

1  
2  
3 **On Improving Seed Germination and Seedling Growth in Rice under Minimal Soil**  
4 **Salinity.**

5  
6 **ABSTRACT**

7  
8 **Aims:** It was assumed that two- way approach i.e. nutrient-priming with potassium salt of the seeds and  
9 later on ammonium sulphate application may be binary beneficial for growth of rice besides evidencing  
10 genetic variability under salt stress

11 **Study Design:** The experiment was laid out in Complete Randomized Design with three replications.

12 **Place and Duration of Study:** The study was conducted in laboratory and glass house of Soil Salinity  
13 Research Programme of Land Resources Research Institute at National Agricultural research Centre,  
14 Islamabad, Pakistan during the period from May to August, 2016.

15 **Methodology:** Seeds of *Oryza sativa* (cv. KS-282 and BAS 385) were primed with potassium nitrate. In  
16 the second phase of the study, the primed seeds were raised in a minimal saline soil with ammonium  
17 sulphate nutrition gradually up to 150 mg Kg<sup>-1</sup>.

18 **Results:** Bas-385 was more responsive for mean germination time than KS-282. In Bas-385 and KS-282  
19 germination was 100 and 90 percent. Germination rate index of Bas-385 was 16 percent higher than that  
20 of KS-282. Biomass of Bas-385 seedlings was higher than that of KS-282 with the treatments. In both the  
21 cultivars of rice, Na<sup>+</sup>/K<sup>+</sup> ratio was in antagonistic relation  $R = (- 0.99)$  with the gradual increase in  
22 ammonium sulphate application. Potassium ion was accordant with sulphate ion and N concentration.  
23 Bas-385 was more tolerant to KS-282 based on Na<sup>+</sup>/K<sup>+</sup> ratio and bio mass.

24 **Conclusion:** BAS 385 (salt sensitive) superseded to KS-282 (salt tolerant) under minimal salt stress due  
25 to nutrient priming and then enhanced nutrition.  
26

27 **Key words:** Ammonium sulphate, priming, rice, salinity, tolerance

28  
29 **1. Introduction**

30 The embryo in the seed seems to be prone to surplus ionic effects from the micro-external  
31 environment. These consequences may change its cellular responses either proceeding to seed  
32 germination or its senescence. Excess of salts in the form of 'salinity monster' is restricting the crop  
33 production, whereas research findings report that seeds when treated with some salts in minute quantity,  
34 a process scientifically known as priming, is beneficial for its germination. After the priming process, once  
35 the seeds are germinated, provided the other processes/microenvironments are favorable and  
36 concomitant, seedling growth may take place in a normal direction under salt stress. Salinity is a major  
37 abiotic stress [1]. Saline soils contain multiple types of soluble salt components, each of which has a  
38 different effect on the initial growth of plants [2]. Soil salinity may affect the germination of seeds either by  
39 creating an osmotic potential external to the seed, preventing water uptake or through the damaging  
40 effects of Na<sup>+</sup> and Cl<sup>-</sup> ions on the germinating seed [3]. Salt and osmotic stresses are responsible for  
41 either inhibition or delayed seed germination and for seedling establishment [4]. NaCl is the predominant  
42 salt causing salinization [5]. Germination parameters and biomass of rice decreased with increasing salt  
43 stress [6]. To have a healthy mature plant, it is vital that its early stages of nourishment such as  
44 germination and seedling must be prone to any set back in a metabolic process as [7] reported that

