

# Original Research Article

## Population dynamics of metazoan parasites of *Rhizoprionodon acutus* from Nellore Coast off Bay of Bengal.

**Short running title:** Population dynamics of metazoan parasites of *Rhizoprionodon acutus*

### Abstract:

*Rhizoprionodon acutus* Rüppell 1837, commonly known as 'Milk shark' is the frequently available elasmobranch from the Nellore (14.43°N 79.97°E.) coast off Bay of Bengal. A total of 152 *R. acutus* were collected from this coast during January, 2014- December, 2015, of which 89 hosts were infected with one or more parasites. A total of eleven species were collected, comprising of 6 cestodes, 2 nematodes, one monogenic, one copepod and one isopod. Various ecological parameters such as prevalence, mean intensity, mean abundance and index of infection were calculated to determine the monthly population dynamics and the seasonal dynamics of the parasites in *R. acutus*. The study was carried out for both overall and groupwise parasitization. Endoparasitic infection predominated the ectoparasitic infection in the host. The study reveals the role of the temperature and season in the recruitment of parasite fauna in the hosts. The present study would be a great contribution to the knowledge of the metazoan parasite fauna of elasmobranch fishes to the future helminthologists from this Nellore Coast, Bay of Bengal.

**KEYWORDS:** Population dynamics, metazoan parasites, *Rhizoprionodon acutus*, prevalence, mean intensity, mean abundance.

### Introduction:

Parasites are an imperative part of a distinct operating ecosystem [1]. Just as every other organism has a part within the ecosystem; parasites also have an ecological niche. Their niche includes the resources and space of the host organism's body and the abiotic conditions they survive in while completing their life cycle. These aquatic parasites are acquiring

potential attention in ecological point of view due to their interrelationship with their hosts. Further, their role as ‘biological tags’ has attracted many scientists to use them as sensitive probes to monitor changes in the environmental factors. But on the other hand, parasites affect fish health, growth, behavior, fecundity and mortality and also regulate host population dynamics and their community structure [1]. As a parasite develops an ecological association with a particular host, there may be host specificity but also an immune reaction by the host. Site specificity within the host indicates parasitic adaptation to its environment. Elasmobranchs serve as a very good host for all types of metazoan parasites especially cestodes. *Rhizoprionodon acutus* commonly known as ‘Milk Shark’ and vernacularly as ‘Kukkasorrah’ and ‘Pala sorrah’ in Southern India especially, Andhra Pradesh is believed to improve the milk production of a human mother in some localities of Southern India. It is a coastal species observed in a broad range of artisanal, survival, profitable fisheries and recurrently spotted in fish markets but it is assessed as ‘Least Concern’ due to their extensive distribution and moderately productive life history [2]. Data on the population dynamics of helminth parasites of sharks are very scarce. Seasonal variations in the parasitic infections are very frequent in tropical waters. The occurrence of some species is throughout the year while some other species are restricted to particular season. [3] Opined that seasonal changes of water such as temperature, pH and conductivity severely influence the incidence of parasites in aquatic hosts. Several eminent authors like [4-18] also discussed the role of certain environmental factors on the incidence of parasitization. The present study was focused on the population dynamics of both ectoparasitic and endoparasitic helminth parasites. Even today there is dearth in our knowledge in this field, mainly the contributions in the ecology of elasmobranch fish parasites are very meager and there is a big lacuna in this field and many more studies are very much required to fill this gap. Hence, the present study is a genuine attempt to interpret the status of parasitic communities of *R. acutus* in terms of prevalence, mean intensity, mean abundance, standard deviation and index of infection and also to study the seasonal impact on the incidence of parasitization.

## Materials and Methods:

In the present study, various ecto and endoparasites of *Rhizoprionodon acutus* were collected from Nellore (14.43°N 79.97°E.) coast Bay of Bengal, India (Plate-1; Fig-1). The collections were made for a period of 2 years i.e., from January, 2014-December, 2015. A total of 152 *R. acutus* were obtained from fishing landing centers and local fish markets along the coast (Fig-1). Monthly an average of 5-10 fishes were collected and brought to the laboratory.

Before dissecting the fish, various morphometric characters such as sex, weight and length were noted and thoroughly examined for ectoparasites from various possible organs like eyes, buccal cavity, skin and gills. The collected ectoparasites like Copepods and isopod parasites were fixed in 10% formalin. For the identification purpose, these parasites were cleared in a few drops of lactic acid for 12– 24 hours. Lactic acid acts as a good clearing solution and other monogenetic trematodes were collected with the help of small pipettes worms are kept in the center of the slide in a small drop of water and a coverslip is placed and then water is removed using filter paper and glycerine is added underneath the coverslip and all the four corners are sealed with a nail enamel. The visceral organs like heart, stomach, liver, intestine and air bladder were also examined thoroughly for the endoparasites. The spiral intestine were kept in petri-dishes filled with physiological saline and were dissected with a longitudinal incision and the intestinal mucosal spirals were unrolled as a flat sheet. Then they were shaken thoroughly in order to dissipate the gut contents. The gut contents were decanted many times to remove the excess mucous and observed under stereozoom microscope (LM-52-3621 Elegant) for parasites. The collected cestode parasites were kept between two slides or with a cover glass for proper pressing. During this process, proper care was taken to avoid any damage to the parasite. Properly pressed parasites were preserved in FAA (Formalin-10ml, Alcohol-85ml, and Acetic acid-5ml). Later, these parasites were washed thoroughly and stained with alum carmine. After proper dehydration in alcoholic grades (70%, 90%, 95% and absolute alcohol), the parasites were cleared in xylene and mounted in Canada balsam by the conventional techniques adopted from [19-20]. Nematodes collected from different locations such as liver, intestine and coelom were preserved, in 70% ethanol or less fluid (9 parts 70% ethanol and 1 part glycerol). These parasites uncoil and killed instantaneously and were finally transferred into bottles containing a mixture of 70% ethanol and glycerol (3:1). Standard biostatistical books by [21-23] were followed. Ecological terminologies were adapted from [24-26].

**Prevalence:** Prevalence is the number of individuals of the hosts infected with particular parasite species (or) with total parasites divided by the number of hosts examined. Prevalence is expressed in terms of percentage (%).

$$\frac{\text{Number of individuals of a host species infected with particular parasite species (or) Number of individuals of a host species infected with total parasites}}{\text{Number of individuals of a host species examined}} \times 100$$

**Prevalence** (usually expressed as %) = \_\_\_\_\_

98 Number of hosts examined

99 **Mean Intensity:** Mean intensity is the average intensity of total number of individuals of  
 100 particular parasite species in a sample of host species or total number of individuals of all  
 101 parasites found in a sample of host species divided by the number of hosts infected with that  
 102 parasite or the total number of parasites.

$$\begin{aligned}
 &\text{Total number of individuals of a particular parasite species in a sample of a host species} \\
 &\quad \text{(or)} \\
 &\text{Total number of individuals of all parasites in a sample of a host species} \\
 107 \text{ **Mean intensity} = & \frac{\quad}{\quad} \\
 108 &\text{Number of infected individuals of the host species in the sample} \\
 109 &
 \end{aligned}**$$

