

**FANCONI ANEMIA GENES AND REACTIVE OXYGEN SPECIES IN CANCER****DEVELOPMENT****ABSTRACT**

Fanconi Anemia (FA) is an autosomal recessive disease of childhood. However, the FA pathway is responsible for the development of leukemia and the other cancers. It has been also demonstrated that FA, an only human genomic instability syndrome is very sensitive to oxidative stress and ROS overproduction. In the present work, we consider major mechanisms of antioxidant protection in FA cells. We showed that there are two types of such mechanisms: the suppression of ROS overproduction by FA genes through the activation of basic FA anemia proteins under the conditions of oxidative stress and the application of free radical scavengers able to react with iron-dependent ROS such as flavonoids rutin and quercetin. The last nontoxic compounds of vitamin P group might be recommended for the treatment of FA anemia patients. Then, we discussed the role of FA anemia proteins in cancer development.

Key words: Fanconi anemia, reactive oxygen species, antioxidants, rutin

Fanconi anemia (FA) is an autosomal recessive disease of childhood characterized by progressive pancytopenia, developmental abnormalities, bone marrow failure, and high disposition to leukemia and other cancers. It has been demonstrated that the FA pathway is an important way in the development of such deadly adult pathologies as breast and cervical cancer [1,2]. These findings sharply increase an interest to studying FA molecular mechanisms. The special interest is attracted to FA because FA is the only human genomic instability syndrome uniquely sensitive to oxidative

26 stress. Earlier, an importance of reactive oxygen species (ROS) overproduction i.e. oxidative stress  
27 has been demonstrated. Joenje *et al.* has shown that erythrocyte superoxide dismutase (SOD)  
28 decreased in FA [3]. These authors also proposed that the formation of chromosomal aberrations in  
29 FA anemia might be explained by the genetic toxicity of oxygen [4] underlining an important role of  
30 oxidative stress in FA development.

### 31 **EARLIER STUDIES**

32 In 1960, medicine doctors from Russian Institute of Hematology for Children (Moscow) asked **us as**  
33 **scientists** working in the field of free radical-associated diseases to study the possible application of  
34 antioxidants for the treatment of FA patients. We hoped that antioxidants and free radical scavengers  
35 could act positively on behalf of FA patients. To study the effects of antioxidants on FA cells, **ROS**  
36 **production especially superoxide in these cells was measured**. Major results of this *in vitro* study  
37 have been summarized as follows [5,6]: ROS formation was measured by lucigenin- and luminol-  
38 dependent chemiluminescence (CL) in non-stimulated and stimulated blood and bone marrow  
39 leukocytes of FA patients. It was found that the FA blood leukocytes produced the enhanced level of  
40 luminol CL in comparison with leukocytes from normal donors. Lucigenin CL was also higher in  
41 FA blood leukocytes, although its effect was not very significant. Similar but smaller effects were  
42 observed for bone **marrow** FA leukocytes. Earlier, we have already demonstrated that flavonoids  
43 rutin and its aglycone quercetin possess both free radical scavenging and chelating properties. We  
44 **had** shown that both flavonoids were the most effective inhibitors of iron-dependent microsomal  
45 lipid peroxidation comparing to lipid peroxidation initiated by carbon tetrachloride [7]. **(This work**  
46 **has been cited more than 800 times at present.)**

47 This work had a good response; at present, it has been cited more than 800 times).

48 We proposed that ROS formation in FA leukocytes was the iron-catalyzed process because it  
49 was characterized by the enhanced luminol CL typical for the formation of iron-dependent ROS.

50 This proposal was supported by studying the effects of various free radical inhibitors on luminol CL  
51 of FA leukocytes [5]: SOD inhibited only slightly luminol CL, while mannitol (the typical hydroxyl  
52 radical scavenger) and rutin were the strongest inhibitors. Later on, it was also demonstrated that  
53 rutin was a strong inhibitor of iron-dependent lipid peroxidation of rat brain homogenates [8] and  
54 the free radical formation in iron-overloaded rats [9]. Moreover, it was found that rutin efficiently  
55 inhibited free radical formation and oxyhemoglobin oxidation in  $\beta$ -thalassemic red blood cells [10].  
56 These findings reassured us to apply the non-toxic flavonoid rutin for the treatment of FA patients.

