

Original Research Article

Phytochemical screening, evaluation of antioxidant and anti-sickling activities of two polar extracts of *Combretum glutinosum* leaves. Perr. ex DC

ABSTRACT

Sickle cell disease is a major public health problem in Africa and many other areas across the world. In Senegal, a lot of plants are proposed by traditional healers to manage the sickle cell disease, among them *Combretum glutinosum*. Antisickling activity of *Combretum glutinosum* leaves extracts was studied in this work. Methanolic and ethyl acetate extracts of *Combretum glutinosum* leaves were evaluated on SS sickles type to determine their anti-sickling potential. Antiradical properties of methanolic extract were evaluated using the DPPH radical as oxidant. Total phenolic content of the methanol extract was determined. Phytochemical screening of the crude extract of methanol revealed the presence of tannins, saponins, phenols and flavonoids. Results show a good antisickling effect of extracts with a maximum antisickling revers of 81% and 69% respectively, of the methanol and ethyl acetate extracts at 10 mg/mL versus 67% for arginine at 120 min of incubation. The measured IC₅₀ were 0.65 and 0.163 for respectively the methanol extract and ascorbic acid. Antiradical powers 0.155 and 0.62 respectively for methanol extract and ascorbic acid were calculated from the effective concentrations. The results of this study confirm the traditional use of *Combretum glutinosum* leaves in the management of sickle cell disease.

Keywords: Sickle cell disease, oxidative stress, medicinal plant, *Combretum glutinosum*, antisickling activity, antioxidant activity

24 1. INTRODUCTION

25 Medicinal plants are used in the whole world and have a growing economic importance. The
26 use of plant parts to treat human disease is as old as disease itself [1]. According to the World
27 Health Organisation (WHO), as many as 80% of world population, particularly in developing
28 countries, have used at least one time in their lives, traditional medicines for their primary
29 health care need today and 25% of drugs are based on plants and their derivatives [2].
30 *Combretum glutinosum* is a very large genus comprising of about 250 species found in both
31 temperate and tropical regions in the world [3]. It is largely distributed in Africa throughout
32 the Sahel belt from Senegal to Cameroon and eastwards to the Sudan. It is a bushy shrub or
33 small tree growing up to 12 m and deciduous species sprouting in the middle of the dry
34 season. The plant parts are used in traditional medicine in Africa to treat many human
35 diseases. In Nigeria, the roots, the stem, the bark and the leaves of *Combretum glutinosum* are
36 used in the treatment of scrotal elephantiasis, dysentery, ring worms, bacterial diseases,
37 syphilis, typhoid fever etc [4]. Antidiarrheal activity of aqueous young leaves extract of
38 *Combretum glutinosum* was demonstrated [5]. Anti-malarial and anti-schistosomal activities
39 of *Combretum glutinosum* were respectively described by Yahaya and Ouattara [2, 6].
40 Therapeutic potential of different parts of *Combretum glutinosum* remains to be explored on
41 several other diseases such as sickle cell disease (SCD).

42 Indeed Sickle cell disease is a form of haemoglobinopathy that is characterized by chronic
43 hemolytic anemia. SCD is a genetic disease in which the SS individual possesses an
44 abnormal β -globin gene. A single base substitution in the gene encoding the human β -globin
45 subunit results in the replacement of $\beta 6$ glutamic acid by valine, which leads to the
46 devastating clinical manifestations of SCD. This substitution is followed by a remarkable
47 inflammatory response and oxidative stress within the blood circulation [7]. It also causes a
48 drastic reduction in the solubility of sickle cell hemoglobin when deoxygenated, frequent

49 chest pain and vaso-occlusive complication [8]. SCD occurs world-wide and the African
50 continent bears the highest burden. Hispanic, Arabic, Mediterranean and some Asian
51 populations are also affected. According to Tim [9], approximately 250,000 children are born
52 each year with sick cell disease and over 70,000 individuals in the U.S. suffer from the
53 disease. In Africa, SCD is a major public health problem with a prevalence of SCD trait
54 between 10 and 40% [10]. In Senegal it is between 8 and 10% in the general population [11].
55 Due to the high mortality rate of sickle cell patients, especially in children, and since
56 chemotherapy has its adverse effects, there is need for rational drug development that must
57 embrace not only synthetic drugs but also natural products. It is acknowledged world-wide
58 that traditional medicine can be explored and exploited for enhanced health management. The
59 ever increasing demand for safer and cheaper herbal recipes in developing country, has led to
60 the extraction and development of several drugs and chemotherapeutic agent from plants, as
61 well as from traditional herbal remedies [5]

