

Proximate and Chemical Evaluations of *Ocimum gratissimum* Plant grown with different Manure

ABSTRACT

The proximate, minerals, and phytochemical composition of *Ocimum gratissimum* plant (OGP) grew with different manure were investigated. OG seeds were planted in experimental plots in a Randomized Complete Block Design (RCBD) laid out in Oyo State College of Agriculture, Igboora with three replicates having plot size 3m x 3m of four treatments namely; Treatment A-No fertilizer or manure -(control), B- Poultry manure, C- N:P:K 15:15:15 and D-Compost manure respectively. Other parameters studied were plant height, leave areas, number of leaves and plant girth. The pre-soil test carried out indicated that the soil had organic matter(OM) content of 1.56%, total N 0.11% and available P 16.92mg/Kg. This low level of nutrients justified the need for fertilizer application for maximum production. It could be concluded that there was significant ($p<0.05$) differences in other parameters considered from 3 to 9 Weeks After Planting (WAP) on all the agronomic characters. Furthermore, the proximate and minerals of the OGP went in favour of Compost raised OG plant with highest values of significance followed by Poultry manure plant while N: P: K and No-manure treatments had the least ($P>0.05$) values respectively. The phytochemicals of the OGP which include Alkaloid, Flavonoid, Saponin and others were also high ($P<0.05$) in values on Compost and Poultry manure and the least value were observed in N. P. K and Control plants.

Keywords: *Ocimum gratissimum*, Manure, Phytochemical, Proximate and Minerals

1. INTRODUCTION

Ocimum gratissimum, a herbaceous plant belonging to the Lamiaceae family, germinate naturally in tropical areas especially, India and it is also found in the Savannah and Coastal areas of Nigeria in West Africa. The leaves have strong aromatic odour and are popularly used to flavour soup and spice meat. It is used in traditional medicine in the treatment of diarrhea [1]. Dalziel, 1956 as a febrifuge and component of anti-malaria remedies, [2], mosquito/insect repellent, stomachic and general tonic, antiseptic, in wound dressing, skin infections, conjunctivitis and bronchitis. An infusion of the leaves, called '*Ocimum* tea,' is dispensed as a remedy for fever and diaphoresis [1]. The roots are used as sedative for children [3]. The plant extract is an excellent remedy for cough. Therefore, the study aimed at analyzing the *Ocimum* raise with different manure in Ibarapa zone of Oyo State in Nigeria.

2. MATERIALS AND METHODS

MATERIALS

O. gratissimum seeds, experimental site, N.P.K fertilizer, compost manure, poultry manure, muffle furnace, soxhlet extractor, atomic absorption spectrophotometer, kjeldahl apparatus e.t.c.

METHODS

Experimental Land Design of *Ocimum gratissimum* Plants

The experimental layout was Randomized Complete Block Design with three replicates having plot size 3m x 3m treatments. The experimental soils were randomly sampled at depth 30cm. These samples were bulked, air dried and sieved using 2mm mesh before physico and chemical analysis. No fertilizer or manure (Control), Poultry manure (five months decomposed chicken droppings) and Compost (farmyard manures) were applied at 5 tonnes/ha while the N.P.K fertilizer was applied at 120kg/ha.

Ocimum gratissimum Plant Cultivation

Poultry manure and compost were applied a week before planting to enhance nutrient use efficiency by the crop while the N.P.K fertilizer was applied a week after transplanting. The plots were weeded manually whenever necessary throughout the experiment and harvested 70th days after planting. Growth parameter considered at 3-9 weeks after planting (WAP) includes plant height, number of leaves, leaf area and plant girth.

PROXIMATE ANALYSIS

Moisture content determination was carried out by drying 1g of each sample in an oven at 105⁰C until constant weights were obtained. Ash Content was analyzed by heating 2g of the sample in a muffle furnace at 600⁰C for 6hrs until it ashed completely. Crude fat was obtained by soxhlets extraction with petroleum ether at 600⁰C boiling range for 6 hrs. Determination of crude fibre was achieved by boiling 2g of the residue obtained from fat extraction with dilute tetraoxosulphate (vi) acid (H₂SO₄) and sodium hydroxide (NaOH), then washed with methylated spirit, thrice with petroleum ether and finally ashed. Crude protein content was estimated using kjeldahl techniques by digesting the samples in hot concentrated tetraoxosulphate (vi) acid (H₂SO₄) and distilled in alkaline medium into a boric acid (H₂BO₃) and finally titrated with hydrochloric acid (HCl), then % N was converted to the percentage crude protein by multiplying by 6.25. The carbohydrate content was calculated by difference, all as described by [4].

