

Original Research Article

Polyphenol content and antioxidant activity of bark hydroethanolic extract of *Piliostigma reticulatum* (DC) Hochst and its fractions

Abstract

Objective: The purpose of this study was to evaluate the total phenol content and the antioxidant capacity of the hydro-alcoholic bark extract of *Piliostigma reticulatum* and its ethyl acetate, methanol and aqueous fractions.

Material and Methods: Powdered barks were extracted with ethanol (80°). From the crude extract, 3 fractions were obtained after fractionation by column chromatography with three different solvents (ethyl acetate, methanol and water). The polyphenol content was determined with Folin Denis reagent. Antioxidant activity was performed using DPPH (2, 2-diphenyl-picrylhydrazyl), ABTS (2,2'-azinobis (3-ethylbenzothiazoline 6-sulfonate) and FRAP (ferric reducing antioxidant potential) assays.

Results: The total extract had higher polyphenol contents with 12.55 ± 0.12 mg tannic acid equivalent/ g of dried extract (TAE/g). For ethyl acetate, methanol and water fractions, respective polyphenol content of 7.67 ± 0.4 - 9.01 ± 0.05 and 5.56 ± 0.2 mg TAE / g were obtained. The DDPH test had revealed that the methanol fraction was more active (IC_{50} : 30.83 ± 0.22 µg/ml), while for ABTS assay the ethyl acetate fraction had shown better activity (IC_{50} : 29.08 ± 0.44 µg/ml). For the FRAP test, at all tested concentrations, the methanol fraction revealed highest ability to reduce ferric ion with respective percentages of reduction of $38 \pm 0.73\%$ (concentration: 7.81 µg/ml) to $93.21 \pm 0.24\%$ (concentration: 250 µg/ml).

Conclusion: These studies showed that the methanol fraction exhibited the best antioxidant activity and that this activity may be related partially to its polyphenols content.

Keywords: *Piliostigma reticulatum*, bark, fractions, polyphenol, antioxidant.

1- INTRODUCTION

Piliostigma reticulatum (DC) Hochst, (Synonym: *Bauhinia reticulata*) is a common plant of Sahelo-Sudanian zone, from Senegal to Sudan. The traditional healers of Senegal mainly use its leaves and bark. These latters are often prescribed against many diseases such as ulcers [1], boils, wounds, syphilitic cancer, toothache, gingivitis and diarrhea [2,3,4]. Phyto-chemical analysis of the acetone bark extract showed the presence of polyphenols [5].

Furthermore, oxidative stress is often involved in the occurrence of many diseases causing irreversible damage to organism components such as lipids, proteins and DNA [6]. Antioxidant supplementation could have benefit health effect.

The aim of this study was to evaluate the total polyphenol content and the antioxidant capacity of the hydro-ethanolic bark extract of *Piliostigma reticulatum* and its ethyl acetate, methanol and water fractions.

2- MATERIAL AND METHODS

2-1. Plant material

Bark of *Piliostigma reticulatum* was collected in August 2015 in the forest of Diourbel, a region in central Senegal. The plant has been identified in the Laboratory of Pharmacognosy and Botanical of the Faculty of Medicine, Pharmacy and Odontology of the University Cheikh Anta Diop of Dakar. The barks were washed, dried at room temperature in an airy room of the laboratory and then reduced to powder.

2-2. Extraction and fractionation

A moderate decoction under reflux of 600 g of powder was carried out with 6 L of water-ethanol mixture (20v / 80v) for 30 minutes. After filtration with Whatmann No. 1 filter paper, the resulting filtrate was evaporated on a rotary evaporator at 60 ° C to obtain a dry extract.

Fractionation of the dry extract thus obtained on a silica column was carried out according to the modified method of Labourel and Péaud-Lenoel [7].