110 **Mean Abundance:** Mean abundance is the total number of individuals of a particular  
 111 parasite species in a sample of particular host species divided by the total number of hosts of  
 112 that species examined (including both infected and uninfected hosts).

$$\begin{aligned}
 &\text{Number of individuals of a parasite in a sample of host} \\
 115 \text{ **Mean abundance} = & \frac{\quad}{\quad} \\
 116 &\text{Total number of individuals of the host species (infected + uninfected)} \\
 117 &
 \end{aligned}**$$

118 **Index of infection:** Index of infection is the product of number of individuals of total  
 119 parasitic species and the number of infected fish in a sample of host species divided by the  
 120 square of total number of fish examined in the sample (infected and uninfected).

$$\begin{aligned}
 &\text{Total number of infected hosts (b) X Total number of parasites in a sample of host species (c)} \\
 123 \text{ **Index of infection} = & \frac{\quad}{\quad} \\
 124 &\text{square of total number of hosts examined (a}^2\text{)} \\
 125 &
 \end{aligned}**$$

126 To determine the seasonal influence on the parasitic infection, each annual cycle was  
 127 catalogued into three seasons: **Summer** (March to June), **rainy** (June to September) and  
 128 **winter** (October to January) for expediency of calculation, though there is no significant  
 129 division between one season and the other. A chi-square test was performed to test the  
 130 significance between the season and the incidence of infection (Prevalence)  
 131 ([www.socscistatistics.com](http://www.socscistatistics.com)).

132

133

134 **Results:**

135 Total 152 fishes were examined, out of which 89 were found to be infected. 5 species of  
 136 metazoan parasites were collected, comprising 6 cestodes, 2 nematodes and each 1 of  
 137 Monogenean, copepod and isopod (Table-1).

138 **Monthly population dynamics of overall parasites in *Rhizoprionodon acutus***

139 The month-wise prevalence, mean intensity, mean abundance and index of infection of  
 140 metazoan parasites in *Rhizoprionodon acutus* were presented graphically.

141 **Prevalence of parasitization with total parasites during the years 2014-15 (Plate-2, Fig.**  
 142 **1a)**

143 The prevalence of infection was nil in the first six months and raised in July and reached to  
 144 the peak in the months of August, September, October and November (100 %) and slightly  
 145 declined in the month of December. However, for the 2015 cycle, first 3 months showed  
 146 highest prevalence (100 %) and after that in the months of May and December it shows  
 147 highest prevalence and it was moderate during the months of July to September and shows  
 148 lowest in the Months of April, June, October and November.

149 **Mean intensity, mean abundance and index of infection of parasitization with total**  
 150 **parasites during the years 2014-15 (Plate-2, Fig. 1b, 1c & 1d)**

151 Mean intensity was highest in the months of August, October to November,  
 152 and reaches to a peak in the month of December (5.9) and shows lowest in the month of July  
 153 for 2014 cycle. While in 2015 cycle Mean intensity was moderate during the months of  
 154 January to April, shows lowest in the month of May and reaches to highest in the months of  
 155 August (7.7) and somewhat declined during September to December. Mean abundance was  
 156 highest in the month of December (5.1) in the year 2014 and after that it shows highest values  
 157 in the month of August and November and lowest in the month of July. In the year 2015 it  
 158 shows highest mean abundance in the month of August (6.8) and it was lowest in the month  
 159 of April. Index of infection was high in the month of August (4.87), and showed highest  
 160 during the months of October to December for 2014 cycle and lowest in the month of July.  
 161 However, for the 2015 cycle, Index of infection was highest in the month of August (5.90)  
 162 and lowest in the month of April.

**Prevalence, Mean intensity, Mean abundance and Index of Infection of parasitization with Monogenean parasites (Plate-3, Fig. 1a, 1b, 1c & 1d)**

Prevalence of Monogenean parasite is highest in the month of December in the year 2015, and it shows zero prevalence from February to July in both the years. In the month of August in both the years and in December 2014 it shows moderate values. Mean intensity of monogenean parasites reaches to a peak in the month of August in the year 2015 and its mean intensity is zero from February to July in both the years. When compared to 2014 in the year 2015 there is slight increase in the mean intensity values. Same like mean intensity, mean abundance also shows highest abundance in the August of 2015 and zero abundance from February to July in both the years. Infection of monogenean parasite is almost zero except for August and December of the year 2014. In the year 2015 its infection reaches to a peak in the month of August and shows moderate infection in the months of January and November and observed zero infection in the remaining months.

**Prevalence, Mean intensity, Mean abundance, Index of Infection of parasitization with cestode parasites (Plate-4, Fig. 1a, 1b, 1c & 1d):**

Prevalence of parasitization with cestodes showed some variations in both the years. Zero infection was recorded during Jan to June in 2014 and was higher during the months of July to December in both the years 2014 and 2015. Besides this, there is a coincidence in both the years during the months of July to December. Mean intensity of cestodes reaches to peak in the month of August in the year 2015 and it was higher from November to December and moderate during September and October in both the years and it shows zero intensity from January to July in the year 2014. Mean abundance of cestodes shows zero abundance from January to June in the year 2014 and reaches to peak in the month of August in the year 2015 and it was higher from November to December and modest during July to October in both the years and there is a coincidence in the month of September in both the years. Index of infection of cestode parasites is zero from January to June in the year 2014 and slowly raises and reaches to a highest value in the months of November and December. In the year 2015 it shows moderate from January to July and reaches to a peak in the month of August.

**Prevalence, Mean intensity, Mean abundance, Index of Infection of parasitization with Nematodes (Plate-5, Fig. 1a, 1b, 1c & 1d):**

Nematodes showed zero prevalence in the months of April to August in both the years and reaches to higher in the months of September and October and moderate during the months of November and December in both the years. Mean intensity of nematodes is higher in the months of January and March in the year 2015 and however it also shows higher mean intensity values in the months of November and December in both the years 2014 and 2015. Zero intensity was observed from April to September in both the years. Mean abundance reaches to a peak in the month of October during the first year and it shows higher in the months of January and November in the year 2015 and shows moderate in the month of December in both the years and mean abundance was zero from April to September in both the years. Infection of nematode parasites is zero from January to September of the year 2014 and where as it reaches to a peak in the October month and shows moderate in the months of November and December, where as in the year 2015 it reaches to higher in the months of January and November and observed moderate in march and December and almost shows zero infection in the remaining months.

**Prevalence, Mean intensity, Mean abundance and Index of Infection of parasitization with copepod (Plate-6, Fig. 1a, 1b, 1c & 1d):**

Prevalence of copepod reaches to a maximum in the month of December in the year 2014 and shows higher in the month of October and November in the year 2015 it was moderate in the months of august and December in the year 2015. And almost observed zero prevalence from April to July in both the years. Zero Intensity of parasitization is observed from April to July in both the years, and reaches to a peak in the month of March & December and it was highest in the month of august and shows moderate intensity from September to November in the year 2015. Except, December it almost shows zero abundance in the year 2014 whereas it reaches to a peak in the month of august and shows moderate from September to November in the year 2015. However it is observed slight difference in abundance in the month of December in both the years. Except, December remaining all months of the year 2014 showed zero infection of copepod parasites. In the year 2015, 5 months shows zero infection of copepod parasites and remaining 7 shows moderate infection of copepod parasite.

