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2 **Plant growth promotion and biocontrol potential of a**
3 ***Streptomyces* sp. strain N3-3b** isolated from the rhizosphere of
4 ***Chakhao*, a black rice variety of Manipur, India**

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12 **Abstract**

13 **Aims:** To isolate and investigate actinomycete strains from rhizospheric soil of *Chakhao* (black rice)
14 in Manipur, India for biocontrol and plant growth promoting activities.

15 **Study design:** Dual culture, plant growth promoting activities and vigor index.

16 **Place and Duration of Study:** Microbial Biotechnology Research Laboratory (MBRL), Department of
17 Biochemistry, Manipur University, India, between August 2010 and July 2013; Re-analyzed and
18 reviewed during August 2015-May 2016.

19 **Methodology:** Isolates were screened for biocontrol activity and one strain which showed significant
20 antagonistic potential was selected for further studies. The selected strain was subjected to plant
21 growth promoting traits such as IAA, ammonia and siderophore production, and inorganic phosphate

22 solubilization. It was further assayed for rice seed germination and growth of rice seedlings under *in*
23 *vitro* conditions. Characterization of the strain was also done.

24 **Results:** Among 122 putative actinomycetes isolates, 9 exhibited antagonistic activity against rice
25 fungal pathogens. N3-3b was selected as the most promising biocontrol strain for further studies. The
26 strain exhibited plant growth promoting traits including IAA production, Phosphate solubilisation and
27 Ammonia production. It promoted seed germination and significantly enhanced growth of seedlings
28 (vigor index) under *in vitro* conditions. Based on the phenotypic characteristics and 16S rRNA gene
29 sequence analysis, the strain N3-3b was characterized as a member of the genus *Streptomyces*.

30 **Conclusion:** Strain N3-3b could be a potential candidate for development as bioinoculant for rice
31 cultivation.

32 **Keywords:** Chakhao, actinomycetes, biocontrol, PGP, *Streptomyces*, Vigor Index, *Chakhao* rice
33 Manipur.

34 1. Introduction

35 Synthetic agrochemicals pose serious risks to human health and the ecosystem and this warrants
36 development of alternative approaches such as microbial biofertilizers and biocontrol agents including
37 the use of bioinoculants for plant growth promotion and disease control (1). Microorganisms may
38 facilitate plant growth directly by nitrogen fixation (N fixation), phosphate solubilization (P
39 solubilization), siderophore and phytohormone production or indirectly by antagonism, HCN
40 production, niche competition, ACC deaminase production and induced systemic resistance (ISR) (2).
41 The importance of native strains and ecological specificity has also been emphasized (3).
42 Rhizospheres of crop plants are generally richer in microbial population due to release of root
43 exudates. Among the rhizobacteria, the genus *Bacillus* and *Pseudomonas* have been intensively
44 studied but actinobacteria have received comparatively lesser attention (4).

45 Among the Actinobacteria, *Streptomyces* spp. have shown great potential as prolific producer of
46 bioactive metabolites (5,6,7). So, native *Streptomyces* strains from rice rhizosphere and other
47 biotopes could potentially be biocontrol agents (BCAs) and plant growth promoting (PGP) agents for

48 rice. Two actinomycete bioinoculants, Mycostop (*Streptomyces griseoviridis* strain K61) and
49 Actinolron and Actinovate (*Streptomyces lydicus*) are commercially available (8).

50 Black rice cultivation has a long history especially in Asia (9). The dark purple color of black rice is
51 due to high anthocyanin content in the pericarp layers (10). More than 200 black rice cultivars are
52 grown in parts of Asia including Manipur in India (11). It was known as 'Emperor's Rice' or 'Forbidden
53 Rice' in ancient China as only the royal family was allowed to consume it then.

