

Aflatoxin M1 in Milk and Milk Products in Jordan and Methods for its Reduction

Khalaf S. Al-Delaimy^{1*} Iman F. Mahmoud²

¹ Department of Clinical Nutrition and Dietetics, Faculty of Pharmacy and Medical Sciences, University of Petra, Amman, Jordan

² Department of Nutrition and Food Technology, The University of Jordan, Amman, Jordan

ABSTRACT

Aims: The aim of this study was to assess Aflatoxin M1 (AFM1) levels in liquid and powdered milk and other dairy products in Jordan using ELISA technique and antiaflatoxin M1 (AFM1 reduction) effect of fermentation, high heat treatments and the addition of plant extracts during milk fermentation.

Place and Duration of Study: Department of Nutrition and Food Technology, The University of Jordan, Amman, Jordan during July 2006 and March 2007.

Methodology: A total of 22 samples of milk and dairy products were randomly collected from local markets and were grouped as follows: raw (n=3), pasteurized (n=4) and powdered milk (n=3), infant formula (n=3), yoghurt (n=3), labaneh (low moisture sour yoghurt) (n=3) and jameed (traditional dry yoghurt) (n=3). Dairy products were analyzed for their AFM1 content.. Anti-aflatoxin treatments of fermentation, high heat, garlic (*Allium sativum*), black cumin (*Nigella sativum*) and carrot (*Ducaus carota sativus*) water extracts were studied to determine their influence on AFM1 content.

Results: The results show that AFM1 was detected in 81.8% of tested samples with varying levels ranging from 25 ppt to no detection. The highest levels were found in 2 labaneh samples purchased from local brands (25.8 and 22 ppt of AFM1). Levels of AFM1 of 19, 15.5, and 14 ppt were detected in a yoghurt, raw milk, and jameed samples respectively. No AFM1 was detected in imported infant formula samples, pasteurized milk and one labaneh brand. Carrot water extract had the highest impact by decreasing AFM1 by 72% of the total toxin followed by heating (56%) and fermentation (40%).

Conclusion: These AFM1 levels found in milk and dairy products in Jordan are considered low and within the European maximum limit of 50 ppt. An important finding for AFM1 reduction was our finding that a natural product such as carrot extract can be effective in lowering AFM1 in milk samples.

Keywords: Aflatoxin M1, ELISA, ,AFM1 Reduction, Plant extracts, Milk products

1. INTRODUCTION

Aflatoxins are a group of mycotoxins, which came to our knowledge in the early 1960s. In the early months of 1960 at total of 100,000 turkey poultries died in mysterious circumstances in the United Kingdom. Thorough investigations showed that the turkeys died from acute liver necrosis and bile duct hyperplasia after they consumed infected groundnuts [1].

Aflatoxins (AFs) are produced primarily by some strains of *Aspergillus flavus* and *A. parasiticus*. These strains produce four major classes of aflatoxins: AFB1, AFB2, AFG1 and AFG2 in addition to major metabolites derivatives such as aflatoxin M1 (AFM1) and aflatoxin Q1 (AFQ1) that are developed either by metabolism in humans, animals, and microorganism or by environmental reactions [2]. AFB1 was found to be the most common and the most hazardous of all mycotoxins found in human food and animal feed. AFs were classified as group 1 carcinogens by IARC [3]. AFB1 is categorized as class 1 carcinogen (definitely carcinogenic) for humans, whereas AFM1 is a group 2B carcinogen (probably carcinogenic) for humans [4]. AFM1 is a hydroxylated metabolite of AFB1, first detected in the milk of lactating cows that had injected feeds contaminated with AFB1 [5]. AFM1 is released with milk produced by cows, sheep, goats and other lactating animals that have ingested feeds contaminated with AFB1. The relationship between the amount of AFB1 ingested by animal and quantity of AFM1 recovered in milk is quite variable and estimated to be between 1-3% [6]. It was also estimated that about 0.3-6.2% of AFB1 in feed consumed by lactating animals is transformed to AFM1 in their milk [7]. The daily intake of AFM1 is estimated to be 6.8ng/person for the European diet, 3.5ng/person for the Latin American diet, 12ng/person for the Far Eastern diet, 0.7ng/person for Middle Eastern diet and 0.1ng/person for the African diet [8, 9]. Several studies from Turkey [10], Iran [11, 12], Morocco [13] and Jordan [14] demonstrated high levels of AFM1 in milk products that reach levels above the accepted maximum levels determined by the European Union. All these studies recommend development of approaches to address this public health problem.

Although the potency of AFM1 is one tenth that of AFB1, AFM1 still represents a potential carcinogen for humans [2]. The contamination of dairy products with AFM1 has been recognized as a significant human health hazard, particularly for babies, since milk is their basic food.

The tolerance limit for AFM1 in milk varies from one country to another. The European Community and Codex Alimentarius Commission put a maximum level of AFM1 in liquid milk not to exceed 50ng/l whereas the USA FDA stretched the safety levels to 500ng/Kg [8].

