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MOLECULAR SPECTROSCOPIC ANALYSIS OF *DAUCUS CAROTA* PLANT PIGMENT EXTRACTS.

3 Abstract

4 Various solvents (distilled water, methanol, ethanol, acetone and chloroform) extracts of Daucus carota 5 were scanned with UV – Visible spectrophotometer (Thermo - spectronic) and Perkin – Elmer FT – IR 6 model (Spectrum BX) infra – red spectrometer. All the extracts though with different pH showed typical 7 orange colour with more polar solvent (H_2O) giving the lighter hue. The UV – Visible spectra analyses of 8 the extracts revealed a good absorption between 202 - 452nm and λmax range of 272 - 340nm. The IR 9 spectrometry revealed various functional groups such as alcohol, arenes, alkanes and particularly, the 10 conjugated dienes. These functional groups obviously are attributable to the structural chemistry of the 11 Daucuscarota plant pigment.

12 Keywords: Molecular Spectroscopy, Pigment, UV- Visible, Infra-Red, Spectra

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14 INTRODUCTION

Plant pigment include a variety of different kinds of molecules, including porphyrins, carotenoids, anthocyanins and betalains. All biological pigments selectively absorb certain wavelength of light while reflecting others. The light that is absorbed may be used by plants to power chemical reactions while the reflected wavelengths of the light determine the colour pigment that will appear to the eye. Pigments also serve to attract pollinators. Plants pigments in fruits and vegetable are mainly chlorophyll (green), carotenoids (red, yellow, orange), and anthocyanin (blue, purple, red). The orange in carrots is due to the carotene[1].

Major chemical constituent of carrots is lycopene which has eleven conjugated double bonds that gives it, its deep red colour and are responsible for its anti-oxidant activity. Due to its non-toxicity, lycopene is used for food coloring[2]. Plants and photosynthetic bacteria naturally produce all-trans lycopene[3]. When exposed to light and heat, lycopene undergo isomerization to number of cis- isomer, which have a bent rather than linear shape.

In synergy with other nutrients, lycopene has been shown to decrease biomarker of oxidativestress and carcinogenesis[2]. In different studies, lycopene was even found to have inhibitory effect on cataract development [4] to several different kinds of cancer including breast cancer [5]. Lycopene may be the most powerful carotenoids quencher of singlet oxygen[6], being 100times more efficient in test-tube studies of singlet oxygen quencher action than vitamin E, which in turn has 125times the quenching action of glutathione (water soluble). Singlet-oxygen produced during exposure to ultra –violet light is the primary cause of skin ageing[7].

Though lycopene is non-toxic and is commonly found in diet, but cases of excessive carotenoid intake have been reported .In a middle aged woman who had prolonged and excessive consumption of tomato juice, her skin and liver were colored orange-yellow and she had elevated levels of lycopene in her blood .Her skin colour returned to normal after three weeks on lycopene –free diet[8].This coloration
of the skin in known as lycopene dermia.

39 In recent years, complementary and alternative medicine, together with their various practices has 40 become increasingly popular and even considered a highly acclaimed discipline in the western world [9]. 41 It is noted that 40-50% of medicine are direct or synthetic copies of plants ingredient [10]. The federal 42 government of Nigeria inaugurated the Presidential Initiative Committee (PICO) in 2006 on the development, promotion and commercialization of Nigerian herbal medicine plants. The committee's blue 43 44 print is yet to be implemented despite the global resurgence of medicinal plants. This Study have, 45 therefore intends to elucidate the pigment chemistry of Daucus carota commonly consumed in Nigeria. It 46 is hoped that this could provide some basis for its utilization in food ,health and related industries.

47 2.0 EXPERIMENTAL

48 2.1 Sample collection and preparation

A sufficient quantity of fresh sample of carrot (*Daucus carota*) was purchased from vegetable market at Airport Road, Benin city, Oredo local government area of Edo state. The vegetable was identified by comparison with Herbarium Reference Material at Department of Botany, Faculty of Life Science, University of Benin. The sample was prepared for extraction by carefully cleaning and slicing them into small portions and thoroughly crushed with mortar and pestle. The crushed vegetable was used as starting material for the extraction.