45 germination and seedling growth are critical stages regarding salt stress. The entry of dissolved ions  
46 except the toxic one, trigger metabolic processes especially the enzymatic activities in water-imbibed  
47 seeds. For such activities, the role of potassium ion is significant. Potassium salts are the widely used  
48 source of seed priming [8]. Potassium ions with different combinations of anions are in use. Nutrient  
49 priming of seeds also improved nutrient supply [9]. Use of potassium source as seed priming mediator  
50 improved germination process and decreased  $\text{Na}^+/\text{K}^+$  ratio in the follow-on seedlings [10]. After seed  
51 germination, seedlings need the optimum environment and balanced nutrition for growth. Since nutrition  
52 priming support germination process, post germination processes also need to overcome salinity effects.  
53 Salt diminution process necessitates decreasing sodium ion impact in the growth medium. To decrease  
54 its shock, it becomes significant to covenant it with a suitable nutrition ion. Potassium ion is the most  
55 suitable ion due to its chemical or physical structure. Its suitability is measured in the form of  $\text{Na}^+/\text{K}^+$  ratio  
56 in plant material under salt stress. The lower this ratio, the lower is the impact of sodium ion on a  
57 glycophytes growth. The sulphur application can be beneficial for rice growth [11]. The sulphur application  
58 is well-known for amino acids formation. It has been observed that sulphur application also decreases  
59  $\text{Na}^+/\text{K}^+$  ratio in plant organ and sulphur is in synergistic relation with potassium ion [12]. Since sulphur and  
60 nitrogen are in synergistic relations and nitrogen has its own importance in the metabolism of plant tissue  
61 build up, therefore application of sulphur in the form of ammonium sulphate could play a better role for the  
62 growth of seedlings under salt stress. To what extent sulphur nutrition could be helpful for plant growth  
63 especially when the seeds of rice are nutrient-primed. It was assumed that two- way approach i.e.  
64 nutrient-priming with potassium salt of the seeds and later on ammonium sulphate application may be  
65 binary beneficial for growth of rice besides evidencing genetic variability under salt stress.

## 66 **2.MATERIALS AND METHODS**

67 Treated seeds of *Oryza sativa* (cv. KS-282 and BAS 385) with 1 % sodium hypochlorite for 15  
68 minutes [13] and then washed these seeds three times with distilled water. For nutrient-priming, placed  
69 the seeds in a 100 ml beaker and applied one percent (w/v) of potassium nitrate salt solution. Kept these  
70 seeds in the solution for five hours. Then dehydrated these seeds with tissue papers, and air dried.  
71 [Placed ten dried seeds on filter paper in Petri dish \(11cm dia.\) in quadruplicates for germination.](#) Counted  
72 germinated seeds on 2<sup>nd</sup> to 7<sup>th</sup> day. Calculated mean germination time (MGT) according to the equation of  
73 Ellis and [14]. Converted observed germination percentage into ASIN form for statistical analysis  
74 purpose as worked by [15]. Computed rate of germination index (RGI) as given by [16]. Aanalyzed the  
75 soil for physicochemical characteristics (Table 1) and applied ammonium sulphate @ 0, 30, 60, 90, 120,  
76 150 mg kg<sup>-1</sup> in this saline soil ( $\text{pH} = 7.84$ ,  $\text{EC} = 4.55 \text{ dSm}^{-1}$ ). [The primed seeds of the varieties were sown in  
77 this soil.](#) The pots were arranged in complete randomized design with three replications and were placed  
78 in glass house under controlled conditions. After 30 days of seedling establishment, excised the aerial  
79 portion of the plants. Rinsed the shoots with deionised water, surface dehydrated with tissue paper and  
80 recorded fresh mass (FM). Dried shoots at 65 °C and recorded dry mass (DM). Cut each sample into  
81 small pieces and digested separately in a perchloric-nitric (1:2) di-acid mixture [17]. Determined sodium

82 and potassium ions in the digested material by flame photometry Analyzed sulphur in the digested  
 83 material as given by [18]. Being a saline soil having silty clay loam texture and with low to moderate  
 84 nutrients, the plants were grown with ammonium sulphate in adequate moisture (Table 1). Computed the  
 85 data statistically according to factorial CRD and compared treatment means using LSD test [19].

86  
 87  
 88

**Table1 . Physico-chemical characteristics of the saline soil.**

89  
 90  
 91  
 92  
 93  
 94  
 95  
 96  
 97  
 98  
 99  
 100  
 101  
 102  
 103  
 104  
 105  
 106