54 Besides being a special food item, black rice has several health benefits and is considered a
55 medicinal food, nutraceutical and functional food. It is richer in protein, fiber, B vitamins, niacin, Vit. E
56 and minerals such as Ca, Mg, Fe, and Zn compared to white rice (12). The high anthocyanin content
57 and associated protective and antioxidant action makes black rice a 'superfood' (13). Black rice has
58 been reported to have cardioprotective, anticancer, and antiatherogenic effects (9).

59 Manipur has several endemic varieties of black rice. The most popular cultivar, *chakhao* (also known
60 as *chakhao amubi*, literally means delicious black rice) is grown at certain parts of the Imphal valley in
61 Manipur. Due to its poor yield (about 2.5 tons/hectare), *chakhao* is grown in very limited acreage by
62 farmers in Manipur for ceremonial and cultural purposes (14). As it is gaining popularity due to its
63 nutraceutical properties, there is urgent need to enhance its productivity. As there are no high
64 yielding varieties yet, research on development of microbial inoculants with biocontrol and PGP
65 potential for *chakhao* is urgently warranted.

66 The rice variety *Chakhao* (black rice) endemic to Manipur is rich in nutritional values and contains 18
67 amino acids, iron, zinc, copper and carotene. The dark purple colour is due to its high anthocyanin
68 content and presence of vitamin E renders antioxidant property. It may help prevent cancer, diabetes,
69 heart disease and Alzheimer's disease (15,16). As the chemical composition of the root exudates of
70 *chakhao* might be different from that of other rice varieties, *chakhao* rhizosphere may harbor different
71 microbial profile. Hence, the present study was aimed at isolation and characterization of bioactive
72 actinomycetes from *chakhao* rhizosphere and screen their PGP and biocontrol activities.

73 **2. Material and methods**

74 **2.1. Rhizosphere sample collection**

75 Rhizospheric soil samples were collected from a rice field in Nambol region of Manipur, India
76 cultivating the local rice variety 'Chakhao' (black rice). The soil was clayey type with a pH of 5.5.
77 Nambol (24.7 °N, 93.84 °E) is located at a distance of 18.1 km from Imphal, the capital city of
78 Manipur.

79 **2.2. Isolation of Rhizospheric Actinomycetes**

80 Soil samples were treated with CaCO₃ (10%) and air dried for one week. They were then suspended
81 in 100 mL of sterilized distilled water and were serially diluted. 0.1 ml of each diluted sample was
82 spread plated on Starch Casein Nitrate Agar (SCNA) medium and kept incubated at 30°C for 4-5
83 days. Morphologically distinct isolates were selected and subcultured on SCNA plates till pure
84 cultures were obtained.

85 **2.3. Screening for Biocontrol activity**

86 The isolates were screened for biocontrol activity by dual culture assay (17) against five rice fungal
87 pathogens viz. *Rhizoctonia solani* (MTCC 4633, Sheath Blight Disease) and *Pyricularia oryzae*
88 (MTCC 1477, Blast Disease), *Bipolaris oryzae* (MTCC 3717, Brown Spot Disease), *Rhizoctonia*
89 *oryzae-sativae* (MTCC 2162, Aggregate Sheath Blight Disease), *Fusarium oxysporum* (MTCC 287,
90 Root Rot Disease). Mycelial growth inhibition was calculated using the formula: (C-T)/C x 100, where
91 C is the colony growth in control (mm), and T is the colony growth of pathogen in dual-culture (mm).
92 The isolate showing highest percentage of mycelial growth inhibition was selected for further studies.

93 **2.4. Screening for PGP traits**

94 **2.4.1. Indole-3-acetic acid**

95 The production of IAA was determined according to the method of Bano and Musarrat (18). The strain
96 was inoculated in SCN broth (SCNB) containing 2 mg/ml of L-tryptophan (trp) (HiMedia) and
97 incubated in a shaker (150 rpm, 30 °C, 6 d). The culture broth was centrifuged at 10,000 rpm for 10
98 min. One ml of the supernatant was mixed with 2 ml of Salkowski reagent. Appearance of pink colour
99 indicated IAA production.

