Free radicals: Health implications and their mitigation by herbals

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Abstract

Free radicals pose a serious threat to tissues and vital organs, especially membrane lipids, proteins and nucleic acids of cells. Overproduction of reactive oxygen/ nitrogen species (ROS/RNS) and other related radicals lead to oxidative stress which has been implicated in aging and a number of diseases. Free radicals react with biomolecules and cause lipid peroxidation, loss of enzyme activity, mutation and carcinogenesis. A number of degenerative diseases including cardiovascular disease, diabetes, and adverse hepatic conditions have been attributed to accumulation of free radicals. Diseases resulting from radical overload might also lead to different types of cancers. However free radicals at low or moderate levels are vital to human health. ROS and RNS produced in a well regulated manner help maintain homeostasis at the cellular level in the normal healthy tissues and play an important role as signaling molecules. Cellular antioxidant enzyme systems including superoxide dismutase, catalase, glutathione peroxidases/reductase, peroxiredoxins along with non enzymatic antioxidants viz., tocopherols, vitamin C, and glutathione etc., apart from several dietary components protect cells and organisms from the lethal effects of excessive ROS production. Natural products of plant origin have been used in traditional medicine for the treatment of diseases resulting from radical overload. The diversity of phytochemicals such as polyphenols, flavonoids, carotenes and saponins etc. present in plants and dietary components provide drug leads for the development of novel therapeutic agents. This review deals with the components of free radical biology, their adverse consequences in humans and amelioration of diseases by botanical therapeutics.

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Key words: Reactive oxygen species, Oxidative stress, Cancer, Aging, Diabetes, Plant Products.

## 36 1. INTRODUCTION

37 Oxidative stress is initiated by free radicals, which seek stability through electron pairing with biological 38 macromolecules in healthy human cells and cause protein and DNA damage along with lipid per-39 oxidation. It may be defined as an imbalance between free radicals and antioxidants in our body (Figure 1). Free radicals are fundamental to any biochemical process and represent an essential part of aerobic 40 life and metabolism [1]. In general, free radicals are very short lived, with half lives in milli, micro or 41 nanoseconds. The most common reactive oxygen species (ROS) include superoxide (O<sup>2-</sup>) anion, 42 43 hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), peroxyl (ROO<sup>-</sup>) radicals, and reactive hydroxyl (OH<sup>-</sup>) radicals. The nitrogen 44 derived free radicals are nitric oxide (NO<sup>-</sup>) and peroxynitrite anion (ONOO<sup>-</sup>). Under physiological 45 conditions, ROS formation and elimination are delicately balanced. However, enhanced activity of oxidant 46 enzymes and/or reduced activity of antioxidant enzymes lead to oxidative stress. Majority of the 47 diseases/disorders are mainly linked to oxidative stress produced due to free radicals [2, 3].

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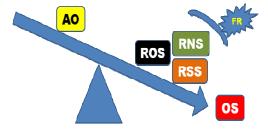
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48 ROS have been implicated in over a hundreds of disease states which range from arthritis, connective 49 tissue disorders to carcinogenesis, aging, physical injury, infection and acquired immunodeficiency 50 syndrome [4, 5]. Pathological conditions that predispose to cardiovascular events, such as hypertension, 51 hypercholesterolemia, and diabetes, are associated with oxidative stress. Antioxidant therapy has gained 52 an immense importance in the treatment of these diseases. Antioxidants have been reported to prevent 53 oxidative damage caused by free radicals and ROS, and may prevent the occurrence of diseases such as 54 cancer and aging. They can interfere with the oxidation process by reacting with free radicals, chelating 55 catalytic metals, and also acting as oxygen scavengers [6, 7, 8]. Many phytochemicals have been found 56 to play as potential antioxidants. Present review summarizes the causes and consequences of free radical generation, antioxidants and use of plants derivatives in controlling diseases. 57

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# 60 **2. FREE RADICALS** 61

62 Free radicals are atoms, molecules or ions with unpaired electrons that are highly unstable, short lived 63 and active towards chemical reactions with other molecules. They may be derived from oxygen, nitrogen 64 and sulfur [9, 10]. Internally, free radicals are produced as a normal part of metabolism within the 65 mitochondria, through xanthine oxidase, peroxisomes, inflammation processes, phagocytosis, 66 arachidonate pathways, ischemia, and physical exercise. External factors that help to promote the 67 production of free radicals are smoking, environmental pollutants, radiation, drugs, pesticides, industrial 68 solvents and ozone. It is paradox that these elements, essential to life (especially oxygen) have 69 deleterious effects on the human body through these reactive species [9].



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71 Fig. 1. Effect of imbalance between antioxidants and free radicals (Abbreviations: AO-antioxidant, ROS-

- reactive oxygen species, RNS-reactive nitrogen species, RSS-reactive sulphur species, FR-free radicals,
   OS-oxidative stress).
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#### 75 **2.1 Reactive oxygen and nitrogen species (ROS and RNS)**

76 Free radicals derived from oxygen and nitrogen are known as reactive oxygen species (ROS) and 77 reactive nitrogen species (RNS), respectively. Formation of ROS and RNS in the cells can occur by 78 enzymatic and/or non-enzymatic reactions. Enzymatic reactions include those involved in the respiratory 79 chain, the prostaglandin synthesis, the phagocytosis, and the cytochrome P450 system [11]. Some of 80 ROS molecules are extremely reactive, such as the hydroxyl radical, while some are less reactive 81 (superoxide and hydrogen peroxide) [5, 12]. The superoxide anion created from molecular oxygen by the 82 addition of an electron is, in spite of being a free radical, not highly reactive. It lacks the ability to 83 penetrate lipid membranes and is therefore enclosed in the compartment where it was produced. The 84 formation of superoxide takes place spontaneously, especially in the electron-rich aerobic environment in

85 vicinity of the inner mitochondrial membrane with the respiratory chain. Superoxide (as well as hydrogen 86 peroxide) is also produced endogenously by flavoenzymes, e.g., xanthine oxidase activated in ischemia-87 reperfusion [13, 14]. Other superoxide-producing enzymes are lipoxygenase and cyclooxygenase [15, 88 16]. Hydrogen peroxide plays a radical forming role as an intermediate in the production of more reactive 89 ROS molecules including hypochlorous acid by the action of myeloperoxidase, an enzyme present in the 90 phagosomes of neutrophils [17]. Most importantly, hydrogen peroxide forms hydroxyl radical in a reaction catalyzed by metal ions (Fe<sup>2+</sup> or Cu<sup>+</sup>), often bound in complex with different proteins or other molecules by 91 92 a reaction known as the Fenton reaction [18, 19].

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94 Nitric oxide (NO) is formed from L-arginine by one of the three NO synthase (NOS) isoforms. The three 95 isoforms are nNOS (identified constitutive in neuronal tissue), iNOS (inducible by cytokines in activated 96 macrophages and liver) and eNOS (identified constitutive in vascular endothelial cells) [20]. NO is rapidly 97 oxidized by oxyhemoglobin to form nitrate, the major end stable oxidation product of NO in the body. NO 98 also reacts with glutathione to form nitrosothiol or with heme to yield heme-NO. Physiologically, 99 nitrosothiol can serve as a vehicle to transport NO in plasma, thereby increasing the biological half-life of 910 physiologic concentrations of NO [21, 22].

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#### 102 **2.2 Physiological functions of free radicals**

103 ROS and RNS are involved in many physiological activities and function as cellular signaling agents. 104 Activation of phagocytes produces ROS in amounts enough to kill intruding bacteria [23]. In this system 105 ROS are produced by the NADPH oxidase complex that converts O2 to O2 - [24, 25]. Superoxide is then 106 reduced in the phagosome by SOD to H<sub>2</sub>O<sub>2</sub> that can be further converted to HOCI by myeloperoxidase 107 [26]. Hypochlorous acid may then spontaneously form hydroxyl radical. The two highly reactive ROS 108 molecules thereby formed in phagosomes (HOCI and •OH) are highly toxic to bacteria ingested by the 109 phagocyte and carry the direct antimicrobial effects of ROS. The hypochlorous acid produced in the 110 myeloperoxidase reaction is also an important part of the antimicrobial defense by destruction of the DNA 111 anchoring at the bacterial membrane, resulting in cessation of DNA replication [27].

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113 ROS can directly affect the conformation and/or activities of all sulfhydryl-containing molecules, such as 114 proteins or GSH, by oxidation of their thiol moiety. This type of redox regulation affects many proteins important in signal transduction and carcinogenesis such as protein kinase C, Ca<sup>2+</sup>-ATPase, collagenase, 115 116 and tyrosine kinases [28], among many other enzymes and membrane receptors [29]. For several 117 transcription factors, ROS function as physiological mediators of transcription control. Well-known 118 examples of redox-sensitive transcription factors are Nuclear Factor-KB (NF-KB) and Activator Protein-1 119 (AP-1) [30]. Activator Protein-1, a dimer of gene products from the Jun and Fos proto-oncogene families, 120 expression is induced by several pro-oxidant conditions, including different types of irradiation [31, 32]. 121 Nitric oxide (NO) is one of the most important signaling molecules. Physiologic levels of NO produced by 122 endothelial cells are essential for regulating the relaxation and proliferation of vascular smooth muscle 123 cells, platelet aggregation, leukocyte adhesion, angiogenesis, vascular tone, thrombosis, and 124 hemodynamics. In addition, NO produced by neurons serves as a neurotransmitter, and NO generated 125 by activated macrophages is an important mediator of the immune response [33, 34].

