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<u>Original Research Article</u> Comparative Effect of Chilli Pepper Extract and Capsaicin on Some Haematological Parameters and Serum Electrolytes in Albino Wistar Rats

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6 ABSTRACT

Aim: Capsaicin is the active ingredient in chilli pepper, and is responsible for the pungency of chilli pepper. This study compared the effect of ethanolic extract of chilli pepper fruit and capsaicin on haematological parameters and serum electrolytes in female albino Wistar rats on the background that they are widely consumed in foods.

Methodology: Fifteen female Wistar rats (140 - 200 g b.w) fed with rat feed and water *ad libitum* were divided into three groups (n = 5) thus: control, chilli pepper and capsaicin groups. The three groups were treated with daily oral administration of 0.2 mL normal saline, chilli pepper extract (5 mg/100g b.w) and capsaicin (3 mg/100g b.w) respectively, for 30 days. Blood samples were collected from each animal via cardiac puncture for assessment of haematological parameters and serum concentration of electrolytes.

Results: Red blood cell (RBC) count, haemoglobin (Hb) concentration and packed cell volume (PCV) in capsaicin group, unlike chilli pepper group, were significantly (p<0.01) reduced compared with control. PCV was significantly (p<0.05) reduced in capsaicin group compared with the chilli pepper group. Platelet count and platelet large cell ratio (P-LCR) were significantly reduced (p<0.01) in capsaicin group compared with the control. Serum Na⁺, Cl⁻, and urea concentrations were not significantly (p>0.05) different among groups, but creatinine level decreased significantly (p<0.05) in the treated groups compared with the control. Serum HCO₃⁻ increased while K⁺ decreased significantly (p<0.05) in capsaicin treated group compared with the control. Furthermore, serum K⁺ increased (p<0.05) in chilli pepper group, compared with the control.

Conclusion: Capsaicin and chilli pepper did not cause serious electrolyte imbalance, but reduced red cell indices. Additionally, capsaicin altered platelet parameters. Therefore, we suggest that capsaicin may be detrimental if used by people with bleeding and/or blood coagulation disorders.

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Keywords: Capsaicin, creatinine, chilli pepper, haematology, serum electrolytes, urea

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11 **1. INTRODUCTION**

12 Chilli pepper is a diploid and self-pollinating crop. It is a member of the family of plants known as Solanaceae. It belongs to the genus Capsicum and has five species namely: frutescens, pubescens, 13 14 baccatum, annuum and chinense [1]. Chilli pepper is one of the world's most widely grown spice and a 15 major ingredient in most foods worldwide. It is used as the active substance in defence repellent, ornaments, colouring agents and for therapeutic purposes [2]. It has a pungent sensation due to the 16 17 presence of a group of compounds called capsaicinoids [3]. Chilli pepper increases gastric activities and 18 enhances blood flow and is used to relieve pain associated with child delivery [4]. However, reports have 19 shown that high doses of chilli pepper can also lead to gastric erosion [5]. Chilli pepper promotes the 20 health of diabetic patients by reducing disaccharides in the intestinal lumen through α -glucosidase and α -21 amylase inactivation [6]. Crude ethanolic extract of chilli pepper may be effective in reducing renal 22 insufficiency as it has been reported to decrease serum creatinine and urea and increase total protein 23 levels [7]. A study conducted by Elamin et al.[8] on broilers showed that chilli pepper significantly reduced 24 serum cholesterol, abdominal fat, and AST enzyme activity, but serum total protein, urea, ALP, calcium 25 (Ca) and phosphorus (P) were unaffected.

Phytochemical screening by Dougnon *et al.*[9] revealed the presence of sterols, alkaloids, mucilage, polyterpenes, and reducing compounds in the powder of chilli pepper (*Capsicum frutescens*). This chilli

- pepper powder fed to Hubbard broilers at 5 g and 10 g/kg diet did not significantly change haematological
- 29 parameters and creatinine levels. Another study by Shahverdi et al.[10] showed that 1 % chilli pepper

powder used as broilers' feed additive improved overall performance but decreased haemoglobin
 concentration and packed cell volume.

