



PART 1:

Journal Name:	Current Journal of Applied Science and Technology
Manuscript Number:	Ms_CJAST_42299
Title of the Manuscript:	Molecular Detection of Antimicrobial Peptide Genes and Identification of Novel Antifungal Compounds of <i>Bacillus</i> sp. against <i>Fusarium oxysporum</i> f. sp <i>cubense</i> Causing Wilt of Banana
New Title of the Manuscript:	Molecular Detection of Antimicrobial Peptide Genes and Identification of Antifungal Compounds of <i>Bacillus</i> sp. against <i>Fusarium oxysporum</i> f. sp <i>cubense</i> Causing Wilt of Banana
Type of Article:	Original Research Article

FINAL EVALUATOR'S comments on revised paper (if any)	Authors' response to final evaluator's comments
Final Review on June 7, 2018: Corrections were made on the original documents and on final evaluation form. Author requires including explanations necessary for various sections that were not completely submitted in the revised manuscript. Editions were made in text directly in gray and major corrections in red. Majority of corrections were observed were in reference, results and methods section that were not in line with SDI author guidelines. The details were briefly jotted down here. □ In Introduction, author did not include the value of the crop to answer the weakness 1 stated before. In Materials and methods The sub title, "Isolation of pathogenic fungi (foc) has some errors. □ I do not understand the sterilisation techniques described which is far from the regular methods used by Tutte, 1969. If it is for isolation of microbes inside the corm or on the surface of the corm, why is it required to use mercuric chloride and streptomycin that are antifungal and antibacterial solutions. Mercuric chloride is harmful chemical and a steriliser which kills microbes and antibiotic streptomycin is very specific to kind of bacteria we are interested to select. Kindly incorporate the one that was followed for isolation of fungal strain FOC from corm bits of banana. □ If the purpose of sterilisation is to isolate the resistant pathogenic fungal strain FOC in rhizome bits after rhizome treatment, then please include that statement. □ Incubation temperature was $27 \pm 2^\circ\text{C}$ instead of $20 \pm 2^\circ\text{C}$ that was reported by Vinod Kumar et al in 2017. Please specify the reasons for modification of incubation temperature for Banana FOC. □ Isolation of bacillus species and screening their activity against FOC was exactly similar to the procedure explained by Vinodh Kumar et al..	I have given the detailed information about the crop and made all corrections suggested by reviewer. In materials and methods, changed the streptomycin chlorite instead of mercuric chloride.



SDI FINAL EVALUATION FORM 1.1

2017, Please delete this subhead and replace it with a single sentence, Please incorporate a sentence for changing the incubation temperature to $27\pm 2^{\circ}\text{C}$ for screening FOC

☐ Reasons for selecting only six AMP genes is missing in subheading "PCR Amplification of antibiotic genes" as well as in introduction to justify the purpose and scope of present study. If there are only six genes identified from the bacillus cereus strains of that location, please incorporate that statement.

☐ PLEASE CHECK THE TEMPERATURES for DENATURATION, ANNEALING AND EXTENSION AND THE TIME IN MINUTES AND NUMBER OF CYCLES for PCR for reference Ramanathan, 2007.

☐ I did not understand why the denaturation was done for 40 cycles at 94 degrees after initial cycle of denaturation at 94 for 3 min. (A BIG REVISION in methods, needs explanation for employing 94 degrees temperature for amplification of a gene

☐ I am sorry for not letting you know in the first review, I over looked and was not expected a mistake in standard PCR protocol and primers.

☐ If the PCR conditions are same for the stated five genes and primers used, then it is a good to include the PCR information in the beginnig para before the primer information of genes. Or the AMP genes can be placed in a separate table with primer information used for amplification of transcript.

☐ Further, if the presented PCR conditions are correct from your past experience in amplification of bacterial AMP genes, please explain why those conditions are required for amplication of bacterial gene in specific with reasons and supportive literature.

☐ Author again did not answer the weaknesses stated in the previous reviews for changing the incubation temperatures for bacterial strains 27 and 28 instead of 20.

☐ Author did not provide significance information for any data set presented in the manuscript either in the text or for the tables.

☐ Author still did not mention the samples, definition of variables, replicates, design of the experiment in methods

☐ Please follow the correct temperature or range that was executed during the experimental incubation time

Made the corrections for incubation temperature of *Fusarium* $27\pm 2^{\circ}\text{C}$.

Corrected the denaturation of some AMP genes. All the corrections were made as suggested by reviewer.