#### 126 **2.3 Molecular damage induced by free radicals**

127 All the biological molecules present in our body are at risk of being attacked by ROS. It is estimated that 128 every day a human cell is targeted by the hydroxyl radical and other such species on an average of 105 129 times inducing oxidative stress [33]. The main targets of ROS and other free radicals are proteins, DNA 130 and RNA molecules, sugars and lipids [34-37]. Membrane lipids present in sub-cellular organelles are 131 highly susceptible to free radical damage. During lipid per-oxidation a large number of toxic byproducts 132 are also formed that can have effects at a site away from the area of generation, behaving as second 133 messengers. The damage caused by lipid peroxidation is highly detrimental to the functioning of the cell 134 [38]. Oxidation of proteins by ROS/RNS can generate a range of stable as well as reactive products such 135 as protein hydroperoxides that can generate additional radicals particularly upon interaction with 136 transition metal ions. Table 1 summarizes the mechanisms involved in free radical damage of 137 biomolecules. Oxidative damage to DNA is a result of interaction of DNA with ROS or RNS. The C4-C5 138 double bond of pyrimidine is particularly sensitive to attack by hydroxyl radical, generating a spectrum of 139 oxidative pyrimidine damage products, including thymine glycol, uracil glycol, urea residue, 5-140 hydroxydeoxyuridine, 5-hydroxydeoxycytidine, hydantoin and others. 8-Hydroxydeoxyguanidine (8-141 OHdG) has been implicated in carcinogenesis and is considered a reliable marker for oxidative DNA 142 damage [38].

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Targets of free radicals	Mode of damage
Proteins	Oxidative modification of a specific amino acid. Free radical-mediated peptide cleavage. Formation of protein cross -linkage due to reaction with lipid peroxidation products [9].
DNA and RNA	Production of base-free sites. Deletions, modification of bases. Frame shifts. Strand breaks. DNA–protein crosslink and chromosomal arrangements. Oxidation of DNA by hydroxyl radicals [39, 40].
Sugars	Formation of oxygen free radicals during early glycation could contribute to glycoxidative damage [40]. Short sugar fermentation products (glycoaldehyde) due to autoxidation produce superoxide radical [40].
Lipids	Lipid peroxidation takes place by the abstraction of hydrogen atom from a methylene carbon of fatty acid side chain resulting into free radical chain reaction producing peroxyl radicals [41].
	Another way to generate lipid peroxides is through the attack on polyunsaturated fatty acids (PUFA) or their side chain by the singlet oxygen which is a very reactive form of oxygen [41].

#### 148 **3. ANTIOXIDANTS**

149 Antioxidants are substances that neutralize free radicals or their actions [42]. The antioxidants acting in 150 the defense systems act at different levels such as preventive, radical scavenging, repair and de novo, 151 and the fourth line of defense, i.e., the adaptation. The first line of defense is the preventive antioxidants, 152 which suppresses the formation of free radicals. The second line of defense is the antioxidants that 153 scavenge the active radicals to suppress chain initiation and/or break the chain propagation reactions. 154 The third line of defense is the repair and de novo antioxidants. The enzymes present in the cytosol and 155 in the mitochondria of mammalian cells recognize, degrade, and remove oxidatively modified proteins and 156 prevent the accumulation of oxidized proteins. There is another important function called adaptation 157 where the signal for the production and reactions of free radicals induces formation and transport of the 158 appropriate antioxidant to the right site [43]. Antioxidants can be classified into two major classes i.e., 159 enzymatic and non-enzymatic.

#### 161 **3.1 Enzymatic antioxidants**

162 Nature has endowed each cell with adequate protective mechanisms against harmful effects of free 163 radicals. Cellular antioxidant enzyme systems serve to protect cells and organisms from the lethal effects 164 of excessive ROS formation. Superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase and 165 glutathione reductase are examples of some antioxidant enzymes.

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167 In eukaryotic cells, O2-- can be metabolized to hydrogen peroxide by two metal containing SOD 168 isoenzymes, tetrameric Mn-SOD present in mitochondria, and dimeric Cu/Zn-SOD present in the cytosol 169 [43, 44]. In the reaction catalyzed by SOD, two molecules of superoxide form hydrogen peroxide and 170 molecular oxygen and are thereby a source of cellular hydrogen peroxide. In mitochondria, superoxide is 171 formed in relatively high concentrations due to the leakage of electrons from the respiratory chain. 172 Expression of Mn-SOD is, in contrast to Cu/Zn-SOD, induced by oxidative stress [44]. Cytosolic Cu/Zn-173 SOD seems less important than Mn-SOD, and transgenic animals lacking this enzyme are able to adapt 174 so that the phenotype appears normal [45].

176 Catalases of many organisms are mainly heme-containing enzymes [46]. The predominant subcellular 177 localization in mammalian cells is in peroxisomes, where catalase catalyzes the dismutation of hydrogen 178 peroxide to water and molecular oxygen. Catalase also has functions in detoxifying different substrates, 179 e.g., phenols and alcohols, via coupled reduction of hydrogen peroxide. One antioxidative role of catalase 180 is to lower the risk of hydroxyl radical formation from  $H_2O_2$  via the Fenton reaction catalyzed by Cu or Fe 181 ions. Catalase binds NADPH, which protects the enzyme from inactivation and increases its efficiency 182 [47].

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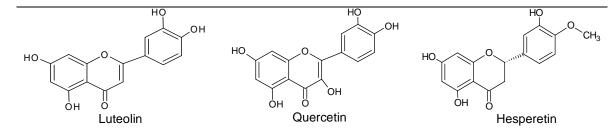
Peroxiredoxins (Prx; thioredoxin peroxidases) are recently discovered enzymes capable of directly reducing peroxides, e.g., hydrogen peroxide and different alkyl hydroperoxides [48]. In mammalian cells, thioredoxin regenerate oxidized Prx formed in the catalytic cycle [49]. In the mitochondria of mammalian cells the mitochondrial thioredoxin system is probably a specific reductant of Prx [50]. Peroxiredoxins have been shown to inhibit apoptosis induced by p53 and by hydrogen peroxide on a level upstream of bcl-2 [51].

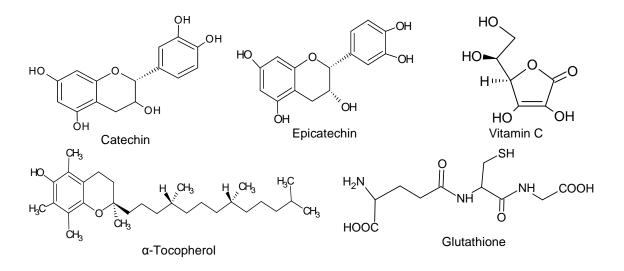
190 There are at least four different Glutathione peroxidases (GPx) in mammals (GPx1-4), all of them

191 containing selenocysteine [52]. GPx1 and GPx4 both are cytosolic enzymes abundant in most tissues. 192 GPx4 has recently been found to have dual functions in sperm cells by being enzymatically active in 193 spermatids but insoluble and working as a structural protein in mature spermatozoa [53]. GPx2 194 (gastrointestinal GPx) and GPx3 (plasma GPx) are mainly expressed in the gastrointestinal tract and 195 kidney, respectively [54]. All glutathione peroxidases may catalyze the reduction of  $H_2O_2$  using 196 glutathione as substrate. They can also reduce other peroxides (e.g., lipid peroxides in cell membranes) 197 to alcohols. Some data has indicated that GPx should be of high antioxidant importance under 198 physiological conditions while others place the enzymes as important only at events of oxidative stress 199 [55]. The function of GPx isoenzymes in antioxidant defense is still unclear, but the kinetic properties and 200 widespread distribution still imply that they constitute major contributors to the total protection against 201 oxidative damage.

### 202 **3.2 Non enzymatic antioxidants**

203 The non-enzymatic antioxidants include tocopherols, carotenoids, ascorbic acid, flavonoids and 204 polyphenols which are obtained from natural plant sources [56]. Some non enzymatic antioxidants are 205 shown in Figure 2. Exposure to DNA by irradiation or hydroxyl radical may leads to the formation of 8-206 hydroxydeoxyguanosine. On this basis Fischer-Nielsen et al. (1992) [57] found that vitamin C at 207 physiological concentration exhibits a protective effect against free radical-induced oxidative damage. 208 Vitamin E and tocotrienols (such as those from palm oil) are efficient lipid soluble antioxidants that 209 function as a chain breaker during lipid peroxidation in cell membranes and various lipid particles 210 including LDL [58, 59]. Animal studies have shown the antioxidant effect of dietary phytochemicals. 211 Among them, phenolic compounds, such as flavonoids exhibit potent antioxidant activities. For example 212 tea polyphenols have capability to enhance red blood cell resistance to oxidative stress; scavenge 213 superoxide and hydroxyl radicals; and inhibition of oxidative modification of low density lipoprotein. 214 Dietary supplementation of polyphenols is also reported to decrease serum concentrations of total 215 cholesterol and malondialdehyde [21].  $\beta$ -Carotene and other carotenoids ( $\alpha$ -carotene,  $\gamma$ -carotene, and  $\beta$ -216 cryptoxanthin) are potent antioxidants of plant origin. They react with a peroxyl radical to form a 217 resonance-stabilized carbon-centered radical within its conjugated alkyl structure, thereby inhibiting the 218 chain propagation effect of ROS. Lycopene, lutein, canthaxanthin, and zeaxanthin also have their 219 antioxidant actions similar to those of  $\beta$  –carotene [60]. A wide range of antioxidants from both natural 220 and synthetic origin have been proposed for use in the treatment of various human diseases [61]. Some 221 synthetic antioxidant compounds commonly used in processed foods have been shown to produce toxic 222 effects like liver damage and mutagenesis [5, 62]. Hence, nowadays search for natural compounds 223 antioxidant source is gaining much importance.





#### Fig.2. Non enzymatic antioxidants

Antioxidant-based drugs/formulations for prevention and treatment of complex diseases like atherosclerosis, stroke, diabetes, Alzheimer's disease (AD), Parkinson's disease, cancer, etc. appeared over the past three decades. There are a number of epidemiological studies that have shown inverse correlation between the levels of established antioxidants/phytonutrients present in tissue/blood samples and occurrence of cardiovascular disease, cancer or mortality due to these diseases.