32 The active ingredient (i.e. the main capsaicinoid) in chilli pepper is capsaicin, and is responsible for the 33 irritation and pungency of various hot peppers [11]. Capsaicin can be extracted from chilli pepper using 34 different solvents and is also available in synthetic form. It acts by binding to transient receptor potential 35 vanilloid-1 (TRPV1), formerly called vanilloid receptor. Basically, TRPV1 is located in nociceptive neurons and widely distributed in brain tissues, intestines, liver, keratinocytes of epidermis and macrophages. 36 37 TRPV1 was first discovered on the beta cells of pancrease. It was discovered that capsaicin could 38 activate TRPV1 with resultant increase in insulin secretion [12]. Capsaicin, administered to healthy and 39 diabetic rats taking high iron diet has been reported to reduce levels of haemoglobin, cholesterol and 40 triglycerides [13]. It was found that capsaicin could decrease plasma glucose level [14, 15] and inhibit 41 glucose absorption into the blood stream [16]. Capsaicin is used to treat osteoarthritis, post-herpatic neuralgia and diabetic neuropathy [17]. It has been reported to have antimicrobial activity [2, 3], 42 43 cardioprotective effect [18], anti-inflammatory effect [19] and anticancer activity [20, 21]. Capsaicin also 44 reduces the severity of headaches [19]. Additionally, it has been reported that capsaicin has anti-obesity 45 effect [22, 23], but the potential side-effects limits its clinical application [24]. Capsaicin has been reported 46 to increase gastric acid secretion and mucosal blood flow [25]. A low dose of capsaicinoids is beneficial to 47 gastrointestinal defense [26] but a high dose may be detrimental to the gastrointestinal tract since it 48 damages capsaicin-sensitive afferent nerves and causes exhaustion of neurotransmitters [27]. In spite of 49 the numerous researches carried out with chilli pepper and capsaicin, very little or none has documented 50 and compared the effect of ethanolic extract of chilli pepper fruit and capsaicin on haematological 51 parameters and serum electrolytes, hence the present study.

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54 2. MATERIAL AND METHODS

55 2.1 Extract Preparation

56 Chilli pepper, bought from Watt market, Calabar, Nigeria, were washed with tap water and dried at 35°C. 57 The dried pepper fruits (including the wall, seed and placenta) were ground to powder using an electric 58 grinder. Grinded chilli pepper (300g) was extracted in 2L of 95 % ethanol by maceration for 72 hours. The 59 extract was filtered using Whatman No1 filter paper and the filtrate was evaporated to dryness in a rotary 60 evaporator, lyophilized and thereafter preserved for use. 61

62 2.2 Experimental Animals

Fifteen female albino Wistar rats (140 – 200 g body weight) handled according to Helsinki's (1964) laid down principles, were used for the study. They were bought from Department of Agriculture, University of Calabar, Nigeria, and housed in well ventilated wooden cages in the animal house of the Department of Physiology, University of Calabar. The animals were given rat feed and water *ad libitum* and exposed to 12/12 hours light/dark cycle. All animals were allowed for seven days to acclimatize before treatment began.

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70 2.3 Animal Grouping and Extract Administration

The fifteen (15) rats were randomly assigned into three (3) groups (n = 5) thus: control, chilli pepper and capsaicin groups. All groups had access to rat feed and water. The control group was treated with 0.2 mL normal saline. Chilli pepper was administered at 5mg/100g body weight daily for 30 days, while capsaicin was administered at 3 mg/100g body weight daily. All treatments lasted for 30 days.

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76 2.4 Collection of blood sample

At the end of the 30 days period, all animals were sacrificed under chloroform anaesthesia. Blood samples were collected through cardiac puncture using 5 ml syringes with 21G needles into sample bottles and EDTA vials for measurement of serum electrolytes concentration and haematological parameters respectively.

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82 2.5 Assessment of Haematological Parameters

Haematological parameters were measured using automated cell counter (Coulter Electronics, Luton, Bedfordshine, UK) having standard calibrations in line with the instructions of the manufacturer. Parameters measured were: red blood cells (RBC) count, haemoglobin (Hb) concentration, packed cell
volume (PCV), mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH), mean
corpuscular haemoglobin concentration (MCHC), platelet count, platelet distribution width (PDW), mean
platelet volume (MPV) and platelet large cell ratio (P-LCR).

