

FANCONI ANEMIA GENES AND REACTIVE OXYGEN SPECIES IN CANCER

DEVELOPMENT

ABSTRACT

Fanconi Anemia (FA) is an autosomal recessive disease of childhood. However, The FA pathway is responsible for the development of leukemia and the other cancers. It has been also demonstrated that FA, an only human genomic instability syndrome is very sensitive to oxidative stress and reactive oxygen species (ROS) overproduction. In the present review, we consider major mechanisms of antioxidant protection in Fanconi anemia cells. We showed that there are two types of such mechanisms: the suppression of reactive oxygen species overproduction by Fanconi anemia genes through the activation of basic Fanconi anemia proteins under the conditions of oxidative stress and the application of free radical scavengers able to react with iron-dependent reactive oxygen species such as flavonoids rutin and quercetin. The last nontoxic compounds of vitamin P group might be recommended for the treatment of Fanconi anemia patients. Then, we discussed the role of Fanconi anemia proteins in cancer development.

Key words: Fanconi anemia, ROS, antioxidants, rutin

Fanconi anemia (FA) is an autosomal recessive disease of childhood characterized by progressive pancytopenia, developmental abnormalities, bone marrow failure, and high disposition to leukemia and other cancers. It has been demonstrated that the FA pathway is an important way in the development of such deadly adult pathologies as breast and cervical cancer [1,2]. These findings sharply increase an interest to studying FA molecular mechanisms. The special interest is attracted

26 to Fanconi anemia because FA is the only human genomic instability syndrome uniquely sensitive
27 to oxidative stress. Earlier, an importance of reactive oxygen species (ROS) overproduction i.e.
28 oxidative stress has been demonstrated. Joenje *et al.* has shown that erythrocyte superoxide
29 dismutase (SOD) **is** decreased in FA [3]. These authors also proposed that the formation of
30 chromosomal aberrations in FA anemia might be explained by the genetic toxicity of oxygen [4]
31 underlining an important role of oxidative stress in FA development.

32 **EARLIER STUDIES**

33 In 1960, medicine doctors from Russian Institute of Hematology for Children (Moscow) asked **us as**
34 **scientists** working in the field of free radical-associated diseases to study the possible application of
35 antioxidants for the treatment of FA patients. We hoped that antioxidants and free radical scavengers
36 could act positively on behalf of FA patients. To study the effects of antioxidants on FA cells, **ROS**
37 **production especially superoxide in these cells was measured**. Major results of this *in vitro* study
38 have been summarized as follows [5,6]: ROS formation was measured by lucigenin- and luminol-
39 dependent chemiluminescence (CL) in non-stimulated and stimulated blood and bone marrow
40 leukocytes of FA patients. It was found that the FA blood leukocytes produced the enhanced level of
41 luminol CL in comparison with leukocytes from normal donors. Lucigenin CL was also higher in
42 FA blood leukocytes, although its effect was not very significant. Similar but smaller effects were
43 observed for bone marrow FA leukocytes. Earlier, **we** have already demonstrated that flavonoids
44 rutin and its aglycone quercetin possess both free radical scavenging and chelating properties. We
45 have shown that both flavonoids were the most effective inhibitors of iron-dependent microsomal
46 lipid peroxidation comparing to lipid peroxidation initiated by carbon tetrachloride [7]. (This work
47 had a good response; ~~at present, to-date~~, it has been cited more than 800 times).

48 **We** proposed that ROS formation in FA leukocytes was the iron-catalyzed process because it
49 was characterized by the enhanced luminol CL typical for the formation of iron-dependent ROS.

50 This proposal was supported by studying the effects of various free radical inhibitors on luminol CL
51 of FA leukocytes [5]: Superoxide dismutase (SOD) inhibited only slightly luminol CL, while
52 mannitol (the typical hydroxyl radical scavenger) and rutin were the strongest inhibitors. (Later on,
53 **it was also demonstrated** that rutin was a strong inhibitor of iron-dependent lipid peroxidation of rat
54 brain homogenates [8] and the free radical formation in iron-overloaded rats [9]. Moreover, **it was**
55 found that rutin efficiently inhibited free radical formation and oxyhemoglobin oxidation in β -
56 thalassemic red blood cells [10]). These findings reassured **us** to apply the non-toxic flavonoid rutin
57 (vitamin P) for the treatment of FA patients.