62 In recent years, the interest in natural antioxidants, in relation to their therapeutic properties,
63 has increased dramatically. Scientific research in various specialties has been developed for
64 extraction, identification and quantification of these compounds from several natural
65 substances namely, medicinal plants and food products [12, 13]. The antioxidant activity of a
66 compound corresponds to its ability to resist oxidation. The best known antioxidants are the
67 β -carotene (provitamin A), ascorbic acid (vitamin C), tocopherol (vitamin E) as well as
68 phenolic compounds. Indeed, most synthetic or naturally occurring antioxidants have
69 phenolic hydroxyl groups in their structures; and antioxidant properties are attributed in part
70 to the ability of these natural compounds to scavenge free radicals such as hydroxyl radicals
71 ($\text{OH} \cdot$) and superoxide ($\text{O}_2^{\cdot -}$) [14, 15]. Several methods are used to evaluate, *in vitro* and *in*
72 *vivo* antioxidant activity by scavenging different radicals such as peroxy radical ($\text{ROO}^{\cdot}$) by
73 ORAC and TRAP methods [16]; ferric ions by FRAP [17]; or the ABTS $\cdot$ radical [18] and

74 the method using free radical DPPH• [19]. Because it takes place at room temperature and
75 can eliminate the risk of thermal degradation of thermolabile molecules, the DPPH• radical
76 test is the most recommended for very hydrophilic extracts which are rich in phenolic
77 compounds [20].

78 To our knowledge, the activities of *Combretum glutinosum* parts have not been investigated
79 against the Sickle cell disease. The objective of this study is on the one hand to assess the
80 potential anti-sickling activity of two polar leaves extracts of *Combretum glutinosum* and on
81 the other hand to measure *in vitro* the antioxidant activity of the most active extract by using
82 the DPPH radical method.

83 2. MATERIALS AND METHODS

84 2.1. Materials

85 2.1.1. Blood sample, chemistry and biochemistry products

86 The sickle cells used in this work were obtained from unused SS sickle blood of patients who
87 undergo a diagnostic test for sickle cell disease. These patients had been neither transfused
88 nor treated with hydroxyurea in the last six months. The age of patients ranged from 18 to 35
89 years in both sexes. Sampling is made at the elbow crease of the patients and the blood was
90 collected in EDTA tubes. Chemical and biochemical products used in this work were
91 obtained from different suppliers (Prolabo, Scharlau, Aldrich or Carlo Erba). All solvents
92 were freshly distilled before use. The human ethics approve was obtained from Cheikh Anta
93 Diop University of Dakar under the number 0228/2017/CER/UCAD on February 03 2017.

94 2.1.2. Plant Material

95 The leaves of *Combretum glutinosum* used in this work were purchased from traditional
96 healers in Tilene market in Dakar. The identification was carried out at the Laboratory of

97 Pharmacognosy of the Medicine, Pharmacy and Odontology Faculty at Cheikh Anta Diop
98 University of Dakar. The dried leaves were crushed and powdered using a Brabender mill.

99 **2.2. Methods**

100 **2.2.1. Extraction of the *Combretum glutinosum* leaves powder**

101 **Leaves powder** 60g of *Combretum glutinosum* were extracted with 300 mL of ethyl acetate.
102 The mixture was stirred at room temperature for 48 hours. The macerated matter was filtered
103 and the filtrate evaporated to dryness using vacuum rotary evaporator. The crude methanol
104 extract was obtained from 60g of *Combretum glutinosum* leaves powder in 300 mL of
105 methanol using the same operation as the one that has **been achieved using** the ethyl acetate
106 extract. The extracts were **labeled** as follows: **EAE** (ethyl acetate extract); **ME** (methanol
107 extract) and stored in a refrigerator at 4°C until use. **The percentage yield of the extracts was**
108 **determined by using the following formula: $Y = X/WI \times 100$, with X = mass of extract and**
109 **WI = initial mass of the powder.**

110 **2.2.2. Phytochemicals screening**

111 Methanol extract was subjected to preliminary phytochemical tests using standard technique
112 to detect the presence of alkaloids, saponins, polyphenols, tannins and flavonoids as
113 described in our previous work [21].