Characteristics	Values
pH (1:1)	7.84
ECe (1:1) (dS m <sup>-1</sup> )	4.55
Cl <sup>-1</sup> (meq L <sup>-1</sup> )	5.13
CO <sub>3</sub> <sup>-2</sup> (meq L <sup>-1</sup> )	0.35
HCO <sub>3</sub> <sup>-1</sup> (meq L <sup>-1</sup> )	1.71
SO <sub>4</sub> <sup>-2</sup> (mg kg <sup>-1</sup> )	6.22
NO <sub>3</sub> <sup>-</sup> -N (mg kg <sup>-1</sup> )	14.3
P (ABDTPA Extractable) ( mg kg <sup>-1</sup> )	5.21
SAR (mol L <sup>-1</sup> ) <sup>1/2</sup>	5.24
Saturation (%)	32
Clay (%)	36
Silt (%)	50
Sand (%)	14
Text. Class	Silty Clay Loam

107  
 108  
 109

### 110 3. RESULTS AND DISCUSSIONS

111  
 112  
 113  
 114

Nutrient-priming of rice seeds with potassium nitrate and post-germination application of ammonium sulphate significantly ( $P = 0.01$ ) affected seed germination parameters and seedling growth respectively.

#### 115 3.1 Germination Parameters

116  
 117  
 118  
 119  
 120  
 121  
 122  
 123  
 124  
 125  
 126

In KS-282 germination percentage of unprimed seeds was 80 percent, but the seeds of the same variety, when primed with potassium nitrate salt, the germination was higher (90 %). In Bas-385 germination of un-primed seeds was 90 percent; however, it was 100 percent when the seeds of the same variety were primed with the same salt (Table 2). Rice varieties physio-genetically responded to seed priming treatment. Genetic traits of Bas-385 supported to seed-surface treatment with potassium nitrate than the hereditary attributes of KS-282 for germination at initial salinity. [20] found that germination is inversed to the increasing salt stress. During the treatment time, useful metabolic processes in the form of enzyme activation passed successfully through the lag phase. Usage of potassium nitrate as the seed priming agent was useful from nutritional aspect also. Priming treatment usability was revealed in the later stages of growth. Germination rate index (GRI) improved 22 percent in the primed seeds than the un-primed one in KS-282 (Table 3). In Bas-385, GRI improved 33 percent

127 in the primed seeds than the un-primed one. The performance of Bas-385 was better than KS-282 by  
 128 16 percent. Mean Germination Time (MGT), remained low in the primed seeds than the un-primed one  
 129 in KS-282. Overall performance difference for MGT of both the varieties was non-significant.  
 130 Germination rate index expresses the speed of the germination. With increasing level of salinity, GRI  
 131 decreased [21]. Therefore seed-priming improved the speed of germination. Higher GRI values  
 132 indicated higher and faster germination. Early germination is reciprocal to MGT [22] as MGT is related  
 133 to the time during which radicle appeared. Priming softens seed coat adherence due to imbibition,  
 134 which may permit to emerge out radicle without resistance as reported by [23].  
 135

136 Table 2 Germination of rice seeds with nutrient -priming using potassium nitrate.

Priming Treatment	Germination (%)		Means	ASIN		Means
	Cv. KS-282	Cv. Bas-385		Cv. KS-282	Cv. Bas-385	
Primed	90 b	100 a	95 A	71.6 b	90.0 a	80.8 A
Un-primed	80 c	90 b	85 B	63.4 c	71.6 b	67.5 B
Means	85 B	95 A		67.5 B	80.8 A	

137 Means sharing similar letter(s) for a parameter do not differ significantly at  $p < 0.01$   
 138 Cv. 0.01 %  
 139

140 Table 3 Effect of potassium nitrate as nutrient halo-priming on GRI and MGT  
 141 of two varieties of rice.

Priming Treatment	Germination Rate Index (GRI)		Means	Mean Germination Time (MGT), (days)		Means
	Cv. KS-282	Cv. Bas-385		Cv. KS-282	Cv. Bas-385	
Primed	10.9 b	13.2 a	12.05 A	4.1 c	4.1 c	4.1 B
Un-primed	8.9 d	9.9 c	9.35 B	4.3 a	4.2 b	4.3 A
Means	9.9 B	11.5 A		4.2 A	4.2 A	

