# 1 Original Research Article 2 EFFECTS OF ETHANOLIC EXTRACT OF MONODORA MYRISTICA 3 SEED (AFRICAN NUTMEG) ON SOME LIVER FUNCTION 4 PARAMETERS USING ALBINO WISTAR RATS

## 5 ABSTRACT

Aim: This study was carried out to Investigate the effects of *Monodora myristica* on the
physiological status of the liver of consumers who consume it for medicinal and nutritive
purposes.

9 Study design: This research was conducted at the Department of Human Physiology, Faculty of
10 Basic Medical Sciences, University of Port Harcourt, Nigeria; between May and October, 2014.

Methodology: Thirty six (36) albino Wistar rats weighing between 180 - 220g were used in the 11 12 study. They were grouped into 3 groups of 12 rats each (2 test groups and a control group). The animals were fed with standard feed and clean water, in addition, those of test groups 2 and 3 13 also received 400mg/kg and 200mg/kg of ethanolic extract of *M. myristica* seed respectively on 14 daily basis for twenty eight days (four weeks). After each week of administration, three rats from 15 each group were sacrificed and blood samples collected by cardiac puncture for biochemical 16 analysis of some liver function parameters (alkaline phosphatase (ALP), alanine transaminase 17 (ALT), aspartate aminotransferase (AST), serum total protein and serum albumin). The liver of 18 the sacrificed animals were also harvested for histological study. 19

Results: The results from the biochemical assay were subjected to statistical analysis and it showed a significant (P<0.05) increase in ALT values, non significant increase in AST and ALP values, and a non significant effect on total serum protein and serum albumin. The histological study of the harvested liver showed hepatocytes degeneration and periportal inflammations, which are indications of alteration on the normal physiological status of the liver.

Conclusion: Findings from this study showed that the extract produced an adverse alteration in
the normal functioning of the liver. Therefore, the plant should be used with caution, especially
when used for medicinal purposes.

Keywords: Ethanolic extract, *Monodora myristica*, liver enzymes, hepatocytes and physiological
status

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#### 31 **INTRODUCTION**

The use of medicinal plants as therapy for health conditions is an age long practice [1]. This 32 usage has gained prominence worldwide over the last three decades and has been estimated that 33 34 at present over two third of the developing countries' population relies on plant preparation as medicines to take care of their health needs [2]. This surge in the use of medicinal plants as 35 therapy is believed to be due to their accessibility and affordability and the perceived failure of 36 synthetic drugs in the treatment of some chronic diseases like hypertension, diabetes, 37 arthrosclerosis etc. [2]. According to Akerele (1993) [3], about 65-80% of the world's population 38 in developing countries, due to poverty and lack of access to modern medicine, depend 39 essentially on plants and their formulations for their primary health care. 40

Medicinal plants and their formulations have continued to attract attention as a result of the strong speculations and belief that they are safe and very efficacious [4] [5]. This strong speculation and assumption has led to the indiscriminate use of medicinal plants and their formulations especially in developing countries, like Nigeria [1].

M. myristica is a species of plant which belong to the family of annonaceae [6]. M. myristica 45 forms a large branching tree with a gray-barked trunk and reach 35m high in nature. It has large 46 leaves (35cm long and 18cm wide) at the end of its branches. The leaves are purple at first but 47 turn a smooth deep green on the upper side with paler green underneath. They are prominently 48 veined and the petiole is purplish [7] [8]. This is widely distributed from Africa to Asia, Central 49 and South America and Australia [9] [10]. It is native to Central, East and West Africa [11]. M. 50 myristica grows very well in the ever green forest of West Africa and in Nigeria are most 51 prominent in the Southern part [12] [13]. Its common names include; Calabash nutmeg, African 52 53 nutmeg, False nutmeg, Jamaican nutmeg, while its local names include Ehuru or Ehiri (Igbo),

