytes (arrow) and multifocal areas of hepatocellular necrosis with mononuclear leucocytic infiltration (Figure 1).

Sections of the liver collected from the animals in the other groups including the control as compared to the MSG-exposed group showed normal hepatic histo-architecture viz: normal hepatic lobules with normal hepatocytes arranged in radiating cords around the central veins (CV) with the hepatocytes (arranged in cords) radiating towards the portal areas characteristic of normal hepatic artery, hepatic vein (HV) and bile duct (B) (Figure, 2,3,4,5).

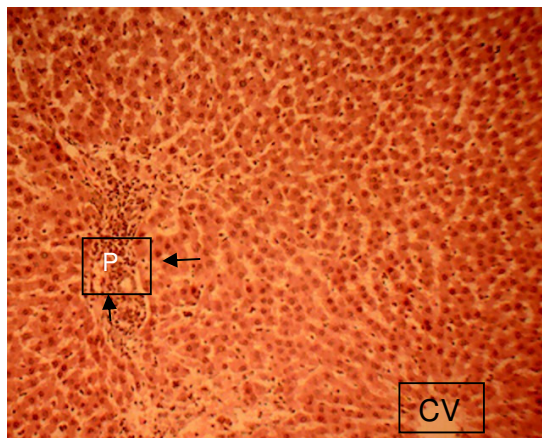


Figure 1. Photomicrograph of liver sections of rats exposed to MSG showing damaged Portal area (P) as mild periportal infiltration of mononuclear inflammatory leucocytes (arrow) and multifocal areas of hepatocellular necrosis with mononuclear leucocytic infiltration. H&E: $\times 100$.

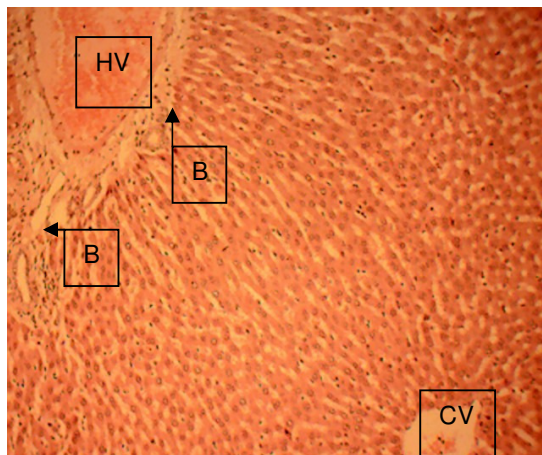


Figure 2. Photomicrograph of liver sections of rats exposed to MSKE alone showing normal central veins (CV) with normal hepatocytes arranged in radiating cords towards the portal areas characteristic of normal hepatic artery, hepatic vein (HV) and bile duct (B). H&E: $\times 100$.

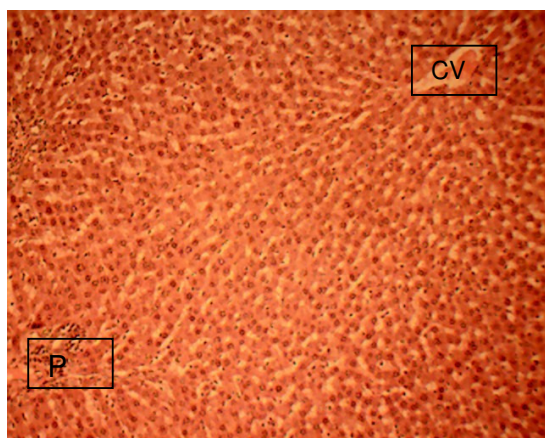


Figure 3. Photomicrograph of liver sections of rats in the control group showing normal Central vein (CV); Portal area (P). H&E: $\times 100$.

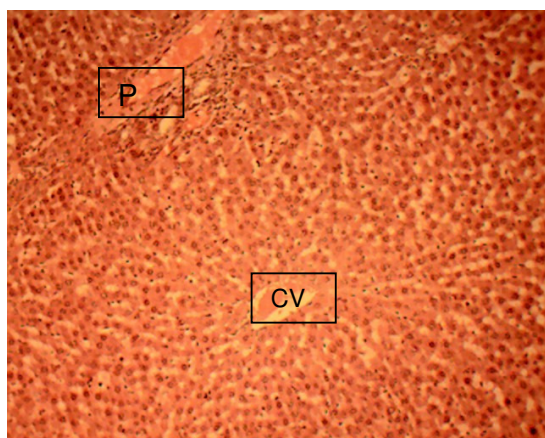


Figure 4. Photomicrograph of liver sections of rats exposed to MSG + Low MSKE showing normal Central vein (CV) and Portal area (P). H&E: $\times 100$.