**Prevalence, Mean intensity, Mean abundance and index of infection of Isopods (Plate-7, Fig. 1a, 1b, 1c & 1d):**

Isopods were obtained only once in the month of August, 2014 and in rest of the months, the infection was nil. Hence, prevalence, mean intensity, mean abundance and index of infection

of Isopod parasites were shown only in the month of August, 2014 and remaining rest of the months it showed zero value. There was no infection of isopods in the year 2015.

## Seasonal dynamics of parasites in *Rhizoprionodon acutus*

### Influence of Seasons on the parasitic infection in *Rhizoprionodon acutus*:

The impact of seasons on the incidence of parasitization showed mixed results as the first the chi-square value ( $\chi^2 = 19.63$ ,  $p=0.000055$ ) for annual cycle 2014 at 5% level of significance and 2 degrees of freedom shows that there might be a significant impact of seasons on the parasitization. However, the chi-square value ( $\chi^2 = 0.1700$ ,  $p=0.918$ ) for annual cycle 2015 at 5% level of significance and 2 degrees of freedom shows that there is no influence of seasons on the parasitization. Prevalence of infection was high during rainy season for the year 2014 and lowest during the summer season whereas, during the year 2015 prevalence was high in the winter season. (Table-2; Plate-8, Fig. 1a & 1b). Recruitment of the parasites may take place after summer and reach their peak periods in the winter months.

### Discussion:

The overall prevalence, mean intensity, mean abundance and index of infection showed very less similarity during the two annual cycles with significant deviations. First six months of the of the first annual cycle showed no infection and remaining six months showed moderate to high infection rate while the second annual cycle showed seasonal changes with highest infection rates in rainy and winter seasons and least in summer season. These disparities may be due to host density and its feeding behavior. Elasmobranchs were infected with large number of parasites during the winter months; parasitization being more in winter months than other seasonal months. Temperature is considered as one of the crucial factors in determining the seasonal periodicity of parasitic infection[13,16,][27-35]. The environmental conditions of tropical waters are quite favorable in winter months where the waters are warm but not too cold. [29] also expressed the same view that infections are more in warm seas than in colder ones. At moderate temperatures the zooplankton fauna may be rich when compared to high temperatures of summer months in tropical areas. The sea is remains quiet and calm with very few disturbances during the winter months; hence the recruitment of infection may take place after summer and reach to its peak in winter months. Thus, temperature and season plays a crucial role in the recruitment of parasite fauna. The present study comes closer to the views of the [27-29] where the prevalence of *R. acutus* was



relatively high in the rainy and winter season than the summer season. In the present study, endoparasitic infection dominated the parasitic communities than the ectoparasites. The present study showed highest cestode infection which was correlated with the experimental studies of [36] who illustrated that the cestode parasites in a fish can survive for longer period at low temperature, but with the increase in water temperature, parasitic infections gets eliminated and thus, the temperature is a major controlling factor of seasonal periodicity of infection. Thus, parasites recruitment within the host is totally dependent on the temperature which might influence the seasonality of parasitic infections either directly or indirectly [37-38]. Thus it can be concluded that the present work showed an evident variations in the population dynamics statistics

## Conclusion

Marine fishes are very significant in commercial fisheries in various parts of the world including India. The study has been conducted for two consecutive years to depict the nature of occurrence of parasites within *R. acutus*. This study has put forward very fine results since the overall parasitization and groupwise parasitic infestation within the host showed less uniformity for both the consecutive years. The temperature and season plays a crucial role in the recruitment of parasite fauna. In the present study, the prevalence of infection, mean intensity, mean abundance, index of infection were high during the months of July to December due to low temperature and low during the months of January-June because of high temperature.

**Significance Statement:** This study discovers the fact that the abiotic factors such as temperature play a significant role in determining the parasitic infestation within the hosts.

## References:

1. Marcogliese DJ. Parasites: Small Players with Crucial Roles in the Ecological Theatre. *Ecohealth*. 2004; 1(2): 151-164.
2. Simpfendorfer CA. Abundance, movement and habitat use of the smalltooth sawfish. Final Report to the National Marine Fisheries Service, Grant number WC133F-02-SE-0247. Mote Marine Laboratory Technical Report. 2003; (929).
3. Dogiel VA, Petrushevski GK, Polyanski Yu I. In Parasitology of fishes. English translation, Oliver and Boyd, Edinburgh and London. 1961.

- 287 4. Bauer ON. Ecology of the parasites of fresh-water fish. Interrelationships between the  
288 parasites and its habitat. Izvestiya Gosudarstvennogo Nauchno-Issledovatel'skogo Instituta  
289 Ozer'nogo i Rechnogo Ribnogo Khozyaistva. 1959; 49, 5-206.
- 290 5. Kennedy CR. Ecological Animal Parasitology. Blackwell Scientific Publications, Oxford,  
291 London, Edinburgh and Melbourne. 1975.
- 292 6. Chubb JC. Seasonal occurrence of helminthes in freshwater fishes. Part I. Monogenea. In  
293 Dawes, Ben (Editor). Advances in parasitology. Vol. 15. London, UK; Academic Press Inc.  
294 (London) Ltd. 1977; 133-199. [En.]
- 295 7. Evans NA. The occurrence of *Sphaerostoma bramae* (Digenea : Allocreadiidae) in the roach  
296 from the Worcester Birmingham Canal. J.Helminthol. 1977; 51: 189-196.
- 297 8. Moller H. The effects of salinity and temperature on the development and survival of fish  
298 parasites. J.Fish. Biol. 1978; 12:311-324.
- 299 9. Mc Cullough JS, Fairweather I, Mont Gomery WI. The seasonal occurrence of *Trilocularia*  
300 *acantiaevulgaris* (Cestoda : Tetraphyllidae) from the spiny dog fish in the Irish Sea. Parasitol.  
301 1986; 93 (1):53-162.
- 302 10. Muralidhar A. Seasonal variation of helminth parasites in marine fishes at east coast of  
303 India. Ind. J. Helminthol. 1989; 41(1):1-4.
- 304 11. Timi JT, Lanfranchi AL, Etchegoin JA. Seasonal stability and spatial variability of  
305 parasites in Brazilian sandperch *Pinguipes brasilianus* from the Northern Argentine Sea:  
306 evidence for stock discrimination. J. Fish Biol. 2009; 74: 1206–1225.
- 307 12. Vidal-Martinez VM, Pech D, Sures B, Purucker ST, Poulin R. Can parasites really reveal  
308 environmental impact? Trends Parasitol. 2010; 26: 44–51.
- 309 13. Gudivada M, Anu prasanna V. Population dynamics of metazoan parasites of marine  
310 threadfin fish, *Polydactylus sextarius* (Bloch and Schneider, 1801) from Visakhapatnam  
311 Coast, Bay of Bengal. The Bioscan. 2010; 5(4): 555-561.
- 312 14. Carvalho AR, Luque JL. Seasonal variation in metazoan parasites of *Trichiurus lepturus*  
313 (Perciformes, Trichiuridae) of Rio de Janeiro, Brazil. Brazilian Journal of Biology. 2011;71  
314 (3): 771-782.