58 The results of rutin therapy for a small group of FA patients were encouraging. Rutin (vitamin P)
59 was permitted for application to patients (FA patients were under the supervision of medical
60 doctors). No toxic side effects were observed in patients. ROS production by FA leukocytes sharply
61 decreased **and** patients' health was essentially improved [6]. Unfortunately, **there was no possibility**
62 to continue our study despite the numerous requests of medical doctors.

63

64 **THE NATURE OF REACTIVE OXYGEN SPECIES IN FA CELLS**

65 **Our** findings demonstrated that ROS formation in FA cells was connected with the iron species. In
66 1992, in accord with the views of that time **we** proposed that the major damage produced by ROS in
67 cells is the direct interaction of ROS with biomolecules. As the superoxide, a major precursor of
68 ROS in cells is unreactive in free radical processes [11,12], we proposed that the reactive iron-
69 contained species could be responsible for free radical damage in FA cells. **It was** suggested that the
70 iron-catalyzed superoxide conversion into reactive hydroxyl radicals (the Fenton reaction) was
71 responsible for their formation in FA cells. Indeed, there are some evidences of hypersensitivity of
72 FA cells **in the presence of** oxygen and iron [13]. Although reactive hydroxyl radicals are probably

73 unable to achieve the target biomolecules, they can be formed during the contacts of superoxide
74 with DNA “iron fingers” [14].

75 A major conclusion from our previous work was that FA **is** characterized by the enhanced
76 production of reactive iron-dependent species, which might be the source of fatal disorders in this
77 hereditary disease. At present, many authors agreed that FA cells exist under the conditions of ROS
78 overloading. Thus, Hadjur *et al.* concluded that the abnormality of FA cells depended on ROS
79 overproduction [15]. Du *et al.* pointed out that FA **is** the only human genomic instability syndrome,
80 which **is** uniquely sensitive to oxidative stress [16]. Therefore, there is no doubt about **the**
81 importance of ROS in Fanconi anemia. However, the damaging mechanisms of ROS activity remain
82 to be investigated.

83

84 **THE FA GENETIC PATHWAY**

85 The most important **discovery in** FA molecular mechanisms was the identification of Fanconi
86 anemia genes responsible for synthesis of special FA proteins FANC (**among them FANCA,**
87 **FANCB, FANCC, FANCE, FANCF, FANCG, FANCL and FANCM**). It was found that eight major
88 FA proteins (FANCA, B, C, E, F, G, L, and M) formed a nuclear complex [16]. In response to DNA
89 damage or DNA replication stress FA complex monoubiquitinates **into** the two FA proteins
90 FANCD2 and FANCI, which then recruit the other downstream FA proteins including FANCD1
91 (which is also named the breast cancer protein BRCA2), FANCI, and FANCN to enter nuclear loci
92 containing the damaged DNA.

93 It has been shown that FA proteins play the critical role in the regulation of oxidative stress.
94 Thus, deficiency in FA genes apparently affect mitochondrial ROS [17]. The redox-sensitive
95 proteins FANCA and FANCF exist as monomers under non-oxidizing conditions but form a new
96 nuclear complex through the intermolecular disulfide bonds in response to oxidative damage [18].

97 FANCA, FANCC, and FANCG participate in redox processes in mice with combined deficiencies of
98 the genes encoding FANCC and Cu/Zn superoxide dismutase [15].

99 FA proteins function through the interaction with some enzymes. Saadatzadeh *et al.* showed that
100 *Fancca*^{-/-} cells were highly sensitive to oxidants (hydrogen peroxide) and underwent enhanced
101 apoptosis [19]. Antioxidative compounds enhanced the survival of these cells. Thus, the redox-
102 dependent ASK1 kinase was hyperactive in hydrogen peroxide-treated *Fancca*^{-/-} cells. Another FA
103 protein FANCG interacted with mitochondrial antioxidant enzyme peroxiredoxin-3 and cytochrome
104 P450 2E1 (CYP2E1) [20]. This member of P450 superfamily is responsible for ROS production and
105 the activation of carcinogens. These findings suggested that the interaction of FANCG with
106 CYP2E1 might increase DNA oxidation.