A cylindrical glass column 3 cm of diameter was filled with 100 g of silica (Scharlau GE 0048, 60A- 0.04-0.06 mm). After being well packed with a pump (50Hz), the silica was washed three times with 200 ml of ethyl acetate. Then 15 g of dry extract triturated before with 15 g of silica, were gently deposited at the top of the stationary phase. Elution was done by polarity gradient eluting successively with 1.5 L of ethyl acetate; 1.5 L of methanol and 1.5 L of distilled water with an average flow of 12.3 mL / min. The different fractions thus obtained were evaporated and dried by the same method as the total extract.

2-3. Total polyphenol content

Total polyphenol contents were evaluated according to the method described by Elgailani and Ishak [8]. To 1 ml of each dry extract in water (250 mg / L), 1 mL of Folin-Denis reagent was added shaking. Then after 3 minutes, 1 mL of 12% sodium carbonate was added and the whole is incubated in the dark for 2 hours at room temperature.

The absorbances of the samples were read at 725 nm against a blank without extract after centrifugation of the tubes at 3000 rpm for 5 minutes. The tests were repeated three times for each sample. A calibration line was established with tannic acid at different concentrations (0.01-0.015-0.02-0.03-0.045 mg / mL). The results were expressed in mg TAE/g.

2-4. Antioxidant activity

2-4-1. DPPH assay

The scavenging activity of free radicals of the samples was evaluated by the DPPH assay according to the method of Molyneux [9]. An ethanolic solution of DPPH[•] was prepared by dissolving 4 mg of this of DPPH reagent in 100 ml of ethanol followed by a cool incubation between 4-8 ° for at least 16 hours. Then, at 50 µL of the sample (extract, fractions or ascorbic acid) at different concentrations (250 - 125 - 62.5 - 31.25 - 15.62 - 7.81 µg / ml) were added 950 µL of the diluted DPPH solution.

Absorbances were measured at 517 nm after 30 mn of incubation in the dark at room temperature. Three tests were performed for each concentration. The antioxidant activity related to the DPPH free radical scavenging effect, is expressed in IC₅₀ (Inhibiting Concentration 50% of free radicals) using Statgraphics Plus 5.0 software

2-4-2. ABTS assay

The method of Leong and Shui [10] was used to evaluate the antiradical activity of the samples by the ABTS test. A 7 mM ABTS solution was prepared with distilled water. To obtain a solution of radical ABTS^{•+}, were mixed at equal volumes, a solution of ABTS at 7 mM and a solution of potassium persulfate at 2.5 mM. This solution ABTS^{•+} thus obtained was diluted with ethanol to obtain an absorbance of 0.700 ± 0.02 to 734 nm before use.

Then, 1.5 mL of the ABTS^{•+} solution was mixed with 50 µL of the sample (extract, fractions and Vitamin C) at different concentrations (250 - 125 - 62.5 - 31.25 - 15.62 - 7.81 µg / ml). Absorbances were measured at 734 nm after incubation for 10 minutes in the dark and at room temperature. Three tests were performed for each concentration and the results were expressed as IC₅₀ as described above for the DPPH test.

2-4-3. Ferric Reducing Antioxidant Power assay (FRAP)

The reducing power of the extract or fractions as well as that of the reference **were** determined by the FRAP method according Bassène [11]. Thus, 400 µL of each sample at different concentrations is mixed with 1 **mL** of phosphate buffer (0.2 M, pH = 6.6) and 1 **mL** of 1% potassium ferricyanide ($K_3[Fe(CN)_6]$). After 30 minutes of incubation at 50 ° C., 1 mL of 10% trichloroacetic acid was added before the tubes were centrifuged at 3000 rpm for 10 mn. Then, 1 **mL** of the supernatant from each tube was mixed with 0.2 **mL** of freshly prepared 0.1% ferric chloride solution. The tubes were incubated thereafter at room temperature away from light for 15 minutes before measuring the absorbances at 700 nm against a blank without sample. The results are expressed as percentage of reduction (PR) according to the following formula:

$$PR = [(A_X - A_B) / A_B] \times 100$$

A_x: absorbance sample tested; A_B: Blank absorbance

2-4-4. Statistical analyses

Significance tests were performed by the Fisher test using the StatView software. A value of $p < 0.05$ was considered statistically significant. Data were expressed as mean \pm SD.