57 The results of rutin therapy for a small group of FA patients were encouraging. Rutin was  
58 permitted for application to FA patients under the supervision of our co-authors medical doctors.  
59 No toxic side effects were observed.” Rutin (vitamin P) was permitted for application to patients  
60 (FA patients were under the supervision of medical doctors). No toxic side effects were observed in  
61 patients. ROS production by FA leukocytes sharply decreased and patients’ health was essentially  
62 improved [6]. Unfortunately, there was no possibility to continue our study despite the numerous  
63 requests of medical doctors.

64

## 65 **THE NATURE OF REACTIVE OXYGEN SPECIES IN FA CELLS**

66 Our findings demonstrated that ROS formation in FA cells was connected with the iron species. In  
67 1992, in accord with the views of that time we proposed that the major damage produced by ROS in  
68 cells is the direct interaction of ROS with biomolecules. As the superoxide, a major precursor of  
69 ROS in cells is unreactive in free radical processes [11,12], we proposed that the reactive iron-  
70 contained species could be responsible for free radical damage in FA cells. It was suggested that the  
71 iron-catalyzed superoxide conversion into reactive hydroxyl radicals (the Fenton reaction) was  
72 responsible for their formation in FA cells. Indeed, there are some evidences of hypersensitivity of  
73 FA cells in the presence of oxygen and iron [13]. Although reactive hydroxyl radicals are probably

74 unable to achieve the target biomolecules, they can be formed during the contacts of superoxide  
75 with DNA “iron fingers” [14].

76 A major conclusion from our previous work was that FA characterized by the enhanced  
77 production of reactive iron-dependent species, which might be the source of fatal disorders in this  
78 hereditary disease. At present, many authors agreed that FA cells exist under the conditions of ROS  
79 overloading. Thus, Hadjur *et al.* concluded that the abnormality of FA cells depended on ROS  
80 overproduction [15]. Du *et al.* pointed out that FA is only human genomic instability syndrome,  
81 which was uniquely sensitive to oxidative stress [16]. Therefore, there is no doubt about  
82 importance of ROS in Fanconi anemia. However, the damaging mechanisms of ROS activity remain  
83 to be investigated.

84

## 85 **THE FA GENETIC PATHWAY**

86 The most important **discovery in** FA molecular mechanisms was the identification of Fanconi  
87 anemia genes responsible for synthesis of special FA proteins FANC (**among them FANCA,**  
88 **FANCB, FANCC, FANCE, FANCF, FANCG, FANCL and FANCM**). It was found that eight major  
89 FA proteins (FANCA, B, C, E, F, G, L, and M) formed a nuclear complex [16]. In response to DNA  
90 damage or DNA replication stress FA complex monoubiquitinate into the two FA proteins FANCD2  
91 and FANCI, which then recruit the other downstream FA proteins including FANCD1 (which is  
92 also named the breast cancer protein BRCA2), FANCI, and FANCN to enter nuclear loci containing  
93 the damaged DNA.

94 It has been shown that FA proteins play the critical role in the regulation of oxidative stress.  
95 Thus, deficiency in FA genes apparently affects mitochondrial ROS [17]. The redox-sensitive  
96 proteins FANCA and FANCF exist as monomers under non-oxidizing conditions but form a new  
97 nuclear complex through the intermolecular disulfide bonds in response to oxidative damage [18].

98 FANCA, FANCC, and FANCG participate in redox processes in mice with combined deficiencies of  
99 the genes encoding FANCC and Cu/Zn superoxide dismutase [15].

100 FA proteins function through the interaction with some enzymes. Saadatzadeh *et al.* showed that  
101 *Fancca*<sup>-/-</sup> cells were highly sensitive to oxidants (hydrogen peroxide) and underwent enhanced  
102 apoptosis [19]. Antioxidative compounds enhanced the survival of these cells. Thus, the redox-  
103 dependent ASK1 kinase was hyperactive in hydrogen peroxide-treated *Fancca*<sup>-/-</sup> cells. Another FA  
104 protein FANCG interacted with mitochondrial antioxidant enzyme peroxiredoxin-3 and cytochrome  
105 P450 2E1 (CYP2E1) [20]. This member of P450 superfamily responsible for ROS production and  
106 the activation of carcinogens. These findings suggested that the interaction of FANCG with  
107 CYP2E1 might increase DNA oxidation.