ELEMENTAL ANALYSIS

2g of each samples were subjected to 6hrs heating at 600⁰C in a muffle furnace until the ash turns to white completely, it was then washed with 10ml 0.1M HCl into a 100ml volumetric flask. Warmed to avoid frothing and filtered with Whatman No 42 (ashless) filter paper. Distilled water was added to the filtrate to make it up to 100ml for mineral determination. Samples for heavy metals were collected separately and acidified with concentrated nitric acid to keep the metals in solution and to minimize their absorption to the walls of the sample bottle. Atomic Absorption Spectrophotometer (AAS) was used to determine the level of metals in the samples; [5].

PHYTOCHEMICAL SCREENING

Tannins Determination

[6] method of determination of tannis was used. 20 ml of 50% methanol was added to 0.2 g weight of each sample in a beaker. The mixture was covered with paraffin and placed in a water bath at 77-80⁰C for 1hr, stirred thoroughly and filtered using Whatman No.1 filter

paper into a 100 ml volumetric flask. This was made up to mark with distilled water and thoroughly mixed. 2.5 ml of Folin-Denis reagent, 10 ml of 17% Na_2CO_3 and 20 ml of distilled water was added and properly stirred and mixed with 1ml of sample extract pipette into 50 ml volumetric flask. The mixture was made up to mark with distilled water, vigorously mixed and left standing for 20 min when a bluish-green colouration developed. Standard Tannic Acid solutions of range 0-10 ppm were treated similarly as 1 ml of sample above. Tannic Acid Standard solutions and the samples absorbances were read after colour development on a Spectrophotometer at a wavelength of 760 nm. Percentage tannin was calculated.

Saponin Determination

Based on the method of [7], 5 g of each plant samples was weighed and 100 ml of 20% ethanol was added and heated over a hot water bath at 55°C for 4 hrs with continuous stirring to ensure uniform mixing. The filtrate and the residue were re-extracted with another 100 ml of 20% ethanol. The combined extracts were concentrated to 40 ml on a water bath at 90°C. The concentrate was transferred into a separating funnel and 20 ml of diethyl ether was added and mixed vigorously. The aqueous layer was recovered while the ether layer was discarded. The aqueous layer was allowed to undergo this purification process again and about 30 ml of n-butanol was added. The combined n-butanol extracts were washed two times with 10 ml of 5% aqueous sodium chloride and heated in a water bath, after evaporation, the samples were dried in the oven to a constant weight. The saponin content was calculated as percentage of the sample.

Alkaloid Determination

This was determined by the method described by [7], 5g of each sample was weighed and addition of 200ml of 20% acetic acid in ethanol was done and allowed to stand for 4hrs. The mixture was filtered and the filtrate (extract) was concentrated on a water bath to evaporate 0.25 of the original volume. Ammonium hydroxide was added drop wise to the extract until the alkaloid precipitation was formed. The precipitate was collected by filtration after the whole solution has been left standing to settle. The residue(alkaloid) was dried and weighed.

Total flavonoid Determination

Using a standard method of [8], 10g of the samples was extracted repeatedly with 100ml of 80% aqueous methanol at room temperature. The whole solution was filtered using Whatman filter paper No. 42 (125mm). The filtrate was transferred into crucible and evaporated into dryness over water bath and weighed to a constant weight.

Steroids Determination

100 ml of distilled water was added to 5g of each plant sample and mix to achieve homogeneity. The filtrate from the homogenized mixture was eluted with normal NH_4OH solution of pH 9. Ice-cold acetic anhydride of 3ml was added to the mixture of 2 ml each of the eluents and chloroform in the flask and 2 drops of conc. H_2SO_4 were gently added to cool. Standard sterol solution was prepared and treated as described above. Spectrophotometer was used to measure the absorbances of standard and prepared samples at 420 nm. This method was described by [9].