114 **2.2.3. Determination of antioxidant activity by the DPPH radical scavenging method**

115 Measurement of the antiradical activity of methanolic extract from *Combretum glutinosum*
116 leaves is performed by the 2,2'-diphenyl-1-picrylhydrazyle (DPPH) test in accordance with
117 the method described by Scherer [19]. **DPPH (4 mg) was** dissolved in 100 mL of methanol,
118 the solution is then stored for 1 hour in the dark. A concentration range (0.25 to 5 mg/mL) of
119 methanolic extract from the *Combretum glutinosum* leaves and (0.09 to 0.5 mg/mL) of

ascorbic acid (reference) was prepared in methanol. A volume of 0.1 mL of *Combretum glutinosum* leaves solution is mixed with 3.9 mL of the DPPH solution. After homogenization, the mixture was incubated in the dark for 30 minutes. The mixture absorbance is measured with a spectrophotometer at 517 nm.

Antioxidant activity is defined as the amount of antioxidant necessary to decrease the initial DPPH radical concentration by 50% [22]. Results are expressed in the first time as the percentage of the Radical Scavenging Activity (%RSA) by using the following expression.

$$\%RSA = \left[\frac{Abs_{control} - Abs_{sample}}{Abs_{control}} \right] \times 100$$

And in the second time, the inhibition concentration, IC₅₀ (concentration of extract that traps 50% of DPPH radical) was determined by projection of the absorbance values of methanol extract and ascorbic acid solution on the graphic representing the trapping percentage as function of the concentration. It is possible to deduced from IC₅₀ values, the efficient concentration (EC₅₀) and the Antiradical Power (ARP) as described in our previous paper [23]

2.2.4. Determination of polyphenols content

Phenols content of methanol extract was determined by the Folin Ciocalteu method according to a protocol written by Bidie et al [24]. To a concentration of 0.1 mg/mL of methanolic *Combretum glutinosum* leaves extract, 0.5 mL of Folin Ciocalteu reagent 10% (v/v) is mixed with 2.5 mL of distilled water and 2 mL at 0.5M of sodium carbonate (Na₂CO₃). After 30 minutes incubation at room temperature, the absorbance is read on a spectrophotometer at 765 nm. Gallic acid was used as the reference antioxidant. Concentrations of 0.001, 0.015, 0.025, 0.05, 0.075, 0.1, 0.15, 0.2 mg/mL were prepared from gallic acid in hemolysis tubes. To each tube is added 2.5 mL of Folin Ciocalteu reagent 10% (v/v) and 2 mL of sodium

carbonate. The tubes are incubated at room temperature for 30 minutes and then absorbance read at 765 nm. Methanol was used as a blank. Results obtained allowed to draw calibration curve of gallic acid. Projection of the absorbance value of methanolic extract on calibration curve has permitted to determine graphically total phenol content expressed in terms of gallic acid equivalent (mg GAE/g of the sample). All analyses were performed in triplicate.

2.2.5. Antisickling activities

In vitro screening of polar extract of *Combretum glutinosum* leaves for antisickling proprieties with blood samples collected from patients with the confirmed SS sickle cell by Emmel test and electrophoresis, was carried out using a protocol described by Imaga [25] and given in our previous work [20]. Morphological analysis of erythrocytes (100 cells in 4 to 5 fields) is performed using an immersion microscope. When extracts are brought into contact with blood, every 30 minutes up to 120 minutes an erythrocyte count is performed.