3- Results

3-1. Extraction and fractionation

From 600 g of bark powder, 70 g of dry hydroethanolic extract were obtained representing a yield of 11.5%. Fractionation of 15 g of crude extract gave the fractionation yields mentioned in Table I.

Table I: Yields of different fractions of bark hydroethanolic extract of *P. reticulatum*.

	Weight (g)	Yield (%)
Hydroethanolic extract	15	
Ethyl acetate fraction	0.6	4
Methanol fraction	11.16	74.4
Aqueous fraction	0.41	2.73

3-2. Total polyphenol content

The polyphenol contents of the crude extract and its fractions were obtained from a tannic acid standard curve ($y = 24.90x - 0.13$; $R^2 = 0.9994$). The total extract was richer in polyphenols with 12.55 ± 0.12 mg TAE / g. Among the fractions, methanol had the highest content at 9.01 ± 0.05 mg TAE / g. For the ethyl acetate and aqueous fractions, respective contents of 7.67 ± 0.4 mg TAE / g and 5.56 ± 0.2 mg TAE / g were obtained (see Figure 1).

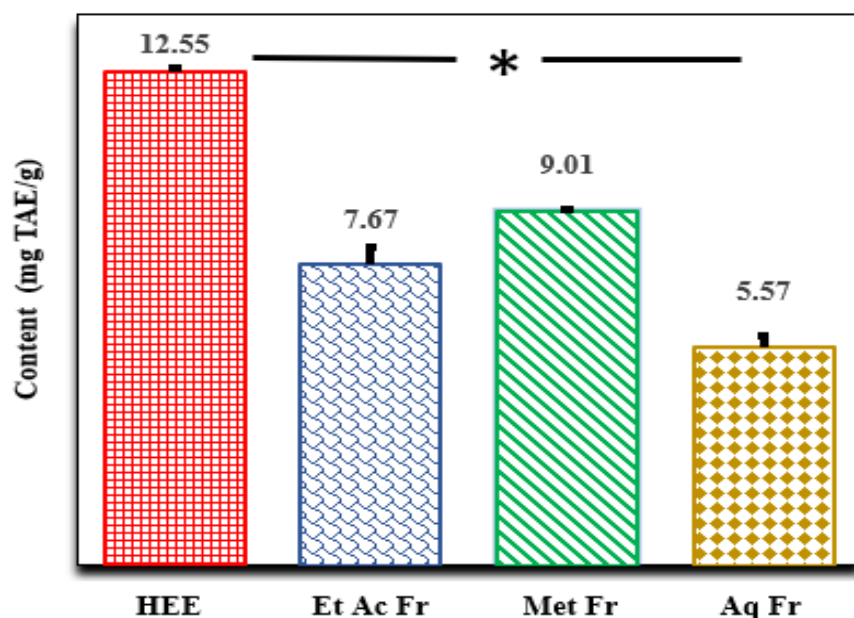


Figure 1 : Total phenol content of samples

HEE : Hydroethanolic extract; Et Ac Fr: Ethyl acetate fraction; Met Fr : Methanol fraction;
Aq Fr: Aqueous fraction; * : $p < 0.05$

3-3. DPPH assay

The root hydro-ethanolic extract of *P. reticulatum* had an IC_{50} of 19.92 ± 1.01 $\mu\text{g/mL}$. Among the fractions, the methanol had shown the lowest IC_{50} value (30.83 ± 0.22 $\mu\text{g/mL}$) comparatively to ethyl acetate fraction (IC_{50} : 52.33 ± 0.36 $\mu\text{g/mL}$) and aqueous fraction (IC_{50} : 103 ± 0.54 $\mu\text{g/mL}$). Ascorbic acid had an IC_{50} value of 18.6 ± 1.01 $\mu\text{g/mL}$ (Figure 2).