142 Means sharing similar letter(s) for a parameter do not differ significantly at  $p < 0.01$   
 143 Cv. 0.86 %  
 144

### 145 3.2 Seedlings Growth

146 With the increasing of ammonium sulphate (AS) application, fresh mass (FM) and dry mass (DM)  
 147 of both the cultivars were affected (Table 4). In KS-282 and Bas-385, FM was increased 4, 6, 8 and 10  
 148 percent; and 6, 9, 12, 15 and 18 percent than the respective control from 30 to 150 mg kg<sup>-1</sup> of AS  
 149 application. In KS-282 and Bas-385, DM was increased 12, 15, 17, 19 and 21 percent; and 14, 18, 21, 24  
 150 and 27 percent than the respective control from 30 to 150 mg kg<sup>-1</sup> of AS application. Biomass of Bas-385  
 151 was higher than that of KS-282 with AS treatments. Fresh mass of plant material is a product of water  
 152 and other chemicals in the form of tissue. Water potential is affected by included salts in a plant. Water  
 153 retention of a plant tissue indicates its health and turgidity [24]. Antagonistic relations between plant  
 154 biomass to increasing salinity stress has been reported [25]. Dry mass of plant material is the net  
 155 outcome of the resultant metabolic activities [24].

156  
157

Table 4 Effect of ammonium sulphate application on fresh and dry mass of shoot of rice grown from nutrient –primed seeds in saline soil

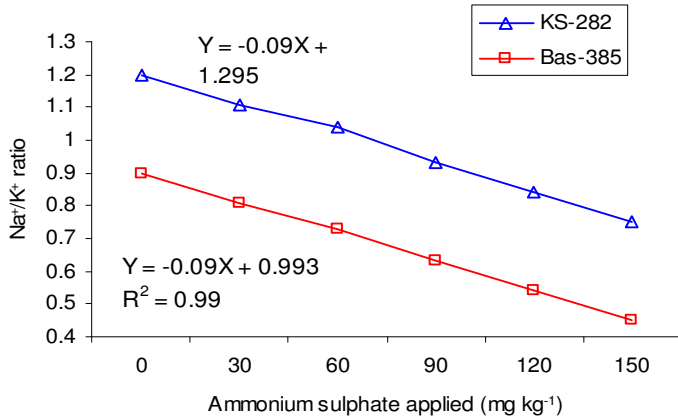
Ammonium sulphate (mgKg <sup>-1</sup> ) applied	Fresh mass (mg plant <sup>-1</sup> )		Dry mass (mg plant <sup>-1</sup> )	
	Cv. KS-282	Cv. Bas-385	Cv. KS-282	Cv. Bas-385
Control	140.2 l	146.1 k	9.81 k	10.1 3j
30	146.3 j	155.2 f	11.07 i	11.53 fg
60	149.2 i	159.6 d	11.23 h	12.03 d
90	152.1 h	164.2 c	11.43 g	12.33 c
120	154.6 g	168.3 b	11.63 f	12.63 b
150	157.3 e	172.5 a	11.8e	13.03 a

Means sharing similar letter(s) of a parameter do not differ significantly at p < 0.01  
Cv. 2.05 %

158  
159  
160

### 161 3.3 Sodium Potassium Ratio

162 In both the cultivars of rice, Na<sup>+</sup>/K<sup>+</sup> ratio was in antagonistic relation (r = - 0.99) with the applied  
163 AS (Fig.1) . In KS-282, Na<sup>+</sup>/K<sup>+</sup> ratio decreased 8, 13, 23, 30 and 38 percent than the control at 30, 60,  
164 90, 120 and 150 mg kg<sup>-1</sup> of AS application respectively. In BAS-385, Na<sup>+</sup>/K<sup>+</sup> ratio decreased 10, 19,  
165 30, 40 and 50 percent than the control in same sequence of AS application as above. Potassium ions  
166 are synergistic to sulphate ion and nitrogen application. These ions are beneficial to crop plants.  
167 Potassium and sulphate ions are inversed to Na<sup>+</sup> in glycophyes. In both the varieties, the linear  
168 equation shows that every 100 units increase in sulphur application decreased 9 units of Na<sup>+</sup>/K<sup>+</sup> ratio. In  
169 addition, KS-282 needed 1.31 times higher the application of AS application than that of Bas-385 to  
170 maintain low Na<sup>+</sup>/K<sup>+</sup> ratio. Sodium ion impedes positive biochemical activities resulting in decreased dry  
171 mass. Therefore sodium ion maneuvers senescence in reduced growth of glycophytes. It has been  
172 evident by [26] that reduction in seedling growth under saline conditions is due to increase in sodium  
173 chloride toxicity. In many species, salt sensitivity is associated with the accumulation of sodium ion in  
174 photosynthetic tissues [27]. Externally decreased in water potential created by NaCl might have affected  
175 fresh mass by preventing water uptake. The capacity of plants to maintain a high K<sup>+</sup>/Na<sup>+</sup> ratio is one of  
176 the key determinants of plant salt tolerance [28]. KS-282 sequester and accumulate K<sup>+</sup> according to the  
177 external application of sulphate ion. According to [29] change in ratios of ions in plants may be a  
178 resultant due to same pathways of Na<sup>+</sup> and K<sup>+</sup>. Na<sup>+</sup> competes with K<sup>+</sup> uptake through Na<sup>+</sup>- K<sup>+</sup> co-  
179 transporters and may also block the K<sup>+</sup> specific transporters of root cells under salinity [30].