100 Quantitative assay of IAA production at different trp concentrations (%) was also studied by
101 inoculating the strain in SCNB containing different concentrations (%) of trp (0, 0.2, 0.4, 0.6, 0.8, 1,
102 1.2, 1.4) and kept incubated under shaking conditions (150 rpm, 30 °C, 6 d). The culture broth was
103 centrifuged at 10,000 rpm for 10 min. One ml of the supernatant was mixed with 2 mL of Salkowski
104 reagent and incubated for 20 min at room temperature. Optical density (OD) was read at 530 nm and
105 the amount of IAA produced was calculated by comparing with the standard IAA (Rankem) curve.

106 **2.4.2. Phosphate (P) solubilization**

107 Phosphate (P) solubilization assay was done using NBRIP-BPB medium (19). A halo zone
108 surrounding the colony after 4 d of incubation at 30 °C indicated P solubilization.

109 Quantitative estimation of P solubilization was done according to Kapri and Tewari (20). Strain was
110 inoculated in 100 ml of NBRIP medium and kept incubated in a shaker (150 rpm, 30 °C, 6 d). The
111 culture broth was centrifuged at 10,000 rpm for 10 min. The amount of P in the culture supernatant
112 was estimated using the method of Fiske and Subbarow (21), and expressed as equivalent P ($\mu\text{g/ml}$).
113 KH_2PO_4 was used as the standard.

114 **2.4.3. Siderophore production**

115 Siderophore production was assayed according to You et al. (22) with few modifications. Agar plug (8
116 mm) of strain N3-3B was inoculated on SCNA (without iron) amended with CAS-substrate and kept
117 incubated at 30 °C for 6 d. Halo zone with orange colour surrounding the colony was considered as
118 positive for siderophore production.

119 **2.4.4. Ammonia production**

120 Ammonia production was screened in peptone water. Strain was inoculated in 10 ml peptone water
121 and kept in a shaker (150 rpm, 30 °C) for 4 d. 0.5 ml of Nessler's reagent was then added in each
122 tube. Development of brown to yellow colour indicated ammonia production (23).

123 **2.5. In vitro seed germination test (Vigor index)**

124 Strain N3-3b was grown on SCNB for 6 d, centrifuged (10,000 rpm, 10 min) and the pellet collected
125 was washed thrice with sterile distilled water (SDW). The pellet was dissolved in SDW and different

126 inoculum sizes were prepared (3×10^7 , 6×10^7 , 1.2×10^8 , 1.8×10^8 , 2×10^8 and 2.4×10^8 cfu/ml). Rice
127 seeds (Variety: *Chakhao*) were surfaced sterilized with 0.2% sodium hypochlorite for 5 min followed
128 by 70 % ethanol for 5 min and rinsed four times with SDW. Sterilized seeds were soaked in the cell
129 suspensions prepared earlier and kept overnight. Sterilized seeds soaked in SDW were taken as
130 control. The seeds were dried under laminar flow and then transferred to sterile plates containing
131 wetted filter papers at the rate of 10 seeds per plate. Plates were incubated at 28-30°C and after 4 d,
132 the number of germinated seeds, root lengths and shoot lengths were noted and compared with
133 controls. Four replications were done per treatment and the experiments were repeated twice. Vigor
134 index was calculated using the formula shown as follows(24):

135 Vigor index = Percent germination x Seedling length (shoot length + root length).

136 **2.6. Biochemical, morphological and molecular characterization**

137 Biochemical tests viz. Catalase production, Gelatin liquefaction, Citrate reduction, Indole production,
138 MR and VP tests, Oxidase production, Nitrate reduction and Sugar fermentation (Glucose, sucrose,
139 fructose, lactose, maltose and mannitol) were performed as described by Cappuccino and Sherman
140 (23). Utilization of sole carbon and nitrogen sources was determined as described by Shirling and
141 Gottlieb (25). Growth morphologies of the strain were observed in different International *Streptomyces*
142 Project (ISP) media (25). The colony colour was determined according to the ISCC-NBS colour chart
143 (26). Growth at different salt concentrations (0-10% NaCl) and pH values (4 to 10) was also
144 evaluated.