#### 233 4. FREE RADICALS AND HUMAN DISEASES

234 Free radicals have different types of reaction mechanisms. They can react with surrounding molecules by 235 (a) electron donation, reducing radicals, and electron acceptance, oxidizing radicals, (b) hydrogen 236 abstraction, (c) addition reactions, (d) self-annihilation reactions, and (e) by disproportionation [63]. These 237 reactions lead to the production of ROS, RNS and other radicals which have been linked to many severe 238 diseases like cancer, cardiovascular diseases including atherosclerosis and stroke, neurological 239 disorders, renal disorders, liver disorders, hypertension, rheumatoid arthritis, adult respiratory distress 240 syndrome, auto-immune deficiency diseases, inflammation, degenerative disorders associated with 241 aging, diabetes mellitus, diabetic complications, cataracts, obesity, autism, alzheimer's, parkinson's and 242 huntington's diseases, vasculitis, glomerulonephritis, lupus erythematous, gastric ulcers, 243 hemochromatosis and preeclampsia, among others [64, 65]. Effects of free radicals on disease 244 occurrence are shown below (Fig. 3).

#### 245 4.1 Cancer

DNA is a major target of free radical damage. The types of damages induced include strand breaks (single or double strand breaks), various forms of base damage yielding products such as 8hydroxyguanosine, thymine glycol or abasic sites, damage to deoxyribose sugar as well as DNA protein cross links. These damages can result in mutations that are heritable change in the DNA that can yield cancer in somatic cells or foetal malformations in the germ cells.

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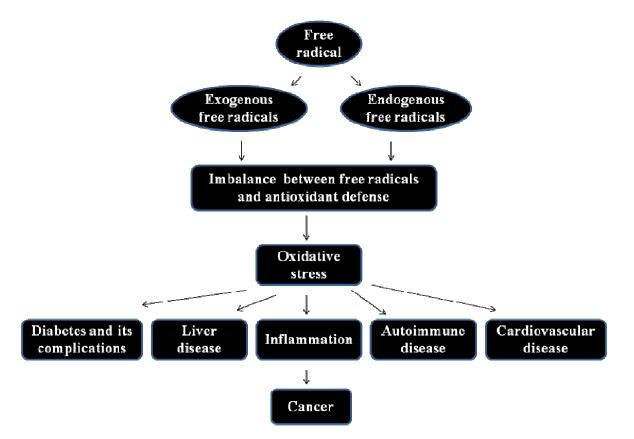
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 Disease
 Cancer

Crohn's disease Ulcerative colitis	Colon [67, 68]	
Barrett's oesophagus	Oesophageal [69]	
Pancreatitis	Pancreatic [70]	
Prostatitis	Prostate [71]	
Human papilloma virus infection	Cervix [72]	
Viral hepatitis B and C Haemochromatosis	Liver [73, 74]	

253 254 The involvement of free radicals with tumor suppressor genes and proto-oncogenes suggest their role in 255 the development of different human cancers [66]. Cancer develops through an accumulation of genetic 256 changes. Initiating agents can be tobacco smoking and chewing, UV rays of sunlight, radiation, viruses, 257 chemical pollutants, etc. Promoting agents include hormones (androgens for prostate cancer, estrogens 258 for breast cancer and ovarian cancer). Inflammation induces iNOS (inducible nitric oxide synthase) as 259 well as COX and LOX. These can initiate carcinogenesis. Table 2 summarizes examples of radical over 260 load diseases. These develop from condition of chronic inflammation and can have an etiology that is 261 primarily inherited or acquired through viral, bacterial and parasitic infection, or acquired through chemical 262 induction. Cancer proneness is frequently a pathological consequence of extensive and sustained free 263 radical stress related damage in these diseases.



#### 266 Fig. 3. Consequences of free radical load 267 268 Experimental as well as epidemiological data indicate that a variety of nutritional factors can act as 269 antioxidants and inhibit the process of cancer development and reduce cancer risk. Some of these 270 include vitamins A, C, E, beta-catotene, and micronutrients [75]. Chemopreventive phytochemicals can 271 block initiation or reverse the promotion stage of multistep carcinogenesis. They can also halt or retard 272 the progression of precancerous cells into the malignant ones. Many molecular alterations associated 273 with carcinogenesis occur in cell-signalling pathways that regulate cell proliferation and differentiation. 274 One of the central components of the intracellular signaling network that maintains homeostasis is the 275 family of mitogen activated protein kinases (MAPKs), they are prime targets of diverse classes of 276 chemopreventive phytochemicals [76]. A number of plants (Table 3) have been found to inhibit cancer 277 progression.

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#### **Table 3:** Phytoconstituents and anti cancer activity

Plant	Family	Compound	Mode of action
Catharanthus roseus	Apocynaceae	Vindesine and Vinorelbine	mitotic block [77]
Catharanthus roseus	Apocynaceae	Vinflunine	mitotic block [78]
Podophyllum peltatum	Berberidaceae	Etoposide	mitotic block [79]
Camptotheca acuminate	Nyssaceae	Topotecan	DNA topoisomerase I inhibition [80]
Berberis amarensis	Berberidaceae	Berbamine	Caspase-3- dependent apoptosis [81]
Hvdrastis canadensis	Ranunculaceae	Berberine	Inhibit bcr/abl gene fusion [82]
Tabebuia avellanedae	Bignoniaceae	Betalapachon e	Inhibition of topoisomerase I and II [83]
Betula alba	Betulaceae	Betulinic acid	Triggers mitochondrial pathway of apoptosis [84]
Colchicum autumnale	Colchicaceae	Colchicine	Anti-mitotic [85]
Curcuma longa	Zingiberaceae	Curcumin	Exact mechanism of action is still unknown [86]
Wikstroemia indica	Thymelaeaceae	Daphnoretin	suppression of protein and DNA synthesis [87]
Psoralea corylifolia	Fabaceae	Psoralidin	enhanced TRAIL-induced (Tumor necrosis factor-related apoptosis- inducing ligand) apoptosis [88]
Vicia faba	Fabaceae	Diadzein and Genistein	Inhibits 3A 4- mediated metabolism and oxidative metabolism [89]
Ochrosia borbonica	Apocynaceae	Ellipticine	DNA intercalation and inhibition of topoisomerase II [90]

Amoora rohituka	Meliaceae	Flavopiridol	Inhibits cell cycle progression at G1 or G2 phase [91]
Cephalotaxus harrintonia	Cephalotaxaceae	Harringtonine	Inhibition of protein synthesis and chain elongation during translation [92]
lpomoeca batatas	Convolvulaceae	4-Ipomeanol	cytochrome P-450 mediated conversion into DNA-binding metabolites [93]
Iridaceaelatea pallasii	Iridaceae	Irisquinone	Acts as a chemosensitizer[94]
Erythroxylum pervillei	Erythroxylaceae	Pervilleines	Inhibitors of Pglycoprotein [95]
Salvia prionitis	Lamiaceae	Salvicine	Inhibition of topoisomerase II [96]
Aglaia foveolata	Meliaceae	Silvestrol	apoptosome/ mitochondrial pathway is involved in triggering extrinsic pathway of programmed cell death of tumor cells [97]

## 282 4.2 Cardiovascular disease

283 Several established risk factors for cardiovascular disease have been linked to excessive generation of 284 ROS. For instance, in animal models of hiperlipidemia, hypertension, and diabetes, the elevated levels of 285 vascular superoxide anion production have been found [98, 99]. The studies strongly suggest that 286 increased oxidative stress is involved in the pathophysiology of cardiovascular disease. Several 287 mechanisms have been proposed to explain how excessive production of ROS leads to vascular 288 pathology. First, ROS are able to promote the oxidation of low-density lipoprotein (LDL) [100]. Uptake of 289 oxidatively modified lipoproteins by macrophages transforms these cells into foam cells, which are a key 290 component of atherosclerotic plaques [101]. Second, superoxide anion rapidly inactivates endothelium 291 derived nitric oxide (NO), a molecule with intrinsic antiatherogenic properties, leading to endothelial dysfunction, which is a hallmark of early atherosclerosis [102]. Moreover, the reaction between 292 293 superoxide anion and NO generates peroxynitrite (ONOO-), which has been found to be cytotoxic to 294 endothelial and vascular smooth muscle cells through a broad range of biological actions, such as lipid 295 oxidation and mitochondrial DNA damage. Third, ROS have been shown to be involved in increased 296 expression of certain vascular pro-inflammatory genes that are pertinent to atherogenesis, such as 297 monocyte chemoattractant protein-1 (MCP-1), vascular cell adhesion molecule-1 (VCAM-1), and 298 intercellular adhesion molecule-1 (ICAM-1) [103, 104].

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Phytochemicals prevent endothelial dysfunction and reduce blood pressure, oxidative stress, and end organ damage in hypertensive animals. Moreover, some clinical studies have shown that phytochemicals can improve endothelial function in patients with hypertension and ischemic heart disease [105]. The effects of individual plant products on the relaxation of isolated arteries from rats have been investigated in many studies. Tetracyclic triterpene saponins, the ginsenosides are often attributed to the effects of *Panax ginseng* (Araliaceae) on the cardiovascular system. Studies show that phytosterols also have 306 effect on the cardiovascular system by lowering cholesterol levels [106].

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#### 308 4.3 Diabetes

Diabetes mellitus is a metabolic disorder characterized by hyperglycemia and insufficiency of secretion or action of endogenous insulin. Although the etiology of this disease is not well defined, viral infection, autoimmune disease, and environmental factors have been implicated [107]. Increased oxidative stress is a widely accepted participant in the development and progression of diabetes and its complications [108]. People suffering from diabetes are not able to produce or properly use insulin in the body and therefore chronic hyperglycemia occurs. Hyperglycemia is also found to promote lipid peroxidation of low density lipoprotein (LDL) by a superoxide-dependent pathway resulting in the generation of free radicals [109].