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90 **2.6 Assessment of Serum Electrolyte Concentration**

91 The blood samples collected were allowed 1h to clot and retract. The serum was obtained after 92 centrifuging blood in the sample bottles at 300rpm at room temperature for 15 minutes using a bucket 93 centrifuge machine (B-Bran Scientific and Instrument Company, England). The serum was used to 94 determine serum Na⁺, K⁺, Cl⁻ and HCO₃⁻ levels using ion-selective electrolyte analyser (Biolyte 2000/ 95 BioCare Corporation, Hsinchu 300, Taiwan).

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2.7 Assessment of Serum Concentration of Creatinine and Urea

Assay for creatinine was carried out using the Reflotron Dry Chemistry Analyzer as described by Estridge
 et al. [28]. Blood urea concentrations were determined using Berthelot's reaction as described by Kaplan
 and Teng [29].

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102 2.8 Statistical Analysis

Results are presented as mean ± standard error of mean (SEM). Data were analysed using One way
 analysis of variance (ANOVA) along with post hoc multiple comparison test (Tukey test) using Statistical
 Package for Social Science (SPSS) (version 17.0). p<0.05 was considered statistically significant.

107 3. RESULTS

1083.1Comparison of haematological parameters in the different experimental groups109following 30 days of treatment with chilli pepper and capsaicin

110111 *Red blood cell indices*

Table 1 shows the RBC count (x10⁶ cell/ μ L), Hb concentration (g/dL), PCV (%), MCV, MCH and MCHC for control, chilli pepper and capsaicin groups. RBC count and Hb concentration were significantly (p<0.01) reduced in chilli pepper and capsaicin groups compared with control. PCV was also significantly (p<0.01) lower in chilli pepper and capsaicin groups compared with the control group. There was no significant difference in MCV and MCH in the different experimental groups. MCHC was significantly (p<0.01) increased in chilli pepper group compared with control and significantly (p<0.05) lower in capsaicin group compared with chilli pepper group.

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120 Table 1: Comparison of red blood cell indices between the different experimental groups

Parameters	Control	Chilli Pepper	Capsaicin
RBC (x10 ⁶ cell/µL)	10.17 ± 0.36	8.22 ± 0.39 ^b	8.16 ± 0.27 ^b
Hb (g/dL)	16.33 ± 0.23	14.26 ± 0.72 ^b	13.76 ± 0.25 ^b
PCV (%)	59.46 ± 1.10	51.96 ± 2.17 ^b	47.28 ± 0.64 ^{b,x}
MCV	58.74 ± 1.48	63.40 ± 1.59 ^{№S}	58.20 ± 1.53 ^{NS}
MCH	16.15 ± 0.51	17.35 ± 0.39 ^{№S}	16.91 ± 0.29 ^{NS}
MCHC	27.48 ± 0.32	40.00 ± 0.42 ^b	29.10 ± 0.35 [×]

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Values are expressed as mean \pm SEM, n = 5. NS = not significant vs control; b = p<0.01 vs control;

p = p < 0.01 vs control; x = p < 0.05 vs chilli pepper.

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• Platelet indices

Table 2 shows platelet count $(x10^3 \text{ cell/}\mu\text{L})$, PDW (fL), MPV (fL) and P-LCR (%) for control, chilli pepper and capsaicin groups. Platelet count was significantly (p<0.01) lower in capsaicin group compared with control and chilli pepper groups. PDW was not significantly changed in the different experimental groups.

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131 MPV was significantly (p<0.05) increased in chilli pepper group compared with control. P-LCR was 132 significantly (p<0.01; p<0.001 respectively) reduced in capsaicin group compared with control and chilli

133 pepper groups.

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135 **Table 2: Comparison of platelet indices between the different groups**

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Parameters	Control	Chilli Pepper	Capsaicin	
Platelet Count (x10 ³ cell/µL)	972.86 ± 50.82	1115.80 ± 52.29 ^{NS}	787.00 ± 78.25 ^{b, y}	
PDW (fL)	8.45 ± 0.24	8.32 ± 0.20 ^{NS}	7.41 ± 0.63 ^{NS}	
MPV (fL)	10.05 ± 0.70	13.44 ± 1.36 ^a	11.92 ± 0.51 ^{№S}	
P-LCR (%)	8.60 ± 0.23	12.42 ± 0.72 ^{NS}	6.54 ± 0.14 ^{b, z}	
Values are expressed as mean \pm SEM, n = 5.				