107 It is known that the tumor suppressor protein p53 plays an important role in the prevention of
108 cancer. Furthermore, some findings demonstrate that p53 deficiency might enhance cancer
109 development in FA patients and FA mice. Therefore, it was suggested that FA proteins could
110 interact with p53 under the conditions of oxidative stress. Freie *et al.* showed that ionizing radiation
111 (IR) induced p53 elevated levels in cells from *Fancc* mutant mice and that the inactivation of p53
112 enhanced TNF-induced apoptosis in myeloid cells from *Fancc*^{-/-} mice [21]. Rani *et al.*
113 demonstrated that FA proteins protected cells from the stress-induced proliferative arrest
114 and tumor evolution through the modulation of signaling pathways which connected FA
115 proteins to p53 [22]. Du *et al.* proposed that two major FA proteins FANCA and FANCC might
116 coordinate with p53 in the regulation of oxidative stress response [16].

117 It has been shown that the *Foxo3a* gene might be involved in ROS formation [23]. For
118 example, Tothova *et al.* showed that ROS levels increased in Foxo-deficient hematopoietic stem
119 cells that correlated with the changes in expression of ROS-regulated genes [24]. Accordingly
120 dingly, *Foxo3a* plays an important role in ROS regulation in FA cells. Thus, Li *et al.* showed that

121 the treatment of FA cells with hydrogen peroxide stimulated the formation of a complex between
122 FANCD2 and FOXO3a with subsequent monoubiquitination of FANCD2 [25]. It was suggested
123 that the overexpression of *Foxo3a* reduced abnormal accumulation of ROS, enhanced cellular
124 resistance to oxidative stress, and increased antioxidant gene expression only in cells corrected by a
125 FANCD2 protein capable of interacting with FOXO3a.

126 It has been shown that ROS accelerated the development of hydrocephalus (abnormal
127 accumulation of cerebrospinal fluid in the brain) in mouse model of FA [26]. The deletion of
128 *Foxo3a* in FA mice increased the ROS accumulation and subsequently deregulated mitosis and
129 ultimately apoptosis in the neural stem and progenitor cells NSPCs, leading to hydrocephalus
130 development. The antioxidants NAC and quercetin reduced ROS formation in both neural stem
131 cell NSCs and in the brain of double knockout (DKO) offspring mice. Importantly, quercetin
132 greatly diminished the synthetic lethality imposed by DKO and completely eliminated
133 hydrocephalus in DKO mice.

134

135 **MECHANISMS OF ANTIOXIDANT PROTECTION IN FANCONI ANEMIA**

136 **ANTIOXIDANTS AND FREE RADICAL SCAVENGERS**

137 As it was aforementioned, it has been previously shown that oxidative stress (ROS overproduction)
138 in FA cells might be diminished by the application of flavonoid rutin, an antioxidant and free radical
139 scavenger [5,6]. It was suggested that rutin might be able to scavenge reactive iron-dependent ROS.
140 Rutin is a nontoxic compound (vitamin P) permitted for the treatment of patients. Therefore,
141 medical doctors were allowed to use rutin for the treatment of FA patients. They observed certain
142 positive effects in patients. (It should be mentioned that several FA families asked us to prolong the
143 treatment of FA children but we were unable to continue our work).

144 Now **we** were encouraged to find out that in subsequent studies flavonoids were used for the
145 suppression of oxidative stress in FA. **It has been shown** that quercetin (the aglycone of rutin) turned
146 out to be an effective antioxidant in FA cells. Antioxidant effect of quercetin was also shown in FA
147 animals. Li *et al.* found that mice deficient for the *Fanca* or *Fancc* genes were diabetes-prone
148 when fed with a high-fat diet [27]. Treatment of FA mice with quercetin diminished diabetes
149 and obesity. It was already demonstrated that quercetin reduced ROS formation and eliminated
150 hydrocephalus in double knockout mice [26]. Ponte *et al.* proposed that the cocktail of antioxidants
151 lipoic acid and NAC might be applied as a prophylactic approach to delay progressive clinical
152 symptoms in FA patients [28].