145 Genomic DNA extraction and PCR amplification of the 16S rRNA gene was performed as described
146 by Li et al. (27). The almost complete 16S rRNA gene sequence of the strain was identified using the
147 EzTaxon-e server database (28) and aligned with the 16S rRNA gene sequences of related species
148 using CLUSTAL X version 2.1 (29). Phylogenetic analyses were performed using the software
149 package MEGA version 5 (30). Phylogenetic distances were calculated with the Kimura two-
150 parameter model (31) and tree topologies were inferred using the neighbour-joining method (32). To
151 determine the support of each clade, bootstrap analysis was performed with 1000 resamplings (33).

152 **2.7. Antibiotic sensitivity**

153 Antibiotic sensitivity tests were performed using a total of six antibiotics viz. neomycin (30 µg),
154 chloramphenicol (30 µg), ampicillin (10 µg), penicillin (10 µg), streptomycin (10 µg) and rifampicin (5
155 µg) (HiMedia) for the sensitivity / resistance pattern of the isolate against the antibiotics by paper disc
156 method.

157 **2.8. Statistical analysis**

158 All data were subjected to one-way ANOVA followed by independent t-test ($P \leq 0.05$) using the SPSS
159 16 software (SPSS Inc).

160 **3. Results**

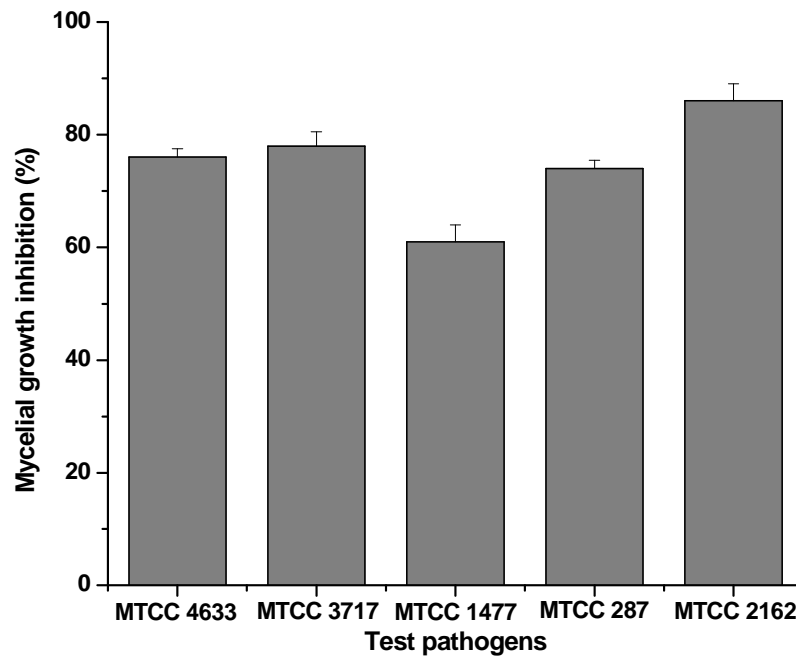
161 **3.1. Isolation and selection**

162 Of 122 actinomycete isolates obtained from Chakhao rhizospheric soils, 9 showed antagonistic
163 activity against the tested fungal pathogens. Strain N3-3b was selected for further studies as it
164 exhibited the highest percentage of mycelial growth inhibition.

165 **3.2. Biocontrol activity**

166 Strain N30-3b could inhibit mycelial growth in the range of 61 to 86%, showing highest inhibition
167 against *Rhizoctonia oryzae-sativae* (86%) and lowest against *Pyricularia oryzae* (61%) (Figure 1).