Studies on the reduction of AFM1 in dairy products by using natural products are very limited. The aim of this study is to assess antiaflatoxin M1 (AFM1 reduction) effect of fermentation, high heat treatments and the addition of plant extracts (Garlic (*Allium sativum*), black cumin (*Nigella sativa*) and carrots (*Daucus carita sativus*)) during milk fermentation. We also assess AFM1 levels in liquid and powdered milk and other dairy products (yoghurt, labaneh and jameed) marketed and consumed in Jordan using the ELISA technique.

2. MATERIAL AND METHODS

2.1 Sampling and sample preparation

A total of 22 samples of raw (n=3), pasteurized (n=4) and powdered milk (n=3); infant formula (n=3); yoghurt (n=3); labaneh (low moisture sour yoghurt produced by straining yoghurt by clean cloth to remove 50% of the water) (n=3); and jameed (traditional dry yoghurt) (n=3) were collected randomly from local markets in Amman, Jordan during the period July 2006 and March 2007. Ten gm (10 gm) of each of powdered milk, infant formula and jameed were suspended in 100 ml of deionized water and homogenized. One ml (1 ml) of each of liquid milk samples and reconstituted powdered milk, infant formula and jameed

* Tel.: 962-65715546 ext. 8413

E-mail address: kaldelaimy@uop.edu.jo

73 were cooled to 10° C and centrifuged at 2500 rpm for 10 min. to separate and remove the fat
74 layer. The upper, creamy layer was aspirated with a Pasteur pipette, and the supernatant
75 (defatted skimmed milk) was used to evaluate AFM1 (100 µl per well). One ml (1 ml) of each
76 sample was prepared for AFM1 assay similar to milk samples.

77 2.2 Quantification of AFM1 by ELISA

78 Samples were analyzed for AFM1 content according to ELISA manufacturer procedure
79 AFM1 standards provided by the same source (R-Biopharm AG, Dermstadt, Germany).

80 The ELISA assay is based on the affinities of the monoclonal or polyclonal antibodies
81 against AFM1. ELISA is a relatively cheap, sensitive and quick method for the determination
82 of AFM1 and is suitable for routine diagnostic application [15, 16]. The sensitivity of ELISA
83 method for AFM1 detection has been investigated by many and compared to other methods,
84 such as HPLC and found to be a more sensitive and reliable method than HPLC [16]. The
85 AFM1 ELISA kit has a quantitation range of 5-100ppt with high reliability but lower levels are
86 detected as the lowest concentration that could be differentiated from zero.

87 The microtitre plate wells are pre-coated with mouse anti- AFM1. Before the first incubation
88 step, AFM1 standards or samples are examined for the presence of AFM1 added to the pre-
89 coated wells. The free AFM1 (in the standard solution or in the sample) is bound by the
90 specific antibody binding sites. Other unbound material of samples are washed away in the
91 washing step by buffer (10 Mm phosphate buffer containing 0.05% Tween 20) leaving only
92 the AFM1 bound to the binding sites. After a washing step and before the second incubation
93 step, enzyme labeled aflatoxin (enzyme conjugate) is added into the wells. The enzyme
94 conjugate binds with free specific antibody binding sites. After the second incubation, the
95 non-bound (enzyme labeled) reagents are removed in a washing step. The amount of bound
96 enzyme conjugate is visualized by the addition of chromogen substrate
97 (tetramethylbenzidine, TMB). The bound enzyme transforms the chromogen into a colored
98 product. The substrate reaction is stopped by the addition of stop solution (sulphuric acid).
99 The color intensity is measured photometrically at 450nm and is inversely proportional to
100 the AFM1 concentration in the sample up to 80.

101 Optical density readings measured by ELISA reader were converted to absorbency % which
102 is :

103
$$\text{Absorbance \%} = (\text{Absorbance of standard (or sample)} \div \text{Absorbance of zero standard}) \times 100$$

104 A standard curve was prepared by plotting % absorbency against 6 concentrations of the
105 standard solution of AFM1 (0, 5, 10, 20, 40 and 80 ppt). Results for each concentration were
106 recorded as the average of duplicates.

107 2.3 Reduction of AFM1

108 Testing for AFM1 reduction was carried out with high heat treatment, fermentation, and the
109 addition of water extracts of garlic, black cumin and carrots added to milk samples before
110 fermentation. AFM1 was artificially added at initial levels of 7.5 - 12.5 ng/ml milk and mixed
111 vigorously for 5 min. by vortex mixer. Five equal portions of milk samples, each sample was
112 containing 3 ml were used in each reduction AFM1 experiment.