153

154

155

156 **ANTIOXIDANT EFFECTS OF FA GENES**

157 In 2001, Hadjur *et al.* showed that encoding *Fancc* and Cu/Zn superoxide dismutase genes might be
158 useful for the treatment of defective hematopoiesis and hepatic steatosis in mice [13]. These findings
159 suggested an important role of FA genes in protection of FA cells from oxidative stress. In
160 subsequent work, it was confirmed that FA genes were able to suppress ROS overproduction. Du *et*
161 *al.* showed that major antioxidant defense genes were downregulated in FA patients due to the
162 increased oxidative DNA damage in the promoters of antioxidant genes [29]. They showed that FA
163 proteins, together with the chromatin-remodeling factor BRG1, protected the promoters of
164 antioxidant defense genes. Oxidative stress activated FA pathway through monoubiquitination of
165 FANCD2. After this, FANCA or FANCD2 proteins formed the ternary complex with BRG1 at the
166 promoters of antioxidant genes. It has been suggested that this complex played essential role **in** the
167 protection of promoters from ROS damage. As it has been mentioned, ROS influenced the

168 development of hydrocephalus in mouse model of FA [26]. Combined deficiency of two FA genes
169 *Fancc* and *Fancd2* led to the inactivation of *Foxo3a* gene, the enhancement of ROS level, and
170 apoptosis of neural stem and progenitor cells. Antioxidants quercetin and NAC reduced ROS
171 formation in the brain of mice, while quercetin completely **suppressed** hydrocephalus.
172 Mukhopadhyay *et al.* identified the FA group G (FANCG) protein in mitochondria, which interacted
173 with the mitochondrial peroxidase peroxiredoxin-3 (PRDX3) [30]. The formation of this complex
174 prevented the destruction of PRDX3 peroxidase and diminished ROS formation.

175

176 **DISCUSSION**

177 Fanconi anemia is a rare chromosome instability syndrome, which is characterized by aplastic
178 anemia in childhood. It is known that FA possesses high disposition to leukemia and other cancers.
179 Now, there are new findings, which make us reconsider more carefully the connection between FA
180 and cancer. Some new contemporary data demonstrate that the FA pathway is an important route for
181 the development of such deadly adult pathologies as breast and cervical cancer. Thus, D'Andrea
182 pointed out that abnormalities in the FA pathway are found, not only in childhood Fanconi
183 anemia, but also in sporadic cancers in adults [1]. Narayan *et al.* investigated the molecular
184 genetic basis and the role of (FA)-BRCA pathway in cervical cancer (CC) [2]. These authors showed
185 that the *fancf* gene was disrupted by either the promoter hypermethylation or the deregulated gene
186 expression in cervical cancer. **It was also** found that gene inactivation in the FA-BRCA pathway by
187 epigenetic alterations **showed** its major role in the development of cervical cancer. Chen *et al.*
188 demonstrated that the regulation of FANCD2 by m-TOR pathway led to the resistance of cancer
189 cells to DNA double-strand breaks [31]. It is not surprising that among **the** thirteen FA genes **one**
190 **gene** is the well-known breast cancer susceptibility gene *brca2*.

191 In normal cells, the FA pathway is not constitutively active but it **activates but it is turned on** by

192 DNA damage. As DNA damage depends on oxidative stress, ROS overproduction should be an
 193 important factor of FA pathway activation. The mechanism of response of FA genes to DNA
 194 damage is not fully established, but it has been shown that FA genes exist as a nuclear complex
 195 before activation. In response to the DNA damage or DNA replication stress FA complex
 196 monoubiquitinates into two FA proteins FANCD2 and FANCI, which then recruit the other
 197 downstream FA proteins including the breast cancer protein BRCA2, FANCI, and FANCN to enter
 198 nuclear loci containing the damaged DNA. Then FANCA and FANCF form a new complex through
 199 the intermolecular disulfide bonds after exposure to oxidative stress [16]. Thus, FA genes
 200 accomplish important defense function, which is confirmed by high sensitivity of *Fanc*^{-/-} cells to
 201 ROS.