55 2.2 Chemical Extraction

56 Cold extraction and maceration was used by taking 800g of the crushed vegetable (carrot), mixed with 57 1200cm³ each of the various analar grade solvents (methanol, ethanol, acetone and chloroform) and 58 distilled water in 250cm³ conical flask. The flask was carefully swirled for 15 minutes after which the 59 mixture was filtered with Whatman filter paper No.1 (11cm). All the pigment extracts were preserved by 60 keeping them in refrigerator temperature for physical and chemical evaluation studies[11].

61 2.3 UV-visible Spectra

The UV-visible spectra of solvents (methanol, ethanol, acetone and chloroform and distilled water) extracts of carrot sample in a rectangular cuvette were scanned respectively, using spectrophotometer (Thermo-Spectronic, England). The extracts were first sufficiently diluted with relevant solvents before introducing into cuvette and then scanned[12]. UV-visible spectroscopy involves the examination of the electronic transition associated with absorption in the UV (180-390nm) and visible (390-700nm) regions of the electromagnetic spectrum[13].

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70 2.4 Infra-Red Spectra (IR)

- 71 Two equal sizes of flat IR transparent circular cells of 2cm in diameter were thoroughly cleaned with
- solvent. Two drops of the sample extracts were added to one of the cells with glass rod and then carefully
- 73 covered with the other cell. The covered cell was then inserted into the Perkin Elmer FT IR model -
- 74 Spectrum BX. The spectra were finally printed out for interpretation [14].

75 The pH meter was standardized by using buffers of pH 4 and pH 10. The pH of the extract were

- 76 determined by using pH meter(Hanna Instrument model H196107), also the pH was altered by adding
- 57 several concentration (0.05M,0.5M & 0.8M) of hydrochloric acid (HCI) and (0.05M,0.5M & 0.8M) sodium
- 78 hydroxide (NaOH). The pH values were finally measured using the pH meter.

79 2.5 Hue

- 80 Hues were visually assessed in conjugation with the λ maximum values obtained from UV visible spectra
- 81 from various spectra.

82 3.0 Results and Discussion

83 **3.1 Effect of varying the types of solvent.**

84 The colours of the extracted pigments obtained using various solvents is presented in table 1

85 Table1: Colours of extracts of carrot using different solvents

86

Extraction Solvent	Colours of extracts		Λmax.
Distilled Water	Light orange		272
Methanol	Orange		268
Ethanol	Orange hue)	(Deeper	294
Acetone	Orange		292
Chloroform	Orange		286

⁸⁷

It would be observed that the colour of the carrot extract using various solvents was basically orange. In other words, the use of the more polar solvents (H_20 , MeOH, C_2H_5OH , CH_3OCH_3) and non-polar solvent (CCI_3) did not affect the colouration of the carrot extract. However, it is obvious in table 1, that the hue of the orange colour was lighter when the extraction was done with distilled water. Of the more polar solvents used in the extraction, water would be considered the most polar. Thus, the most polar solvents gave the least hue of colour. The inference, is that all the solvents extracted the same type of pigment from carrots (orange – colour pigment) but the most polar solvent (H_2O) gave a lighter hue of orange,

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which would tantamount to a less intensive extraction of orange pigments. This can be ascribed to
differences in the affinities of the orange pigment in the carrot to the various types of solvents used in
these studiesUkhun and Azi, 1991 reported a similar phenomenon with meat extracts in which various
λmax values were observed with meat extracts when different solvents were used in the extraction.

99 The orange colouration of the extracts observed in this study, is probable due to primarily, vitamin A 100 (retinol) which is known to be present significantly in carrots. Carrot is the major source of vitamin A [15]. 101 The largely hydrocarbon and therefore hydrophobic nature of vitamin A would make it more soluble in 102 less polar solvents.