Ariwo (Yoruba) [9], Erhe (Urhobo), Ehinawosin (Ikale), Uyengben (Edo) [11]. M. mvristica 54 seed is oblongoid inshape and pale brown in colour with a thin seed coat and hard kernel. 55 Phytochemically, M. myristica seeds have been reported to contain secondary metabolites like 56 saponins, tannins, flavonoids, glycosides, alkaloids and steroids [13] [14] [15]. They also contain 57 minerals like potassium, sodium, magnesium, phosphorus and iron [13]. They have also been 58 reported to contain amino acids like phenylalanine, tyrosine, arginine, glutamic acid and 59 asparagines and vitamin C and E and sugars [16] [17]. *M myristica* has been used as herbal plant 60 and spices since ancient times. It is used in the treatment of hemorrhoids stomach ache, fiber 61 pain, and constipation. [18] [9], control passive uterine hemorrhage after childbirth [19] [20] [9] 62 and has also been associated with antisickling effectiveness [17]. As spices, the seeds are grinded 63 and used in cooking pepper soup and stew [13] [21]. The seeds are also used as an aromatic and 64 stimulating addition to medicines and snuff [7]. In Central African Republic, the seeds are used 65 as condiment and drug in the treatment of headache and hypertension [22]. The direct action of 66 these plant extracts is on the liver, which is central to drug metabolism [23]. 67

The liver is the largest internal and very vital organ in the body, constituting about 2.5% of an 68 adult's body weight. The liver plays an important role in maintaining blood glucose levels. It 69 also regulates the circulating blood lipids by the amount of very low density lipoproteins 70 (VLDLs) it secretes. Liver takes up numerous toxic compounds and drugs which may include 71 medicinal plant formulations from the portal circulation [24]. Many drugs and metabolites are 72 hydrophobic, and the liver converts them into hydrophilic compounds and in the process some 73 may adversely affect the liver. Such drugs or metabolites are said to be hepatotoxic, and their 74 effects on the liver are determined by measuring the plasma concentration of some biochemical 75 76 compounds and enzymes produced by the liver (liver makers). Such liver makers include total

serum protein, albumin, triglycerides, total cholesterol, high and low density lipoproteins, and 77 liver enzymes like alkaline phosphatase, aspartate transaminase and alanine transaminase, etc. 78 Presently, some research studies have showed that some of these used medicinal plants adversely 79 80 affect some vital organs in the body while exhibiting their therapeutic potentials [25]. Consequently, it has become imperative to ascertain the effects of plants used as herbs on the 81 physiological status of vital organs. Hence this study which is aimed at investigating the effects 82 of the ethanolic extract of *M. myristica* seed (African Nutmeg) on the physiological status of the 83 liver using albino Wistar rats. 84



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Figure 1: Fruit (pod) and seeds of African nutmeg.

#### 87 METHODS

88 This research was conducted at the Department of Human Physiology, Faculty of Basic Medical

89 Sciences, University of Port Harcourt, Choba, Nigeria; between May and July, 2014.

#### 90 Experimental Animals

91 Thirty six (36) male albino Wistar rats weighing (180 – 220g) were purchased and kept at the 92 animal house, Department of Human Physiology, College of Health Sciences, University of Port 93 Harcourt, Rivers State, Nigeria. The animals were kept in a spacious and well ventilated cage at 94 room temperature; under 12 hours light and dark cycle and acclimatize for 14 days. They were allowed free access to feed diet (Top Feeds, Broiler finisher – Product of Eastern premier feed
mills Ltd.) and water *ad libitum*.

All animal experiments were conducted in compliance with NIH guidelines for Care and Use of
Laboratory Animals (Pub No. 85-23, Revised 1985).

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#### 100 Collection and Identification of Plant Materials

Dried seeds of *M. myristica* were purchased from Abacha Market along Udu road, Udu LGA.,
Delta State. The plant sample was correctly identified and authenticated in the Herbarium Unit of
the Department of Plant Science and Biotechnology, University of Port Harcourt.

#### 104 Preparation of Ethanolic Extracts of Monodora Myristica Seed

The ethanolic extract of *M. myristica* seeds was prepared according to [9]. The seeds of *M. myristica* were dehulled (the coat removed), and rid of bad seeds and dirt. Thereafter, the seeds were milled to fine powder using manual engine grinder (Model Corene, A.5 lander YCIA S.A). The milled sample of the plant were soaked in 5L of 80% Ethanol for 48 hours, thereafter filtered with Whatman No. 1 filter paper to separate the filtrate from the residue. The filtrate was concentrated under reduced pressure in a vacuum at 45°C using a rotary evaporator (Searl Instruments Ltd. England) to produce a brownish gel like fluid called ethanolic extract.