108 It is known that the tumor protein p53 plays an important role in the prevention of cancer.  
109 Furthermore, some findings demonstrate that p53 deficiency might enhance cancer development in  
110 FA patients and FA mice. Therefore, it was suggested that FA proteins could interact with p53  
111 under the conditions of oxidative stress. Freie *et al.* showed that ionizing radiation (IR) induced p53  
112 elevated levels in cells from *Fancc* mutant mice and that the inactivation of p53 enhanced TNF-  
113 induced apoptosis in myeloid cells from *Fancc*<sup>-/-</sup> mice [21]. Rani *et al.* demonstrated that FA  
114 proteins protected cells from the stress-induced proliferative arrest and tumor evolution  
115 through the modulation of signaling pathways which connected FA proteins to p53 [22]. Du *et*  
116 *al.* proposed that two major FA proteins FANCA and FANCC might coordinate with p53 in the  
117 regulation of oxidative stress response [16].

118 It has been shown that the *Foxo3a* gene might be involved in ROS formation [23]. For  
119 example, Tothova *et al.* showed that ROS levels increased in Foxo-deficient hematopoietic stem  
120 cells that correlated with the changes in expression of ROS-regulated genes [24]. Correspondingly,  
121 *Foxo3a* plays an important role in ROS regulation in FA cells. Thus, Li *et al.* showed that the

122 treatment of FA cells with hydrogen peroxide stimulated the formation of a complex between  
123 FANCD2 and FOXO3a with subsequent monoubiquitination of FANCD2 [25]. It was suggested  
124 that the overexpression of *Foxo3a* reduced abnormal accumulation of ROS, enhanced cellular  
125 resistance to oxidative stress, and increased antioxidant gene expression only in cells corrected by a  
126 FANCD2 protein capable of interacting with FOXO3a.

127 It has been shown that ROS accelerated the development of hydrocephalus (abnormal  
128 accumulation of cerebrospinal fluid in the brain) in mouse model of FA [26]. The deletion of  
129 *Foxo3a* in FA mice increased the ROS accumulation and subsequently deregulated mitosis and  
130 ultimately apoptosis in the neural stem and progenitor cells NSPCs, leading to hydrocephalus  
131 development. **N-acetylcysteine** (NAC) and quercetin reduced ROS formation in both neural stem  
132 cell NSCs and in the brain of double knockout (DKO) offspring mice. Importantly, quercetin  
133 greatly diminished the synthetic lethality imposed by DKO and completely eliminated  
134 hydrocephalus in DKO mice.

135

## 136 **MECHANISMS OF ANTIOXIDANT PROTECTION IN FANCONI ANEMIA**

### 137 **ANTIOXIDANTS AND FREE RADICAL SCAVENGERS**

138 As it was aforementioned, **it has been previously shown** that oxidative stress (ROS overproduction)  
139 in FA cells might be diminished by the application of flavonoid rutin, an antioxidant and free radical  
140 scavenger [5,6]. **It was suggested** that rutin might be able to scavenge reactive iron-depend ROS.  
141 Rutin is a nontoxic compound permitted for the treatment of patients. Therefore, medical doctors  
142 were allowed to use rutin for the treatment of FA patients. They observed certain positive effects in  
143 patients. (It should be mentioned that several FA families asked us to prolong the treatment of FA  
144 children but we were unable to continue our work).