Determination of Terpenoids (Salkowski Test)

5ml of each plant sample extract was mixed in 2 ml of chloroform, and 3 ml of conc. H_2SO_4 was cautiously added to form a layer. The presence of terpenoids was indicated by a reddish brown coloration formed at the interface [10].

Cardiac Glycosides Determination

0.5g of each sample was refluxed in methanol for about 2hrs. The extract obtained was treated with HCl, then further re-extracted using water and dried under anhydrous Na_2SO_4 . The weight of the extract was calculated as percentage of the sample. Modified method of [11].

Oxalate Determination

Oxalate extraction was done with dilute HCl at 50°C and was first treated with ammonium hydroxide and glacial acetic acid and re- treated with CaCl_2 solution. Precipitate of calcium oxalate formed was solubilized with hot dilute H_2SO_4 and titrated against KMnO_4 as equivalent to 2.2mg of oxalate. A method described by [12].

Statistical Analysis

Data from parameters investigated were analyzed in a completely randomized design using one way

Analysis of variance (ANOVA) procedure.

3. Results and Discussions

Results of the pre-soil analysis (Table 1) revealed the organic matter content of 1.56%, total nitrogen 0.11%, available phosphorus 16.92mg/Kg and potassium 1.14%. The soil was low in organic matter, nitrogen and calcium; this justified the need for fertilizer application for maximum yield. Table 2 revealed the total aggregates of the pre-soil analysis, gave sand, silt and clay ratio to be 78.4:11.6:10% respectively which indicated that the soil is sandy loam. However, the proximate analysis of manure (Table 3) indicated that compost had highest value of total nitrogen content of 3.35% and average phosphorus of 21.47 ppm compared with poultry manure which had 3.31% and 12.31ppm respectively and higher values of potassium, 9.50cmol/Kg. The ability of the compost to perform better may be due to appropriate decomposition of the composting materials and mineralization of nitrogen [13].

In Table 4 significant ($P < 0.05$) treatment effects were observed in all the agronomic traits considered. The replenishment of the depleted experimental soil nutrients was achieved through the application of different manure and this corresponds to report made by [14]. The experiment showed that compost based manure performed better in most of the characters considered than the use of poultry manure. The highest response was recorded with the addition of 120Kg/ha of N.P.K. The observed trend showed that the inorganic fertilizer gave an instant response to the crop need compared to organic manure with slow release of nutrients. The above assertion agrees with the findings of [15].

Table 1: Pre-soil analysis of the experimental site

Element	Values
pH	6.76
T°	28.4
Organic Matter(%)	1.56
Total Nitrogen(%)	0.11
Average Phosphorus	16.92
Potassium(cmol/kg)	1.14
Sodium(cmol/kg)	0.31
Calcium(cmol/kg)	0.16

Magnesium(cmol/kg)	0.38
--------------------	------

Table 2: Pre soil analysis

Soil	Texture
Sand	78.4
Silt	11.6
Clay	10
Sandy Loam	

Table 3: Proximate analysis of organic materials

	Poultry Manure	Compost (maize base)
% Total Nitrogen	3.31	3.35
Average Phosphorus (ppm)	12.31	21.47
Potassium (cmol/kg)	9.80	8.50
pH	8.74	9.32
Electric charge	7399	3125
Temperature (°C)	27.1	27.2