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NS = not significant vs control;

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a = p < 0.05, b = p < 0.01, vs control;

y = p < 0.01, z = p < 0.001 vs chilli pepper.

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3.2 Comparison of serum concentration of electrolytes in the different experimental groups following 30 days of treatment with chilli pepper and capsaicin

Results for serum concentrations of Na⁺ (mmol/L), K⁺ (mmol/L), Cl⁻ (mmol/L), HCO₃⁻ (mmol/L), creatinine 145 (µmol/L) and urea (µmol/L) for control, chilli pepper and capsaicin groups are shown in table 3. Na⁺, Cl 146 147 and urea did not differ significantly in the different experimental groups. K⁺ was significantly increased (p<0.001) in chilli pepper group and reduced (p<0.01) in capsaicin group compared with control. K⁺ was 148 also significantly (p<0.001) reduced in capsaicin group compared with chilli pepper group. HCO₃ was 149 significantly (p<0.05) increased in capsaicin group compared with control but did not change significantly 150 151 between control and chilli pepper groups. Creatinine concentration was significantly reduced in chilli 152 pepper (p<0.01) and capsaicin (p<0.05) groups compared with control (Table 3).

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154Table 3: Comparison of serum concentration of electrolytes, creatinine and urea between the155different experimental groups

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Control	Chilli Pepper	Capsaicin
147.14 ± 0.34	147.60 ± 0.43 ^{NS}	145.60 ± 1.02 ^{NS}
8.60 ± 0.23	12.42 ± 0.72 °	6.54 ± 0.14 ^{b, z}
103.14 ± 0.56	103.20 ± 1.05 ^{NS}	98.00 ± 0.46 ^{NS}
19.43 ± 0.48	22.40 ± 0.43 ^{NS}	28.00 ± 0.27 ^a
82.71 ± 1.98	70.40 ± 3.69 ^b	71.20 ± 2.19 ^a
2.70 ± 0.07	2.34 ± 0.06 ^{NS}	2.54 ± 0.07 ^{NS}
	$\begin{array}{r} \textbf{Control} \\ 147.14 \pm 0.34 \\ 8.60 \pm 0.23 \\ 103.14 \pm 0.56 \\ 19.43 \pm 0.48 \\ 82.71 \pm 1.98 \\ 2.70 \pm 0.07 \end{array}$	$\begin{tabular}{ c c c c c } \hline Control & Chilli Pepper \\ \hline 147.14 \pm 0.34 & 147.60 \pm 0.43 & \end{tabular} \\ \hline 8.60 \pm 0.23 & 12.42 \pm 0.72 & \end{tabular} \\ \hline 103.14 \pm 0.56 & 103.20 \pm 1.05 & \end{tabular} \\ \hline 19.43 \pm 0.48 & 22.40 \pm 0.43 & \end{tabular} \\ \hline 82.71 \pm 1.98 & 70.40 \pm 3.69 & \end{tabular} \\ \hline 2.70 \pm 0.07 & 2.34 \pm 0.06 & \end{tabular} \\ \hline \end{tabular}$

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Values are expressed as mean ± SEM, n = 5. NS = not significant vs control;

a = p < 0.05, b = p < 0.01, c = p < 0.001 vs control;

z = p < 0.001 vs chilli pepper.

161 162 **4. DISCUSSION**

163 Chilli pepper is a spice that is widely grown worldwide and used as a basic ingredient in most global 164 cuisine. Its effects are elicited mainly by its active substance, capsaicin, which also gives chilli pepper its 165 pungent sensation. Chilli pepper and capsaicin have been reported to affect the physiology of the human 166 body in different ways. This study investigated and compared the effect of ethanolic extract of chilli 167 pepper and capsaicin on blood parameters and serum electrolytes concentration in female Wistar rats.