316 Auto-oxidation of glucose involves spontaneous reduction of molecular oxygen to superoxide and 317 hydroxyl radicals, which are highly reactive and interact with all biomolecules. They also accelerate 318 formation of advanced glycation end products (AGEs). AGEs such as pyrroles and imidazoles tend to 319 accumulate in the tissue. Crosslinking AGE-protein with other macromolecules in tissues results in 320 abnormalities in the cell and tissue function. Due to protein glycation capacity of antioxidant enzymes is 321 also reduced. Free radicals generated also react with nitric oxide in endothelial cells leading to loss of 322 vasodilation activity. Long lived structural proteins, collagen and elastin, undergo continual non-enzymatic 323 crosslinking during ageing and in diabetic individuals [110]. This abnormal protein crosslinking is 324 mediated by AGEs generated by nonenzymatic glycosylation of proteins by glucose.

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326 Up to now, many kinds of antidiabetic medicines have been developed for the patients and most of them 327 are chemical or biochemical agents aiming at controlling or/and lowering blood glucose to a normal level. 328 Despite the impressive advances in health sciences and medical care, there are many patients who are 329 using alternative therapies alone or complementary to the prescribed medication. Traditional plant 330 remedies or herbal formulations exist from ancient times and are still widely used, despite all the 331 controversy concerning their efficacy and safety to treat hypoglycemic and hyperglycemic conditions all 332 over the world. To date, metformin (a biguanide) is the only drug approved for treatment of type II 333 diabetes mellitus [111]. It is a derivative of an active natural product, galegine, isolated from the plant 334 Galega officinalis L. [112]. Table 4 summarizes the herbs with active components having anti diabetic 335 property.

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Table 4: Anti diabetic activity of plant products

Plant	Family	Active compounds	Mode of action
Abelmoschus moschatus	Malvaceae	Myricetin	enhances glucose utilization to lower plasma glucose with deficient insulin levels. [113]
Achyrocline satureioides	Asteraceae	Dibenzofuran Achyrofuran	lowers blood glucose levels[114]
Psacalium	Asteraceae	Maturine	lowers blood glucose levels [115]

decompositum			
Acourtia thurberi	Asteraceae	benzoquinone perezone	lowers blood glucose levels [116]
Allium sativum	Liliaceae	Allicin	decreases the concentration of serum lipids, blood glucose and activities of serum enzymes [117]
Allium cepa	Liliaceae	S-methyl cysteine sulfoxide	stimulation of insulin secretions and partly due to its antioxidant activity [118]
Bauhinia forficata	Leguminosae	Kaempferitrin	decreases lipid peroxidation in liver cells [119]
Bryonia alba	Curcubiaceae	Trihydroxy octadecadienoic acid	restores the disordered lipid metabolism [120]
Caesalpinia ferrea	Leguminosae	Ellagic acid	ALR2 inhibitor [121]
Dioscorea dumetorum	Dioscoreaceae	Dioscoretine	Lowers glucose level [122]
Eucalyptus macrocarpa	Myrtaceae	Macrocarpals (A, B, C and D)	inhibitory activity against porcine lenses ALR2 [123]
Ficus bengalensis	Moraceae	Leucopelargonidin	serum insulin raising [124]
Galega officinalis	Leguminosae	Guanidine	blood glucose-lowering activity[125]
Gentiana olivieri	Gentianaceae	Isoorientin	Antihyperlipidemic [126]
Hydnocarpus wightiana	Arcariaceae	Hydnocarpin	alpha-glucosidase and moderate N-acetyl-beta- D- glucosaminidase inhibitory activities [127]

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## 340 **4.4 Oxidative stress and metabolic changes in the liver**

341 Hepatocyte plays a central role in the metabolism of alcohol or drugs which may enhance the ROS 342 production [128]. Under some consequences a large amount of free fatty acids (FFAs) from the 343 visceral fat tissue, as well as from dietary glucose and fat, flows directly into the liver [129]. Due to 344 these mitochondria, peroxisomes, and endoplasmic reticulum metabolize the excessive amount of fatty 345 acid, resulting in overproduction of ROS and oxidative stress in the hepatocytes. Excessively high 346 levels of iron are stored in the hepatocytes of patients with fatty liver, alcoholic hepatitis, or hepatitis 347 type C. Such over accumulation of iron also causes oxidative stress in the hepatocytes [8]. The reason 348 hepatocytes have the highest antioxidant function as compared with the cells of other organs is 349 probably that oxidative stress is easily induced in the hepatocytes.

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Herbal medicines have been used in the treatment of liver diseases for a long time. A number of herbal preparations are available in the market. Some commonly used herbal preparations are *Phyllanthus*, *Silybum marianum* (milk thistle), glycyrrhizin (licorice root extract), and Liv52 (mixture of herbs). *Phyllanthus* appears to be promising in patients with chronic hepatitis B virus (HBV) infection [130]. Liu *et al.* (2001) [131] published a meta-analysis of the effect on and safety of genus *Phyllanthus* for chronic HBV infection. None of the trials reported mortality or incidence of liver cirrhosis and/or hepatocellular carcinoma. *Phyllanthus* has a positive effect on clearance of HBV markers. There are no major adverse effects. Though the active compound remains to be identified, significant progress has already taken place in standardization of the extract to ensure the bioefficacy of *P. amarus* [132].

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361 Silybum marianum is the most well researched plant in the treatment of liver disease. In Roman times, 362 Pliny the El-der (A.D. 77), a noted naturalist, reported that milk thistle was excellent for carrying off bile. 363 Culpeper (1650) described its effectiveness in removing obstruction of the liver and spleen [133]. The 364 active complex in mile thistle is a lipophilic extract from the seeds of the plant and is composed of three 365 isomer flavonolignans-silybin, silydianin, and silychrstine collectively known as silymarin [134]. Silymarin 366 acts as an antioxidant by reducing free radical production and lipid peroxidation, has antifibrotic activity, 367 and may act as a toxin blockade agent by inhibiting binding of toxins to heptocyte cell membrane 368 receptors [135]. In animals, silymarin reduces liver injury caused by acetaminophen, carbon tetrachloride, 369 radiation, iron overload, phenylhydrazine, alcohol, cold ischemia, and Amanita phalloides [136].

370

371 Glycyrrhizin is an aqueous extract of the licorice root, *Glycyrrhizin glabra*. Its major constituents are 372 glycyrrhetic acid, multiple flavonoids, isoflavonoids, hydroxycoumarins and sterols, including  $\beta$ -sitosteroid, 373 which may have glucocorticoid and mineralocorticoid activities [137]. Glycyrrhizin prevents several forms 374 of experimental liver injury in animals [138]. This compound has anti-inflammatory and antioxidant 375 activities.

376

Liv52 is considered to be an Ayurvedic hepatoprotective medicine that contains the Capparis spinosa 377 378 (Himsara), Cichorium intybus (Kasani), Mandur bhasma, Solanum nigrum (Kakamachi), Terminalia arjuna 379 (Arjuna), Cassia occidentalis (Kasamarda), Achillea millefolium (Biranjasipha), and Tamarix gallica 380 (Jhavaka). Liv52 has been on the market for over 50 years and has been claimed to be useful in the 381 prevention and treatment a variety of conditions such as viral hepatitis, alcoholic liver disease, protein 382 energy malnutrition, loss of appetite, and radiation and chemotherapy induced liver damage [139]. 383 Experimental data suggest that Liv52 inhibits lipid peroxidation, may have a protective effect on alcohol 384 induced fetotoxicity, and inhibits TNF activity. Liv52 has been claimed to be useful as an adjuvant to 385 hepatotoxic drugs [140-142].

386

### 387 4.5 Free radical and aging

The aging process has been shown to result in an accelerated functional decline. The exact mechanisms that cause this functional decline are unclear. The free radical theory of aging, however, has gained strong support because it is able to explain some of the processes that occur with aging and the degenerative diseases of aging. This theory proposes that an increase in oxygen radical production with age by mitochondria produce an increase in cellular damage [143-145]. Aerobic organisms are wellprotected against oxidative challenges by sophisticated antioxidant defense systems. However, it appears that during the aging process an imbalance between oxidants and antioxidants balance may occur. Oxidative damage of biomolecules increases with age and is postulated to be a major causal factor of cellular biochemical senescence [146-148]. Resveratrol, a phytoalexin, is synthesized in the leaf epidermis and the skin (pericarp) of grape berries and has potential antioxidant and anti-aging property [149]. Some plants and their parts having anti aging activity are given in Table 5.

- 399
- 400

 Table 5: Some of the plants and their part used for anti aging activity

Part used	Plant	Family
Leaves	Adansonia digitata	Bombacaceae [150]
	Alstonia boonei	Apocynaceae [151]
	Bambusa vulgaris	Poaceae [152]
	Elaeis guineensis	Palmae [153]
	Ficus capensis	Moraceae [154]
	Harungana madagascariensis	Harungaceae [80]
	Spondias mombin	Anacardiaceae [155]
	Tectona grandis	Verbanaceae [156]
	Zea mays	Poaceae [157]
Seed	Aframomum melegueta	Zingiberaceae [158]
	Garcinia kola	Gutiferae [159]
Whole plant	Baphia nitida	Papilinionaceae [160]
	Lophira alata	Ochnaceae [161]
Root	Montandra guineensis	Apocynaceae [162]
	Cocos nucifera	Palmae [163]
Stem bark	Cordia millenii	Boraginaceae [164]
	Khaya ivorensis	Meliaceae [165]
Fruits	Milicia excels	Moraceae [166]

401

402 The main function of mitochondria is energy production. During oxidative phosphorylation, however, 403 highly reactive oxygen radicals are generated. One major site of oxidant production occurs in the 404 mitochondrial electron transport chain in which  $O_2$  is reduced to  $H_2O$ . Several studies have investigated 405 age associated increase in the generation of oxidants by mitochondria [167, 168]. Experiments using

406 intact muscle mitochondria from house flies have shown that the rate of H<sub>2</sub>O<sub>2</sub> generation progressively 407 increases 2-fold as the house fly ages [169]. The enhanced generation of oxidants by older 408 mitochondria may itself be caused by oxidative damage to mitochondrial membranes and proteins [170]. 409 Miquel and his colleagues have widely promulgated the mitochondrial mutation theory of aging [170]. In 410 this theory, senescence is linked to mutations of mitochondrial DNA (mtDNA) in differentiated cells. 411 Mitochondrial DNA lacks excision and recombination repair mechanisms, it has been postulated that 412 these mutations would lead to problems in replication, leading to a decline in physiological performance 413 and the pathogenesis of many age-related diseases [169, 170]. In addition, mtDNA is not protected by 414 histones or DNA-binding proteins and, therefore, is directly exposed to a high steady state level of 415 reactive oxygen and nitrogen species. Thus, oxidative modification and mutation of mtDNA may occur 416 with great ease. During the aging process, protein oxidation is increased in a wide variety of human and 417 animal tissues. The exact pathways for oxidative cellular damage are poorly understood because the 418 reactive metabolites are very short-lived and difficult to detect directly in vivo. The quantification of 419 oxidative damage to proteins has been studied almost exclusively by assessing the total carbonyl 420 content [171]. The oxidants responsible for carbonyl formation within the proteins in vivo are believed to 421 be radicals, such as, hydroxyl radicals. Indeed, hydroxyl radicals can be generated by metal-catalyzed 422 oxidation systems, and different metal catalyzed oxidation systems convert several amino acid residues 423 to carbonyl derivatives [169-171].