202 We now can visualize the complete defense scheme of ROS inhibition in FA cells. It has been
 203 shown that the oxidative stress (ROS overproduction) initiates the activation of FA genes, which
 204 remain inactive under unstressed conditions. After activation, FANCA and FANCF proteins form a
 205 complex through the intermolecular disulfide bonds, and we might propose that the disulfide
 206 complex is formed by the reaction of superoxide with these proteins:



208 This proposal supported by the effect of SOD on the redox processes of FANCA, FANCC, and
 209 FANCG in mice [13].

210 Another mode of antioxidant activity of FA proteins is the protection of antioxidant gene
 211 promoters from ROS overproduction. Du *et al.* pointed out that oxidative stress is an important
 212 pathogenic factor in leukemia-prone marrow diseases such as Fanconi anemia [29]. It has been
 213 proposed that the FA pathway plays a crucial role in protecting major antioxidant defense genes
 214 from oxidative damage. This protection is probably accomplished, in response to oxidative stress, by
 215 the interaction with the chromatin-remodeling machinery. Indeed, it has been shown that the

216 oxidative stress-induced activation of FA pathway (FANCD2 ubiquitination) **is** required for the
 217 formation of the FA-BRG1-promoter complex. This complex is essential for the protection of the
 218 antioxidant gene promoters from oxidative damage.

219 Thus, the activation of FA pathway under the conditions of oxidative stress led to cellular
 220 protection through various ways. Taking into account the above-mentioned consideration, these
 221 ways might be presented as follows:

222

223 **Table 1**

224 **FA PATHWAYS OF CELLULAR PROTECTION**

225

226 **1. Protection of antioxidant gene promoters**

227

228 FANCD2 ubiquitination

229 ↓

230 FA-BRG1-promoter complex

231 ↓

232 Protection of antioxidant gene promoters

233

234

235 **2. Formation of the disulfide complex**

236

237 $FANCA + FANCF + O_2^- \rightarrow FANCA-SS-FANCF$

238

239

240 **3. Suppression of DNA double-strand breaks**

241

242 FANCD2 → m-TOR pathway → DNA double-strand breaks

243

244

245 **4. Suppression of ROS overproduction**

246

247 Antioxidants (quercetin and NAC) → ROS↓ (diabetes and hydrocephalus in FA patients)

248

249 **5. Enhancement of gene antioxidant activity**

250

251 The interaction of genes with enzymes (Foxo3a, p50, p450)

252

253

254 **CONCLUSIONS**

255 (1) Direct interaction of FANCA and FANCF with ROS (superoxide) forms a complex through the
256 intermolecular disulfide bonds.

257 (2) The regulation of FANCD2 by the m-TOR pathway might lead to the resistance of cells to DNA
258 double-strand breaks as in cervical cancer.

259 (3) FANCD2 ubiquitination following the formation of FA-BRG1-promoter complex **is** capable of
260 protection of antioxidant gene promoters.

261 (4) Antioxidant activity of FA genes **is** enhanced by the interaction with various enzymes such as
262 Foxo3a, p50, p450.

263 (5) Application of antioxidants and free radical scavengers (quercetin and NAC) **may lead to**
264 **decreased** ROS overproduction in such diseases as diabetes or hydrocephalus in FA patients.

265

266 **REFERENCES**

267 [1] D'Andrea AD. The Fanconi anemia and breast cancer susceptibility pathways. *N Engl J Med.*

268 2010; 362: 1909-19.

269 [2] Narayan G, Arias-Pulido H, Nandula SV, Basso K, Sugirtharaj DD, Vargas H, et al. Promoter
270 hypermethylation of *FANCF*: Disruption of Fanconi Anemia-BRCA pathway in cervical cancer.