2.2.6. Data analyses

The results of antisickling activities were evaluated in percentage of residual sickles. For the negative control, since the number of sickle cells increases with time, it was considered that the rate of 100% corresponds to the number of sickle cells obtained in 120 minutes. For samples tested, this number decreases over time; so the rate of 100% is the number of sickle cells at the initial time T0. Evolution of the percentage of residual sickled cells in function of time is given by the following equation:

$$\text{PDR} = \frac{\text{Mean of sickles at Tx}}{\text{Mean of sickles at T0}} \times 100$$

PDR = Percentage of residual sickles

163 $T_x = 0, 30, 60, 90$ et 120 minutes

164 T_0 = initial time

165 3. RESULTS

166 3.1. Yield of extracts

167 Extraction results in **Table 1** show that the percentage yield of methanol extract of
168 *Combretum glutinosum* leaves is higher (7.45%) than the ethyl acetate extract (5.00%). This
169 difference may be justified by the greater polarity of the methanol solvent.

170 **Table 1. Yield of extracts**

Solvents	Methanol	Ethyl acetate
Extracted Plant powder (g)	4.47	3.00
Percentage yield (%)	7.45	5.00

171

172 3.2. Phytochemical screening result

173 The preliminary phytochemical screening of *Combretum glutinosum* leaves shows presence
174 of various bioactive components which are given in **Table 2**. Results provide evidence of the
175 presence of saponins, phenols, tannins and flavonoids and show the absence of alkaloids.
176 These results are confirmed by Yahaya and Albagouri [2, 26] in their phytochemical
177 screening of *Combretum glutinosum* study, they show among other things the presence of
178 saponins, tannins, flavonoids, phenolic compounds and alkaloids. However, the difference is
179 they show the presence of alkaloids. This difference could be explained by a difference of
180 nature of solvent used, the harvest period or the nature of the soil where the plant grows.

181

182

183 **Table 2: Results of the phytochemical screening**

Alkaloids	Saponins	Polyphenols	Tannins	flavonoids
-	+	+	++	++

184 - Absence, + Presence

185 **3.3. Antiradical activity**186 Antiradical activity of methanolic extract of *Combretum glutinosum* leaves is given in **Figure**187 **1**. Activity is determined on an average of three measurements per concentration using the

188 statview statistical software by performing a normal analysis of variance (ANOVA) followed

189 by Fischer test. Difference is considered significant if $p < 0.05$ versus negative control. The190 results give $IC_{50} = 0.65$ and 0.163 respectively for methanol extract and ascorbic acid.191 Antiradical power calculated from the effective concentration (EC_{50}) informs on the ability of

192 active compounds to inhibit the free radical activities. Indeed the more important the

193 antioxidizing power value of the concerned product is, the higher is its activity. Thus we

194 calculated the anti-radical power of 0.155 and 0.62 for respectively, methanol extract of195 *Combretum glutinosum* leaves and ascorbic acid **Table 3**.

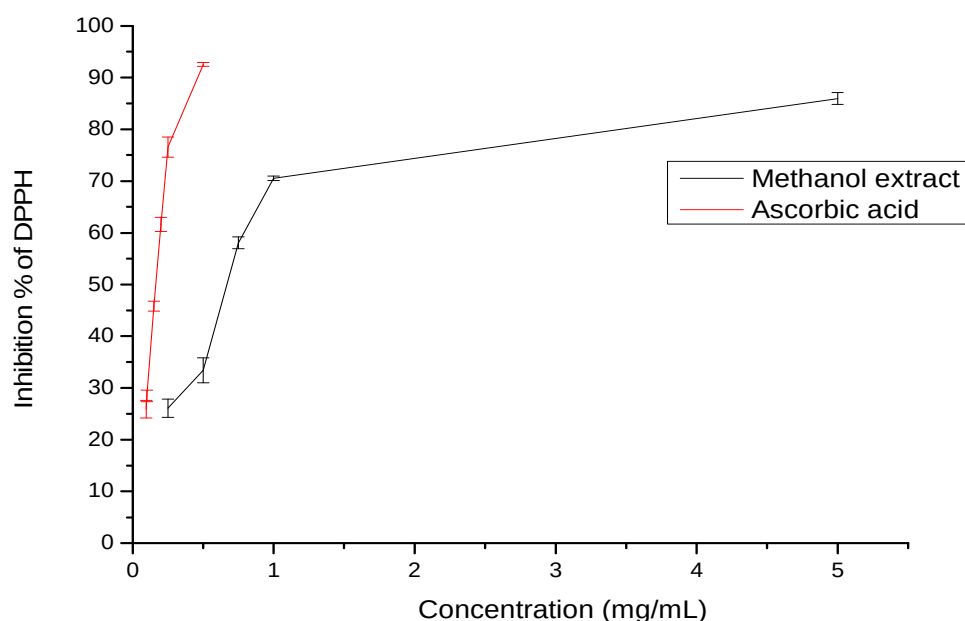


Figure 1. Percentage inhibition of DPPH radical by *C. glutinosum* and ascorbic acid