- 315 15. Palm HW. Fish Parasites as biological indicators in a Changing World: Can We Monitor  
316 Mehlhorn, H., (ed.), Environmental Impact and Climate Change?. Progress in Parasitology,  
317 Parasitology Research Monographs. 2011; 2:223-255.
- 318 16. Gudivada M, Anu prasanna V, Vijayalakshmi C. Population dynamics of metazoan parasites  
319 of marine threadfin fish, *Eleutheronema Tetradactylum* (Shaw, 1804) from Visakhapatnam  
320 Coast, Bay of Bengal. Cibttech Journal of Zoology. 2012; 1(1): 14-32.
- 321 17. Pickering M. and. Caira JN. Seasonal dynamics of the cestode fauna in spiny dogfish,  
322 *Squalus acanthias* (Squaliformes: Squalidae) Parasitology. 2014; 141(7 ):940-947.  
323
- 324 18. Schade FM, Raupach MJ, Wegner KM. Seasonal variation in parasite infection patterns of  
325 marine fish species from the Northern Wadden Sea in relation to interannual temperature  
326 fluctuations. Journal of Sea Research. Online. 2015;1-12.
- 327 19. Hiware CJ, Jadhav BV, Mohekar AD. Applied parasitology. A practical manual.  
328 Mangaldeep Publications, Jaipur. 2003.
- 329 20. Madhavi R, Vijayalakshmi C, Shyamasundari K. Collection, staining and identification of  
330 different helminth parasites. A manual of the workshop on fish parasites-taxonomy capacity  
331 building. Andhra University Press, India. 2007.  
332
- 333 21. Sundar Rao PSS, Richard J. In An Introduction to Biostatistics – a Manual for Students in  
334 Health Sciences. Asoke K Gosh Publishers, New Delhi. 1999.
- 335 22. Daniel WW. Biostatistics: A foundation for analysis in the Health sciences (VIII Ed.), John  
336 Wiley and Sons Int., New York. 1998.
- 337 23. Sokal RR. and Rohlf, F.J., 2000. Biometry. The Principles and practice of statistics in  
338 biological research, 2nd ed. W.W. Freeman and company, San Francisco. 859 p.
- 339 24. Margolis L, Esch GW, Holmes JC, Kuris AM, Schad GA. The use of ecological terms in  
340 parasitology (Report of an Ad hoc Committee of the American Society of Parasitologists).  
341 J.Parasitol. 1982; 68 (1):142-144.
- 342 25. Grabda-Kazubski B, Baturo-Warsza-wska B, Pojmanska T. Dynamics of parasite  
343 infestations of fishes in Lakes Dgal Wielki and Warnaik in connection with introduction of  
344 phytophagous species. Acta Parasitol. 1987; 32:1–28.
- 345 26. Bush AO, Lafferty KD, Lotz JM, Shostak AW. Parasitology meets ecology on its own  
346 terms : Margolis et al., Revisted. J.Parasitol. 1997; 83(4):575-583.

- 347 27. Kennedy CR. The regulation of fish parasite populations. In: Regulation of parasite  
348 populations. 1977a; 61-109.
- 349 28. Kennedy CR. Freshwater fish parasites and environmental quality: an overview and caution.  
350 Parassitologia. 1997; 39:249-254.
- 351 29. Rohde K. "Ecology of Marine Parasites". An Introduction to Marine Parasitology ;  
352 Department of Zoology, Univ. of New England, Armidale, NSW 2351, Australia CAB  
353 International ISBN 0851988458. 1993.
- 354 30. Rodrigues AA, Saraiva A. Spatial distribution and seasonality of *Pseudodactylogyrus*  
355 *anguillae* and *P.bini* (Monogenea) on the gills of the European eel *Anguilla anguilla* from  
356 Portugal. *Bull. of the European Asso. of fish Pathologists*. 1996; 63(3):85-88.
- 357 31. Turner HM. Seasonality of *Alloglossoides cardiacola* (Trematoda :Macroderiodea) infection  
358 in the cray fish, *Procambarus acutus*. *South western Naturalist*. 2000; 45(1):69-71.
- 359 32. Wang GT, Yao WJ Nie P. Seasonal occurrence of *Dollfustrema vaneyi* (Digenea :  
360 Bucephalidae) mataceraria in the bull head cat fish *Pseudobagnus fluvidraeo* in the reservoir  
361 in china. *Diseases of Aquatic organisms*. 2001;44(2): 127-131.
- 362 33. Gupta N, Singhal P, Gupta DK. Population dynamics of a parasite *Pallisentis* in two species  
363 of fish *Channa punctatus* and *Channa striatus*. *Journal of Environmental Biology*. 2012; 35,  
364 195-199.
- 365 34. Anu prasanna V, Srinivasa Kalyan C, Hemalatha M. Aspects of ecology of tetraphyllid  
366 cestodes from the Slender Bamboo Shark, *Chiloscyllium indicum* Gmelin, 1789  
367 (Orectolobiformes: Hemiscyllidae) from Nellore Coast, Bay of Bengal, India. *South Asian*  
368 *Journal of Life Sciences*. 2015; 3(2): 42-50.
- 369 35. Dhanya P, Amina S. Parasitic infestation on different regions of *Tilapia mossambica*  
370 collected from Kayamkulam Estuary, Kerala. *International journal of fisheries and Aquatic*  
371 *Studies*. 2017; 5(4): 265-267.
- 372 36. Kennedy, C.R., 1971. The effect of temperature upon the establishment and survival of the  
373 cestode *Caryophyllaeus laticeps* in orfe, *Leuciscus idus*. *Parasitol*. 63, 59-66.
- 374 37. Sanchis V, Roig JM, Carretero MA, Roca V, Llorente GA. Host-parasite relationships of  
375 *Zootoca vivipara* (Sauria: Lacertidae) in the Pyrenees (North Spain). *Folia Parasit*. 2000:  
376 47,118-122.

38. Mas-Coma S, Valero MA, Bargues MD. Fascioliasis. Adv Exp Med Biol. 2014;766, 77-114.

# PLATE-1



Fig-1: Fish landing centers of Nellore Coast

PLATE-2

Monthly population dynamics of overall parasites in *Rhizoprionodon acutus*

Fig.1a-Prevalence (%), Fig.1b- Mean intensity, Fig.1c- Mean abundance, Fig.1d- Index of infection