271 *Cancer Res.* 2004; 64: 2994–97.

272

273 [3] Joenje H, Frants RR, Arwert F, de Bruin GJ, Kostense PJ, van de Kamp JJ, et al. Erythrocyte
274 superoxide dismutase deficiency in Fanconi's anaemia established by two independent methods of
275 assay. *Scand J Clin Lab Invest.* 1979; 39: 759-64.

- 276 [4] Joenje H, Arwert F, Eriksson AW, de Koning H, Oostra AB. Oxygen-dependence of
277 chromosomal aberrations in Fanconi's anaemia. *Nature*. 1981; 290: 142-3.
- 278 [5] Korkina LG, Samochatova EV, Maschan AA, Suslova TB, Cheremisina ZP, Afanas'ev IB.
279 Release of active oxygen radicals by leukocytes of Fanconi anemia patients. *J Leukocyte Biol*. 1992;
280 52: 357- 62.
- 281 [6] Korkina LG, Samochatova EV, Maschan AA, Suslova TB, Cheremisina ZP, Afanas'ev IB. The
282 use of rutin for the treatment of Fanconi anemia patients. *Drugs of Today*. 1992; 28: (Suppl A)165-
283 9.
- 284 [7] Afanas'ev IB, Dorozhko AI, Brodskii AV, Kostyuk VA, Potapovitch AI (1989). Chelating and
285 free radical scavenging mechanisms of inhibitory action of rutin and quercetin in lipid peroxidation.
286 *Biochem Pharmacol*. 1989; 38: 1763-9.
- 287 [8] Kozlov AB, Ostrachovitch EA, Afanas'ev IB. Mechanism of inhibitory effects of chelating
288 drugs on lipid peroxidation in rat brain homogenates. *Biochem Pharmacol*. 1994; 47: 795-9.
- 289 [9] Afanas'ev IB, Ostrachovitch EA, Abramova NE, L.G.Korkina LG. Different antioxidant
290 activities of bioflavonoid rutin in normal and iron-overloading rats. *Biochem Pharmacol*. 1995; 50:
291 627-35.
- 292 [10] Afanas'ev IB, Afanas'ev II, Deeva IB, Korkina LG. Free radical formation and oxyhemoglobin
293 oxidation in β -thalassemic red blood cells in the presence prooxidants: effects of free radical
294 scavenger rutin and oral chelator L1. *Transfusion Sci*. 2000; 23: 237-8.
- 295 [11] Afanas'ev IB. On mechanism of superoxide signaling under physiological and
296 pathophysiological conditions. *Med Hypotheses*. 2005; 64: 127- 9.
- 297 [12] Afanas'ev IB. Competition between superoxide and hydrogen peroxide signaling in heterolytic
298 enzymatic processes. *Med Hypotheses*. 2006; 66: 1125-8.