#### 112 Experimental Design

113 Thirty six (36) male albino Wistar rats weighing (160 - 220g) were randomized into three (3) 114 groups of 12 rats each. Group I: Served as the Control, and received water and normal feed. 115 Group II: Received 400mg/kg of the extract. Group III: Received 200mg/kg. The doses of the 116 extract (400mg/kg and 200mg/kg) were chosen based on a previous work that determined the 117 LD<sub>50</sub> of *M. myristica* seed to be >5000mg/kg [26]. The extracts were administered orally in the 118 early hours of each day (within 8am to 10am) throughout the period of administration.

Three rats from each group were sacrificed after every seven days, that is, on day 8, day 15, day 22 and day 29 day. After each sacrifices, blood was collected by cardiac puncture into lithium heparin bottles for biochemical analysis, while the liver of each rat was collected via abdominothoracic dissection into plain bottle containing buffered formalin for histological study.

#### **123 Biochemical Parameters**

The collected blood samples were centrifuged at 5000rpm for 10 minutes to obtain clear serum 124 for the biochemical analysis. The serum supernatant was then carefully aspirated with needled 125 syringe and stored in a plain sample bottle for biochemical analysis. The biochemical analysis 126 for serum alanine aminotransferase (ALT), serum aspartate aminotransferase (AST), serum 127 alkaline phosphatase (ALP), total serum protein and serum albumin were done using Mindray 128 Auto-analyzer machine (Model: BS – 800M) in the laboratory of the Department of Chemical 129 Pathology at the University of Port Harcourt Teaching Hospital following standard laboratory 130 131 procedures.

#### 132 Histopathological Examination

Histopathological examination was done following the method outlined by Ekeanyanwu andNjoku (2014).

#### 135 Statistical Analysis

The data were statistically analyzed using SPSS VERSION 20.0, and was analyzed for statistical significance by one way Analysis of Variance (ANOVA) followed by Dunnett's post-test for comparison with control group. The difference was considered to be significant at 5% level (P<0.05).

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#### 142 **RESULTS**

- 143 Results are presented in tables 1-5 and figure 1-5 below;
- 144 Table 1: Effects of ethanolic extracts of *M. myristica* on serum ALT (IU/L) in albino

Groups	ALT (IU/L) Week 1	ALT (IU/L) Week 2	ALT (IU/L) Week 3	ALT (IU/L) Week 4
Control	72.67±8.76	76.28±4.18	71.73±6.34	69.97±2.88
400mg/kg <i>M</i> . <i>myristica</i>	197.33±42.08*	198.00±4.73*	212.67±7.27*	205.00±15.18*
200mg/kg <i>M</i> . <i>myristica</i>	162.67±32.2	146.00±7.77	198.33±112.10*	154.67±2.33

Values are expressed as Mean±SEM; n=3; \*: Significant at P<0.05

146 The result of serum ALT concentrations of rats administered 400mg/kg and 200mg/kg doses of

147 the ethanolic extract of *M. myristica* is presented in table 1 above. The result showed a dose

- dependent significant P < 0.05 increase in the test groups in comparison with the control group.
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#### Table 2: Effects of ethanolic extracts of *M. myristica* on serum AST (IU/L)

Groups	AST (IU/L) Week 1	AST (IU/L) Week 2	AST (IU/L) Week 3	AST (IU/L) Week 4
Control	323.33±22.85	333.45±8.47	325.63±19.65	320.43±21.58
400mg/kg M. <i>myristica</i>	501.67±204.6	388.33±54.23	480.67±26.59	632.33±88.89
200mg/kg M. <i>myristica</i>	328.33±141.21	378.33±6.89	383.67±511.36	432.67±37.55

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Values are expressed as Mean ± SEM; n=3; \*; Significant at P<0.05

The result of serum AST concentrations of rats administered 400mg/kg and 200mg/kg doses of the extract is presented in table 2 above. The result showed a non significant P>0.05 increase in the test groups in comparison with the control group.