424

#### 425 5. CONCLUSION AND FUTURE PROSPECTS

426 Free radicals are known to play a definite role in a wide variety of pathological manifestations. 427 Antioxidants fight free radicals and protect us from various diseases. They exert their action either by 428 scavenging the reactive oxygen species or protecting the antioxidant defense mechanisms. They can 429 greatly reduce the damage due to oxidants by neutralizing the free radicals before they can attack the 430 cells and prevent damage to lipids, proteins, enzymes, carbohydrates and DNA. Phytochemicals 431 including polyphenols, flavonoids and others have potential to provide defense against oxidative damage. 432 Newer approaches are further required for identification and characterization the specific 433 phytoconstituents from diverse flora for providing protection against oxidative stress.

434

#### 435 **REFERENCES**

- 436
- 437 1. Gutteridgde J M C. Free radicals in disease processes: A complication of cause and consequence.
   438 Free Rad Res Commun. 1995;19:141-58

439 2. Mishra A, Kumar S, Bhargava A, Sharma B, Pandey A K. Studies on *in vitro* antioxidant and 440 antistaphylococcal activities of some important medicinal plants. Cell Mol Biol. 2011;57:16-25

3. Mishra A, Sharma A K, Kumar S, Saxena A K, Pandey A K. *Bauhinia variegata* leaf extracts exhibit
considerable antibacterial, antioxidant and anticancer activities. BioMed Res Int. 2013; (2013):Article ID
810734

- 444 4. Joyce D A. Oxygen radicals in disease. Aust Adv Drug React Bull. 1987;127:476-79
- 5. Kumar S, Pandey A K. Chemistry and biological activities of flavonoids: an overview. Sci World J 2013;
  2013:Article ID 162750
- 447 6. Kumar S, Sharma U K, Sharma A K, Pandey A K. Protective efficacy of Solanum xanthocarpum root

- 448 extracts against free radical damage: phytochemical analysis and antioxidant effect. **Ce**ll Mol Biol. 449 2012;58:171-78
- 450 7. Kumar S, Pandey A K. Phenolic content, reducing power and membrane protective activities of 451 Solanum xanthocarpum root extracts. Vegetos. 2013;26:301-07
- 452 8. Kumar S, Pandey A K. Antioxidant, lipo-protective and antibacterial activities of phytoconstituents 453 present in *Solanum xanthocarpum* root. Int Rev Biophys Chem. 2012;3:42-47
- 454 9. Kumar S, Chashoo G, Saxena A K, Pandey A K. *Parthenium hysterophorus*: a probable source of 455 anticancer, antioxidant and anti-HIV agents. BioMed Res Int. 2013;2013:Article ID 810734
- 456 10. Pandey A K, Mishra A K, Mishra A. Antifungal and antioxidative potential of oil and extracts derived
   457 from leaves of Indian spice plant *Cinnamomum tamala*. Cell Mol Biol. 2012;58:142-147
- 458 11. Pham-Huy, Pham-Huy H He. Free radicals, antioxidants in disease and health. Int J Biomed Sci.
  459 2008;4:89-96
- 460 12. Mishra A K, Mishra A, Kehri H K, Sharma B, Pandey A K. Inhibitory activity of Indian spice plant
   461 *Cinnamomum zeylanicum* extracts against *Alternaria solani* and *Curvuluria lunata*, the pathogenic
   462 dematiaceous moulds. Ann Clin Microbiol Antimicrob. 2009;8: Article 9
- 463 13. Kuppusamya P, Zweier J L. Characterization of free radical generation by xanthine oxidase. Evidence
   464 for hydroxyl radical generation. J Biol Chem. 1989;264:9880-84
- 465 14. Zimmerman B J, Granger D N. Mechanisms of reperfusion injury. Am J Med Sci. 1994;307:284-92
- 466 15. Kontos H A, Ellis E P, Jenkins E F, Povlishock L W, Rowe J T, Hess M L. Appearance of superoxide
   467 anion radical in cerebral extracellular space during increased prostaglandin synthesis in cats. Circ Res.
   468 1985;57:142–51
- 469 16. Mcintyre M, Bohr D F, Dominiczak A F. Endothelial function in hypertension. Hypertension
  470 1999;34:539–45
- 471 17. Winterbourn C C, Vissers M C, Kettle A J. Myeloperoxidase. Curr Opin Hematol. 2000;7: 53-58
- 472 18. Lipinski B. Hydroxyl radical and its scavengers in health and disease. Oxid Med Cell Longev. 2011;
  473 DOI: 10.1155/2011/8099696.
- 474 19. Mishra A, Kumar S, Pandey A K. Scientific validation of the medicinal efficacy of *Tinospora cordifolia*.
  475 Sci World J 2013;2013:Article ID 292934
- 476 20. Wu G, Morris S M. Arginine metabolism: nitric oxide and beyond. Biochem J. 1998;336:1-17
- 477 21. Fang Y Z, Yang S, Wu G. Free radicals, antioxidants, and nutrition. Nut 2002;18:872–79
- 478 22. Rassaf T, Preik M, Kleinbongard P. Evidence for in vivo transport of bioactive nitric oxide in human
  479 plasma. J Clin Inv. 2002;109:1241-48
- 480 23. Thomas E L, Lehrer R I, Rest R F. Human neutrophil antimicrobial activity. Clin Infect Diseases.
  481 1998;10:S450–S456
- 482 24. Babior B. NADPH oxidase An update. Blood. 1999;93:1464-76
- 483 25. Nauseef W F. The NADPH dependent oxidase of phagocytes. Proc. Assoc. Am. Physicians.
   484 1999;111:373–382
- 485 26. Rossi F, Bellavite P, Berton G, Grzeskowiak M, Papini E. Mechanism of production of toxic oxygen
  486 radicals by granulocytes and macrophages and their function in the inflammatory. Path Res Prac.
  487 1985;180:136–42
- 488 27. Rosen H J, Orman R M, Rakita B R, Michel D, VanDevanter R. Loss of DNA-membrane interactions
   489 and cessation of DNA synthesis in myeloperoxidase-treated *Escherichia coli*. Prod Nat Acad Sci USA.
   490 1990;87:10048–10052
- 491 28. Dalton T P, Shertzer H G, Puga A. Regulation of gene expression by reactive oxygen. Ann Rev 492 Pharmacol Toxicol. 1999:39:67–101
- 493 29. Babior B M. Superoxide: a two-edged sword. Braz J Med Biol Res. 1997;30:141–55
- 494 30. Arrigo A P. Gene expression and the thiol redox state. Free Rad Biol Med. 1999;27:936-44