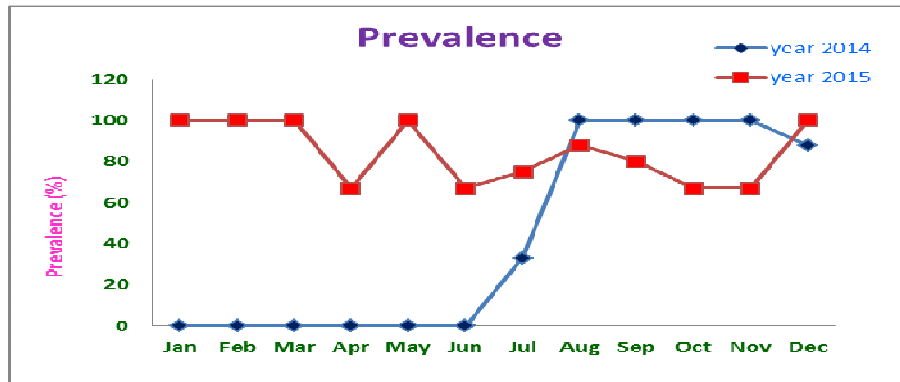


Fig.1a

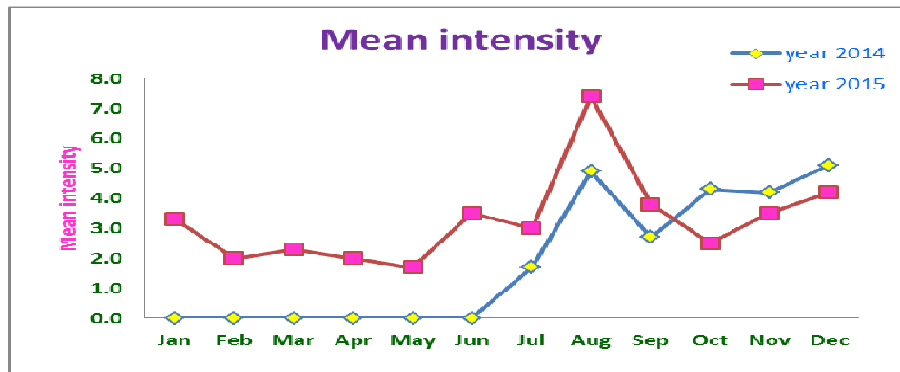


Fig.1b

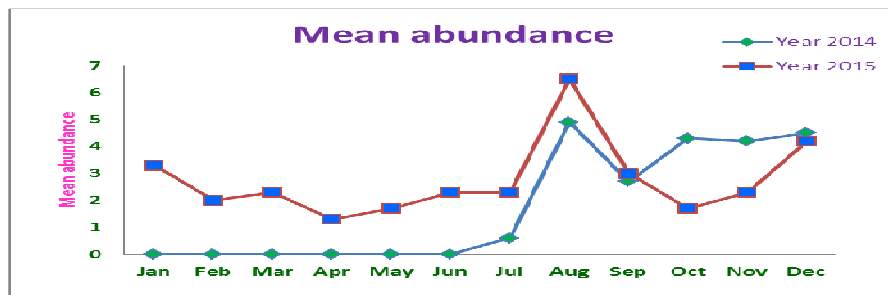


Fig.1c

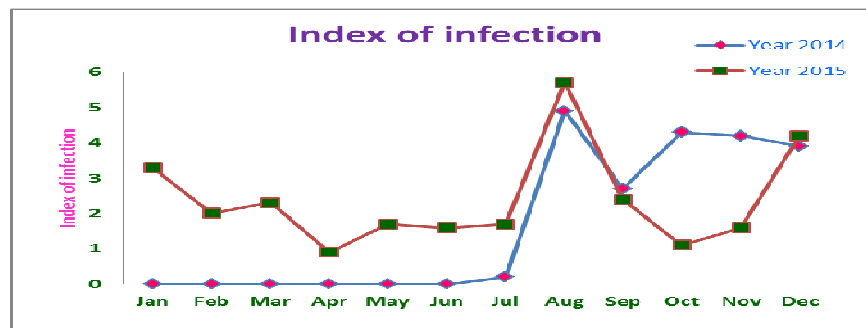


Fig.1d

### Plate-3

#### Monthly population dynamics of monogenean parasites in *Rhizoprionodon acutus*

Fig.1a- Prevalence (%), Fig.1b- Mean intensity, Fig.1c- Mean abundance, Fig.1d- Index of infection

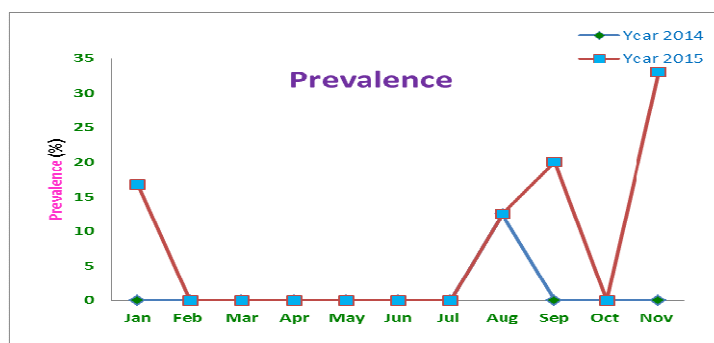


Fig.1a

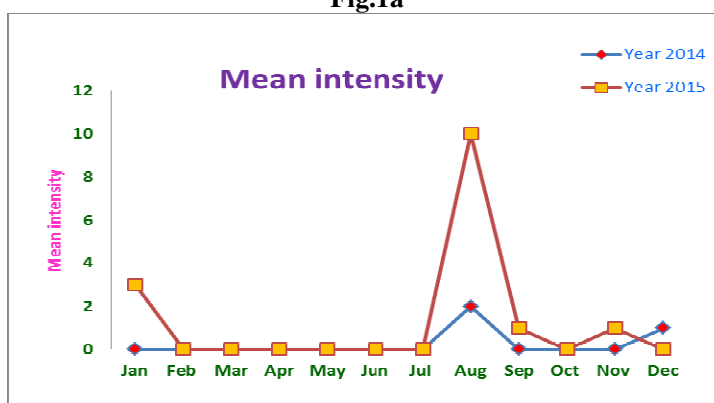


Fig.1b

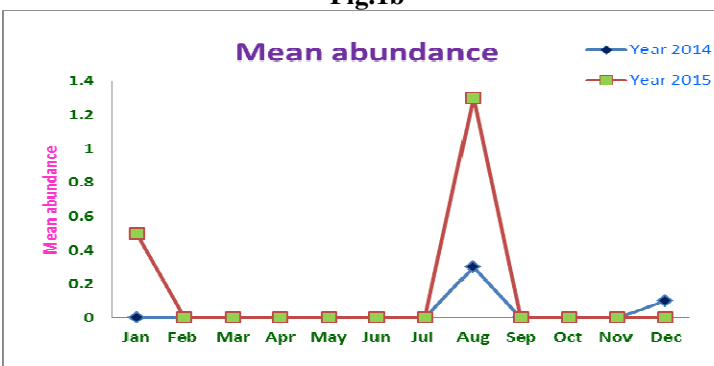


Fig.1c

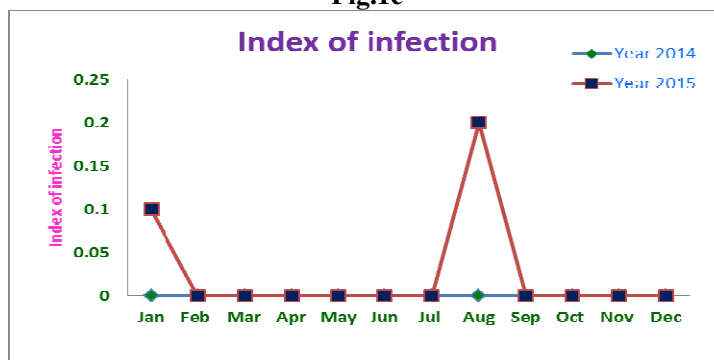


Fig.1d

Plate-4

Monthly population dynamics of cestode parasites in *Rhizoprionodon acutus*  
 Fig.1a- Prevalence, Fig.1b- Mean intensity Fig.1c- Mean abundance, Fig.1d- Index of infection