113 2.3.1 Effect of heat treatment

* Tel.: 962-65715546 ext. 8413

E-mail address: kaldelaimy@uop.edu.jo

114 The 3 milk sample containing AFM1 is divided into 4 portions. They are H1 (0.5 ml) control
115 (no heat treatment), H2 (0.5 ml) pasteurization at 72°C for 15 sec., H3 (1.0 ml) boiling for 30
116 sec. and H4 (1.0 ml) sterilization for 15 min. Each sample is heated in a water bath tempered
117 at each given temperature, and the last sample is autoclaved. All samples were cooled
118 immediately after heat treatment before ELISA assay.

119 2.3.2 Effect of fermentation

120 Three ml (3 ml) of fresh milk with AFM1 was fermented to yoghurt then divided into 4
121 portions in test tubes (F1 & F2, 0.5 ml each; F3 & F4, 1.0 ml each) and incubated at 42°C
122 for 0 times as control, 3hrs, 6hrs, 24 hrs and 48 hrs, respectively. Samples were prepared as
123 in paragraph 2.1 for ELISA assay for possible effect of fermentation process on AFM1
124 reduction.

125 2.3.3 Effect of plant extracts during milk fermentation

126 Water extracts of garlic (*Alium sativum*), black cumin (*Nigella sativum*) and carrots (*Daucus*
127 *carota sativus*) were used for an attempt to detoxify AFM1 by these extracts. Garlic extracts
128 were prepared according to the methods used by Al-Delaimy and Ali; Yin and Change [17,
129 18]. Milk samples with AFM1 were prepared as described above and designated as G0, G1,
130 G2, G3 and G4. Ten percent (10 %) of garlic extracts was added to each of 4 milk samples
131 (except the control G0) before fermentation at 42° C and incubated for 3, 6, 24 and 48 hrs,
132 respectively. Samples were assayed for AFM1 by ELISA technique to observe the AFM1
133 reduction effect of garlic extract.

134 Black cumin seed extracts were prepared as was described by Garcia et al [19] and
135 experimented with similar procedure as was for garlic water extract.

136 Fresh carrots roots produced in Jordan were washed thoroughly with tap water and three
137 times with sterilized water. The ends of the roots were removed then peeled and cut into
138 pieces by a sharp flame sterilized knife. Fifty grams of carrot pieces were blended with 100
139 ml distilled sterilized water for 3 min. Extract was filtered and 10% of the extract was used for
140 each of the 5 samples of fermented milk as was in garlic and black cumin water extract.

141 3. RESULTS AND DISCUSSION

142
143 Standard curve (Fig. 1) of the absorbency % against known AFM1 concentrations was
144 plotted for the determination of AFM1 level by ELISA technique in all samples in this study.

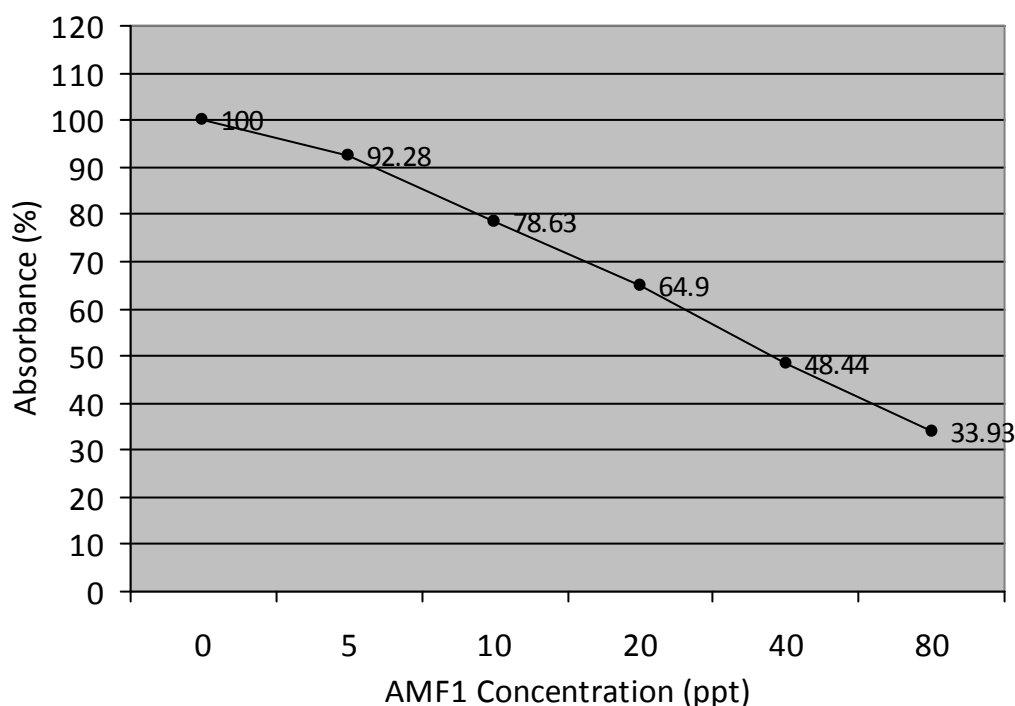


Figure 1: Standard curve of aflatoxin M1 contents in six standard concentrations.