- 31. Timblin C R, Janssen Y M, Goldberg J L, Mossman B T. GRP78, HSP72/73, and cJun stress protein
  levels in lung epithelial cells exposed to asbestos, cadmium, or H<sub>2</sub>O<sub>2</sub>. Free Rad Biol Med. 1998;24: 632–
  42
- 32. Nose K M, Shibanuma, Kikuchi K, Kageyama H, Sakiyama S, Kuroki T. Transcriptional activation of
  early response genes by hydrogen peroxide in a mouse osteoblastic cell line. Eur J Biochem.
  1991;201:99–106
- 501 33. Valko M, Izakovic M, Mazur M, Rhodes C J, Telser J. Role of oxygen radicals in DNA damage and cancer incidence. Mol Cell Biochem. 2004;266:37–56
- 503 34. Kumar S, Gupta A, Pandey A K. Calotropis procera root extract has the capability to combat free 504 radical mediated damage. ISRN Pharmacol 2013;2013:Article ID 691372
- So5 35. Kumar S, Mishra A, Pandey A K. Antioxidant mediated protective effect of *Parthenium hysterophorus* against oxidative damage using *in vitro* models. BMC Comp Alt Med. 2013; 2013:Article 120
- 507 36. Lu J, Lin P H, Yao Q, Chen C. Chemical and molecular mechanisms of antioxidants: experimental
   508 approaches and model systems. J Cell Mol Med. 2010;14:840–60
- 509 37. Craft B D, Kerrihard A L, Amarowicz R, Pegg R B. Phenol-based antioxidants and the in vitro 510 methods used for their assessment. Compreh Rev Food Sci Food Safety. 2012;11:148–73
- 511 38. Devasagayam T P A, Boloor K K, Ramsarma T. Methods for estimating lipid peroxidation: Analysis of 512 merits and demerits (mini review). Ind J Biochem Biophys. 2003;4:0300-08
- 513 39. Dizdaroglu M, Jaruga P, Birincioglu M, Rodriguez H. Free radical-induced damage to DNA: 514 mechanisms and measurement. Free Rad Biol Med. 2002;32:1102-15
- 40. Benov L, Beema A F. Superoxide -dependence of the short chain sugars induced mutagenesis. Free
   Rad Biol Med. 2003;34:429-33
- 41. Halliwell B, Chirico S. Lipid peroxidation: its mechanism, measurement, and significance. Am J Clin
   Nut. 1993;57:715-24
- 42. Pandey A K, Mishra A K, Mishra A, Kumar S, Chandra A. Therapeutic potential o f *C. zeylanicum*extracts: an antifungal and antioxidant perspective. Int J Biol Med Res. 2010;1:228-233
- 43. Niki E. Antioxidant defenses in eukaryotic cells, In Free radicals: From basic science to medicine (Poli
  G, Albano E, Dianzan M U, eds), 1993; pp 365–373, Basel, Switzerland: Birkhauser Verlag
- 44. Yost F J J, Fridovich I. An iron-containing superoxide dismutase from *Escherichia coli*. J Biol Chem.
   1993;248:4905–08
- 43. Fridovich I. J Biol Chem. Superoxide anion radical (O<sub>2</sub><sup>---</sup>), superoxide dismutases, and related matters. 1997;272:18515-17
- 44. Das K C, Lewis-Molock Y, White C W. Am J Res Cell Mol Biol. Elevation of manganese superoxide dismutase gene expression by thioredoxin. 1997;17:713 –26
- 529 45. Ohlemiller K K, McFadden S L, Ding D L, Flood D G, Reaume A J, Hffman E K, Scott R W, Wright J
- 530 S, Putcha G V, Salvi R J. Targeted deletion of the cytosolic Cu/Zn -superoxide dismutase gene (Sod1) 531 increases susceptibility to noise-induced hearing loss. Aud Neurotol. 1999;4:237–46
- 46. Aebi H. In Methods of Enzymatic Analysis (Bergmayer H U, ed) 2<sup>nd</sup> edn, 1974;pp. 673-77, Academic
   Press New York, USA
- 47. Kirkman H N, Gaetani G F. Catalase: a tetrameric enzyme with four tightly bound molecules of NADPH. Prod Natal Acad Sci, USA. 1984;81:4343–47
- 48. Chae H Z, Kim H J, Kang S W, Rhee S G. Characterization of three isoforms of mammalian peroxiredoxin that reduce peroxides in the presence of thioredoxin. Diab Res Clin Prac. 1999;45:101–12
- 49. Poole L B, Godzik A, Nayeem A, Schmitt J D. AhpF can be dissected into two functional units:
   tandem repeats of two thioredoxin-like folds in the N-terminus mediate electron transfer from the
   thioredoxin reductase -like C-terminus to ahpC. Biochem. 2000;39:6602-6615
- 541 50. Miranda-Vizute A D A, Spyrou G. The mitochondrial thioredoxin system. Ant Redox Signaling. 542 2000;4:801–81
- 543 51. Zhou Y K, Kok H, Chun A C, Wong C M, Wu H W, Lin M C, Fung P C, Kung H, Jin D Y. Mouse

- 544 peroxiredoxin V is a thioredoxin peroxidase that inhibits p53-induced apoptosis. Biochem Biophys Res 545 Commun. 2000;268:921–27
- 546 52. Ursini F, Maiorino M, Brigelius-Flohe R, Aumann K D, Roveri A, Schomburg D, Flohe F. Diversity of 547 glutathione peroxidases. Methods Enzymol. 1995;252:38–53
- 548 53. Ursini F, Heim S, Kiess M, Maiorino M, Roveri A, Wissing J, Flohe L. Dual function of the 549 selenoprotein PHGPx during sperm maturation. Science. 1999;285:1393–1396
- 550 54. Dreher I, Schmutzler C, Jakob F, Kohrle J. Expression of selenoproteins in various rat and human
   tissues and cell lines. J Trace Elem Med Bio. 1997;11:83–91
- 55. Jones D P, Eklow L, Thor H, Orrenius S. Metabolism of hydrogen peroxide in isolated hepatocytes:
- relative contributions of catalase and glutathione peroxidase in decomposition of endogenously
- 554 generated  $H_2O_2$ . Arch Biochem Biophys. 1981;210:505–16
- 555 56. Lee J, Koo N. In, Reactive oxygen species, aging and antioxidative nutraceuticals. Comphr Rev Food 556 Sci Food Safety. 2004;3:21-33
- 557 57. Fischer-Nielsen A, Poulsen H E, Loft S. 8-Hydroxydeoxyguanosine in vitro: effects of glutathione, 558 ascorbate, and 5-aminosalicylic acid. Free Rad Biol Med. 1992;13:121-26
- 559 58. Packer L, Ong A S H. Eds. Mechanisms and Health Effects. AOCS Biological Oxidants and 560 Antioxidants, Molecular Press, Champaign. 1998
- 561 59. Kagan V E, Kisin E R, Kawai K, Serinkan B F, Osipov B N, Sertbinova E A, Wolinsky I, Shvedova A
   562 A. Towards mechanism based antioxidant interventions. Ann N Y Acad Sci. 2002;959:188-98
- 60. Aruoma O I. Free radicals, oxidative stress, and antioxidants in human health and disease. J Am
   Chem Soc.1998;75:199-212
- 565 61. CuzzocreaS, Riley D P, Caputi A P, Salvemini D. Antioxidant therapy: A new pharmacological approach in shock, inflammation and ischemia/reperfusion injury. Pharmacol Rev. 2001;53:135-59
- 567 62. Wichi H P. Enhanced tumour development by butylated hydroxyanisole (B HA) from the prospective 568 of effect on fore stomach and oesophageal squamous epithelium. Food Chem Toxicol. 1998;26:717-23
- 569 63. Slater T F. Free-radical mechanis ms in tissue injury. J Biochem. 1984;222:1-15
- 570 64. Rahman K. Studies on free radicals, antioxidants and cofactors. Clin Interven Aging. 2007;2:219-36
- 571 65. Singh A K, Gupta A, Mishra A K, Gupta V, Bansal P, Kumar S. Medicinal Plant for curing 572 Alzheimer's disease. Int J Pharmaceut Biol Arch. 2010;1:108-14
- 573 66. B, Halliwell, O. I. Aruoma. Eds. "DNA and Free Radicals", Boca Raton Press, 1993
- 574 67. Gillen C D, Prior P, Andrews H A, Allan R N. Ulcerative colitis and Crohn's disease: a comparison of 575 the colorectal cancer risk in extensive colitis. J Gasteroenterol. 1994;35:1590-92
- 576 68. Ekbom A, Helmick C, Zack M, Adami H O. Ulcerative colitis and colorectal cancer: a population 577 based study. New Eng J Med. 1990;323:1228-33
- 578 69. Streitz J M. Barrett's esophagus and esophageal cancer. Chest Sur Clin North Am. 1994;4:227-40
- 579 70. Bansal P, Sonnenberg A. Pancreatitis is a risk factor for pancreatic cancer. Gastroenterol. 580 1995;109:247-51
- 581 71. Dennis L K, Lynch C F, Torner J C. Epidemiologic association between prostatitis and prostate cancer. Journal of Urology. 2002;60:78–83
- 583 72. Mitchell H, Drake M, Medley G. Prospective evaluation of risk of cervical cancer after cytological
   584 evidence of human papilloma virus infection. Lancet. 1986;15:573-575
- 73. Hai-rim S, Chae-un L, Hyung-jong P, Sang-young S, Jungmyeong C, Ha-chin C, Yoonok A, Takao S.
   Hepatitis B and C Virus, *Clonorchis sinensis* for the Risk of Liver Cancer: A Case-Control Study in Pusan,
   Korea. Int J Epidemiol. 1996;25:933-40
- 74. Niederau C, Fischer R, Sonnenberg A, Stremmel W, Trampisch H A, Strohmeyer G. Survival and
   causes of death in cirrhotic and in noncirrhotic patients with primary hemochromatosis. New Eng J
   Med. 1985;313:1256-62
- 591 75. Croce C M. How can we prevent cancer? Prod Natl Acad Scie USA. 2001;98:10986-88