	Leaf Num ber			Plant Gro wth			Leaf Area			Plant Height		
	3W AP	6WA P	9W AP	3W AP	6W AP	9W AP	3WA P	6W AP	9W AP	3WA P	6WAP	9WAP
CONT ROL	8.33 ^a	13.20 _a	31.0 _{0^d}	0.20 ^b	0.37 ^c	0.53 ^c	0.79 ^a	3.95 ^d	7.97 ^a	1.33 ^a	1.80 ^c	9.38 ^c
PM	6.67 ^a	21.33 _a	40.6 _{7^c}	0.30 ^a _b	0.53 ^b _c	0.67 ^b _c	1.80 ^a	10.3 _{3^c}	12.0 _{4^c}	2.17 ^a	5.03 ^{bc}	13.29 ^b
COMP OST	6.67 ^a	25.30 _a	49.6 _{0^b}	0.26 ^b	0.67 ^a _b	0.80 ^a _b	1.00 ^a	12.8 _{3^b}	17.6 _{0^b}	2.57 ^a	7.37 ^{ab}	15.30 ^b
N:P:K	6.33 ^a	26.67 _a	63.6 _{7^a}	0.37 ^a	0.77 ^a	0.97 ^a	1.34 ^a	15.0 _{9^a}	25.9 _{7^a}	3.53 ^a	8.97 ^a	21.65 ^a
SE	2.04	3.3	0.62	0.02	0.04	0.05	0.45	1.21	0.59	0.56	1.21	0.59
STD	0.81	0.70	1.05	0.01	0.06	0.50	1.24	1.62	0.46	0.62	1.41	0.91
LSD	3.20	3.05	3.20	3.20	3.20	3.10	3.20	3.20	3.05	3.20	3.20	3.10

188 **Table 4: Growth parameters on Leave number, Plant growth, Leaf area and Plant height**

189 Key: PM = Poultry Manure; NPK 15:15:15; WAP = Week after Planting
190 abcd – means on the same column with different superscripts are significantly different
191 (P<0.05)

192 **Table 5: Proximate analysis of *Ocimum gratissimum* plant raised with different manure**

193

Parameters	TA (No manure)	TB (Poultry manure)	TC (N:P:K)	TD (Compost)
Moisture	77.17 ^b	77.53 ^{ab}	77.73 ^{ab}	78.76 ^a
Ash	5.93 ^c	7.00 ^b	5.83 ^d	7.03 ^a
Crude fibre	3.00	3.20	3.03	3.80
Crude protein	6.60 ^d	8.17 ^b	7.97 ^c	8.33 ^a
Ether Extract	2.20 ^d	3.17 ^b	2.23 ^c	3.33 ^a

194 ^{abcd} – means on the same column with different superscripts are significantly different
 195 (P<0.05)
 196

197 In table 5, proximate compositions of OGP raised with different manure evaluated shows that
 198 the highest moisture content (%) with 78.76 value in TD-compost manure was significantly
 199 (P<0.05) difference to other treatments, this was followed by TB-poultry manure with 77.53
 200 while the least value was found in the control (TA-no manure) with 77.17. The moisture
 201 contents were relatively high and the active ingredients are much higher in the fluid, therefore
 202 hindered micro-organisms growth and increase stored samples life span. This enhanced
 203 preservation thereby, prevent early spoilage [16]. Furthermore, as reported by [17], OG crude
 204 protein contents(%) contain 9.10, 9.80 and 9.22 respectively, which were higher in values
 205 than 7.00% reported by [18], however, lower than 29.78% reported by [19] and 23.74% [20]
 206 while the crude protein(%) in this study were relatively lower 6.60 (TA-No manure), 8.17
 207 (TB-Poultry manure), 7.97 (TC-N.P.K) and 8.33 (TD-Compost) than that reported by [17].
 208 Though it was cleared that TD-Compost (%) and TB- Poultry manure (%) Crude protein were
 209 significantly (P<0.05) different to all other manure grown, OGP however, the least crude
 210 protein mean value was in TA-No manure (Control). The Ash content (%) mean values were
 211 TA-Control (5.93), TB-Poultry manure (7.00), TC–N.P.K (5.83) and TD-Compost (7.03).
 212 These Ash mean values were lower than the values reported by [20] for *Ocimum gratissimum*
 213 leaves 22.84 and 15.09 by [21]. The values of the crude fat (%) for the leaves of OG reported
 214 by [17] were also relatively higher than the mean values reported in this research but almost
 215 equal to those reported as 4.80 by [19] and 3.15 by [16].