3-4. ABTS assay

ABTS test revealed that the lower IC_{50} value was obtained with ethyl acetate fraction (29.08 ± 0.44 $\mu\text{g/mL}$). The methanol and aqueous fractions had respectively IC_{50} values of $48.83 \pm$

0.74 $\mu\text{g/mL}$ and $58 \pm 1.46 \mu\text{g/mL}$, while that of crude extract was at $78.17 \pm 0.80 \mu\text{g/mL}$. Ascorbic acid had an IC_{50} value of $19.66 \pm 1.18 \mu\text{g/mL}$.

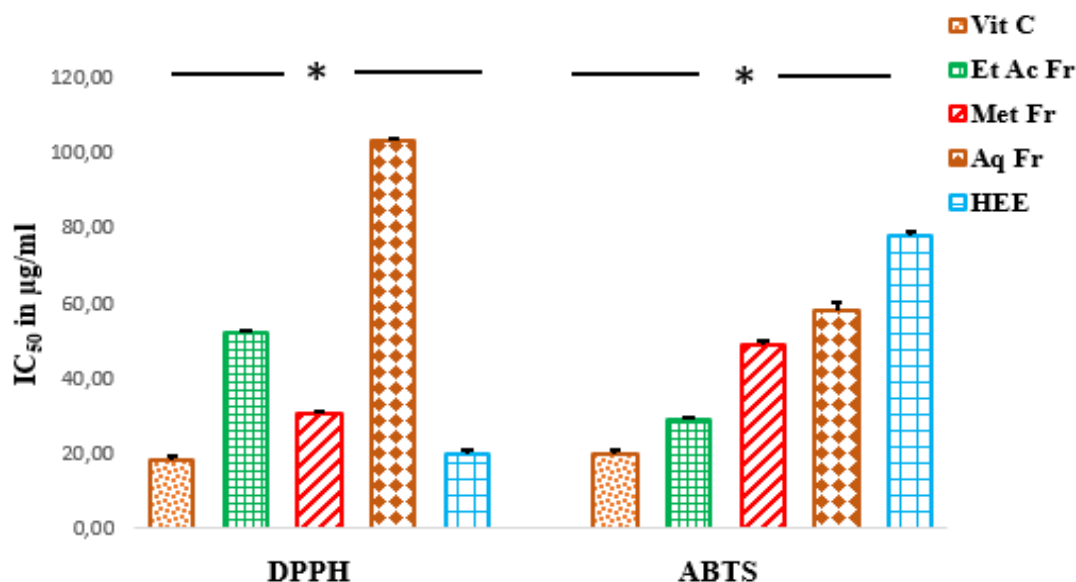


Figure 2 : IC_{50} of samples by DPPH and ABTS assays

Vit c : Vitamin C ; HEE: Hydroethanolic extract; Et Ac Fr: Ethyl acetate fraction; Met Fr : Methanol fraction ; Aq Fr: Aqueous fraction; * : $p < 0.05$

3-5. FRAP assay

Among all fractions, at all the concentrations tested, the methanol fraction showed the best ability to reduce ferric iron with PR from $38 \pm 0.73\%$ to $93.21 \pm 0.24\%$ respectively from concentrations of $7.81 \mu\text{g} / \text{mL}$ to $250 \mu\text{g} / \text{mL}$ (Figure 3). It is followed respectively by the ethyl acetate fraction (PR: $25.74 \pm 0.21\%$ and $87.21 \pm 0.01\%$ respectively at $7.81 \mu\text{g/mL}$ and $250 \mu\text{g/mL}$), the hydroethanolic extract (PR : $14.45 \pm 2.66 \%$ and $88.78 \pm 0.51\%$ respectively at $7.81 \mu\text{g/mL}$ and $250 \mu\text{g/mL}$) and aqueous fraction (PR: $14.17 \pm 1.12\%$ and $83.2 \pm 0.15\%$ respectively at $7.81 \mu\text{g} / \text{mL}$ and $250 \mu\text{g} / \text{mL}$). Vitamin C, showed the best activity of all samples tested (PR: $88.42 \pm 0.01\%$ and $97.09 \pm 0.01 \%$ respectively at $7.81 \mu\text{g/mL}$ and $250 \mu\text{g/mL}$).