Table 3: Values of IC_{50} , EC_{50} and ARP from *C. glutinosum* and ascorbic acid

Studied parameters	<i>C. glutinosum</i> methanol extract	Ascorbic acid
IC_{50} (mg/ml)	0.65	0.163
EC_{50} (CI_{50} / mol of DPPH).	6.43	1.61
ARP	0.155	0.62

IC = Inhibition Concentration, EC = Effective Concentration, ARP = Antiradical Power

3.4. Polyphenols content determination

Determination of phenol content of *Combretum glutinosum* methanolic leaves extract is performed using the regression equation of the calibration curve of gallic acid (**Figure 2**), where **Y** is absorbance of methanolic extract (0.233).

Phenol content is then equal to **X/concentration of methanolic extract**. Calculation gives total polyphenol content equal to **150 mg GAE/g sample**.

This result is better than those obtained by Yahaya [2]. For the determination of total phenols content from different extracts solvent of *Combretum glutinosum*, the best result, 130 mg **GAE/g** of extract was given by methanol extract.

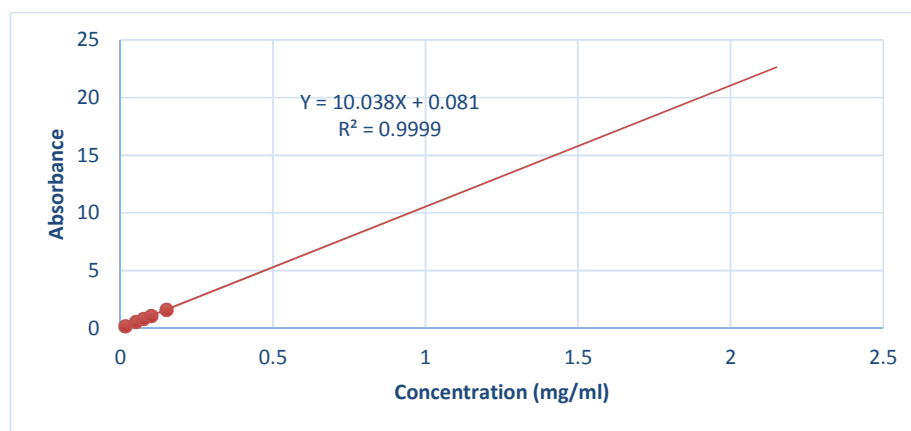
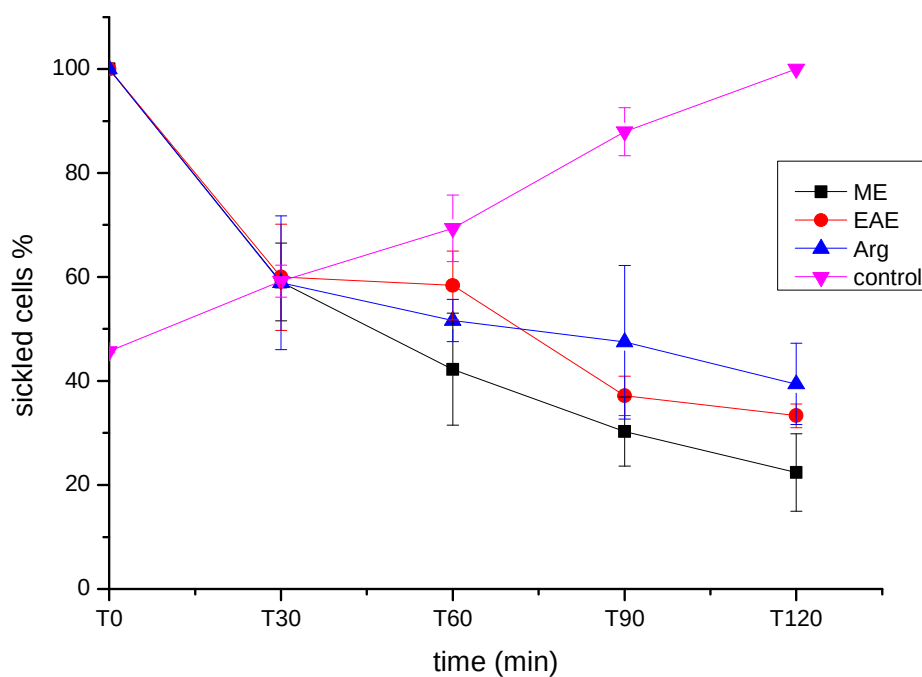