Table 1 shows AFM1 content in milk and dairy products samples collected from different locations in Amman, Jordan by using the ELISA technique. The highest levels were found in 2 labaneh samples purchased from local brands (25.8 and 22 ppt of AFM1). Levels of AFM1 of 19, 15.5, and 14, ppt were detected in yoghurt, raw milk, and jameed samples respectively. No AFM1 was detected in imported infant formula samples (Guigoz and Similac) which may reflect the effective standards and regulations exercised by manufacturers. In addition, samples of each of pasteurized milk (University of Jordan Dairy Production) and labaneh (Baladna) also did not have detectable levels of AFM1. These AFM1 levels found in milk and dairy products in Jordan are considered low and within the European maximum limit of 50 ppt. Most or all tolerance level of AFM1 in different countries use the maximum tolerance levels of either European countries levels of 50 ppt, or USA levels of 500 ppt but there are also some countries with country specific levels that are in between these two levels.

Table 1: Aflatoxin M1 contents (ppt) of different dairy products collected in Amman.

No.	Dairy product	AFM1 (ppt)	No.	Dairy product	AFM1 (ppt)
1	Raw milk	2.0	12	Infant formula	0.5

* Tel.: 962-65715546 ext. 8413

E-mail address: kaldelaimy@uop.edu.jo

2	Raw milk	15.5	13	Infant formula	0.0
3	Raw milk	9.8	14	Yoghurt	19.0
4	Pasteurized milk	0.0	15	Yoghurt	2.0
5	Pasteurized milk	8.2	16	Yoghurt	12.4
6	Pasteurized milk	6.8	17	Labaneh	25.8
7	Pasteurized milk	6.1	18	Labaneh	22.0
8	Powder milk	5.2	19	Labaneh	0.0
9	Powder milk	7.8	20	Jameed	14.0
10	Powder milk	3.5	21	Jameed	13.0
11	Infant formula	0.0	22	Jameed	8.0

160

161 The AFM1 levels in samples from Jordan were lower than the levels in milk samples
162 consumed in Kuwait [20, 21], Iran [22], Turkey [23], Syria [24] and Pakistan [25].
163 Approximately 82% of the dairy samples from our study had detectable AFM1 (with 85% in
164 milk samples). A study in Turkey found that 55% of raw milk, 25% of ultra-heat treated milk,
165 20% white pickled milk, and 20% of yoghurt samples were contaminated with AFM1 above
166 the acceptable Turkish legal limit (250 ng/kg) [10]. In India, for example, AFM1 was found to
167 contaminate up to 99% of milk samples tested and reaching extremely high levels of 48000
168 ppt [26]. In Thailand the levels reached 6600 ppt [27]. In China, using the ELISA assay
169 method to determine the occurrence of AFM1 in milk, it was found that 48.1% of milk
170 samples and 4.5% of yoghurt samples had detectable levels of AFM1. These levels were
171 found to be within the current regulatory limit in China [28]. These results indicate that the
172 health risks associated with AFM1 in China are relatively low. In Japan AFM1 was detected
173 in 99.5% of the tested samples, although the levels were very low (ranging between 1-29
174 ppt) [9]. In our study similar AFM1 levels in milk and dairy products were found as in Japan.
175 It is believed that the higher levels of AFM1 in milk and dairy products in some countries are
176 probably due to poor hygienic and storage conditions with high humidity. These factors are
177 favorable for the growth of *Aspergillus flavus* and *Aspergillus parasiticus* in animal feeds.
178 There was variability in AFM1 levels in the samples were collected and this reflects different
179 production and hygienic practices that requires further investigation and research. This might
180 be more relevant for local products of labaneh and jameed that are more unique to the
181 country of Jordan than other countries and have established procedures for production [29,
182 30].

183 Reduction of AFM1 by high heat treatments (Fig. 2) shows no significant effect after
184 pasteurization and boiling treatments. There was a decrease in AFM1 from 12.5 ppt to 11.5
185 ppt after pasteurization at 72°C for 15 seconds, while boiling at 100°C for 30 seconds had
186 no effect by decreasing AFM1 from 12.5 ppt to only 12 ppt. A significant decrease in AFM1
187 (nearly 56% reduction) was observed after sterilization treatment using the autoclave (121°
188 C for 15 min) which decreased AFM1 from 12.5 ppt to 5.5 ppt. The autoclaving may have
189 partially destroyed the toxin.