145 Now we were encouraged to find out that in subsequent studies flavonoids were used for the  
146 suppression of oxidative stress in FA. It has been shown that quercetin (the aglycone of rutin) turned  
147 out to be an effective antioxidant in FA cells. Antioxidant effect of quercetin was also shown in FA  
148 animals. Li *et al.* found that mice deficient for the *Fanca* or *Fancc* genes were diabetes-prone  
149 when fed with a high-fat diet [27]. Treatment of FA mice with quercetin diminished diabetes  
150 and obesity. It was already demonstrated that quercetin reduced ROS formation and eliminated  
151 hydrocephalus in double knockout mice [26]. Ponte *et al.* proposed that the cocktail of antioxidants  
152 lipoic acid and NAC might be applied as a prophylactic approach to delay progressive clinical  
153 symptoms in FA patients [28].

154

155

156

## 157 **ANTIOXIDANT EFFECTS OF FA GENES**

158 In 2001, Hadjur *et al.* showed that encoding *Fancc* and Cu/Zn superoxide dismutase genes might be  
159 useful for the treatment of defective hematopoiesis and hepatic steatosis in mice [13]. These findings  
160 suggested an important role of FA genes in protection of FA cells from oxidative stress. In  
161 subsequent work, it was confirmed that FA genes were able to suppress ROS overproduction. Du *et*  
162 *al.* showed that major antioxidant defense genes were downregulated in FA patients due to the  
163 increased oxidative DNA damage in the promoters of antioxidant genes [29]. They showed that FA  
164 proteins together with the chromatin-remodeling factor BRG1 protected the promoters of  
165 antioxidant defense genes. Oxidative stress activated FA pathway through monoubiquitination of  
166 FANCD2. After this, FANCA or FANCD2 proteins formed the ternary complex with BRG1 at the  
167 promoters of antioxidant genes. It has been suggested that this complex played essential role in the  
168 protection of promoters from ROS damage. As it has been mentioned, ROS influenced the

169 development of hydrocephalus in mouse model of FA [26]. Combined deficiency of two FA genes  
170 *Fancc* and *Fancd2* led to the inactivation of *Foxo3a* gene, the enhancement of ROS level, and  
171 apoptosis of neural stem and progenitor cells. Antioxidants quercetin and NAC reduced ROS  
172 formation in the brain of mice, while quercetin completely **suppressed** hydrocephalus.  
173 Mukhopadhyay *et al.* identified the FA group G (FANCG) protein in mitochondria, which interacted  
174 with the mitochondrial peroxidase peroxiredoxin-3 (PRDX3) [30]. The formation of this complex  
175 prevented the destruction of PRDX3 peroxidase and diminished ROS formation.

176

## 177 **DISCUSSION**

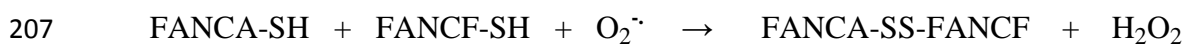
178 Fanconi anemia is a rare chromosome instability syndrome, which is characterized by aplastic  
179 anemia in childhood. It is known that FA possesses high disposition to leukemia and other cancers.  
180 Now, there are new findings, which make us reconsider more carefully the connection between FA  
181 and cancer. Some new contemporary data demonstrate that the FA pathway is an important route for  
182 the development of such deadly adult pathologies as breast and cervical cancer. Thus, D'Andrea  
183 pointed out that abnormalities in the FA pathway are found not only in childhood Fanconi  
184 anemia but also in sporadic cancers in adults [1]. Narayan *et al.* investigated the molecular  
185 genetic basis and the role of (FA)-BRCA pathway in cervical cancer (CC) [2]. These authors showed  
186 that the *fancf* gene was disrupted by either the promoter hypermethylation or the deregulated gene  
187 expression in cervical cancer. **It was also** found that gene inactivation in the FA-BRCA pathway by  
188 epigenetic alterations **showed** its major role in the development of cervical cancer. Chen *et al.*  
189 demonstrated that the regulation of FANCD2 by m-TOR pathway led to the resistance of cancer  
190 cells to DNA double-strand breaks [31]. It is not surprising that among thirteen FA genes **one gene**  
191 is the well-known breast cancer susceptibility gene *brca2*.

