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Journal Name:	British Microbiology Research Journal
Manuscript Number:	Ms_BMRJ_22959
Title of the Manuscript:	OVERPRODUCTION OF XYLANASE FROM MUTANTS OF <i>Bacillus subtilis</i> USING BARLEY HUSK AS THE PRIME CARBON SOURCE IN SUBMERGED FERMENTATION AFTER RANDOM MUTAGENESIS OF ETHYL METHANE SULFONATE (EMS) AND ACRIDINE ORANGE (AO)
Type of the Article	Original research Articles

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This journal's peer review policy states that **NO** manuscript should be rejected only on the basis of '**lack of Novelty**', provided the manuscript is scientifically robust and technically sound.

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PART 1: Review Comments

	Reviewer's comment	Author's comment (if agreed with reviewer, correct the manuscript and highlight that part in the manuscript. It is mandatory that authors should write his/her feedback here)
<u>Compulsory</u> REVISION comments	1. Present study does not increase or contribute to the current scientific knowledge on xylanase production from <i>Bacillus subtilis</i>. Furthermore, there are previous reports (for example: Ho HL (2015) Xylanase Production by <i>Bacillus subtilis</i> Using Carbon Source of Inexpensive Agricultural Wastes in Two Different Approaches of Submerged Fermentation (SmF) and Solid State Fermentation (SsF). J Food Process Technol 6:437. doi: 10.4172/2157-7110.1000437; Irfan M, Nadeem M, syed Q, Baig S. Effect of Medium Composition on Xylanase Production by <i>Bacillus subtilis</i> using Various Agricultural Wastes American-Eurasian J. Agric. & Environ. Sci., 12 (5): 561-565; and many others) on xylanase production from <i>B. subtilis</i> with very high production of xylanase. 2. An extensive and careful editing is required for improvement of the English language	



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	throughout the manuscript. 3. Authors should follow journals reference style and therefore reference list in the manuscript should be modified according to the example given in author guidelines section of the BMRJ. 4. In submerged fermentations, it is a usual practice to keep the medium volume $1/5^{\text{th}}$ of the vessel volume. Authors has used 250 mL medium in 500 mL flask i.e. $1/2$ the volume of the flask, moreover, rate of agitation has been maintained 150 rpm. What is the rational of using high volume and slow speed for an aerobic bacterium like <i>B. subtilis</i>? 5. What is the reason behind using sterile distilled water for dilution of the bacterium instead of normal saline? (page no. 4, line no 116-117). Use of water can result in lowering the actual number of cells in inoculum thus causing methodological error. 6. Which of the methods described in Bailey et al (1992) has been used by authors, as this research article has not given any of its own method but compared methods used in 20	
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	different laboratories? 7. Authors nowhere have presented enzyme activity with $\pm$ standard deviation value. 8. If Authors have checked for presence of cellulase, β-xylosidase and other carbohydrate degrading enzymes in the culture supernatant? Xylanase activity can just be a non-specific (additional) activity of a cellulase or any other carbohydrate degrading enzyme present in the culture supernatant. 9. Authors indicated that there was a direct relationship between secretion of extracellular xylanase and cell count of <i>B. subtilis</i> (page no. 9, line no. 315-316). It has also been shown that highest cell count was recorded in case of wild type <i>B. subtilis</i> (page no. 9, line no 301) as compared to mutant strains, then why the production was lower in case of wild type organism? 10. Incorrect use of terminology for indicating clear zone of hydrolysis because of xylanolytic activity of enzyme (authors have used ring zone of discoloration).	
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<u>Minor</u> REVISION comments	1. In table 1, column 1 should be adjusted as column 3 and labelled as Mutants generated instead of Mutant 2. Title of the table 1 should be changed to 'mutagens concentrations used for generation of <i>Bacillus</i> mutants'	
<u>Optional/General</u> comments		

Reviewer Details:

Name:	Anonymous
Department, University & Country	Central University of Gujarat, Gandhinagar, India