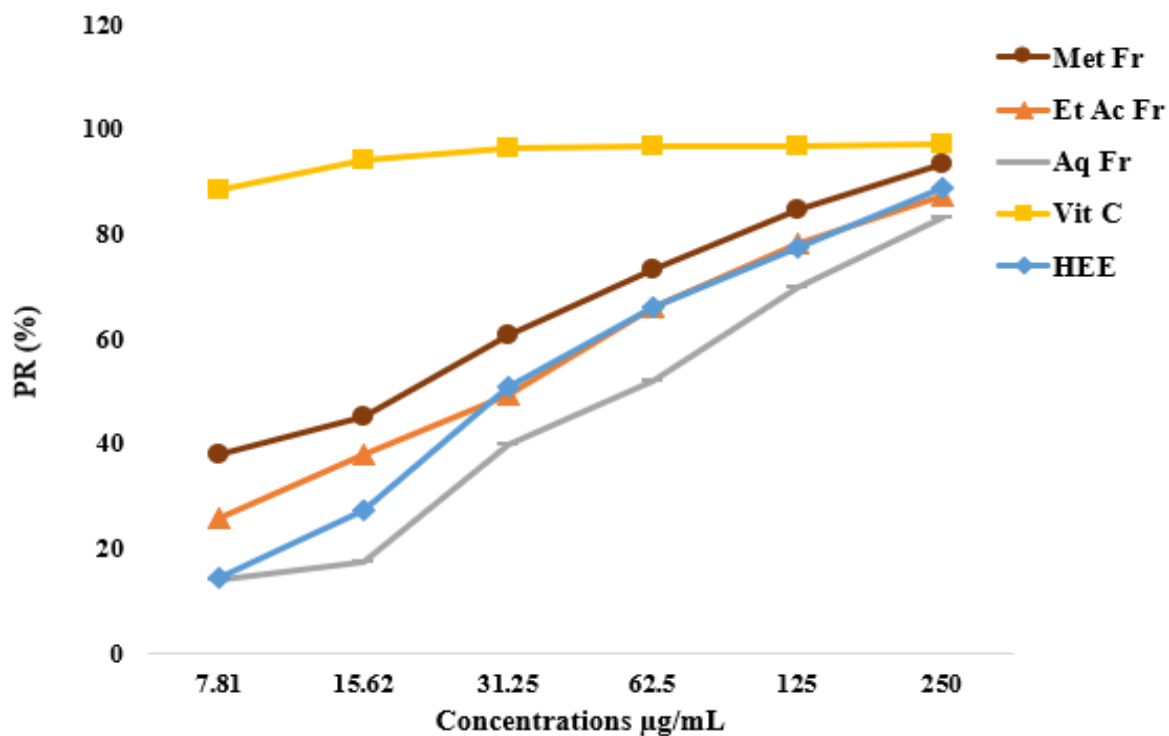


Figure 3 : Percentage reduction of different samples by FRAP

Vit c : Vitamin C ; HEE: Hydroethanolic extract; Et Ac Fr: Ethyl acetate fraction; Met Fr : Methanol fraction ; Aq Fr: Aqueous fraction

4. DISCUSSION

The hydroethanolic extraction of *Piliostigma reticulatum* barks was aimed to extract the maximum of polar compounds such as polyphenols because according to some studies [11,12,13], a compound tends to dissolve better in a solvent similarly polarity. Fractionation of the crude extract showed that the methanolic fraction presented the best yield (74.4%). This suggests that the majority of polar compounds have been eluted by methanol.

This seems to be confirmed by the total polyphenol contents of the different fractions with 9.06 ± 0.05 mg EAT / g dry extract for the methanolic fraction against 7.67 ± 0.4 and 5.56 ± 0.2 mg EAT / g for the ethyl acetate and aqueous fractions respectively ($p < 0.0001$). Compared with the Dieng studies [14], these results show that the methanolic and ethyl acetate fractions contain more polyphenolic compounds than the ethanolic bark extract of *Piliostigma thonningii* (Caesapiniaceae) which had a content equal to 6.16 mg EAT / g dry extract.