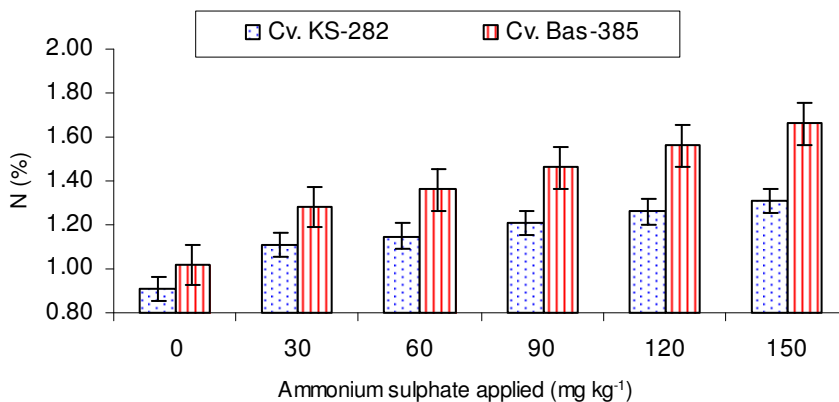


180

181 Figure 1 Relation between ammonium sulphate applied and Na<sup>+</sup>/K<sup>+</sup> ratio in rice.

182 **3.4 Relation between sulphur and nitrogen**

183 Under minimal soil salinity conditions, with the application of ammonium sulphate @ 30, 60, 90,  
 184 120 and 150 mg kg<sup>-1</sup> soil, N concentration in shoot of Cv. Bas-385 was higher 15, 18, 21, 24 and 27  
 185 percent than that in KS-282 ( Fig. 2). Availability of sulphur along with S-containing compounds such as  
 186 ATP-sulfurylase is considered as the first rate-limiting enzyme of the sulphur assimilation pathway and  
 187 is up-regulated under salinity stress [31]. In salt-treated plants, sufficient sulphur supply allows  
 188 glutathione synthesis necessary to prevent the adverse effects of Reactive oxygen species on  
 189 photosynthesis. Plants with higher levels of thiol were more salinity tolerant [32]. It seems K ion  
 190 accordant with sulphate ion increases tolerance against Na<sup>+</sup>. For rice growth, application of nitrogen is  
 191 beneficial under salt stress [33]. In addition it was observed that cultivar response may be variable as  
 192 Bas- 385 was more responsive than KS-282 for N- concentration in such conditions.



193

194 Figure 2 Level of total -N in shoot of two varieties of rice under salt stress.

195

196 **4. CONCLUSIONS**

197 Under salt stress, nutrient-primed with potassium nitrate eased germination 100 and 90 percent in  
198 rice varieties i.e. Bas-385 and KS-282. For germination rate index, the performance of Bas-385 was  
199 better than that of KS-282 by 16 percent. Biomass of Bas-385 was higher than that of KS-282 with  
200 sulphur treatments. In both the cultivars of rice,  $\text{Na}^+/\text{K}^+$  ratio was in antagonistic relation ( $R = - 0.99$ ) with  
201 the applied sulphur, however, Bas-385 accumulated more potassium ions than that of KS-282.