Table 3: Effects of ethanolic extracts of *M. myristica* on serum ALP (IU/L)

Groups	ALP (IU/L) Week 1	ALP (IU/L) Week 2	ALP (IU/L) Week 3	ALP (IU/L) Week 4
Control	298.33±18.32	293.72±06.24	289.94±14.12	298.33±12.98
400mg/kg M. myristica	326.33±41.86	312.00±14.93	353.67±34.37	356.00±89.50
200mg/kg M. myristica	321.00±134.97	294.00±14.57	317.00±151.24	307.67±11.20

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Values are expressed as Mean±SEM; n=3; \*: Significant at P<0.05

The effects of 400mg/kg and 200mg/kg doses of the extract on serum ALP of rats is presented in the table above. The result showed a non significant increase in the test groups in comparison with the control group.

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## Table 4: Effects of ethanolic extracts of *M. myristica* on serum total protein (g/L)

Groups	Total	Total	Total	Total
	Protein(g/L)	Protein(g/L)	Protein(g/L)	Protein(g/L)
	Week 1	Week 2	Week 3	Week 4
Control	64.00±0.58	65.03±1.35	62.14±0.33	65.32±0.29
400mg/kg				
M. myristica	62.67±2.60	65.00±2.08	65.67±3.84	63.33±2.73
200mg/kg			64.33±4.70	
M. myristica	61.67±1.20	66.00±1.73		59.33±4.06

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Values are expressed as Mean±SEM; n=3; \*: Significant at P<0.05

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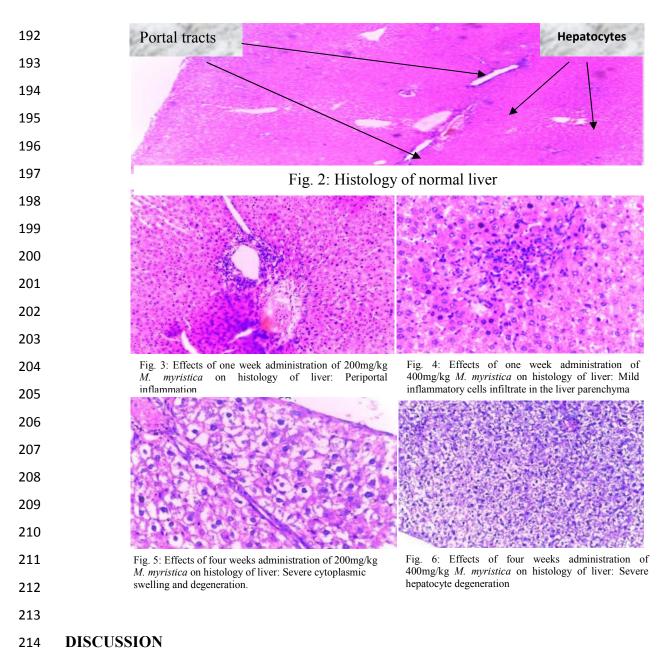
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	Groups	Albumin(g/L)	Albumin(g/L)	Albumin(g/L)	Albumin(g/L)
		Week 1	Week 2	Week 3	Week 4
	Control	29.00±0.58	29.68±0.18	30.00±0.47	33.53±1.08
	400mg/kg				
	M. myristica	34.00±1.53	30.00±0.58	27.67±0.67	33.00±1.73
	200mg/kg <i>M. myristica</i>	31.33±1.45	30.33±0.67	26.33±3.48	30.67±0.88
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171 172		v alues are express	eu as mean±SEM;	n=3; *: Significant a	at r < 0.03
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174	doses of the ext	ract are presented in	n tables 4 and 5 res	pectively. The result	ts showed non signif
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Table 5: Effects of ethanolic extracts of *M. myristica* on serum albumin (g/L)