216

217 **Table 6: Elemental analysis of *Ocimum gratissimum* plant raised with different manure**
 218 **(mg/g)**

Parameters	TA (No manure)	TB (Poultry manure)	TC (N:P:K)	TD (Compost)
K ⁺	61.6 ^c	65.33 ^{ab}	62.67 ^b	68.33 ^a
Na ⁺	787.00 ^b	823.33 ^{ab}	795.33 ^b	853.33 ^a

Ca ²⁺	348.67 ^d	363.33 ^c	395.33 ^a	383.33 ^b
Mg ²⁺	45.67 ^b	42.67 ^c	41.33 ^c	48.33 ^a
Cl ⁻	185.00 ^b	186.00 ^b	175.33 ^c	193.33 ^a
Fe ²⁺	5.87 ^b	6.17 ^a	5.87 ^b	6.50 ^a
Cu ²⁺	0.33	0.40	0.30	0.40
Zn ²⁺	4.00 ^c	5.20 ^b	4.90 ^d	5.90 ^b
Cd ²⁺	0.10	0.10	0.10	0.10
Pb ²⁺	0.10	0.12	0.10	0.13

abcd – means on the same Column with different superscripts are significantly different (P<0.05)

The mineral composition in table 6 (mg/g) of *O. gratissimum* plants were analysed accordingly, the Potassium in TD-Compost grown was significantly (P<0.05) difference to others, followed by mean values of TB-Poultry manure grown *O. gratissimum* and least value was shown in Control-No manure. The Sodium, Magnesium and Chlorine follows the same trend of significance (P<0.05). However, Iron, an important trace element for haemoglobin, useful in the normal functioning of the central nervous system and in oxidation of carbohydrates, protein and fat [22], varies in mean values as detected in this analysis with highest value recorded in TD-Compost and TB-Poultry manure *O. gratissimum* raised plants while TA- Control has the lowest recorded mean value. The values of Zinc content in this study seems low compare to [23] reports, however, Zinc is involved in the normal functioning of immune system [24] and is associated with protein metabolism. The leaves are good source of Zinc because it relatively contains quantity as recommended by RDA [25]. The Lead values in this study range between 0.10mg/g – 0.13mg/g, which are lesser than the suggested concentration of 2mg/g – 6mg/g in the plant species [26]. This suggest that *O. gratissimum* does not contribute or rather cannot be used as a substitute for other blood forming leafy vegetables but can be used in boosting the immune system and are antioxidants micronutrient [27]. The Copper values in different OGPs grown manure ranges between 0.33mg/g to 0.40mg/g. It has been reported that copper deficiency could cause cardiovascular disorders as well as anaemia, bone disorder and nervous systems disorder. Also, Lead and Copper are highly toxic even at low concentrations. Finally, Cadmium concentration values in this study was 0.10mg/g in all the treatments though tolerable amount, although Cadmium is considered non-essential elements of the human being and existence, even in small amounts may cause some health problems such as kidney failure and heart disease [28];[29] and [30].

Table 7: Phytochemical component of *Ocimum gratissimum* plant raised with different manure

Parameters	TA (No manure)	TB (Poultry manure)	TC (N:P:K)	TD (Compost)
------------	----------------	---------------------	------------	--------------

Alkaloid	1299.67 ^b	1313.33 ^{ab}	1331.00 ^{ab}	1356.67 ^a
Saponin	475.00 ^a	473.67 ^a	462.67 ^c	468.33 ^b
Flavonoid	1332.67 ^b	1345.00 ^b	1245.00 ^c	1431.67 ^a
Tannis	835.33 ^b	875.33 ^{ab}	860.00 ^{ab}	935.00 ^a
Terpenoid	1868.33	1875.33	1815.00	1878.33
Oxalate	1.37 ^d	1.45 ^b	1.38 ^c	1.51 ^a
Steroid	11.67 ^b	9.33 ^d	10.00 ^c	12.67 ^a
Cardiac glycoside	5.90 ^c	7.71 ^a	6.60 ^b	7.83 ^a

abcd – means on the same Column with different superscripts are significantly different (P<0.05)