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170 **Figure 1. Percent mycelial growth inhibition of various rice fungal pathogens by *Streptomyces***
 171 **sp. N3-3b**

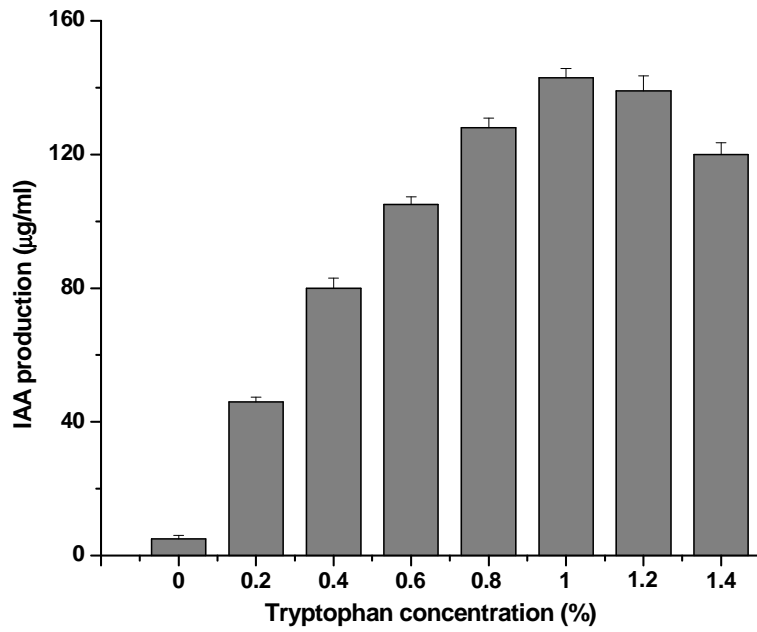
172 Note: MTCC 4633, *Rhizoctonia solani*; MTCC 3717, *Bipolaris oryzae*; MTCC 1477, *Pyricularia oryzae*,
 173 MTCC 287, *Fusarium oxysporium*, MTCC 2162, *Rhizoctonia oryzae-sativae*

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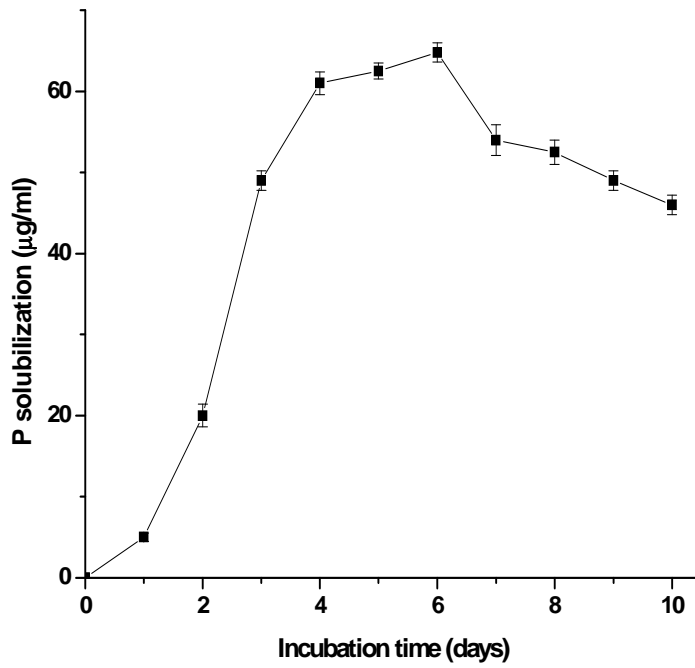
176 3.3. Plant growth promotion activity

177 Plant growth promotion assays indicated that the strain N3-3b was positive for P solubilization, IAA
 178 and ammonia production but negative for siderophore production. Strain N3-3b produced maximum
 179 amount of IAA when amended with 1% trp (143.7µg/ml) (**Figure 2**). It could also solubilize maximum
 180 amount of P (64.80 µg/ml) after 6th d of incubation (**Figure 3**).