215 This study is aimed at investigating the effects of ethanolic extract of *M. myristica* seed on the

- 216 physiological status of the liver.
- 217 The physiological status (functionality or health state) of the liver is mainly determined by
- 218 measuring the plasma levels of some enzymes called the liver enzymes which include alkaline
- 219 phosphatase (ALP), aspartate transaminase (AST) and alanine transaminase (AST), etc [27];
- Ekeanyanwu and Njoku 2014) and other biochemicals produced by the liver which include total

serum protein, albumin, bilirubin, triglycerides, total cholesterol, high and low density 221 lipoproteins, etc. Normally ALT and AST are mainly present and in high concentrations within 222 the liver cells (hepatocytes). Damage or destruction of the hepatocytes leads to the release of 223 224 these enzymes into circulation thereby increasing their plasma levels. Thus, increase in the plasma or serum level of any of these enzymes is an indication of hepatocytes damage. ALP is an 225 enzyme in the cell lining of the biliary ducts of the liver. Its plasma level increases when liver 226 bile duct obstruction is present or there is intrahepatic cholestasis or infiltrative diseases 227 (Angelico *et al.*, 2010). The result from the biochemical analysis of this study showed that the 228 400mg/kg and 200mg/kg dose of the extract significantly (P<0.05) increased the serum ALT, 229 and in a dose dependent manner (table 1). This increase is an indication of liver (hepatocytes) 230 damage and is confirmed by the histological study of the harvested liver organs. 231

Though, both administered doses cause general increase in the serum AST and ALP, but the increase was not significant as shown in tables 2 and 3.

From the results, it was observed that the ethanolic extract of *M. myristica* do not have any effect on the total serum protein and albumin as shown in tables 4 and 5.

The histological study of the harvested liver shows that the ethanolic extract of *M. myristica* negatively alters the physiological status of the liver in a dose and time depended manner, as various degrees of hepatocytes and periportal inflammation and degeneration was observed.

In conclusion, this study reveals that *Monodora myristica* seed have negative effects on the anatomical physiology of the liver. Therefore, we advised that caution should be taken in employing their medicinal effects. Further studies should be done in order to ascertain the biochemical components of the plant responsible for this impairment of the liver and the mechanisms.

244	RI	EFERENCES
245	1.	Mbaka CO, Adeyemi OO and Orumosu AA. Acute and subchronic toxicity studies of the
246		ethanolic extract of the leaves of Sphenocentrum jollyyanum (Menispermaceae). Agric. Biol.
247		J N Am. 2010; 1(3):265-272.
248	2.	Okpuzor J, Ogbunugafor HA and Kareem GK. Hepatic and Hematologic Effects of Fractions
249		of Globimetula Braunii in Normal Albino Rats. EXCLI Journal 2009; 8: 182-189.
250	3.	Akerele O. Summary of WHO guidelines for the assessment of herbal medicines.
251		HerbalGram. 1993; 28: 13-19.
252	4.	Said O, Khalil K, Fulder S and Azaizeh H. Ethnobotanical survey of medicinal herbs of the
253		Middle Eastern region. J. Ethnopharmacol. 2002; 83: 251-265.
254	5.	Farnsworth NR and Soejarto DD. Potential consequence of plant extinction in the United
255		States on the current and future availability of prescription drugs. Econ Bot. 1985; 39: 231-
256		240.
257	6.	Ekeanyanwu RC and Njoku OU. Acute and subacute oral toxicity study on the flavonoid rich
258		fraction of Monodora tenuifolia seed in albino rats. Asian Pacific ournal of Biomedicine.
259		2014; 4(3): 194- 202.
260	7.	Burkill HM. The useful plants of west Tropical Africa. 1985; Vol.1
261	8.	Barwick M. Tropical and subtropical Trees; a Worldwide Encyclopaedic Guide. Thames and
262		Hudson, London. 2004.
263	9.	Akinwunmi Femi Feyisayo and Oyedepo Oluboade Oluokun. Evaluation of antioxidant
264		potentials of Monodora myristica (Gaertn) dunel seeds. African Journal for Food Science.
265		2013; Vol. 7(9) pp 317-324.