Table 7 showed the phytochemicals of OGPs raised with different manure. The plants contained terpenoid which have excellent activity against *Bacillus subtilis* and *Staphylococcus aureus* and lesser activity against gram negative bacteria as well as *Candida albicans*. Alkaloids has been found to have microbiocidal effect and the major anti-diarrheal effect is probably due to their effects on small intestine and also possess antihypertensive, antifungal, anti-inflammatory, antifibrogenic effect [31]. The presence of alkaloids in this different manure grown OGPs, especially compost manure make them recommendable for patient with the above listed challenges. Tannin is non-toxic and can generate physiological responses in animals that consume them [32] but toxic to filamentous fungi, yeast and bacterial. However, the quantity discovered in this study had highest quantity in compost grown followed by poultry and least was found in no manure. Saponins showed a positive result in the leaves of *O. gratissimum*, in the order of TA-Control (475.00mg/g), TB-Poultry manure (473.67mg/g), TD-Compost (468.33mg/g) and TC–N.P.K (462.67mg/g). This compound has been reported to kill or inhibit cancer cells without killing the normal cells in the process [33]. Steroid also showed a positive result in OG leaves, TD-Compost (12.67), TA-Control (11.67), TC–N.P.K (10.00) and TB-Poultry manure (9.33) which are of importance in pharmacy as sex hormones [34], promote immune function in the skin and also reduce inflammation [35]. In this study, the leaves of OG in all treatments also contained flavonoid which modifies the body's reaction to allergens, virus and carcinogens. It has been reported to show anti-inflammatory and antimicrobial activity [36]. [37] reported the presence of flavonoids in *A. hybridus*. Cardiac glycoside showed positive result in the leaves, [38] justifies the plants in treatment and management of hypertension. The Oxalate content of the OG for all the treatments are TD-compost (1.51mg/g), TB-Poultry manure (1.45mg/g), TC-N: P: K (1.38mg/g) and TA-Control (1.37mg/g) respectively. These values are higher than *Musa sapentum* with 0.72mg/g [39] and almost equal to *Butryospermum parkii* with 1.5mg/g [38], *Spondias mombin* with 0.9mg/g [40] and lower than the reported values for cotton leaves of 22mg/g [38].

4. CONCLUSION

It is established that high content of anti-nutrients prevent the absorption of essential elements necessary for body metabolism. Reduction of anti-nutrients in foods may be necessary especially when their levels are higher than those generally regarded as safe for human consumption. This can be accomplished through different hydrothermal treatments, which also enhances the nutritional qualities: increase palatability and digestibility of foods [41].

As a rich source of proximate, minerals and phytochemicals, OG is a potential source of nutrients, drugs, good health and longevity. The study also verified the usefulness of OG for nutritional purposes and partly explains its uses in herbal medicine. The agronomy traits of OG raised with N: P: K fertilizer had the highest significant values. Also, the proximate and minerals analysis went in favour of compost raised OGP followed by poultry manure plant while the N: P: K and No manure treatment had the least values. The phytochemical of OGP were also high in values in compost and poultry manure while the least values were observed in N: P: K and Control grown plant samples. This research conclusively proved that most of the analysis carried out went in favour of OG raised with compost manure. However, OGP can be used for target purpose because the levels of anti-nutrient do not reach lethal dosages. [42].

REFERENCES

1. Iwu M. M. 1993. *Handbook of African medicinal plants*. Florida: CRC Press Inc. Boca Raton.
2. Oliver, B. 1960. Medicinal plants in Ibadan, Nigeria: *Nigerian College of Arts, Sci. and Tech.*
3. Di Stasi, L. C., Oliveira, G. P., Carvalhaes, M. A., Queiroz, M., Jr, Tien, O. S. and Kakinami, S.H. 2002. Medicinal plants popularly used in the Brazilian Tropical Atlantic Forest. *Fitoterapia*, vol. 73: pp. 69–91.
4. A.O.A.C. 1996. Association of Official Analytical Chemists. *Official Methods of Analysis*, 15th ed. AOAC press, Gaithersburg, USA.
5. A.O.A.C. 1996. Association of Official Analytical Chemists. *Official Methods of Analysis*.
6. Swain T. Tannins and Lignins. In: Rosenthal G.A, Jansen DH. (ed.) *Herbivores, their interaction with secondary plant metabolites*. Academic Press. New York. 1979;65:74-82.
7. Obadoni, B. O. and Ochuko, P. O. 2001. Phytochemical studies and comparative efficacy of the crude extracts of some homeostatic plants in Edo and Delta States of Nigeria. *Global. Pure Appl. Sci.*, vol. 8: pp. 203-208.
8. Boham, A. B. and Kocipai, A. C. 1994. Flavonoid and condensed tannins from leaves of *Hawaiian vaccinium vaticulum* and *vicalyinium*. *Pacific Sci.*, vol. 48: pp. 458-463