103 **3.2 UV visible Spectral analysis of extracts**

104 These spectral studies were done with a view to gaining more insight into the physical attributes of the 105 pigments.

Extracting solvent sample	and Region of Absorption (nm)	Λmax. (nm)
Carrot/ distilled water	202 – 388nm	272nm (0.253A)
Carrot / methanol	208 – 448nm	294nm (0.426A)
Carrot / Ethanol	204 – 452nm	268nm (0.410A)
Carrot / Acetone	204 – 422nm	340nm (6.000A)
Carrot / Chloroform	206 – 368nm	286nm (0.463A)

106 Table 2: UV – visible spectral analysis of carrot extracts

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From table 2, the carrot extracts showed absorption from about 200nm to 450nm; absorption in both uv and visible regions of electromagnetic spectrum. Ultraviolet – visible (UV - visible) spectroscopy involves the examination of the electronic transitions associated with absorption in the UV (180 -390nm) and visible (390 – 780nm) regions of theelectromagnetic spectrum [13]. The energies associated with these regions are capable of promoting the outer electrons of a molecule from one electronic energy level to a higher level. The part of the molecule containing the electrons involved in electronic transition responsible for the observed absorption is called the chromophore.

115 The types of transitions that results UV – visible absorption consists of excitation of an electron from the 116 highest occupied molecular orbital (usually of non – bonding p or bonding π orbital) to the next lowest 117 unoccupied molecular orbital (an anti – bonding π^* or α^* molecular orbitals) (Stuart,2013).

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Generally, the study revealed that the various solvent extracts gives differentλ max. (Water extract,
272nm; methanol extract, 294nm; ethanol extract, 268nm; acetone extract 340nm and chloroform extract
280nm). UV – visible spectra are sensitive to differences in solvent, pH and conjugation [16]. Many of the

- 121 benzene based pigments such as anthocyanins, flavonoids etc[17] could show this uv absorption
- because of the conjugation in their structures.

123 Infra – red spectroscopy (IR).

The IR spectroscopy was carried out on the extracts in an attempt to gain further insight into their physico - chemical properties. Infra-red spectroscopy is a technique based on the vibrations within a molecule [18][19][20][21]. In this study, various functional groups are inferred in the ethanol extracts in accordance with the use of spectroscopy. Conjugation lowers the bond order because of contributions of the resonance forms with lower bond orders.

Among the IR bands inferred includes 3080 cm⁻¹ of C – H stretch, coupled with C = C stretch with IR band

130 of 1633cm⁻¹, strong band around 3350cm⁻¹ which shows presence of hydroxyl group and 1200cm⁻¹

131 which is due to C – O stretch of alcohol. The IR band of 1623 cm⁻¹ is lower due to effect of conjugation

132 which reduces the adsorption frequencies of the conjugated groups.

A sharp band at 3600cm⁻¹ tend to suggest, there are free O – H stretch. Bonded O – H stretch appears at

- 134 lower frequency than free O H stretch. Finally, a strong band at 3000cm-1 was seen, which is typical of
- 135 C H stretching of the aromatics.

136 <u>Conclusion</u>

137 Though Daucus Carota pigments extracted with various solvents showed typical orange color, the 138 most polar solvent (H₂0) gave a lighter hue of orange; indicating less intensive extraction of orange pigment. Similarly, the UV. visible spectral analysis of the extracts gave λ max range (272mm-340mm) 139 140 with the more polar solvent (H₂O) recording the least λ max (272nm or 10253A). Various functional groups are inferred in the extracts in accordance with the use of IR spectroscopy in functional group 141 142 detection and analysis. The infra-red region (4000-650cm⁻¹) is of prime importance for the study of 143 organic compounds. Possibility of two different compounds having the same IR spectrum is exceedingly 144 small. Thus IR could be used for identification of organic molecule. In this study, various functional groups 145 like OH, CO, C-H and effect of conjugation on the absorption bands were inferred.

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