202

203 **REFERENCES**

204

- 205 1. Rueda-Puente EO, Garcia-Hernandez JL, Preciado-Rangel P, Murillo-Amador B, Tarazon-  
206 Herrera MA, Flores-Hernandez A, Holguin-Pena J, Aybar AN, Barron-Hoyos JM, Weimers  
207 D, Mwandemele O, Kaaya G, Larrinaga-Mayoral J, Troyo-Diequez E. Germination of  
208 *Salicorniabigelowii* ecotypes under stressing conditions of temperature and salinity and  
209 ameliorative effects of plant growth-promoting bacteria. *J. Crop Sci.* 2007; 193:167-176.
- 210 2. Tobe K, Zhang L, Omasa K. Alleviatory effects of calcium on the toxicity of sodium,  
211 potassium and magnesium chlorides to seed germination in three nonhalophytes. *Seed Sci.*  
212 *Res.* 2003. 13, 47–54.
- 213 3. Khaje-hosseini M, Powell AA, Bingham IJ. The interaction between salinity stress and seed  
214 vigor during germination of soybean seeds. *Seed Sci. Technol.* 2003; 31: 715–725.
- 215 4. Almansouri M, Kinet M, Lutts S. Effect of salt and osmotic stresses on germination in durum  
216 wheat (*Triticum durum* Desf.). *Plant Soil* 2001; 231: 243-254.
- 217 5. Munns R, Tester M. Mechanisms of salinity tolerance. *Annu Rev Plant Biol.* 2008; 59: 651–  
218 681.
- 219 6. Anbumalarnathi J, Mehta P. Effect of salt stress on germination of *indica* Rice varieties. *eJ.*  
220 *Biol. Sci.* 2013; 6 (1): 1-6.
- 221 7. Geraldine LD, Donovan LA. Water potential and ionic effects on germination and seedling  
222 growth of two cold desert shrubs. *Am. J. Bot.* 1999; 86:1146-1153.
- 223 8. Fixen PE. Crop responses to chloride. *Adv. Agron.*1993; 50:107-150.
- 224
- 225 9. Al Mudaris AM, Jutzi SC. The influence of fertilizer-based seed priming treatment on  
226 emergence and seedling growth of *Sorghum bicolor* L., and *Pennisetum glaucum* L., in pot  
227 trials under greenhouse conditions. *J. Agron. Crop Sci.* 1999; 182: 135-141.
- 228 10. Badar Z, Ali A, Hyder SI, Arshadullah M, Bhatti SU. Potassium chloride as a nutrient seed  
229 primer to enhance salt- tolerance in maize. *Braz. J. Agri. Res.*2012; 47(8): 1181-1184.
- 230 11. Kiros H, Mersha S, Habtu S. Nitrogen and sulphur fertilizers effects on yield, nitrogen uptake  
231 and nitrogen use efficiency of upland rice variety on irrigated Fulvisols of the Afar region,  
232 Ethiopia . *J. Soil Sci. Environ. Manage.* 2013; 4(3): 62-70.