9. Okeke, C. U. and Elekwa, I. Z. 2003. Phytochemical study of the extracts of *Gongronema latifolium* (Asclepiaceae). *J. Health Visual Stud*, vol. 5: pp. 47–55.
10. Harborne J. B. 1973. Phytochemical methods: A guide to Modern techniques of plant analysis. Chapman A and Hall, London. Pp 279.
11. Siddique, S. Hafeez, F. and Begum (1987). Isolation and structure of two cardiac glycosides from the leaves of *Nerium olender*. *Phytochemistry* 26(1): pp237-241.
12. Munro and Basir, O. 1969. Oxalate in Nigeria vegetables. *W. Afri. J. Biol. Appl. Chem.*, vol, 12: pp 14-18.
13. Esuf Zai, A. K., Horiuchi, T., Matsui, T. 2008. Effects of green manure and compost of pea plant with chicken manure and rapeseed oil residues on growth, yield and efficiency of nutrient uptake by wheat (*Triticum aestivum*) *Compost Sci.util.*, vol. 16, No. 4: pp. 20-25.
14. Adeniyi, O. N. and Ojeniyi S. O. 2005. Effect of poultry manure and NPK 15:15:15 and combination of their reduced levels on maize growth and soil chemical properties. *Nig. J. Soil Sci.*, vol. 15: pp. 34-41.
15. Adeniyi, O. N., Ojo, A. O., Akinbode, O. A. and Adediran, J. A. 2011. Comparative study of different Organic manures and NPK fertilizer for improvement of soil chemical properties and dry matter yield of maize in two different soils. *J. of Soil and Environmental Management*, vol. 2, No. 1: pp. 9-13.
16. Abolaji, O. A., Adebayo, A. N. And Odesanmi, O. S. 2007. Nutritional qualities of three medicinal plants. *Xylopi aethiopica*, *Blighia sapida* and *Parinari polyandra*. *Pakistan J. of Nutr.*, vol. 6: pp. 665-668.
17. Mlitan, A. M., Sasi, M. S. and Alkherraz, A. M. 2014. Proximate and minor mineral content in some selected basil leaves of *Ocimum gratissimum* L in Libya. *International J. of Chemical Engr. and Application*, vol. 5, No. 6.
18. Isong, E. U. and Idan, U. U. 1997. Comparative studies on the nutritional and toxic composition of three varieties of *Lieanthera africana*, *Plants Food and Human Nutr*, vol. 51: pp. 79-84.
19. Akindahunsi, A. A. and Salawu, S. O. 2005. Phytochemical screening and anti-nutrient composition of selected tropical green vegetables. *African J. Biotech.*, vol. 4: pp. 497– 502.
20. Pandey, M., Abidi, A. M., Singh, S. and Singh, R. P. I. 2006. Nutritional evaluation of leafy vegetables paratha, *J. Human Ecology*, Vol.19: pp 155-156.