168 Red blood cell count, Hb concentration and PCV were significantly decreased in all treated groups

169 compared with the control. PCV was significantly decreased in capsaicin group compared with chilli

170 pepper group. These results contradict the report of Dougnon *et al.* [9], which showed that chilli pepper 171 fed to Hubbard broilers did not significantly change haematological parameters but corroborates that of 172 Shahverdi et al. [10] which showed decreased PCV and Hb concentration following treatment with chilli 173 pepper on broilers. Our results suggest that both chilli pepper and capsaicin may have inhibitory effect on 174 erythropoiesis which is marked by the decreased RBC count, but the effect is greater with capsaicin. PCV 175 was decreased probably because of the decreased RBC count. It is also likely that heme biosynthesis 176 was impaired along with erythropoiesis which led to decreased Hb concentration. Capsaicin has been 177 reported to reduce the levels of haemoglobin despite taking high iron diet [13]. From our study, it is also likely that chilli pepper and capsaicin may have suppressed the synthesis of iron which may have led to 178 179 the presence of microcytic erythrocytes resulting in the reduced Hb concentration observed.

180 MCV and MCH were not significantly changed in all experimental groups. MCHC of capsaicin group was 181 not significantly changed compared with control. These results indicate that the effect which chilli pepper 182 and capsaicin may have on Hb concentration is a mild one. But MCHC was significantly increased in chilli 183 pepper group compared with control, and significantly reduced in capsaicin group compared with chilli pepper group. The non-significant change in MCV indicates that chilli pepper and capsaicin did not alter 184 185 the Na⁺ transport across cell membrane since Na⁺ transport into a cell is accompanied by water to 186 increase the intracellular volume. This is evident in our result which shows no significant change in Na⁺ 187 levels in all experimental groups. This may also mean that both capsaicin and chilli pepper at the 188 administered dosage, do not affect the renin angiotensin aldosterone system.

189 Alterations in some serum electrolytes were observed. Increased K^{+} elicits antihypertensive effect [30]. 190 From our results, chilli pepper increased K^{+} levels while capsaicin decreased K^{+} levels compared with 191 control. This shows that chilli pepper exhibit antihypertensive effect unlike capsaicin. But this 192 antihypertensive effect of chilli pepper must be very mild since Na⁺ was not significantly affected. Na⁺ is the major extracellular electrolyte implicated in hypertension. High levels of Na⁺ causes contraction of 193 194 blood vessels to increase such that a great force is required to pump blood with a consequent hypertension [31]. HCO₃ was significantly increased in capsaicin group compared with the control, but, it 195 was not significantly different between chilli pepper and control groups. HCO₃ is a marker for measuring 196 197 the pH of blood. It acts as a buffer to maintain the pH of blood and body fluids. The results of this study 198 shows that capsaicin has the capacity to increase the pH of blood, while chilli pepper has no significant 199 effect on blood pH.

200 Serum creatinine levels were significantly decreased in chilli pepper and capsaicin groups compared with 201 control group, but was not significantly different between the treated groups. Serum urea levels were not 202 significantly changed in all experimental groups. Our result for decreased creatinine levels is consistent, 203 unlike that of urea, with [7] who reported that crude ethanolic extract of chilli pepper decreased serum 204 creatinine and urea levels. Our results show that the reduction of RBC count, Hb concentration and PCV 205 observed is not due to destruction of RBC since erythrocytes destruction may be accompanied by 206 increased blood urea levels. Also, Chilli pepper and capsaicin, administered at the dosage used, may not 207 cause any damage to functional nephron since decreased levels of creatinine were observed in our study. 208 An increase in serum creatinine levels is basically observed if there is a marked damage to functional 209 nephrons [32, 33].

Platelet count was significantly reduced in capsaicin group compared with control and chilli pepper groups. Platelet count in chilli pepper group was not significantly different from that of control although an increase was observed. These results indicate that capsaicin may have an adverse effect on the clotting mechanism of the body unlike chilli pepper. However, chilli pepper and capsaicin may not have any effect on platelet size as indicated by the non-significant changes in PDW (Table 3). The decreased P-LCR observed in capsaicin group in comparison with control and chilli pepper groups shows that capsaicin affects platelet aggregation while chilli pepper has no significant effect on platelet aggregation.

218 **5. CONCLUSION**

From the results of our study, we therefore conclude that both chilli pepper and capsaicin do not cause serious electrolyte imbalance, but inhibit erythropoiesis, with capsaicin having a greater effect. However, their effects are neither haematoxic nor nephrotoxic. Capsaicin decreases platelet indices while chilli pepper does not have significant effect on platelet indices. Patients with blood coagulation disorders and bleeding disorders should use chilli pepper instead of capsaicin as they may worsen their condition with intake of capsaicin.