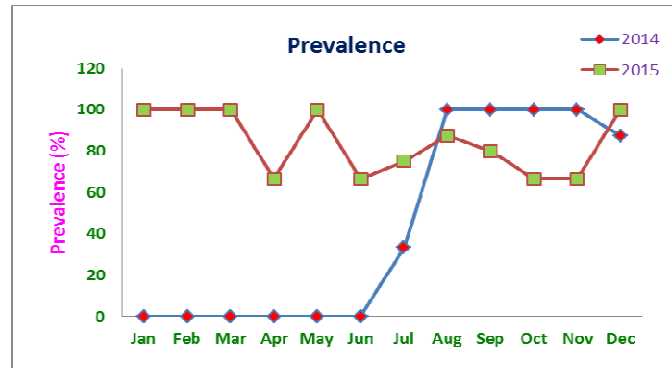


Fig.1a

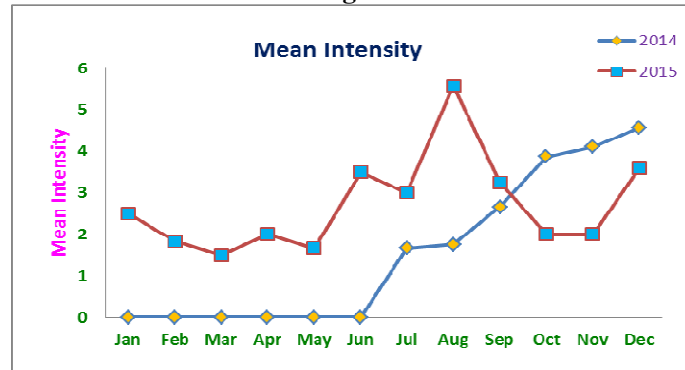


Fig.1b

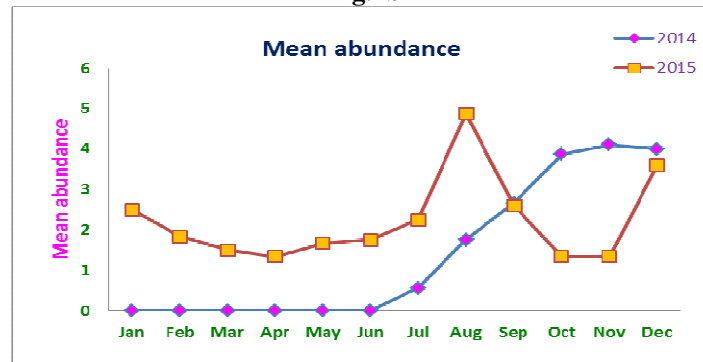


Fig.1c

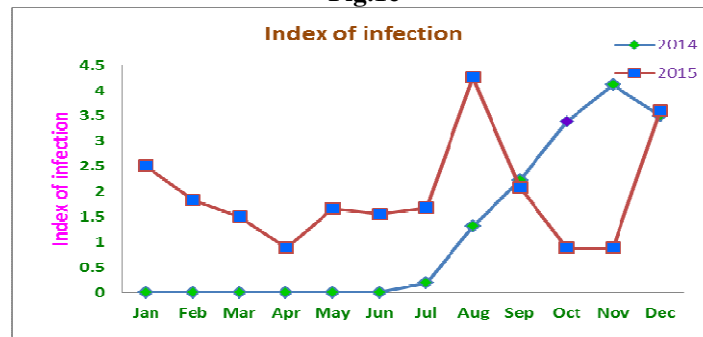


Fig.1d



Plate-5

Monthly population dynamics of nematode parasites in *Rhizoprionodon acutus*  
 Fig.1a- Prevalence, Fig.1b- Mean intensity, Fig.1c- Mean abundance, Fig.1d- Index of infection

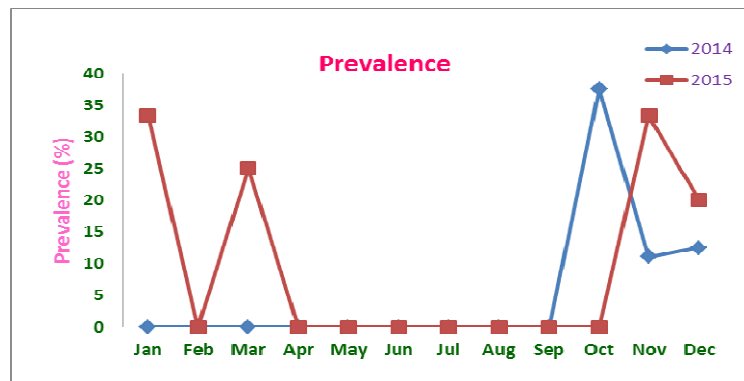


Fig.1a

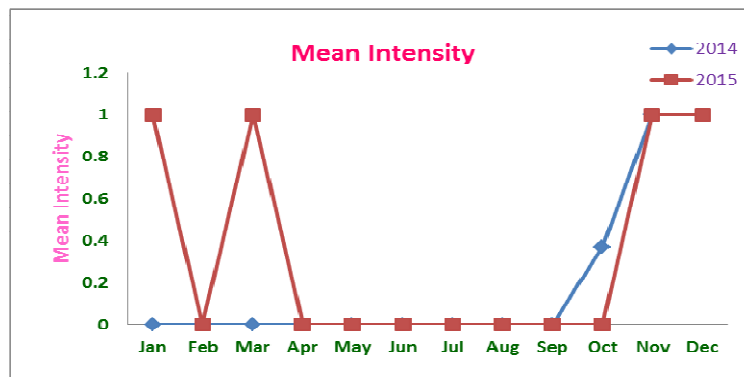


Fig.1b

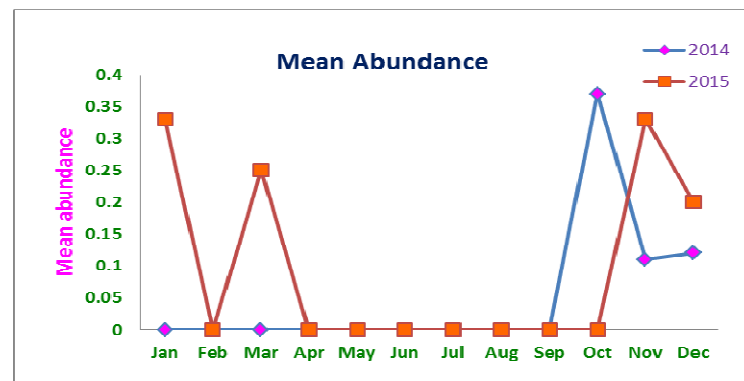


Fig.1c

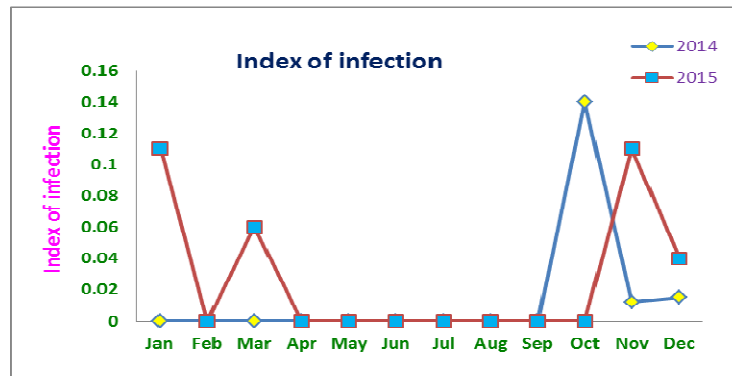


Fig.1d

# Plate-6

## Monthly population dynamics of copepod parasites in *Rhizoprionodon acutus*

Fig.1a- Prevalence, Fig.1b- Mean intensity, Fig.1c- Mean abundance, Fig.1d- Index of infection