Figure. 2. Calibration curve of the gallic acid ($y = 10.038x + 0.081$; $R^2 = 0.99947$)

3.5. Antisickling effect of *Combretum glutinosum* leaves extracts

Results of the antisickling activity of *Combretum glutinosum* leaves extracts are presented in **Figures 3 and 4**. Results show that the two studied polar fractions have interesting activities on sickle cells compared to the negative control for which the number of sickle cell increases over time. We also find that methanol extract which has higher polarity as ethyl acetate

220 extract, presents the best results. In both studied concentrations, methanolic and ethyl acetate
 221 extracts have more reversibility of sickling cells than arginine used as positive control.



222 **Figure 3. Antisickling activities of *C. glutinosum* leaves extracts at 5 mg/mL**

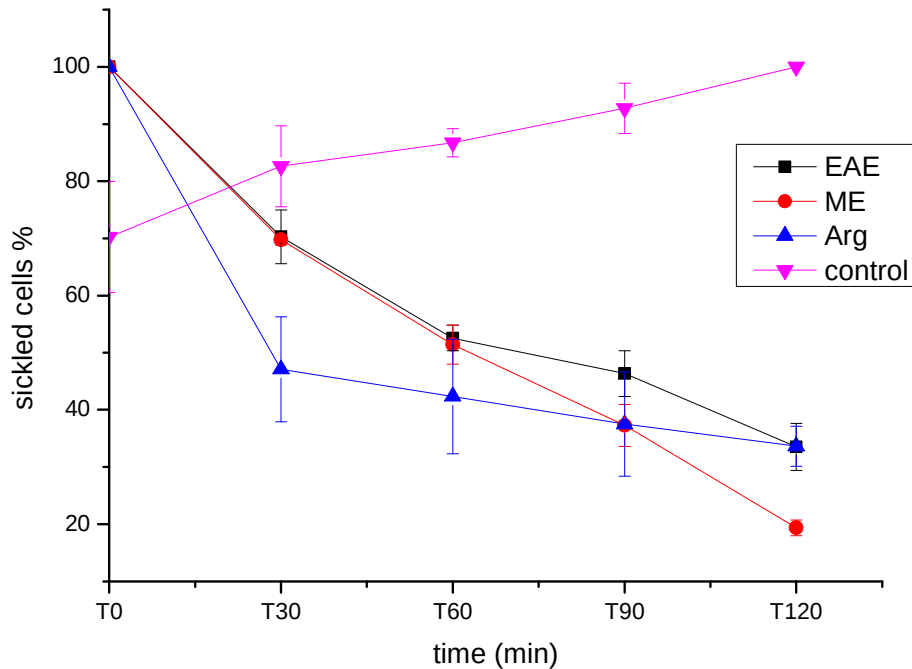


Figure 4, Antisickling activities of *C. glutinosum* leaves extracts at 10 mg/mL

4. DISCUSSION

Today research is booming in Africa in the development of herbal medicines, drugs and dietary supplements, in the management of tropical diseases and genetic disorder, especially in sickle cell disease where science is still struggling to establish a sustainable treatment. These plant treatments would be for most of the population an alternative to modern medicine because they are not expensive and are accessible to all. Thus, it is after an ethnobotanical study conducted by the traditional healers that *Combretum glutinosum* leaves were selected to study their effects on the SS type sickle.