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182 **Figure 2. IAA production by N3-3b at different tryptophan concentrations**



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184 **Figure 3. P solubilization by N3-3b at different time intervals**

185 **3.4. In vitro seed germination test (Vigor index)**

186 Among the different inoculum densities, inoculum size corresponding to 6×10^7 cfu/ml showed the
187 highest vigor index. N3-3b treated seeds showed higher germination percentage, vigor index and
188 significant increases in shoot and root lengths ($P \leq 0.05$) over the control. Treatment of seeds with
189 inoculum size corresponding to 6×10^7 cfu/ml showed significant increase in root and shoot length (P
190 ≤ 0.05) of rice seedlings over other inoculum sizes (**Table 1**).

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198 **Table 1. In vitro seed germination test (Vigor Index)**

| <i>Treatment</i> | <i>Inoculum size</i> <i>($\times 10^8$ cfu/ml)</i> | <i>Germination</i> <i>percent</i> | <i>Root length*</i> <i>(cm)</i> | <i>Shoot length*</i> <i>(cm)</i> | <i>Vigor index</i> |
|------------------|--|--------------------------------------|------------------------------------|-------------------------------------|--------------------|
| Control | | 95 | 4.07±0.34a | 1.2±0.59a | 500.65 |
| N3-3b | 0.3 | 95 | 4.41±0.11b | 1.24±0.04a | 536.75 |
| | 0.6 | 100 | 4.8±0.06c | 1.75±0.08c | 655 |
| | 1.2 | 100 | 4.58±0.05b | 1.48±0.08b | 606 |
| | 1.8 | 95 | 4.50±0.07b | 1.6±0.04b | 610 |
| | 2 | 100 | 4.57±0.04b | 1.47±0.07b | 604 |
| | 2.4 | 95 | 4.29±0.13a | 1.18±0.04a | 519.65 |

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200 *Values with the same letter within a column are not significant at $P \leq 0.05$

201 **3.5. Biochemical, morphological and molecular characterization**

202 The strain N3-3b was positive for nitrate reduction, indole production, citrate utilization, catalase
 203 activity, lipase production and gelatin liquefaction tests. It utilized galactose, lactose, ribose, mannitol,
 204 mannose, maltose, raffinose and fructose as sole carbon source. It could utilize asparagine,
 205 tryptophan, adenine, arginine, leucine, aspartic acid, glutamic acid and tyrosine as sole nitrogen
 206 source. The strain N3-3b was able to grow at a wide range of pH (5 - 10) and could tolerate up to 5%
 207 NaCl (**Table 2**) The cultural characteristics of N3-3b on different ISP media were observed using the
 208 ISCC-NBS colour chart (**Table 3**).

209 Strain N3-3b showed highest 16S rRNA gene sequence similarity (99.68%) with *Streptomyces*
 210 *castelarensis*. Based on the phylogenetic and genomic data, the strain was found to represent a strain
 211 of the genus *Streptomyces* which has now been referred to as *Streptomyces* sp. strain N3-3b (**Figure**
 212 **4**).

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217 **Table 2. Biochemical and physiological characteristics of the strain N3-3b**

| Test | Result | Test | Result | Test | Result |
|---------------------------|--------|----------------------------|--------|----------------------------|--------|
| Citrate utilization | + | Sole C- source utilization | | Sole N- source utilization | |
| Indole production | + | Galactose | + | Asparagine | + |
| Gelatin liquefaction | + | Lactose | + | Tryptophan | + |
| | | Ribose | + | Adenine | + |
| Methyl Red (MR) Test | - | Mannitol | + | Arginine | + |
| Voges Proskauer (VP) test | - | Mannose | + | Leucine | + |
| Catalase production | + | Maltose | + | Aspartic acid | + |
| Nitrate reduction | + | Raffinose | + | Glutamine | + |
| Lipid hydrolysis | + | Fructose | + | Tyrosine | + |
| Oxidase production | - | pH tolerance | 5-10 | NaCl tolerance (%) | 0-5 |