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228 Not applicable.229 .

230 ETHICAL APPROVAL (WHERE EVER APPLICABLE)

All authors hereby declare that "Principles of laboratory animal care" (NIH publication No. 85-23, revised 1985) were followed, as well as specific national laws where applicable. All experiments have been examined and approved by the appropriate ethics committee

236 237 **REFERENCES**

- Rohanizah AR, Ishak M. Phytochemical contents of *Capsicum Frutescens* (Chili Padi), *Capsicum Annum* (Chili Pepper) and *Capsicum annum* (Bell Pepper) aqueous extracts. International Conference on Biological and Life Sciences IPCBEE vol.40 (2012) © (2012) IACSIT Press.
- Omolo MA, Wong Z, Mergen AK, Hastings JC, Le NC. Antimicrobial Properties of Chili Peppers. J Infect Dis Ther. 2014; 2: 145. doi:10.4172/2332-0877.1000145
- Jones NL, Shabib S, Sherman PM. Capsaicin as an inhibitor of the growth of the gastric pathogen Helicobacter pylori. FEMS Microbiol Lett. 1997; 146: 223-227.
- Wee YC, Hsuan K. An illustrated Dictionary of Chinese Medicinal Herbs. Times Edition, Eu Yan Seng Holdings Ltd 1990.
- 5. Jolayemi ATE, Ojewole JAO. Effects of Capsaicin on Coagulation: Will this be the New Blood Thinner. Clinical Medicine Research. 2014; 3(5): 145-149.
- Hanhineva K, Torronen R, Bondia-Pons I, Pekkinen J, Kolehmainen M, Myk-kanen H, Poutanen K.
 Impact of dietary polyphenols on carbohydrate metabolism. International Journal of Molecular
 Sciences. 2010; 11: 1365–1402.
- 252 7. Ogbonnaya EA, Muritala IK. Effect of crude ethanolic extract of *Capsicum frutescens* fruit on cisplatin-induced renal insufficiency in rats. J. Phys. Pharm. Adv. 2014; 4(2): 332-336.
- 8. Elamin HMS, Mohamed KA, Mukhtar MA. Effect of hot red pepper (*Capsicum frutescens*) on performance, abdominal fat and blood serum parameters of broiler. Journal of Global Biosciences. 2015; 4(5): 2251-2257.
- Dougnon TJ, Kiki P, Dougnon TV, Youssao I. Evaluation of Capsicum frutescens powder effects on the growth performances, biochemical and haematological parameters in Hubbard broiler. Journal of Applied Pharmaceutical Science. 2014; 4 (10): 38-43.
- Shahverdi A, Kheiri F, Faghani M, Rahimian Y, Rafiee A. The effect of use red pepper (Capsicum annum L) and black pepper (Piper nigrum L) on performance and hematological parameters of broiler chicks. Euro J Zool Res. 2013; 2(6): 44-48
- Al-Kassie GAM, Al-Nasrawi MAM, Ajeena SJ. The Effects of Using Hot Red Pepper as a Diet
 Supplement on Some Performance Traits in Broiler. Pakistan Journal of Nutrition. 2011; 10(9): 842 845.
- Akiba Y, Kato S, Katsube K, Nakamura M, Takeuchi K, Ishii H. Transient receptor potential vanilloid subfamily 1 expressed in pancreatic islet beta cells modulates insulin secretion in rats. Biochem Biophys Res Commun. 2004; 321: 219-225.
- Márquez-Ibarra A, Huerta M, Villalpando-Hernández S, Ríos-Silva M, Díaz-Reval MI, Cruzblanca H.
 The effects of dietary iron and capsaicin on haemoglobin, blood glucose, insulin tolerance, cholesterol, and triglycerides, in healthy and diabetic Wistar rats. PLoS ONE 2016; 11(4): e0152625.
 doi:10.1371/journal.pone.0152625
- 14. Chaiyata P. Effect of chili pepper (Capsicum frutescens) ingestion on glucose response, metabolic
 rate, lipid profile, lipid peroxidation, thrombogenic and fibrinolytic activities in hyperlipidemic Thai
 women. Doctoral dissertation. Bangkok: Research Unit Nutrition Faculty of Medicine Ramathibodi
 Hospital Mahidol University; 2003.
- 277 15. Chaiyasit K, Khovidhunkit W, Wittayalertpanya S. Pharmacokinetic and the effect of capsaicin in 278 *Capsicum frutescens* on decreasing plasma glucose level. J Med Assoc Thai. 2009; 92 (1): 108-113.
- In Jonietz P. Effect of red pepper and capsaicin on rat intestinal disaccharidases. J Sci Soc Thailand.
 1982; 8: 53-57.