Methanol and ethyl acetate extracts from *Combretum glutinosum* leaves were active on the reversibility of the sickling of sickle cells. Thus methanol is the most active extract and has 81% sickling reversal in 120 minutes of incubation. Arginine used as positive reference present in the same conditions 67% sickle reversibility. Also arginine is recommended in the

239 management of vaso-occlusive crises. Indeed nitric oxide which plays an important role in
240 the sickle cell disease pathophysiology, has arginine as substrate. It is considered as a
241 vasodilator of blood vessels by acting on the endothelial functions. Therefore, it can reduce
242 the vaso-occlusion crises [27]. Ethyl acetate extract presents a less important activity than
243 methanol, its reversibility rate is 69% at 10 mg/mL in 120 minutes of incubation. The most
244 important polarity of methanol as ethyl acetate explains the most important activity of the
245 former. Methanol extract would contain polar secondary metabolites whose synergetic effects
246 justify this most important activity on the reversibility of the sickling of sickle cells.
247 Phytochemical screening performed on methanol extract, showed the presence of
248 polyphenols, tannins, saponins and flavonoids and the absence of alkaloids. According to
249 Iweala [28], the synergistic effects of a wide variety of biologically active substances and
250 amino acids contained in plants are responsible for the observed antisickling activities.
251 Antisickling results of the methanolic extract of *Combretum glutinosum* leaves (81%) in
252 comparison with those obtained in our previous work on *Leptadenia hastata* [21] and on the
253 *Maytenus senegalensis* [23] which are respectively 77 and 80% sickling reversibility of sickle
254 cells, are more interesting and give bright prospects.

255 One of the main characteristics of sickle cell disease is the production of a large amount of
256 free radicals leading to severe oxidative stress and consumption of nitrogen monoxide by
257 oxygen free radicals. Oxidative stress also affects the ratio $\text{Fe}^{3+}/\text{Fe}^{2+}$, quite high in the sickle
258 cells, is involved in sickle cell hemolysis [29]. Identification of plant antioxidant properties,
259 would indicate its favourable action on the management of sickle cell disease. Indeed, we
260 have demonstrated *in vitro* that the methanol extract of *Combretum glutinosum* leaves inhibits
261 the activity of DPPH radical with an IC_{50} of 0.65 versus 0.163 for ascorbic acid. Polyphenol
262 content determined from the calibration curve of gallic acid is 150 mg GAE/g. These results

show that the methanol extract acts also on the reduction of the oxidative stress generated in sickle cell disease.

Oxidative processes, which trigger the production of free radicals, resulting in tissue damage, are a major contributor to diminished health, and manifested in a wide spectrum of disease [30]. The medicinal value of plants is related to their phytochemical component content and secondary metabolites, including: phenolic compounds, flavonoids, alkaloids, tannins, and other stress gene response products. The flavonoids present in the *Combretum glutinosum* leaves represent a large family of polyphenols and the involvement of the latter in the treatment of oxidative stress has been proven [31]. Antioxidant activity of phenolic compounds is mainly due to their redox properties, which allow them to act as reducing agents, hydrogen donors, singlet oxygen quenchers and metal chelators [32]. The information presented above suggests to us that *Combretum glutinosum* leaves with high phenol content could, not only be a solution in the management of sickle cell disease, but also beneficial to all other pathologies generated by oxidative stress.

5. CONCLUSION

Combretum glutinosum leaves have been reported by the traditional medicine healers in Senegal to have antisickling activities. This necessitated scientific evaluation of the concerned part of the plant against the homozygous sickle. Methanol and ethyl acetate extracts of *Combretum glutinosum* leaves have a significant sickle cell reversal activities. These results confirm the traditional use of the *Combretum glutinosum* leaves in the management of sickle cell disease. This alternative therapeutics using phytotherapy has proven to, not only reverse sickling cells but also acts on the oxidative stress. Future studies should isolate and study the bioactivities compounds responsible for the potential antisickling in this plant.

287

288 **Ethical approval**

289 **0228/2017/CER/UCAD, from Cheikh Anta Diop University of Dakar**

290

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Picture of *Combretum glutinosum* Perr.ex DC

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390 This Picture was taken recently, 02-02-2017 in Kounghoul Forest in Senegal

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