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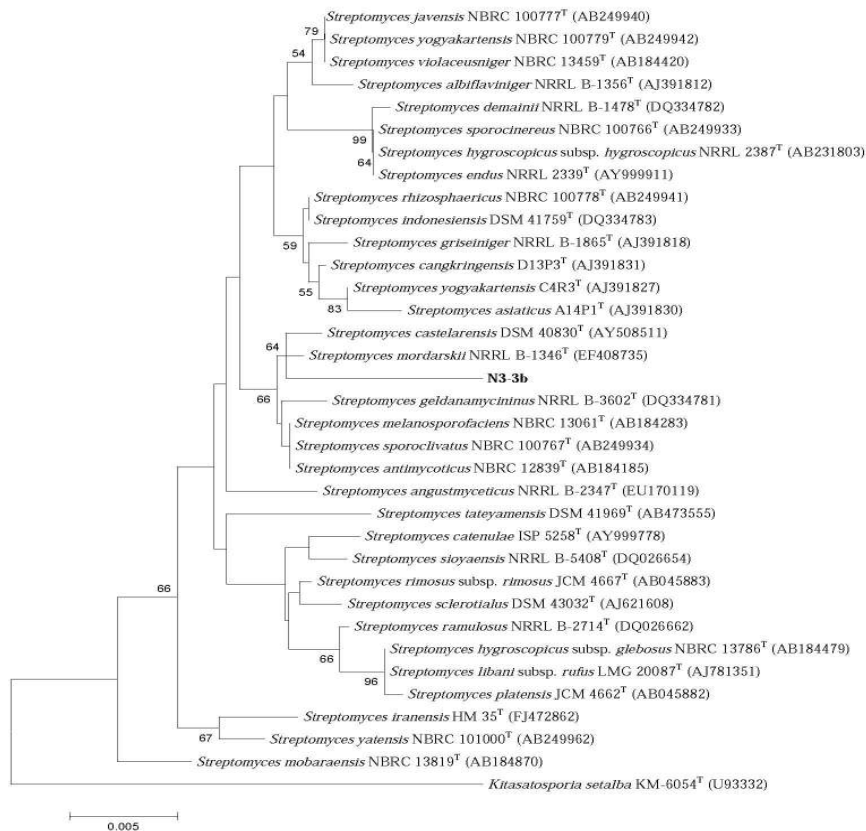
219 **Table 3. Cultural characteristics of N3-3b on different ISP media as observed using ISCC-NBS**
 220 **colour chart (Kelly, 1964)**

| Medium | Growth | Colour of the mycelium | | Pigmentation |
|--------|--------|------------------------|-----------------------|--------------|
| | | Aerial | Substrate | |
| ISP2 | ++ | Yellowish grey | Pale yellow | - |
| ISP3 | +++ | White grey | Yellowish white | - |
| ISP4 | +++ | Light yellow | Mild orange yellow | - |
| ISP5 | - | - | - | - |
| ISP6 | + | Yellow white | Yellow white | - |
| ISP7 | + | Bluish white | - | - |

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225 **Figure 4. Neighbour-joining tree showing phylogenetic relationship of strain N3-3b with its**
 226 **closely related strains**

227 **3.6. Antibiotic sensitivity**

228 The *Streptomyces* sp. N3-3b was sensitive to neomycin and streptomycin, but resistant to
 229 chloramphenicol, ampicillin, penicillin and rifampicin.