* Tel.: 962-65715546 ext. 8413

E-mail address: kaldelaimy@uop.edu.jo

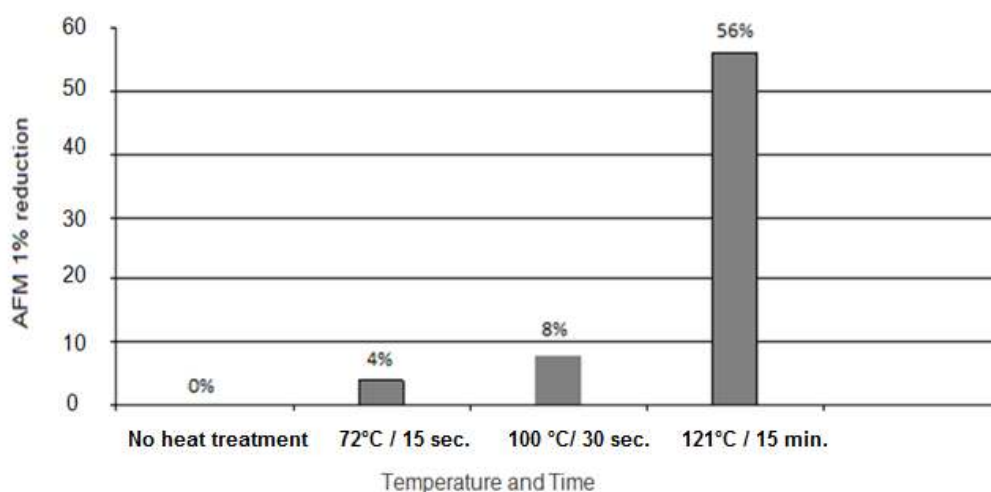


Figure 2: The effect of different heat treatments on the detoxification of aflatoxin M1 in milk measured by percentage reduction for each temperature starting with 12.5 ppt level with no heat treatment and then increasing to pasteurization temperature (72 °C), boiling temperature (100 °C) and autoclaving (121 °C for 15 minutes).

Previous studies [31, 32, 33, 34] also pointed out that pasteurization, boiling and sterilization did not reduce AFM1 concentration significantly. Deveci and Sezgin [35] on the other hand reported a reduction of 16% of AFM1 concentration after pasteurization of cow's milk contaminated artificially with 1.5 µg/L of the toxin. A recent study from India confirmed the inability of boiling to reduce AFM1 in several local milk types [36]. AFM1 in this study and other studies is considered a heat stable toxin.

The effect of fermentation time on the reduction of AFM1 in milk with artificially added toxin is presented in Figure 3. Progressive reduction of AFM1 from 11.5 ppt before fermentation and as the time of fermentation increased to 6 hrs it was reduced to 10 ppt (a 13% reduction), reduced to 9 ppt after 24 hrs (a 22% reduction) and reduced to 7.5 ppt after 48 hrs (a 35% reduction). Partial reduction of AFM1 by milk fermentation was also reported by other workers. The reduction ranged between 12% [37] and up to 97% of total initial AFM1 (600 ppb) added before fermentation of milk to yoghurt. One explanation of the effect of fermentation on AFM1 is the presence of *Lactobacillus* spp in milk that binds to the toxin [38]. Abbes et al. [38] reported that *Lactobacillus rhamnosus* at a level of Log 8/ml in reconstituted milk could remove AFM1 after 24hrs at 30 °C in phosphate buffer saline. *Lactobacillus rhamnosus* has more ability to remove the toxin than *Lactobacillus plantarum*. Similar to our finding, Galvano et al. [39] also found that AFM1 in yoghurt were low. A possible explanation is that low levels of AFM1 in yoghurt samples as well as in jameed are due to the effect of lactic acid formation by lactic acid bacteria acting as an anti-AFM1 during fermentation processes [39]. Results of yoghurt AFM1 content from our study and that of others supports this explanation.

The effect of 10% of each garlic, black cumin and carrot water extracts on the reduction of AFM1 levels during milk fermentation to yoghurt was studied (Fig. 3). Garlic extracts reduced AFM1 from 12 ppt to 9 ppt, a 25% reduction, after a fermentation period of 48 hrs (Fig. 3).

* Tel.: 962-65715546 ext. 8413

E-mail address: kaldelaimy@uop.edu.jo

Garlic extract was reported to significantly reduce the AFB1 [40]. Limited data exists on the effect of garlic on the reduction of AFM1 content in milk and/or during fermentation. The effect of black cumin on the reduction of AFM1 was found to be slightly less than that of garlic extract and at around 20% after fermentation periods of 48 hrs (Fig. 3). This reduction may be partially attributed to the effect of the interaction between fermentation activities and black cumin extract content or the fact that the baseline levels at the start of fermentation was low at only 7.5 ppt. Lower anti-aflatoxin M1 activity with the addition of both garlic and black cumin water extracts compared to the effect of fermentation process alone was observed. It may be speculated that those extracts may have caused metabolic interferences with AFM1 reduction activities. Garlic for example contain alicin, sulfur containing compound as the active ingredient agent. Carrot water extracts seems to be more effective on the reduction of AFM1 than any of garlic and black cumin extracts (Fig. 3). The initial level of AFM1 (12.5 ppt) added to milk before fermentation was reduced to 3.5 ppt (a reduction of 72%) after 48 hrs. Less reduction (8% and 20%) was observed after fermentation periods of 6 hrs and 24 hrs, respectively. It is possible to indicate that the carotenoids compounds in carrots as antioxidants may interact and bind the AFM1 configuration. In agreement with this finding, a study in Egypt [41] showed that adding 5% and 20% carrot juice to yoghurt resulted in a reduction of 20% and 70% of the initial concentration of AFM1, respectively.