Polyphenols are among the major **plants** compounds presenting antioxidant activity [15, 16]. Thus, they could be partly responsible for the activity of the different fractions. With the DPPH test, the methanolic fraction had the best antiradical activity with an IC_{50} of $30.83 \pm 0.38 \mu\text{g} / \text{mL}$ whereas for the ABTS test, the activity was better with the ethyl acetate fraction ($29.08 \pm 0.76 \mu\text{g} / \text{mL}$) with a statistically significant difference from the other fractions for both methods ($p < 0.0001$).

Both DPPH and ABTS tests have similar mechanisms of action but with different wavelengths. The DPPH test is a simple and rapid **test but** often poses a problem of interpretation because some compounds may have overlapping absorbances at 517 nm like carotenoids [17]. While the ABTS test eliminates most of these interferences because of its 734 nm reading wavelength [18]. This could partly explain why the methanol fraction, although **richer** in polyphenols, exhibited a lower activity than the ethyl acetate fraction by the ABTS test. Moreover, natural antioxidants can act differently *in vitro* with respect to the radicals used or oxidizing sources because they have different chemical characteristics. Furthermore, many antioxidants react more slowly with DPPH or may even be inactive [17].

The antioxidant power of the samples, by their ability to reduce oxidants such as iron, seems to be corollary of the antiradical activity and their polyphenol content. The methanolic fraction presented, at all concentrations tested, the best reducing activity with a significant difference ($p < 0.0001$) compared to the other fractions. This activity is probably related to its polyphenols content because they are known for their radical scavenging and antioxidant properties [16, 18, 19]. Several plants or parts of plants are used for several reasons. Recent studies have shown a relationship between the use or consumption of products rich in polyphenols and prevention of many diseases such as cancer, coronary heart disease, inflammation, thrombosis and other [20]. This is linked in part by their ability to inhibit free radicals and thus the oxidation of some compounds such as LDL-cholesterol [3, 21]. This could explain the fairly frequent use of *P. reticulatum* bark extracts in Senegalese traditional medicine.

5-Conclusion

These studies showed that the methanol fraction exhibited the best antioxidant activity and that this activity could be partially linked by the presence of polyphenols. A subsequent study aimed to evaluate the anti-inflammatory, antimicrobial activity and isolation and identification of the compounds of the methanol fraction could be considered.

REFERENCES

- 1- Salawu OA, Tijani AY, Obidike IC, Rafindadi HA and Emeje M. Anti-ulcerogenic properties of methanolic root extract of *Piliostigma reticulatum* (DC) Hoechst (Syn. *Bauhinia reticulata* DC) -Leguminosae in rats. African Journal of Pharmacy and Pharmacology. 2009;3(5):252-258.
- 2- Arbonnier M. Arbres, Arbustes et Lianes des Zones Sèches d'Afrique de l'Ouest. CIRAD-MNHN, Paris; 576p. 2002.
- 3- Aderogba MA, Okoh EK, Okeke IN, Olajide AO and Ogundaini AO. Antimicrobial and Anti-inflammatory Effects of *Piliostigma reticulatum* Leaf Extract. International Journal of Pharmacology. 2006;2(1) :70-74.
- 4- Yelemou B, Bationo BA, Yameogo G, Millogo-Rasolodimby J. Gestion traditionnelle et usages de *Piliostigma reticulatum* sur le Plateau central du Burkina Faso. Bois et Forêts des Tropiques. 2007; 291 (1) : 55-66.
- 5- Sereme A, Millogo-Rasolodimby J, Guinko S, Nacro M. Propriétés thérapeutiques des plantes à tanins du Burkina Faso. Pharmacopée et Médecine traditionnelle Africaines. 2008; 15: 41 – 49
- 6- Barouki R. Stress oxydant et vieillissement. Médecine/sciences. 2006;22 (3):266-272.
- 7- Labourel G et Péaud-Lenoel C. Séparation par Chromatographie sur colonne de silice des glucofructosanes de la série inuline de D. P. entre 1 et 20. Chem. Zvesti. 1969;23 : 765-769.