230 **4. Discussion**

231 Microorganisms isolated from rhizosphere soil may be better adapted to crop plants and provide
 232 better disease control and growth promotion than organisms isolated from the other sources such as

233 composts, harsh environment etc. as rhizosphere isolates are already closely associated with the
234 plant system as well as adapted to the local environment (34).

235 In the present study, we explored the potential of rhizospheric actinomycetes from rhizosphere of an
236 endemic black rice variety “chakhao”, for biocontrol and plant growth promotion potential.
237 *Streptomyces* sp. N3-3b exhibited good antagonistic activity against rice fungal pathogens.
238 Antagonistic property could be due to production of hydrolytic enzymes or antibiotics. Antifungal
239 antibiotics and fungal cell wall degrading enzymes produced by *Streptomyces* species have been
240 reported to inhibit the growth of pathogens and protect plants from infection (35, 36).

241 *Streptomyces* sp N3-3b showed plant growth promotion traits including IAA production, phosphate
242 solubilization and ammonia production. It produced maximum IAA (143.7 µg/ml) when supplemented
243 with 1% of tryptophan which was similar to the report of Khamna et al. (37). The strain produced
244 much higher levels of IAA than those reported for bacterial strains reported by various authors,
245 Khamna et al. (37), Shrivastava et al. (38) and Harikrishna et al. (39). This seems to be a positive
246 feature for N3-3b to be developed as a bioinoculant for rice cultivation. Growth promotion of rice
247 plants by *Streptomyces* sp. N3-3b may be due to the production of IAA. When *Streptomyces* sp. En-1
248 which produce IAA inoculum was applied onto *Arabidopsis*, it significantly increased the biomass
249 indicating the distinct phytostimulating effects; however, when administrated with another
250 *Streptomyces* sp. IFB-A02 or IFB-A03 which does not produce IAA there was no significant
251 improvement of growth as compared to control (40).

252 N3-3b could solubilize a significant amount of inorganic phosphate (64.8 µg/ml). This is comparable to
253 the report of Sadeghi et al. (41) who observed that *Streptomyces* sp. C solubilized inorganic
254 phosphate up to 92 µg/ml. The present strain from *Chakhao* rice solubilized much higher levels of P
255 than those reported by Passari et al. (42) (3.2-32.6 µg/ml of P). Hamdali et al. (43) reported a P
256 solubilizing *Streptomyces griseus* as a PGP bacterium.

257 N3-3b also promoted seed germination under *in vitro* conditions. The highest vigor index was found at
258 inoculum size of 6×10^7 cfu/ml of the cell suspension. The culture filtrate of *Streptomyces* sp. S-580
259 have been reported to promote the germination of rice seeds (44). Several *Streptomyces* spp. have
260 been reported to enhance seed vigour indices and seedling growth promotion. The results clearly

261 indicated that *Streptomyces sp* N3-3b has great potential to be a plant growth promoting and
262 biocontrol agent. The significant improvement in seedling length and plant growth may be due to the
263 high production of IAA by *Streptomyces sp* N3-3b. IAA stimulates cell division and enhances cell
264 enlargement and extension and plays a major role in normal shoot and root growth (45), and seed
265 bacterization with IAA producing bacteria significantly enhanced early seedling germination and
266 seedling vigor(46). Other factors that might have contributed to rice growth promotion by strain N3-3b
267 could be P solubilization, ammonia production and fungal antagonism.

268 5. Conclusions

269 The results of the present study indicated that *Streptomyces sp. strain N3-3b* from *Chakhao* (Manipuri
270 black rice) rhizospheric soil holds promise for development as a biocontrol and PGP agent for
271 *Chakhao* rice cultivation. The utilization of such beneficial rhizobacterial actinomycetes may lead to
272 increased crop yields while reducing the use of agrochemicals. Such an approach is indeed an
273 attractive trend towards introducing sustainable, green, and eco-friendly agriculture. Further work on
274 PGP potential of N3-3b on *chakhao* and other rice cultivars under pot trial and field conditions are
275 underway and shall be published separately.

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