21. Antia, S., Akpan, E. J., Okon, P. A. And Umoren, I. U. 2006. Nutritive and antinutritive evaluation of sweet potatoes (*Ipomea batata*) leaves. *Pakistan J. of Nutr*, vol.5: pp. 166-168
22. Adeleye, E. and Olokiki, M. K. O. 1999. Proximate composition and some nutritionally valuable minerals of two varieties of *Capsicum annu*. *Discovery innovations*, vol. 11: pp. 75-81.
23. Elegbede, J. A., Eka, A. U. and Osagie, O. U. 1998. Legumes, nutritional quality of plant foods: Post Harvest Research Unit, University of Benin. Pp. 53 - 83
24. Ibrahim, N. D. G., Abdulrahman, E. M. And Ibrahim, G. 2001. Elemental analysis of the leaves of *Veronia anygodaleina* and its biological evaluation in rats, *Nigeria Journal of natural products Med*, vol. 5: pp. 13-16.
25. Borgert, G. M. Saunder, W. B. Calloway, H. 1975. Nutritional and physical fitness. Philadelphia, U. S. A. Pp. 34-50.
26. Broyer, T. C., Johnson, C. N. and Pull, R. 1972. Some aspect of lead in plant nutrition. *Plant soil*, vol. 36: pp. 301 – 305.
27. Talwar, G. P., Striva, L. M. and Mudgi, K.O. 1989. *Textbook of biotechnology and human biology*, 2nd ed prentice, half of Indian private Ltd. pp. 100-105
28. Asaolu, S. S., Ipinmoroti, K. O., Adeyinwo, C. E. and Olaofe, 1997. Seasonal variation in heavy metals distribution sediments of Ondo State Coastal region, *Ghana Journal of Chemistry*, vol.3: pp. 11- 16.
29. Melcarz, G. W., Howard, A. N., Williams, N. R., Kinsman G. D., Mizushima, Y. S. and Yamori, Y. 1997. Copper and zinc status as a risk factor for ischemic heart diseases. *The Journal of Trace Elements in Experimental Medicine*, vol. 10, issue 1, pp. 29-35
30. Fagbohun, E. D., Lawal, O. U and Ore, M. E. 2012. The proximate, mineral and phytochemical analysis of the leaves of *Ocimum gratissimum* L. *International Journal of Applied Biology and Pharmaceutical Technology*, vol. 3: pp. 15-22.
31. Ghosal, S., Krishna-Prasad, B. N. and Laksmi V. 1996. Anti-amoebic activity of *Piper longum* fruits against *Entamoeba histolytica* in vivo. *Journal of Ethno-pharmacology*, Vol. 50: pp. 167-170.
32. Scalbert, A. 1991. Antimicrobial properties of tannis. *Phytochemistry* . Vol. 30: pp 3875-3882.
33. Lewis, W. H. and Elvin-Lewis, M. P. 1995. Medicinal plants as sources of new therapeutics. *Ann. Mo. Bot. Gard*, vol. 82: pp.16-24
34. Okwu, D. E. 2001. Evaluation of chemical composition spices and flavouring agents. *Global J. Pure Applied Sci.*, vol. 7: pp. 455-459

- 432
- 433 35. Bell, E. 2008. Vitamin D₃ promotes immune function on the skin Available:
434 <http://www.signalinggateway.org/update/updates/2007>.
435
- 436 36. Cushnie, T. P. and Lamb A. J. 2005. Antimicrobial activity of flavonoids: *International*
437 *Journal of Antimicrobial Agents*, vol. 26, No 5: pp. 343-356
438
- 439 37. Akubugwo, E., Obasi N. A., Chinyere, G.C. and Ugobogu, A. E. 2007. Nutritional and
440 chemical value of *Amaranthus hybridus*. L. *African J. of Biotech.*, vol. 6, No. 24: pp.
441 2883-2889.
- 442 38. Taiwo, C., Abidemi, J., Oyedepo, B., Adebayo, I., Oluwadare, D. and Agboto, D. 2009.
443 Nutrient content and antinutritional factor in Shea butter (*Butryospermum parkii*) leaves.
444 *African J. of Biotech.*, Vol. 8, No. 21:pp 5888-5890.
445
- 446 39. Baiyeri, P. 2000. Effects of nitrogen fertilization on mineral concentration in plaintain
447 (Musasp) fruit peel and pulp at unripe and ripe stages. *Plant Product Research J.*, vol.
448 5:pp. 38-43.
449
- 450 40. Igwe, C. U., Onyeze, G. O. C., Onwuliri, V. A., Osuagwu, A. O. and Ojiako, C. G.
451 2010.
452 Evaluation of chemical composition of leaf of *Sponias mombin* Lin. *Nigeria Journal of*
453 *Basic and Applied Science*, vol. 4, No. 5: pp. 706-710.
454
- 455 41. Adeniji, T. A L., Sanni, I. O., Barimala, I. S. and Hart A.D. 2007. Antinutrients and heavy
456 metals in some new plaintain and banana cultivars. *Nigerian Food J.*, vol.25: p. 2.
457 .
- 458 42. A.O. A. C. 2005. Association of Official Analysis Chemist, *Official methods of analysis*.
459 15th
460 ed. Washinton D.C. pp. 774-784.