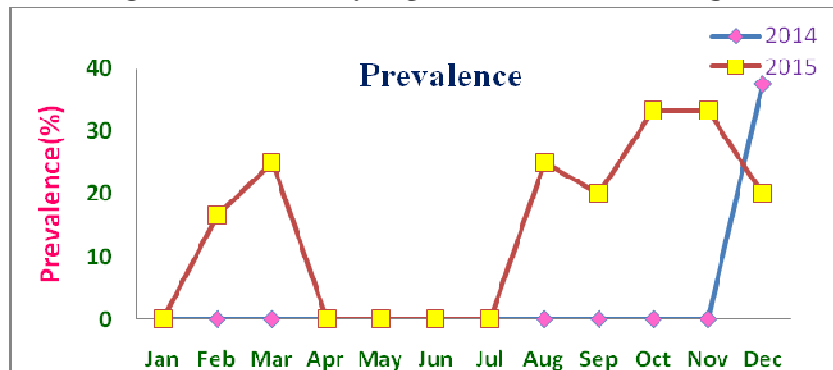


Fig.1a

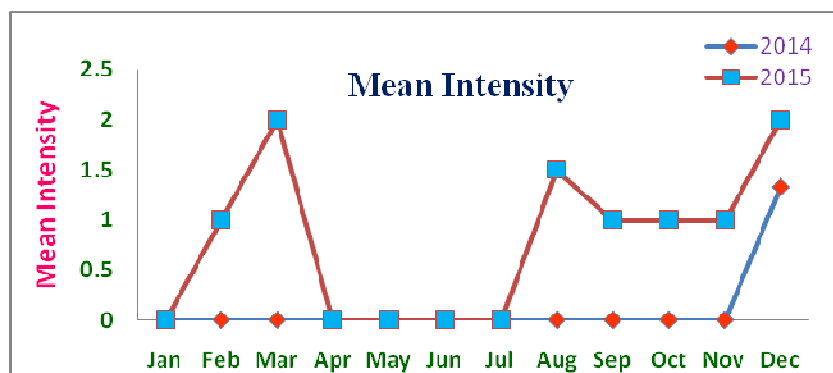


Fig.1b

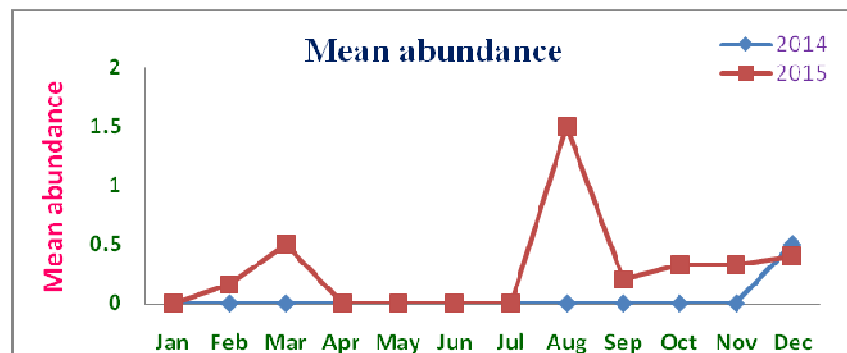


Fig.1c

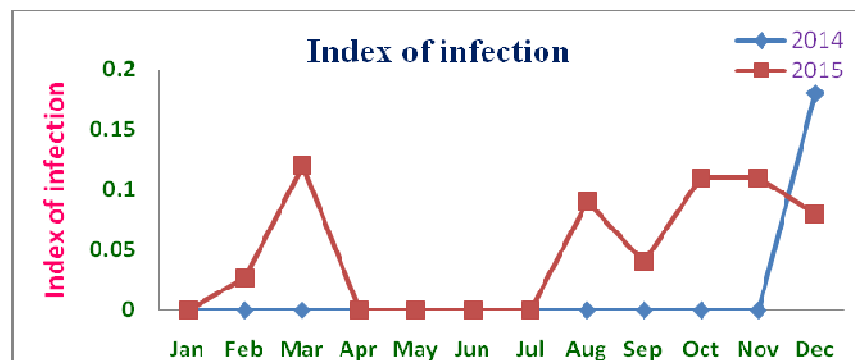


Fig.1d

Plate-7

Monthly population dynamics of isopod parasites in *Rhizoprionodon acutus*  
 Fig.1a- Prevalence, Fig.1b- Mean intensity, Fig.1c- Mean abundance, Fig.1d- Index of infection