- 8- Elgailani IEH, Ishak CY. Determination of Tannins of Three Common *Acacia* Species of Sudan. *Advances in Chemistry*, 2014;192708: 1-5.
- 9- Molyneux P. The use of the stable free radical diphenylpicrylhydrazyl (DPPH) for estimating antioxidant activity. *Songklanakarin J. Sci. Technol.* 2004 ; 26(2):211-219.
- 10- Leong LP, Shui G. An investigation of antioxidant capacity of fruits in Singapore markets. *Food Chemistry*. 2002; 76:69-75.
- 11- Bassène E. *Initiation à la Recherche sur les Substances Naturelles : Extraction- Analyse- Essais Biologiques*. Presse Universitaire de Dakar;140p. 2002.
- 12- Agban A, Gbogbo KA, Hoekou YP, Atchou K, Tchacondo T, Batawila K, De Souza C et Gbeassor M. Évaluation de l'activité antifongique des extraits de *Cassia alata* L. et de *Piliostigma thonningii* (Schumach.) Milne Redh. (Fabaceae) sur *Candida albicans*. *Int. J. Biol. Chem. Sci.* 2013;7(3):1041-1047.
- 13- Millogo-Koné H, Kini BF, Yougbaré Z, Yaro MB, Sawadogo M. Etudes de la phytochimie et de l'activité antimicrobienne *in vitro* des feuilles de *Moringa oleifera* (Moringaceae). *Revue CAMES - Série Pharm. Méd. Trad. Afr.* 2012 ;16 : 2-16.
- 14- Dieng SIMB, Fall AD, Diatta-Badji K, Sarr A, Sene M, Sene M, Mbaye A, Diatta W et Bassene E. Evaluation de l'activité antioxydante des extraits hydro-ethanoliques des feuilles et écorces de *Piliostigma thonningii* Schumach. *Int. J. Biol. Chem. Sci.* 2017.11(2): 768-776.
- 15- Akanni OO, Owumi SE, Adaramoye OA. *In vitro* studies to assess the antioxidative, radical scavenging and arginase inhibitory potentials of extracts from *Artocarpus altilis*, *Ficus exasperata* and *Kigelia africana*. *Asian Pac. J. Trop. Biomed.* 2014;4(1): S492-S499

- 16- Rice-Evans C, Miller N, Paganga G. Antioxidant properties of phenolic compounds. *Trends in Plant Science*.1997;2(4):152-159.
- 17- Prior RL, Wu X and Schaich K. Standardized methods for the determination of antioxidant capacity and phenolics in foods and dietary supplements. *Journal of Agricultural and Food Chemistry*.2005;53:4290-4302.
- 18- Li HB, Wong CC, Cheng KW, Chen F. Antioxidant properties *in vitro* and total phenolic contents in methanol extracts from medicinal plants. *LWT-Food Sci. Tech.* 2008;41: 385-390.
- 19- Maisuthisakul P. Phenolic Constituents and Antioxidant Properties of some Thai Plants. *Phytochemicals – A Global Perspective of Their Role in Nutrition and Health*. InTech, Rijeka.538p. 2012.
- 20- Latte KP, Kolodziej H. Antioxidant Properties of Phenolic Compounds from *Pelargonium reniforme*. *J. Agric. Food Chem.* 2004;52: 4899-4902.
- 21- Auddy B, Ferreira M, Blasina F, Lafon L, Arredondo F, Dajas F, Tripathi PC, Seal T, Mukherjee B. Screening of antioxidant activity of three Indian medicinal plants, traditionally used for the management of neurodegenerative diseases. *Journal of Ethnopharmacology*. 2003;84 : 131-138.