- 592 76. Surh Y J. Cancer chemoprevention with dietary phytochemicals. Nature Rev Cancer. 2003;3:768-80
- 593 77. Cragg G M, Newman D J. Plants as a source of anti-cancer agents. J Ethnopharmacol. 2005;100:72 594 79
- 595 78. Okouneva T, Hill B T, Wilson L, Jordan M A. The effects of vinflunine, vinorelbine, and vinblastine on centromere dynamics. Mol Cancer Ther. 2003;2:427-436
- 597 79. Simoens C, Lardon F, Pauwels B, De Pooter C M J, Lambrechts H A J, Pattyn G G O, Breillout F,
  598 Vermorken J B. Comparative study of the radiosensitising and cell cycle effects of vinflunine and
  599 vinorelbine, in-vitro. BMC Cancer. 2008;8:65
- 80. Shoeb M, MacManus S M, Jaspars M, Trevidadu J, Nahar L, Thoo-Lin P K, Sarker S D. Montamine,
   a unique dimeric indole alkaloid, from the seeds of *Centaurea montana* (Asteraceae), and its in-vitro
   cytotoxic activity against the CaCo2 colon cancer cells. Tetrahedron. 2006;62:11172-11177
- 81. Creemers G J, Bolis G, Gore M, Scarfone G, Lacave A J, Guastalla J P, Despax R, Favalli G,
  Kreinberg G, Van Belle S. Topotecan, an active drug in the second -line t reatment of ep ithelial ovarian
  cancer: results of a largeEuropean phase II study. J Clin Oncol. 1996;14:3056-61
- 606 82. Xu R, Dong Q, Yu Y, Zhao X, Gan X, Wu D, Lu D, Xu X, Yu X F. Berbamine: a novel inhibitor of 607 bcr/abl fusion gene with potent anti-leukemia activity. Leukemia Res. 2006; 30:17-23
- 608 83. Lei H, Wang B, Li W P, Yang Y, Zhou A W, Chen M Z. Aging effect of astragalosides and its 609 mechanism of action. Acta Pharmacol Sinica. 2003;24:230-34
- 84. Li Y, Li C J, Yu D, Pardee A B. Potent induction of apoptosis by β-lapachone in human multiple
   myeloma cell lines and patient cells. Mol Med. 2000;6:1008-15
- 612 85. Fulda S. Betulinic acid for cancer treat ment and prevention. Int J Mol Sci. 2000;9:1096-107
- 613 86. Dubey K K, Ray A R, Behera B K. Production of demethylated colchicines through microbial 614 transformation and scale-up process development. Process Biochem. 2008;43:251-57
- 87. Sa G, Das T, Banerjee S, Chakraborty J. Curcumin: from exotic spice to modern anticancer drug. Al
   Ameen J Med Sci. 2010;3:21-37
- 617 88. Bronikowska J, Szliszka E, Jaworska D, Czuba Z P, Krol W. The Coumarin Psoralidin Enhances 618 Anticancer Effect of Tumor Necrosis Factor-Related Apoptosis-Inducing Ligand 619 (TRAIL). Molecules. 2012; 17:6449-6464
- 89. Kaufman P B, Duke J A, Brielmann H, Boik J, Hoyt J E. A comparative survey of leguminous plants
  as sources of the isoflavones, genistein and daidzein: implications for human nutrition and health. J Alt
  Comp Med. 1997;3:7-12
- 90. Moon Y J, Wang X, Morris M E. Dietary flavonoids: effects on xenobiotic and carcinogen metabolism.
   Toxicol in Vitro. 2006;20:187-210
- 625 91. Kuo Y C, Kuo P L, Hsu Y L, Cho C Y, Lin C C. Ellipticine induces apoptosis through p53-dependent 626 pathway in human hepatocellular carcinoma Hep G2 cells. Life Sci. 2006;78:2550-57
- 92. Mans D R A, Da Rocha A B, Schwartsmann G. Anti-cancer drug discovery and development in Brazil:
   targeted plant collection as a rational strategy to acquire candidate anti-cancer compounds. Oncol.
   2000;5:185-98
- 630 93. Ancuceanu R V, Istudor V. Pharmacologically active natural compounds for lung cancer. Alt Med Rev.
   631 2004;9:402-19
- 632 94. Hazra B, Sarma M D, Sanyal U. Separation methods of quinonoid constituents of plants used in
   633 oriental traditional medicines. J Chromatography B. 20042;812:259-75
- 634 95. Mi Q, Cui B, Silva G L, Lantvit D, Lim E, Chai H, You M, Hollingshead M J, Mayo J G, Kinghorn A D.
  635 Pervilleine A, a novel tropane alkaloid that reverses the multidrug resistanc Phenotype. Cancer Res.
  636 2001;61:4030-4037
- 637 96. Deng F, Lu J J, Liu H Y, Lin L P, Ding J, Zhang J. Synthesis and antitumor activity of novel salvicine
   638 analogues. Chinese Chem Lett. 2011;22:25-28
- 639 97. Kinghorn D, De Blanco E J C, Chai H B, Orjala J, Farnsworth J R, Soejarto D D, Oberlies N H, Wani
  640 N C, Kroll D J, Pearce C J. Discovery of anticancer agents of diverse natural Origin. Pure Applied Chem.

- 641 2009;81:1051-63
- 98. Miller F J, Gutterman D D, Rios C D, Heistad D D, Davidson B L. Superoxide production in vascular
  smooth muscle contributes to oxidative stress and impaired relaxation in atherosclerosis. Cir Res.
  1998;82:1298–1305
- 645 99. Morawietz H, Weber M, Rueckschloss U, Lauer N, Hacker A, Kojda G. Up regulation of vascular
- NAD(P)H oxidase subunit gp91phox and impairment of the nitric oxide signal transduction pathway in
   hypertension. Biochem Biophys Res Commun. 2001;85:1130–1135
- 100. Hiramatsu H, Rosen H, Heinecke J W, Wolfbauer G, Chait A. Superoxide initiates oxidation of low
   density lipoprotein by human monocytes. Arteriosclerosis. 1987;7: 55-60
- 101. Berliner J A, Heinecke J W. The role of oxidized lipoproteins in atherogenesis. Free Rad Biol Med.
   1996;20:707–27
- 102. Darley-Usmar V, Wiseman H, Halliwell B. Nitric oxide and oxygen radicals: A question of balance.
   FEBS Lett. 1995;369:131–35
- 103. Chen Y N, Chang Y J, Jiang M J. Monocyte chemotactic protein-1 gene and protein expression in atherogenesis of hypercholesterolemic rabbits. Atherosclerosis. 1999;143:115–23
- 104. Boring L, Gosling J, Cleary M, Charo I F. Decreased lesion formation in CCR2 -/-mice reveals a role
   for chemokines in the initiation of atherosclerosis. Nature. 1998;394:894–97
- 105. Perez-Vizcaino F, Ibarra M, Cogolludo A L, Duarte J, Zaragoza-Arnaez F, Moreno L, Lo´pez-Lo´pez
   G, Tamargo J. Endothelium-independent vasodilator effects of the flavonoid quercetin and its methylated
   metabolites in rat conductance and resistance arteries. J Pharmacol Exp The. 2006;302:66–72
- 106. Klingberg S, Ellegård G, Johansson I, Hallmans G, Weinehall L, Andersson H, Winkvist A. Inverse
   relation between dietary intake of naturally occuring plant sterols and serum cholesterol in northern
   Sweden. Am J Clin Nut. 2008;87:993-1001
- 107. Paik S G, Blue M L, Fleischer N, Shin S. Diabetes susceptibility of BA LB/cBOM mice treated with
   streptozotocin. Inhibition by lethal irradiation and restoration by splenicly mphocytes. Diabetes.
   1982;31:808–15
- 108. Baynes J W, Thorpe S R. Role of oxidative stress in diabetic complications: A new perspective on
   an old Paradigm. Diabetes. 1999;48:1-9
- 109. Kawamura M, Heinecke J W, Chait A. Pathophysiological concentrations of glucose promote
  oxidative modification of low density lipoprotein by a superoxide dependent pathway. J Clin Interven.
  1984;94:771–78
- 110. Vasan S, Foiles P, Founds H. Therapeutic potential of breakers of advanced glycation end products
   -protein crosslinks. Arch Biochem Biophys. 2009;419:89-96
- 111. Huxtable R J. The harmfull potential of herbal and other plant products. Drug Safety. 1990;5:126-36
- 675 112. Witters L. The blooming of the French lilac. J Clin Interven. 2001;108:1105-07
- Liu I M, Liou S S, Lan T W, Hsu F L, Cheng J T. Myricetin as the active principle of Abelmoschus
   moschatus to lower plasma glucose in STZ-diabetic rats. Planta Medica. 2005;71:617-21
- 114. Caney J R, Krenisky J M, Williamson R T, Luo J. Achyrofuran, a new antihyperglycemicd
  ibenzofuran from the South A merican medicinal plant *Achyrocline satureioides*. J Nat Prod.
  2005;65:203-05
- 115. Alarcon-Aguilar F J, Roman-Ramos R, Jimenez-Estrada M, Reyes-Chilpa R, Gonzalez-Paredes B,
   Flores-Saenz J L. Effects of three Mexican medicinal plants (Asteraceae) on blood glucose levels in
   healthy mice and rabbits. J Ethnopharmacol. 1997;55:171-77
- 116. Alarcon-Aguilar F J, Hernandez-Galicia E, Campos-Sepulveda A E, Xolalpa-Molina S, Rivas-Vilchis
   G F, Vazquez-Carrillo L I, Roman-Ramos R. Evaluation of the hypoglycemic effect of *Cucurbita ficifolia* Bouche' (Cucurbitaceae) in different experimental models. J Ethnopharmacol. 2002;82:185-89
- 117. Sheela C G, Augusti K T. Antidiabetic effects of S -allylcysteine sulphoxide isolated from garlic
   *Allium sativum* Linn. Ind J Exp Biol. 1992;30:523-26
- 689 118. Kumari K, Augusti K T. Antidiabetic and antioxidant effects of S-methyl cysteine sulfoxide isolated