- 299 [13] Poot M, Gross O, Epe B, Pflaum M, Hoehn H. Cell cycle defect in connection with oxygen and
300 iron sensitivity in Fanconi anemia lymphoblastoid cells. *Exp Cell Res.* 1996; 222: 262-8.
- 301 [14] Afanas'ev I. Nucleophilic mechanism of ROS/RNS signaling in cancer epigenetic
302 modifications. *Am J Biomed Sci.* 2012; 4: 282-306.
- 303 [15] Hadjur S, Ung K, Wadsworth L, Dimmick J, Rajcan-Separovic E, Scott RW, Buchwald M, Jirik
304 FR. Defective hematopoiesis and hepatic steatosis in mice with combined deficiencies of the genes
305 encoding Fance and Cu/Zn superoxide dismutase. *Blood.* 2001; 98: 1003-11.
- 306 [16] Du W, Adam Z, Rani R, Zhang X, Pang Q. Oxidative stress in Fanconi anemia hematopoiesis
307 and disease progression. *Antioxidants & Redox Signaling.* 2008;10: 2108-29.
- 308 [17] Kumari U, Jun WY, Bay BH, Lyakhovich A. Evidence of mitochondrial dysfunction and
309 impaired ROS detoxifying machinery in Fanconi Anemia cells. *Oncogene.* 2014; 33:165-172.
- 310 [18] Park SJ, Ciccone SLM, Beck BD, Hwang B, Freie B, Clapp DW, Lee S-H. Oxidative
311 stress/damage induces multimerization of Fanconi anemia proteins. *J Biol Chem.* 2004; 279:
312 30053-9.
- 313 [19] Saadatzaheh MR, Bijangi-Vishehsaraei K, Hong P, Bergmann H, Haneline LS. Oxidant
314 hypersensitivity of Fanconi anemia type C-deficient cells is dependent on a redox-regulated
315 apoptotic pathway. *J Biol Chem.* 2004; 279: 16805-12.
- 316 [20] Futaki M, Igarashi T, Watanabe S, Kajigaya S, Tatsuguchi A, Wang J, Liu JM.
317 The FANCG Fanconi anemia protein interacts with CYP2E1: possible role in protection against
318 oxidative DNA damage. *Carcinogenesis.* 2002; 23: 67-72.
- 319 [21] Freie B, Li X, Ciccone SL, Nawa K, Cooper S, Vogelweid C. et al. Fanconi anemia type C
320 and p53 cooperate in apoptosis and tumorigenesis. *Blood.* 2003; 102: 4146-52.
- 321 [22] Rani R, Li J, Pand Q. Differential p53 engagement in response to oxidative and oncogenic
322 stresses in Fanconi Anemia mice. *Cancer Res.* 2008; 68: 9693-702.

- 323 [23] Afanas'ev I. Reactive oxygen species and age-related genes p66Shc, Sirtuin, Foxo3 and Klotho
324 in senescence. *Oxid Med Cell Longevity*. 2010; 3:1-9.
- 325 [24] Tothova Z, Kollipara R, Huntly BJ, Lee BH, Castrillon DH, Cullen DE, et al. FoxOs are critical
326 mediators of hematopoietic stem cell resistance to physiologic oxidative stress. *Cell*. 2007; 128:
327 325-39.
- 328 [25] Li J, Du W, Maynard S, Andreassen PR, Pang O. Oxidative stress specific interaction between
329 FANCD2 and FOXO3a. *Blood*. 2010; 115: 1545-8.
- 330 [26] Li X, Li L, Li J, Sipple J, Schick J, Mehta PA, et al. Concomitant inactivation of Foxo3a and
331 Fanccl or Fancd2 reveals a two-tier protection from oxidative stress-induced
332 hydrocephalus. *Antioxid Redox Signal*. 2004; 21:1675-92.
- 333 [27] Li J, Sipple J, Maynard S, Mehta PA, Rose SR, Davies SM, Pang O. Fanconi anemia link
334 reactive oxygen species to insulin resistance and obesity. *Antioxid Redox Signal*. 2012; 8: 1083-98.
- 335 [28] Ponte F, Sousa R, Fernandes AP, Goncalves C, Barbot J, Carvalho F, Porto B. Improvement of
336 genetic stability in lymphocytes from Fanconi anemia patients through the combined effect of α -
337 lipoic acid and N-acetylcysteine. *J Rare Diseases*. 2012; 7: 28, 1-11.
- 338 [29] Du W, Rani R, Sipple J, Schick J, Myers KC, Mehta P, et al. The FA pathway counteracts
339 oxidative stress through selective protection of antioxidant defense gene promoters. *Blood*. 2012;
340 119: 4142 - 51.
- 341 [30] Mukhopadhyay SS, Leung KS, Hicks MJ, Hastings PJ, Youssoufian H, Plon SE. Defective
342 mitochondrial peroxiredoxin-3 results in sensitivity to oxidative stress in Fanconi anemia.
343 *J Cell Biol*. 2006;175: 225-35.
- 344 [31] Shen C, Oswald D, Phelps D, Cam H, Pelloski CE, Pang O, Houghton PF. Regulation of
345 FANCD2 by the mTOR pathway contributes to the resistance of cancer cells to DNA double-strand
346 breaks. *Cancer Res*. 2013; 73(11) June 1.

347

348