This is an important and significant finding that requires further investigation about the potential for carrot juice as an effective anti-aflatoxin M1 (AFM1 reduction).

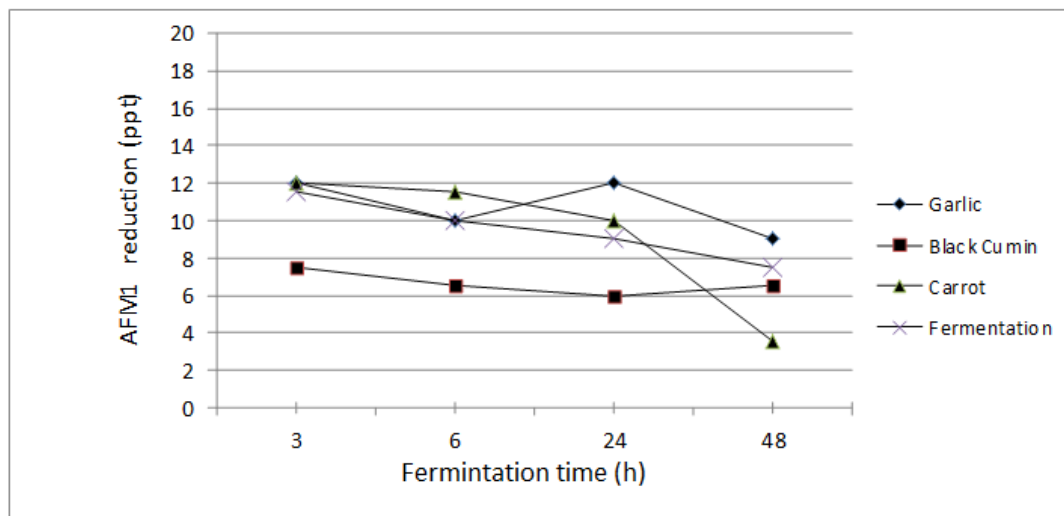


Figure 3: The effect of fermentation time alone, and fermentation plus garlic, black cumin or carrot water extracts on the detoxification of an initial aflatoxin M1 concentration of 12 ppt with garlic, 7.5 ppt with black cumin, 12.5 ppt with carrot and 11.5 ppt with fermentation alone (as control) .

4. CONCLUSION

The AFM1 levels in milk and milk products samples collected in Amman, Jordan were considered to be low (ranging between 25.8-nill ppt.). They were less than the maximum European standard for AFM1 of 50 ppt. Fermentation of milk does decrease AFM1 levels, but carrot water extract added to the fermentation process seemed to have the highest

impact on AFM1 levels after 48 hrs and clearly exceeds that of other extracts of garlic or black cumin. This finding from this preliminary study deserves further studies to determine its use in dairy food technology, especially in areas and countries where very high levels of AFM1 have been found in milk and dairy products.

ACKNOWLEDGEMENTS

This work was self-funded by the authors and the Department of Nutrition at the University of Jordan. No other funding or support was provided by any other entity.

COMPETING INTERESTS

NONE

AUTHORS' CONTRIBUTIONS

“ ‘KA’ DESIGNED THE STUDY, FINALIZED THE MANUSCRIPT AND OVERSAW THE LABORATORY ANALYSES. ‘IM’ PERFORMED THE LABORATORY ANALYSES, MANAGED LITERATURE SEARCHERS AND WROTE THE FIRST DRAFT OF THE MANUSCRIPT. ALL AUTHORS READ AND APPROVED THE FINAL MANUSCRIPT.”

CONSENT (WHERE EVER APPLICABLE)

NOT APPLICABLE

ETHICAL APPROVAL (WHERE EVER APPLICABLE)

NOT APPLICABLE

REFERENCES

1. Asao T, Buchi G, Abdel-Kader, MM, Chang SB, Wick EL, Wogan GN. Aflatoxins B and G. Journal of the American Chemical Society. 1965; 87: 882-86.
2. Williams JH, Phillips TD, Jolly PE, Stiles JK, Jolly CM, Aggarwal D. Human aflatoxicosis in developing countries: a review of toxicology, exposure, potential health consequences, and interventions. American Journal of Clinical Nutrition. 2004; 80: 1106-22.
3. International Agency for Research on Cancer (IARC). Some traditional herbal medicines, some mycotoxins, naphthalene and styrene. Summary of data reported and evaluation.. IARC Monographs on the Evaluation of the Carcinogenic Risk to Humans. Vol. 82. International Agency for Research on Cancer, Lyon, France. 2002.
4. International Agency for Research on Cancer (IARC). Aflatoxins. Some Naturally Occurring Substances: Food Items and Constituents, Heterocyclic Aromatic Amines and Mycotoxins. IARC Monographs on the Evaluation of the Carcinogenic Risk to Humans. Vol. 56. International Agency for Research on Cancer, Lyon, France. 1993.