- from onions (*Allium cepa* Linn) as compared to standard drugs in alloxan diabetic rats. Ind J Exp Biol.
   2002;40:1005-09
- 692 119. De Sousa E, Zanatta L, Seifriz I, Creczynski-Pasa T B, Pizzolatti M T, Szpoganicz B, Silva F R.
  693 Hypoglycemic effect and antioxidant potential of kaempferol-3, 7- O-(alpha)-dirhamnoside from *Bauhinia*694 *forficata* leaves. J Nat Prod. 2004;67:829-32
- Karageuzyan K G, Vartanyan G S, Agadjanov M I, Panossian A G, Hoult J R. Restoration of the
   disordered glucose-fatty acid cycle in alloxan-diabetic rats by trihydroxyoctadecadienoic acids from
   *Bryonia alba*, a native Armenian medicinal plant. Planta Medica. 1998;64:417-22
- 121. Ueda H, Kawanishi K, Moriyasu M. Effects of ellagic acid and 2-(2,3,6- t rihydro xy-4-carboxyphenyl)
   ellagic acid on sorbitol accumulation in vitro and in vivo. Biol Pharm Bull. 2004;27:1584-87
- 122. Iwu M M, Igboko O A, Okunji C O, Tempesta M S. Antidiabetic and ALR2 activities of biflavanones of
   *Garcinia kola.* J Pharm Pharmacol. 1990;42:290-92
- Murata M, Yamakoshi Y, Homma S, Arai K, Nakamura Y. Isolation and characterization of
   macrocarpals B-G antibacterial compounds from *Eucalyptus macrocarpa*. Bio Biotechnol Biochem.
   1992;56:2062-2065
- 124. Cherian S, Augusti K T. Antidiabetic effects of a glycoside of leucopelargonidin isolated from *Ficus bengalensis* Linn. Ind J Exp Biol. 1993;31:26-29
- 125. Vuksan V, Sievenpiper J L, Koo V Y, Francis T, Beljan-Zdravkovic U, Xu Z, Vidgen E. American
   ginseng (*Panax quinquefolius* L) reduces postprandial glycemia in nondiabetic subjects and subjects with
   type II diabetes mellitus. Arch Int Med. 2000;160:1009-13
- 126. Ekrem S, Mustafa A, Erdem Y. Hypoglycaemic activity of *Gentiana olivieri* and isolation of the active
   constituent through bioassay-directed fractionation techniques. Life Sciences. 2005;76:1223-38
- 712 127. Reddy S V, Tiwari A K, Kumar U S, Rao R J, Rao J M. Free radical scavenging, enzyme inhibitory
   713 constituents from antidiabetic Ayurvedic medicinal plant *Hydnocarpus wightiana* Blume. Phytother Res.
   714 2005:19:277-81
- 715 128. Morita Y, Ueno T, Sata M. Acta Hepatol Jap. Comparison in liver histology among NASH, ASH and
   716 chronic hepatitis type C. 2003;44:A526
- 717 129. Ratziu V, Giral P, Charlotte F, Bruckert E, Thibault V, Theodorou I, Khalil L, Turpin G, Opolon,
  718 Poynard T. Liver fibrosis in overweight patients. Gastroenterol. 2000;118:1117–1123
- Thyagarajan S P, Subramanian S, Thirunaksundari , Venkateswaran P S, Blumberg B S. Effects of
   *Phyllanthus amarus* on chronic carriers of hepatitis B virus. Lancet. 1998;2:764–66
- 131. Liu J, Lin H, McIntosh H. Genus Phyllanthus for chronic hepatitis B virus infection: a systematic
   review. J Viral Hep. 2001;8:358–66
- 132. Thyagrajan S P, Jayaram S, Gopalakrishnan V, Han R, Jayakumar P, Sripathi M S. Herbal
   medicines for liver diseases in India. J Gastroenterol Hepatol. 2000;17:S370–376
- 133. Luper S. A review of plants used in the treatment of liver disease: Part 1. Alt Med Revi. 1998;3:410–
  21
- 134. Feher J, Lang I, Nekam K J, Gergely P, Muzes G. In vivo effect of free radical scavenger
  hepatoprotective agents on superoxide dismutase (SOD) activity in patients. Tokai J Exp Clin Med.
  1990;15:129–34
- 135. Faulstich H, Jahn W, Wieland T. Silybin inhibition of amatoxin uptake in the perfused rat liver.
   Arzneimittelforschung. 1980;30:452–54
- 136. Jacobs B P, Dennehy C, Ramirej G, Sapp J, Lawrence V A. Milk thistle for the treatment of liver
   diseases: a systematic review and meta-analysis. Am J Med. 2002;113:506-15
- 137. Seef L B, Lindsay K L, Bacon B R, Kresina T F, Hoofnagle J H. Complementary and alternative
   medicine in chronic liver disease. Hepatol. 2001;34:595-603
- 138. Van Rossum T G, Vulto A G, deMan R A, Browner J T, Schalm S W. Review article: glycyrrhizin as
   a potential treatment for chronic hepatitis C. Alimentary Pharmacol Therap. 1998;12:199–05
- 139. Sandhir R, Gill K D. Hepatoprotective effects of Liv52 on ethanol induced liver damage in rats. Ind J

- 739 Exp Biol. 1999;37:762–66
- 140. Pandey S, Gujrati V R, Shanker K, Singh N, Dhawan K N. Hepatoprotective effect of Liv-52 against
   CCl4 induced lipid peroxidation in liver of rats. Ind J Exp Bio. 1994;32:674-75

141. Kumar S, Kumar R, Diwedi A, Pandey A K. *In vitro* antioxidant, antibacterial, cytotoxic activity and *in vivo* efficacy of *Syngonium podophyllum* and *Eichhornia crassipes* leaf extracts on isoniazid induced oxidative stress and hepatic markers. BioMed Res Int. 2014; Article ID 459452:11

142. Sharma A K, Kumar S, Pandey A K. Cell cycle inhibitory activity of *Piper longum* against A549 cell
line and its protective effect against metal-induced toxicity in rats. Ind J Biochem Biophys. 2014;51:358364

- 143. Harman D. Aging: a theory based on free radical and radiation chemistry. J Gerontol. 1956;11:298 300
- 750 144. Harman D. The biologic clock: the mitochondria? J Am Geriatrics Society. 1972;20:145-47
- 751 145. Harman D. Free radicals in aging. Mol Cell Biochem. 1998;84:155-61
- Ames B N, Shigenaga M K, Hagen T M. Oxidants, Antioxidants, and the degenerative diseases of
   aging. Prod Natl Acad Sci USA. 1993;79:15-22
- 147. Dean R T, Fu S, Stocker R, Davies M J. Evidence for roles of radicals in protein oxidation in advanced human atherosclerotic plaque. J Biochem. 1997;324:1-18
- 148. Beckman K B, Ames B. The free radical theory of aging matures. Physiol Rev. 1998;78:547-81
- 757 149. Orr W C, Sohal R S. Extension of life-span by over expression of superoxide dismutase and catalase in *Drosophila melanogaster*. Science. 1994;263:1128-1130
- 150. Chintawar S D, Somani R S, Kasture V S, Kasture S B. Nootropic activity of *Albizia lebbeck* in mice.
   J Ethnopharmacol. 2002;81:299-05
- 151. Shiva kumar L, Gouda S T, Venkat R N, Verma R. Evaluation of memory enhancing activity of SR 105 in Experimental animals. Int J Res Ayur Pharm. 2011;2:973-77
- 152. Chiba T, Yamaza H, Higami Y, Shimokawa I. Anti-aging effects of caloric restriction: involvement of
   neuroendocrine adaptation by peripheral signaling. Micro Res Technol. 2002;59:317-24
- 153. Bastianetto S, Quirion R. Natural extracts as possible protective agents of brain aging. J NeurolAging. 2002;23:891-97
- 154. Chang I M. Anti-aging and health-promoting constituents derived from traditional oriental herbal
   remedies: information retrieval using the TradiMed 2000 DB. Ann N Y Acad Scie. 2000;928:281-6
- 769 155. Yu M S, SaranaKa-Yang L, Sau-Wan L, Chi-Ming C, Sze-Yong Z, Mattson M P. Emerging
   770 neuroprotective strategies for Alzheimer's disease: dietary restriction, telomerase activation, and stem
   771 cell therapy. J Exp Gerontol. 2000;35:489-02
- 156. Schmid D C, Schürch P, Blum E, Belser F, Zü I. Plant stem cell extract for longevity of skin and hair.
   Int J App Sci. 2008;134:30-35
- 157. Oladele A T, Alade G O, Omobuwajo O R. Medicinal plants conservation and cultivation by
   traditional medicine practitioners (TMPs) in Aiyedaade Local Government Area of Osun State, Nigeria.
   Agri Biol J N Am. 2011;2:476-87
- 158. Schulz V, Hansel R, Tyler V. Rational phytotherapy: A physician's guide to herbal medicine. 3<sup>rd</sup> ed,
   1998; pp. 107-123, Berlin, Germany: Springer-Verlag
- 159. Hanumanthachar J, Navneet K, Jyotibala C. Evaluation of nootropic effect of Argyreiaspeciosa in
   mice. J Health Sci. 2000;53:382-88
- 160. Singh H K, Dhawan B N. Neuropsycho pharmacological effects of the Ayurvedic nootropic *Bacopa monniera* Linn. (Brahmi). Ind J Pharmacol. 1997;29:359-65
- 161. Vollala V R, Upadhya S S. Nayak S. Effect of *Bacopa monniera* Linn. (Brahmi) extract on learning
   and memory in rats: A behavioral study. J Vet Behavior. 2010;5:69-74
- 162. Singh P P, Chandra A, Mahdi F, Ray A, Sharma P. Reconvene and reconnect the antioxidant
   hypothesis in Hu man health and disease. Ind J Clin Biochem. 2010;25:225-43

- 163. Sudharani D, Krishna K L, Deval K. Safia A K. Pharmacological profiles of *Bacopa monniera* : A
   review. Int J Pharmacol. 2011;1:15-23
- 164. Deval K, Vaibhav S & Krishna K L. Effect of bacopaon memory deficit produced by chronic admin
   istration of Topiramatein Rats. Int J Pharmacol. 2011;1:118-24
- 165. Hagen T M, Yowe D L, Bartholomew J C, Wehr C M, Do K L, Park J Y, Ames B N. Mitochondrial
  decay in hepatocytes from old rats: membrane potential declines, heterogeneity and oxidants increase.
  Prod Natal Acad Sci USA. 1997;94:3064-69
- 166. Sastre J, Pallardo F V, Pla R, Pellin A, Juan G, O'Connor J E, Estrela J M, Miquel J, Vina Aging of
   the liver: Age-associated mitochondrial damage in intact hepatocytes. J. Hepatol. 1996;24:1199-05
- 167. Sohal R S. Sohal B H. Hydrogen peroxide release by mitochondria increases during aging. Mec
   Aging Dev. 1991;57:187-202
- 168. Miquel J. Fleming J E. A two-step hypothesis on the mechanisms of in vitro cell aging: cell
   differentiation followed by intrinsic mitochondrial mutagenesis. Exp Gerontol. 1984;19:31-36
- 800 169. Iquel J M. An update on the oxygen stress -mitochondrial mutation theory of aging: genetic and 801 evolutionary implications. J Exp Gerontol. 1988;33:113-26
- 170. Levine R L, Williams J A, Stadtman E R, Shacter E. Carbonyl assays for determination of oxidatively
   modified proteins. Methods Enzymol. 1991;233:346-56
- 171. Lett B S, Stadtman E. Protein oxidation in aging, disease, and oxidative stress. J Biol Chem.
   1997;272:2031-36