192 In normal cells, the FA pathway is not constitutively active but it **activates** by DNA damage. As



193 DNA damage depends on oxidative stress, ROS overproduction should be an important factor of FA  
 194 pathway activation. Mechanism of response of FA genes to DNA damage is not fully established,  
 195 but it has been shown that FA genes exist as nuclear complex before activation. In response to the  
 196 DNA damage or DNA replication stress FA complex monoubiquitinate into two FA proteins  
 197 FANCD2 and FANCI, which then recruit the other downstream FA proteins including the breast  
 198 cancer protein BRCA2, FANCI, and FANCN to enter nuclear loci containing the damaged DNA.  
 199 Then FANCA and FANCF form a new complex through the intermolecular disulfide bonds after  
 200 exposure to oxidative stress [16]. Thus, FA genes accomplish important defense function, which is  
 201 confirmed by high sensitivity of *Fanc*<sup>-/-</sup> cells to ROS.

202 We now can visualize the complete defense scheme of ROS inhibition in FA cells. It has been  
 203 shown that the oxidative stress (ROS overproduction) initiates the activation of FA genes, which  
 204 remain inactive under unstressed conditions. After activation, FANCA and FANCF proteins form a  
 205 complex through the intermolecular disulfide bonds, and we might propose that the disulfide  
 206 complex formed by the reaction of superoxide with these proteins:



208 This proposal supported by the effect of SOD on the redox processes of FANCA, FANCC, and  
 209 FANCG in mice [13].

210 Another mode of antioxidant activity of FA proteins is the protection of antioxidant gene  
 211 promoters from ROS overproduction. Du *et al.* pointed out that oxidative stress is an important  
 212 pathogenic factor in leukemia-prone marrow diseases such as Fanconi anemia [29]. It has been  
 213 proposed that the FA pathway plays a crucial role in protecting major antioxidant defense genes  
 214 from oxidative damage. This protection probably accomplished in response to oxidative stress by  
 215 the interaction with the chromatin-remodeling machinery. Indeed, it has been shown that the  
 216 oxidative stress-induced activation of FA pathway (FANCD2 ubiquitination) required for the

217 formation of the FA-BRG1-promoter complex. This complex is essential for the protection of the  
 218 antioxidant gene promoters from oxidative damage.

219 Thus, the activation of FA pathway under the conditions of oxidative stress led to cellular  
 220 protection through various ways. Taking into account the above-mentioned consideration, these  
 221 ways have been presented in Table 1.

## 222 **Table 1**

### 223 **FA PATHWAYS OF CELLULAR PROTECTION**

#### 224 225 **1. Protection of antioxidant gene promoters**

226 FANCD2 ubiquitination  
 227 ↓  
 228 FA-BRG1-promoter complex  
 229 ↓  
 230 Protection of antioxidant gene promoters  
 231

#### 232 233 234 **2. Formation of the disulfide complex**

235  
236  $FANCA + FANCF + O_2^- \rightarrow FANCA-SS-FANCF$   
 237

#### 238 239 **3. Suppression of DNA double-strand breaks**

240  
241  $FANCD2 \rightarrow m-TOR \text{ pathway} \rightarrow \text{DNA double-strand breaks}$   
 242

#### 243 244 **4. Suppression of ROS overproduction**

245  
246 Antioxidants (quercetin and NAC)  $\rightarrow$  ROS↓ (diabetes and hydrocephalus in FA patients)

#### 247 248 **5. Enhancement of gene antioxidant activity**

249  
250 The interaction of genes with enzymes (Foxo3a, p50, p450)  
 251

252

253

### 253 **CONCLUSIONS**

- 254 (1) Direct interaction of FANCA and FANCF with ROS (superoxide) forms a complex through the  
255 intermolecular disulfide bonds.
- 256 (2) The regulation of FANCD2 by the m-TOR pathway might lead to the resistance of cells to DNA  
257 double-strand breaks as in cervical cancer.
- 258 (3) FANCD2 ubiquitination follows the formation of FA-BRG1-promoter complex capable of  
259 protection of antioxidant gene promoters.
- 260 (4) Antioxidant activity of FA genes has been enhanced by the interaction with various enzymes  
261 such as Foxo3a, p50, p450.
- 262 (5) Application of antioxidants and free radical scavengers (quercetin and NAC) decreases ROS  
263 overproduction in such diseases as diabetes or hydrocephalus in FA patients.

264

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