* Tel.: 962-65715546 ext. 8413

E-mail address: kaldelaimy@uop.edu.jo

- 303 5. Allcroft R, Carnaghan RBA. Ground nut toxicity: *Aspergillus flavus* toxin (Aflatoxin) in
304 animal product. *Veterinary Research*. 1963; 24: 863-64.
- 305 6. Fremy JM, Gautier JP, Herry MP, Terrier C, Calet C. Effects of ammonization on the
306 'carry-over' of aflatoxins into bovine milk. *Food Additives and Contaminants*. 1988; 5:
307 39-44.
- 308 7. Creppy EE. Update of Survey, regulation and toxic effects of mycotoxins in Europe.
309 *Toxicology Letters*. 2002; 127: 19-28.
- 310 8. Codex Alimentarius. Draft maximum level for aflatoxin M1 in milk, Alinorm 97/37
311 report of the twenty-second session of the Joint FAO/WHO Codex Alimentarius
312 Commission, Geneva, 23-28 June, 1997. 2001
- 313 9. Nakajima M, Tabata S, Akiyama H, Itoh Y, Sunagawa H, Tyonan T, et al.
314 Occurrence of aflatoxin M1 in domestic milk in Japan during winter season. *Food*
315 *Additives and Contaminants*. 2004; 21: 472-78.
- 316 10. Temamogullari F, Kanici A. Short communication: Aflatoxin M1 in dairy products
317 sold in Şanlıurfa, Turkey. *Journal of Dairy Science*. 2014; 97: 162-65.
- 318 11. Riahi-Zanjani B, Balali-Mood M. Aflatoxin M1 contamination in commercial
319 pasteurized milk from local markets in Fariman, Iran. *Mycotoxin Research*. 2013; 29:
320 271-74.
- 321 12. Rahimi E. Survey of the occurrence of aflatoxin M1 in dairy products marketed in
322 Iran. *Toxicology and Industrial Health*. 2014; 30: 750-54.
- 323 13. El Marnissi B, Belkhou R, Morgavi DP, Bennani L, Boudra H. Occurrence of
324 aflatoxin M1 in raw milk collected from traditional dairies in Morocco. *Food and*
325 *Chemical Toxicology*. 2012; 50: 2819-21.
- 326 14. Omar SS. Incidence of aflatoxin M1 in human and animal milk in Jordan. *Journal of*
327 *Toxicology and Environmental Health A*. 2012; 75: 1404-9.
- 328 15. Gurses M, Erdouan A, Cetin B. Occurrence of aflatoxin M1 in some cheese types
329 sold in Erzurum, Turkey. *Turkish Journal of Veterinary and Animal Science*. 2004;
330 28: 527-30.
- 331 16. Colak H, Hampikyan H, Ulusoy B, Ergun O. Comparison of a competitive ELISA with
332 an HPLC method for the determination of aflatoxin M1 in Turkish White, Kasar, and
333 Tulum cheeses. *European Food Research and Technology*. 2006; 217: 258-64.
- 334 17. Al-Delaimy KS, Ali SH. Antibacterial action of vegetable extracts on the growth of
335 pathogenic bacteria. *Journal of the Science of Food and Agriculture*. 1970; 21: 110-
336 12.
- 337 18. Yin M, Change W. Inhibition of *Aspergillus niger* and *Aspergillus flavus* by some
338 herbs and spices. *Journal of Food Protection*. 1998; 61: 123-25.
- 339 19. Garcia S, Araiza M, Gemez M, Heredia N. Inhibition of growth, enterotoxin
340 production and spore formation of *Clostridium perfringens* by extracts of medicinal
341 plants. *Journal of Food Protection*. 2002; 65: 1667-69.