UNDER PEER REVIEW

- In R, Pan J, Xie H, Zhou B, Xia X. Separation and quantitative analysis of capsaicinoids in Chili
 peppers by reversed-phase argentation LC. Chromatographia, 2009; 70(5–6), 1011.
- Peng J, Li YJ. The vanilloid receptor TRPV1: role in cardiovascular and gastrointestinal protection.
 Eur. J. Pharmacol. 2010; 627: 1–7.
- 19. Mortensen JM, Mortensen JE. The power of capsaicin. Journal of Continuing Education Topics &
 Issues. 2009; 11(1), 8-12.
- 287 20. Surh YJ. More than spice: capsaicin in hot chili peppers makes tumor cells commit suicide. J. Natl
 288 Cancer Inst. 2002; 94: 1263–1265.
- 289 21. Yang ZH, Wang XH, Wang HP, Hu LQ, Zheng XM, Li SW. Capsaicin mediates cell death in bladder
 290 cancer T24 cells through reactive oxygen species production and mitochondrial depolarization.
 291 Urology. 2010; 75: 735–741.
- 292 22. Leung FW. Capsaicin-sensitive intestinal mucosal afferent mechanism and body fat distribution. Life
 293 Sci. 2008; 83: 1–5.
- 23. Reinbach HC, Smeets A, Martinussen T, Moller P, Westerterp-Plantenga MS. Effects of capsaicin,
 green tea and CH-19 sweet pepper on appetite and energy intake in humans in negative and
 positive energy balance. Clin. Nutr. 2009; 28: 260–265.
- 297 24. Joo JI, Kim DH, Choi JW, Yun JW. Proteomic analysis for antiobesity potential of capsaicin on white
 298 adipose tissue in rats fed with a high fat diet. J. Proteome Res. 2010; 9: 2977–2987.
- 299 25. Limlomwongse L, Chaitauchawong C, Tongyai S. Effect of Capsaicin on Gastric Acid Secretion and Mucosal Blood Flow in the Rat. J. Nutr. 1979; 109: 773-777.
- Nishihara K, Nozawa Y, Nakano M, Ajioka H, Matsuura N. Sensitizing effects of lafutidine on CGRP containing afferent nerves in the rat stomach. Br. J. Pharmacol. 2002; 135: 1487–1494.
- Wang L, Hu CP, Deng PY, Shen SS, Zhu HQ, Ding JS, Tan GS, Li YJ. The protective effects of rutaecarpine on gastric mucosa injury in rats. Planta Med. 2005; 71: 416–419.
- 28. Estridge BH, Reynolds AP, Walters NJ. Basic medical laboratory techniques, 4th ed, Thomson
 Learning, USA, 2000.
- 307 29. Kaplan A, Teng LL. In Selected Methods of Clinical Chemistry, Vol. 9, Ed. By W.R. Faulkner and S.
 308 Meites, AACC, Washington, 1982; 357-363.
- 309 30. Cappucio FP, MacGregor GA. Does potassium supplementation lower blood pressure? A meta-310 analysis of published trials. J. Hypertens, 1991; 9: 465-473.
- 31. Vasudevan DM, Sreekumari S. Textbook of Biochemistry for Medical Students, 5th Ed. New Delhi:
 312 Jaypee Brothers Medical Publishers 2007; 239-246.
- 313 32. Davis ME, Berndt WD. Renal methods for toxicology. In: Hayes, A.W. (eds). Principles and methods of toxicology, 3rd Ed. New York Raven 1994; 871-894.
- 315 33. Mukinda JT, Eagles FK. Acute and subchronic oral toxicity profiles of the aqueous extract of 316 *Polygala fruticosa* in female mice and rats. J. Ethnopharmacol. 2010; 128: 236–240.