- 233 12. Badr Z, Ali A, Salim M, Niazi BH. Role of sulphur for potassium/sodium ratio in sunflower  
234 under saline conditions. *Helia* 2002; 25(37): 69-78.
- 235 13. Britto DT, Kronzucker HJ.  $\text{NH}_4^+$  toxicity in higher plants: a critical review. *J Plant Physiol.*  
236 2002; 159: 567-584.
- 237 14. Ellis RH, Roberts EH. Towards a rational basis for testing seed quality. In Hebblethwaite, P.D.  
238 (ed.). *Seed Production*. Butterworths, London 1980; pp. 605-635.
- 239 15. Dezfuli PD, Sharif-zadeh F, Janmohammadi M. Influence of priming techniques on seed  
240 germination behaviour of maize inbred lines (*Zea mays* L). *ARPN J. Agri. Biol. Sci.* 2008;  
241 3(3): 22-25.
- 242 16. Islam MS, Jahan QSA, Bunnarith K, Viangkum S, Merca SD. Evaluation of seed health of  
243 some rice varieties under different conditions *Bot. Bul. Acad. Sinica.* 2000; 41:293-297.
- 244 17. Chapman, HD, Pratt, PF. *Methods of Analysis of Soils, Plants and Water*. Div of Agric.  
245 Science. Univ. of California. Davis. CA. 1961; pp 56-65.
- 246 18. Verma BC, Swaminathan KS, Sud KS. An improved turbidimetric method for sulphur  
247 determination in plants and soils. *Talanta.* 1977; 24, 49-50.
- 248 19. Gomez KA, Gomez AA. *Statistical Procedure for Agricultural Research*. John Wiley and  
249 Sons., N.Y. 2<sup>nd</sup> Edition. 1984; pp 20-28.
- 250 20. Sedghi M, Nemati A, Amanpour-Balaneji B, Gholip A. Influence of Different Priming Materials  
251 on Germination and Seedling Establishment of Milk Thistle (*Silybum marianum*) under  
252 Salinity Stress. *World Appl. Sci. J.* 2010; 11 (5): 604-609.
- 253 21. Omar B, Pistorale SM, Andrés AN. Salinity tolerance during seed germination from  
254 naturalized populations of tall wheatgrass (*Thinopyrum ponticum*). *Cienc. investig.*  
255 *agrar.* 2008; 35(3): 231-238.
- 256 22. Mavi K, Demir I, Matthews S. Mean germination time estimates the relative emergence of  
257 seed lots of three cucurbit crops under stress conditions. *Seed Sci. Technol.* 2010; 38: 14-25.
- 258 23. Ibrahim, D. and Mavi, K. The effect of priming on seedling emergence of differentially  
259 matured watermelon (*Citrullu slanatus* (Thunb.) Matsum and Nakai) seeds. *Sci. Horti.* 2004;  
260 102: 467-473.
- 261 24. Badar Z, Ali A, Mahmood IA, Arshadullah M, Shahzad A, Khan AM. Potassium  
262 consumption by rice plant from different sources under salt stress. *Pak. J. Sci. Ind. Res.*  
263 2010; 53(5): 271-277.
- 264 25. Badr Z, Rehana A, Salim M, Safdar A, Niazi BH, Arshad A, Mahmood IA. Growth and ionic  
265 relations of *Brassica campestris* and *B. juncea* (L.) Czern & Coss. under induced salt stress.  
266 *Pak. J. Agri. Sci.* 2006; 43(3-4): 103-107.
- 267 26. Okcu G, Kaya MD, Atak M. Effects of salt and drought stresses on germination and seedling  
268 growth of pea (*Pisum sativum* L.). *Turk J Agric For.* 2005. 29:237-242.



- 269 27. Davenport R, James RA, Zakrisson-Plogander A, Tester M, Munns R. Control of sodium  
270 transport in durum wheat. *Plant Physiol.* 2005; 137: 807-818.
- 271 28. Frans, JMM, Amtmann A. K<sup>+</sup> Nutrition and Na<sup>+</sup> Toxicity: The Basis of Cellular K<sup>+</sup>/Na<sup>+</sup> Ratios.  
272 *Ann. Bot.* 1999; 84: 123-133.
- 273 29. Blumwald, E. Sodium transport and salt tolerance in plants. *Curr. Opin.Cell Biol.* 2000; 12:  
274 431–434.
- 275 30. Zhu JK. Regulation of ion homeostasis under salt stress. *Curr. Opin.Plant Biol.* 2003; 6: 441–  
276 445.
- 277 31. Ruiz JM, Blumwald E. Salinity-induced glutathione synthesis in *Brassica napus*. *Planta* 2002;  
278 214: 965-969.
- 279 32. Astolfi S, Zuchi S. Adequate S supply protects barley plants from adverse effects of salinity  
280 stress by increasing thiol contents. *Acta. Physiol. Plant* 2013; 35: 175-181.
- 281 33. Ghulam M, N Hussain, A Ghafoor. Growth Response of Rice (*Oryza sativa* L.) to Fertilizer  
282 Nitrogen in Salt-Affected Soils. *Int. J. Agric. Biol.* 2000; 2(3): 204-206.