* Tel.: 962-65715546 ext. 8413

E-mail address: kaldelaimy@uop.edu.jo

- 342 20. Srivastava V P, Bu-Abbas A, Basuny A, Al-Johar W, Al-Mufi S, Siddiqui MK.
343 Aflatoxin M1 contamination in commercial samples of milk and dairy products in
344 Kuwait. Food Additives and Contaminants. 2001; 18: 993-97.
- 345 21. Dashti B, Al-Hamli S, Alomirah H, Al-Zanki S, Bu-Abbas A, Sawaya W. levels of
346 aflatoxin M1 in milk cheese consumed in Kuwait and occurrence of total aflatoxin in
347 local and imported animal food. Food Control. 2009; 20: 686-90.
- 348 22. Kamkar A. A study on the occurrence of aflatoxin M1 in raw milk produced in Sarab
349 city of Iran. Food Control. 2005; 16: 593-99.
- 350 23. Unusan N. Occurance of aflatoxin M1 in UHT milk in Turkey. Food and Chemical
351 Toxicology. 2006; 44: 1897-1900.
- 352 24. Orfi GM. Aflatoxin M1 in raw, pasteurized and powdered milk available in the Syrian
353 Market. Food Control. 2009; 20: 603-605.
- 354 25. Hussain I, Anwar J, Asi MR, Munawar MA and Kashit M. Aflatoxin M1 contamination
355 in Milk from five dairy species in Pakistan. Food Control. 2010; 21: 122-24.
- 356 26. Thirumala-Devi K, Mayo MA, Hall AJ, Craufurd PQ, Wheeler TR, Waliyar F, et al.
357 Development and application of an indirect competitive enzyme-linked immunoassay
358 for aflatoxin M1 in milk and milk-based confectionery. Journal of Agricultural and
359 Food Chemistry. 2002; 50: 933-37.
- 360 27. Saitanu K. Incidence of aflatoxin M1 in Thai milk products. Journal of Food
361 Protection. 1997; 60: 1010-12.
- 362 28. Guo Y, Yuan Y, Yue T. Aflatoxin M1 in milk products in china and dietary
363 assessment. Journal of Food Protein. 2013; 76: 849-53.
- 364 29. Abu Lehia IH. The chemical composition of jameed cheese. Ecology of Food and
365 Nutrition 1987; 20: 231-239.
- 366 30. Nsabimana C, Jiang B, Kossah R. Manufacturing, properties and shelf life of labneh:
367 a review. International Journal of Dairy Technology. 2005; 58:129-137.
- 368 31. Van Egmond HP. Aflatoxins in milk. In: D.L. Eaton, and J.D. Groopman, (Eds.). The
369 Toxicology of Aflatoxin: Human Health, Veterinary, and Agricultural Significance..
370 San Diego, USA: Academic Press. 1994 (pp. 364-381).
- 371 32. Diaz DE, Hagler WM, Blackwelder, JT, Eve JA, Hopkins BA, Anderson KL, et al.
372 Aflatoxin binders II: reduction of aflatoxin M1 in milk by sequestering agents of cows
373 consuming aflatoxin in feed. Mycopathologia. 2004; 157: 233-41.
- 374 33. Park DL. Effect of processing on aflatoxin. Advances in Experimental Medicine and
375 Biology. 2002; 504: 173-79.
- 376 34. Lorezini G, Martini A, Contini C, Omodei Zorini, L, Riccio F, Cervelin F, et al.
377 Mycotoxins in the milk from organic farms in the Florence. In: Proceedings of the 3rd
378 SAFO Workshops 16-18 September 2004, Falenty, Poland. 2005;PP. 69-82

* Tel.: 962-65715546 ext. 8413

E-mail address: kaldelaimy@uop.edu.jo

- 379 35. Deveci O, Sezgin E. Changes in concentration of aflatoxin M1 during manufacture
380 and storage of skim milk powder. *Journal of Food Protection*. 2006; 69: 682-85.
- 381 36. Awasthi V, Bahman S, Thakur LK, Singh SK, Dua A, Ganguly S. Contaminants in
382 milk and impact of heating: an assessment study. *Indian Journal of Public Health*.
383 2012;56 :95-99.
- 384 37. Govaris A, Roussi V, Koidis PA, Botsoglou NA. Distribution and stability of aflatoxin
385 M1 during production and storage of yoghurt. *Food Additives and Contaminants*.
386 2002;19:1043-50.
- 387 38. Abbès S, Salah-Abbès JB, Sharafi H, Jebali R, Noghabi KA, Oueslati R. Ability of
388 *Lactobacillus rhamnosus* GAF01 to remove AFM1 in vitro and to counteract AFM1
389 immunotoxicity in vivo. *Journal of Immunotoxicology*. 2013;10:279-86.
- 390 39. Galvano F, Galofaro V, De Angelis A, Galvano M, Bognanno M, Galvano G. Survey
391 of the occurrence of aflatoxin M1 in dairy products marketed in Italy. *Journal of Food*
392 *Protection*. 1998;61:738-41.
- 393 40. Belloir C, Singh V, Daurat C, Siess MH, Le Bon AM. Protective effects of garlic
394 sulfur compounds against DNA damage induced by direct- and indirect acting
395 genotoxic agents in HepG2 cells. *Food and Chemical Toxicology*. 2006;44: 827-34.
- 396 41. Salwa A, Aly-Galal EA, Elewa NA. Carrot yoghurt: sensory, chemical,
397 microbiological properties and consumer acceptance. *Pakistan Journal of Nutrition*.
398 2004;3:322-30.

399

* Tel.: 962-65715546 ext. 8413

E-mail address: kaldelaimy@uop.edu.jo