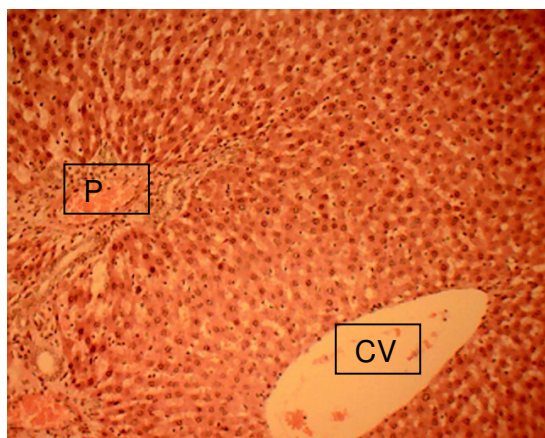


Figure 5. Photomicrograph of liver sections of rats exposed to MSG + high MSKE showing normal Central vein (CV); Portal area (P). H&E: ×100.

Discussion

Monosodium glutamate, MSG, a widely used flavour enhancer could induce hepatic injury in humans while mango (*Mangifera indica*) seed kernel, MSK, a common fruit waste could be nutritive and therapeutic. Thus, this study investigated the proximate properties of MSK and the effect of MSK on normal and monosodium glutamate-hepato-compromised Proximate content of MSK were: total carbohydrate (71.99%), moisture (10.53%), crude protein (5.68%), ash (4.31%), fat (4.29%), and crude fiber (3.20%) which suggest that MSK could have dietary potential. In particular, the total carbohydrate content of MSK compared with the value range (70.12% - 77.90%) obtained in previous studies [27-28] and far higher than that (30.85%) in *Terminalia catappa* endocarp [29]. This indicated that mango seed could serve as a good alternative carbohydrate-energy source. The crude fibre content compared with that (5.72%) reported by Nzewi and Egbunu [30] and higher than that (2.40 and 2.02, respectively) reported by Fowomola [27] and Nzikou *et al.* [31] but far lower than that (36.33%) reported for *Terminalia catappa* endocarp [29] indicating that MSK could not offer high satiety (appetite satisfaction) or enhanced peristaltic movement of food through the alimental canal that could prevent constipation [32]. The moisture content of mango seed obtained from this work was slightly higher when compared to the value range (5.00% - 9.8%) reported in previous studies [29-34] though lower than that reported by Nzikou *et al.* [31]. The observed moisture content was considered moderately high to ensure ease transportation of nutrients and other necessary metabolic reactions and low to ensure long shelf life for the mango seed kernel flour [34].

The ash percentage obtained (4.31%) was slightly higher than that (3.07%) reported by Egbunu [9] for *Citrullus lanatus* rind and seed but lower than that (9.05%) reported by Abulude *et al.* [32] and that (11.03%) reported by Anuforo *et al.* [29] implying moderate mineral supplying potential of MSK. The protein content of this study (5.68%) was within the range (5.20-8.28%) obtained from many varieties of mango in previous studies [34,35] and *Terminalia catappa* endocarp [29] suggesting that MSK could be a good dietary protein source. Also, MSK may be a good source of fat and oil as the fat and oil content obtained in this study (4.29%) was in the upper limit of the range (1.94% - 5.68%) obtained in previous studies [27,32,34,36] though lower compared to that (8.53) in *Terminalia catappa* endocarp [29].

The determined serum bio-indicators of hepatic function (alanine aminotransferase, aspartate aminotransferase and alkaline phosphatase activity and total bilirubin concentration) in the rats exposed to MSG alone which were significantly ($p<0.05$) higher than those in the control were reduced ($p<0.05$) dose dependently following concomitant exposure of the rats to MSG and increasing concentration of MSKE. Bilirubin is the catabolic product of haem metabolism via the activities of the liver enzyme UDP-glucuronyltransferase [37] hence an indicator of liver function and integrity. Increased activity of ALP aside indicating liver damage could be indicating bone diseases [37,5]. Thus, the results indicated MSG-related hepatotoxicity and possibly bone dysfunctions in the rats which perhaps were significantly and dose-dependently ameliorated by MSKE. Toxic assault on the liver was associated with an increase in the studied indicators of liver function [38,39] and MSG-induced hepatic damage was reported earlier [5]. Multifocal necrosis with periportal infiltration of mononuclear inflammatory leucocytes in the liver histology of MSG-treated rats contrasted the normal hepatic histo-architecture in that of the control and MSKE-exposed groups. This seemingly confirmed the seric chemistry results that suggested MSKE-related amelioration of MSG-induced hepatic assault and damage.

4. CONCLUSION

The study highlighted the dietary potential of MSK and confirmed a definite MSG-induced adverse effect on the liver histology and serum functional indices of rats, probably ameliorated on concomitant exposure with MSKE. Further studies to exploit MSK in diets and drugs could reduce its waste status hence warranted.

ETHICAL APPROVAL (WHERE EVER APPLICABLE)

All authors hereby declare that "Principles of laboratory animal care" (NIH publication No. 85-23, revised 1985) were followed, as well as specific national laws where applicable. All experiments have been examined and approved by the appropriate ethics committee

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