266	10. Omobuwajo TO, Omobuwajo OR, Sanni LA. Physical properties of calabash nutmeg
267	(Monodora myristica) seeds. J. Food Eng. 2003; 57: 375-381.
268	11. Keay RWJ. Trees of Nigeria. Claridon Press Oxford. 1989; Vol II issue 3. pp. 5.
269	12. Adegoke E and Akinsanya A. Studies of Nigerian Medicinal Plants. J. West Afr. Sci. Assoc.
270	1970; 13(2):15-65.
271	13. Bassey ME, Johnny II and Okoro BI. Lesser known spices of Akwa Ibom; their nutritional,
272	antinutritional, mineral and phytochemical analysis. Archives of Applied Science Research,
273	2011; 3(3); 553-559.
274	14. Uhegbu FO, Iweala EJ, Kanu I. Studies on the chemical and anti nutritional content of some
275	Nigerian Spices. Inter. J. Nutri. Metab. 2011; 3(6):72-76.
276	15. Ekeanyanwu RC, Etienajirhevwe OF. Invitro antihelmintic potentials of Xylopia aethiopica
277	and Monodora myristica from Nigeria. Afr. J. Biochem. Res. 2012; 6(9):115-120.
278	16. Nwachukwu N. Nutritional and Antinutritional substances in some selected indigenous
279	spices Ph.D Thesis, FUTO, Nigeria. 2000.
280	17. Uwakwe AA, Nwaoguikpe RN. In vitro antisickling effects of Xylopia aethiopica and
281	Monodora myristica on sickle cell blood. Journal of Medicinal Plant Research. 2008;
282	2(6):119-124.
283	18. Gill LS. Ethnomedical uses of plants in Nigeria. University of Benin Press, Benin city,
284	Nigeria. 1992; Pp. 165, 248.
285	19. Nwaoguikpe RN, Ujowundu CO, Emejulu AA. The Antioxidant and Free Radical
286	Scavenging Effects of Extracts of Seeds of Some Neglected Legumes of South-East Nigeria.
287	Sch. Acad. J. Biosci. 2014; 2(1): 51-59.

299

288	20. Okafor JC. Edible indigenous woody plants in the rural economy of the Nigerian forest zone.
289	Forest Ecology and Management. 1981; 3: 48-55.
290	21. Okafor JC. Development of Forest tree crops for food supply in Nigeria. Forest Ecology and
291	Management. 1987; 1: 235-247.
292	22. Koudou J, Etou Ossibi AW, Aklikokou K, Abena AA, Gbeassor M and Bessiere JM.
293	Chemical Composition and Hypotensive Effects of Essential Oil of Monodora myristica
294	Gaertn. Journal of Biological Sciences. 2007; 7 (6): 937-942.
295	23. Guyton A and Hall JA. Textbook of Medical Physiology. 9th Edition; W.B. Saunders's
296	Company. 1996.
297	24. Maton Anthea, Jean Hopkins, Charles McLaughlin, Susan Johnson, Maryanna Warner,
298	David LaHart, et al. Human Biology and Health. Englewood Cliffs, New Jersey, USA:

25. Chan K. Some aspect of toxic contaminants in herbal remedies. A review. Chemosphere.
2003; 52: 1361-1371.

Prentice Hall. 1993; ISBN 0-13-981176-1.

- 302 26. Ezenwali MO, Njoku OU and Okoli CO. Studies on the anti-diarrheal properties of seed
- extract of *Monodora tenuifolia*. International Journal of Applied Research in Natural
  Products. 2010; Vol. 2(4), pp. 20-26.
- 27. Ekeanyanwu RC, Njoku O and Ononogbu IC. The Phytochemical Composition and Some
  Biochemical Effects of Nigerian Tigernut *(Cyperus esculentus L.)* Tuber. Pakistan Journal of
  Nutrition. 2010; 9 (7): 709-715.
- 28. Aliyu R, Adebayo AH, Catsing D and Garba IH. The effects of ethanolic extract leaf of
  Commiphora Africana (Burseraceae) on rat liver and kidney function. J. Pharmacol Toxicol.
  2006; 2: 373-379.

- 311 29. Angelico F and Del-Ben M. Toward predicting therapeutic response in patients with hepatitis
- C: author's reply. Ailment pharmacol ther. 2010; 31: 339-340.