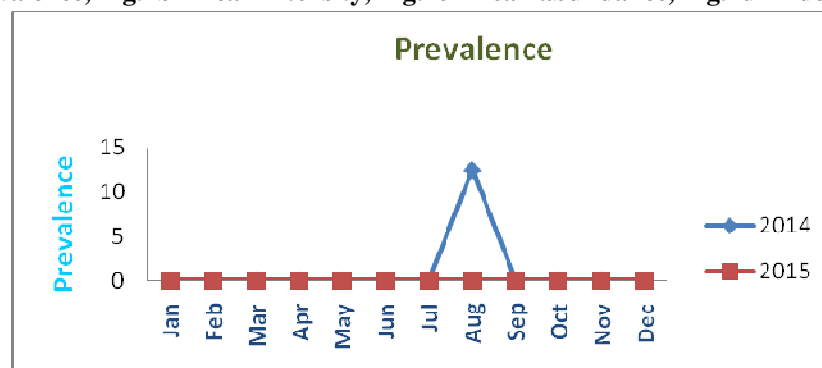


Fig.1a

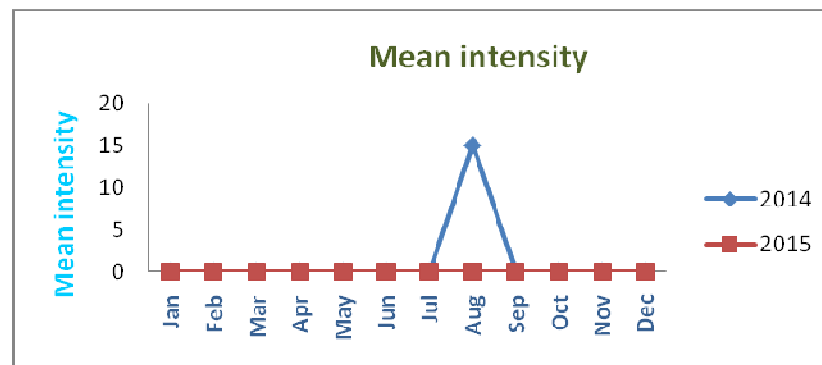


Fig.1b

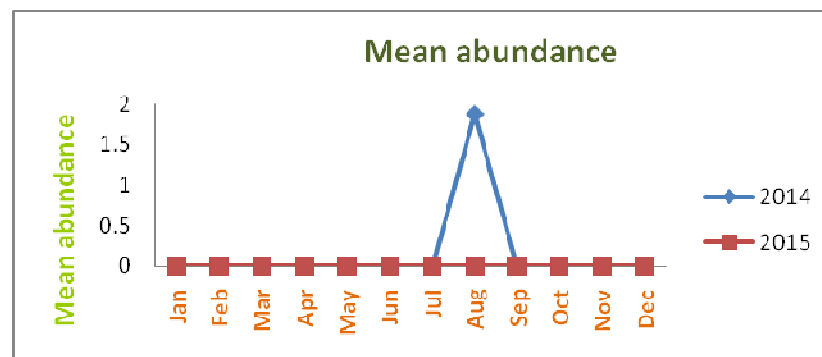


Fig.1c

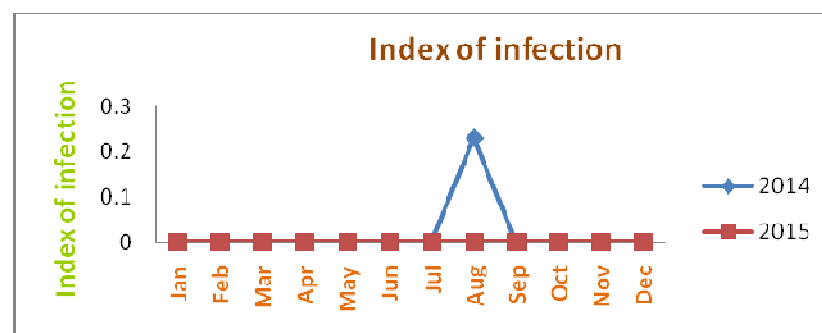


Fig.1d

Plate-8

Fig.1: Seasonal dynamics of overall parasitization in *R.acutus*

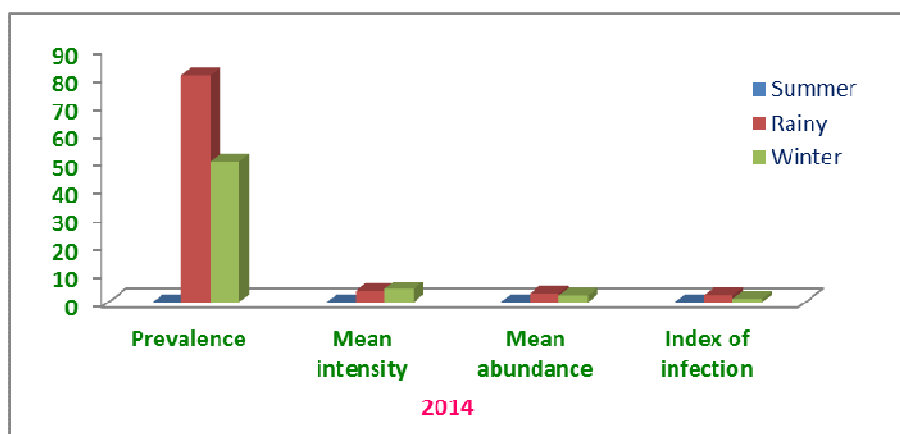


Fig. 1a

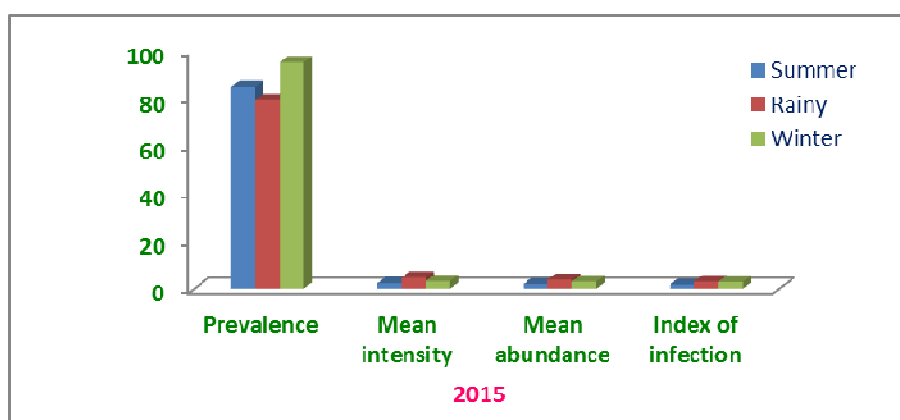


Fig. 1b

Table-1

Metazoan parasites of *Rhizoprionodon acutus* Rüppell, 1937

Sl. No.	Name of the host	Name of the parasite	Number of parasites collected
1.	<i>Rhizoprionodon acutus</i> Rüppell, 1937	<b>Monogenean:</b> 1. <i>Loimos secundus</i> Chauhan and Bhalerao, 1945	18
		<b>Cestodes:</b> 2. <i>Paraorygmatobothrium floraformis</i> (Southwell, 1912) Ruhnke, 2011	149
		3. <i>Nybelinia lingualis</i> (Cuvier, 1817) Dollfus, 1927	24
		4. <i>Heteronybelinia peridareus</i> Shipley et Hornell, 1906	58
		5. <i>Nybelinia indica</i> Chandra, 1986	6
		6. <i>Phoreobothrium</i> sp.	3
		7. <i>Poecilancistrum ilishae</i> , Southwell et	

		Prashad,1918	45
		<b>Nematodes:</b>	
		8. Larva of <i>Anisakis</i> sp.	6
		9. Larva of <i>Hysterothylacium</i> sp.	4
		<b>Copepod:</b>	
		10. <i>Kroyeria minuta</i> Pillai, 1968	15
		<b>Isopod/Amphipod:</b>	
		11. <i>Lafystius sturionis</i> Kroyer, 1842	15

492

493

**Table-2**

494 **Seasonal changes in overall prevalence, mean intensity, mean abundance and Index of**  
 495 **infection of parasites of *R. acutus***

496

Seasons	No. of examined fishes (a)	No. of infected fishes (b)	No. of parasites (c)	Prevalence = b/a*100	MI c/b	MA c/a	Index of infection b*c/a <sup>2</sup>	χ <sub>2</sub> Value (at 5% level of significance and 2 degrees of freedom)
R.acutus (Jan, 2014-Dec, 2014)								
Summer	32	0	0	0	0	0	0	χ <sub>2</sub> = 19.63; p=0.000055 (The result is significant)
Rainy	31	25	94	80.6	3.8	3	2.4	
Winter	32	16	74	50	4.6	2.3	1.2	
R.acutus (Jan, 2015-Dec, 2015)								
Summer	13	11	25	84.6	2.3	1.9	1.62	χ <sub>2</sub> = 0.170; p=0.918 (The result is not significant)
Rainy	24	19	90	79.2	4.9	3.9	3.06	
Winter	20	19	60	95	3.3	3.1	2.94	

497