

Enhancement of seed zinc and iron density in cowpea (*Vigna unguiculata*): A Physiological Approach

Abstract

Aims: The present study was conducted to enhance the zinc (Zn) and iron (Fe) concentration in cowpea (*Vigna unguiculata* L.) var. Rituraj using physiological approach. **Methods:** Cowpea was grown in soil-pot culture subjected to soil application at presowing and foliar application of Zn, Fe and their combination at different time period of plant growth. In soil application, the rate of $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$ and $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ application was 10 mg kg^{-1} soil each. Zinc and Fe were also applied by single and combined foliar treatment with 0.5% aqua solution at 35, 45 and 55 days of plant growth. **Results:** The concentration of photosynthetic pigments, specific activities of catalase, peroxidase and carbonic anhydrase in leaves and tissue Zn and Fe concentration in different plant parts were significantly increased as compared to control. **Conclusion:** Zinc translocation from flag leaves to seed and its accumulation was more significant in combined treatment of Zn and Fe foliar application at 35 days.

Keywords: zinc, iron, translocation, accumulation, foliar application

1. Introduction

Zinc (Zn) and iron (Fe) deficiencies are major micronutrient disorder and an important soil constraint for crop yield (Cakmak 2010). There is great need to focus on availability of Zn and Fe in soil because main reason for Zn and Fe deficiencies are low availability of this micronutrient to plant roots rather than low percentage of that nutrient in soil. Sometimes plant roots cannot absorb required amount of essential nutrients to plants. In that condition foliar application is more effective and supportive technique for supplying nutrients to plants more easily, when soil conditions are not suitable to plant growth (Kolata et al. 2001; Kinaci et al. 2007).

Zinc is an important micronutrient that affects flowering, floral development, fertilization and seed maturation in crop. Insufficient supply of zinc affects more the seed yield than vegetative growth because zinc has a specific role in pollen fertilization. Iron is another micronutrient that is directly involved in photosynthetic activity (Brait et al. 2007), chlorophyll synthesis, respiration (Vigani et al. 2009) and nitrogen metabolism. In enzyme system iron serves as a cofactor that catalyzes biochemical reaction and occurs with a heme group in some enzymes like catalase, peroxidase and cytochrome oxidase (Balakrishnan 2000).

In the present study effort has been made to focus on improving cowpea yield, biomass and tissue Zn and Fe concentration in edible plant parts (seed) by applying foliar application of Zn, Fe and their combination at different time points of plant growth in addition to soil Zn and Fe application.

38 2. Material and methods

39 Cowpea (*Vigna unguiculata* L.) var. Rituraj plants were grown in pots containing 10 kg soil in a glass
40 house at an ambient temperature (25-30°C) and subjected to treatment of iron and zinc singly and
41 their combination in addition to soil application at pre-sowing and foliar application at different time
42 period of plant growth. Experimental soil was loamy- sand with pH 7.30, electrical conductivity 0.303
43 dSm⁻¹ and Diethylenetriaminepentaacetate (DTPA)–extractable iron (Fe), and Zinc (Zn) was 7.93
44 and 1.16 mg kg⁻¹ soil respectively before mixing. In soil application, ZnSO₄.7H₂O and FeSO₄.7H₂O
45 application were applied at the rate of 10 mg kg⁻¹ soil each. Foliar application was applied at the rate
46 of 0.5% ZnSO₄ and 0.5% FeSO₄ singly and their combination at 35, 45 and 55 days of plant growth
47 after sowing. Pots were divided into four lots: control, Zn, Fe and Zn+Fe treatment and arranged in
48 complete randomized block design (CRD). In each treatment, there were three replicates. After
49 emergence, seedlings in each pot were uniformly thinned to maintain two plants per pot.

50 The concentration of photosynthetic pigment (chl a, chl b and carotenoids) and activities of
51 certain enzyme catalase (CAT), peroxidase (POX), carbonic anhydrase (CA) were studied in young
52 leaves at day 35, 45 and 55. Total chlorophyll, chlorophyll a, chlorophyll b and carotenoid were
53 measured according to Lichtenthaler (1987). 100 mg leave were crushed in 10 ml 80 % chilled
54 acetone. Extract were centrifuged at 2000xg for 10 minutes. Absorbance of supernatant was
55 estimated spectrophotometer at 662 nm, 645 nm and 470 nm. Chlorophyll contents were expressed
56 in terms of mg chlorophyll present g⁻¹ fresh weight of tissue.

57 Catalase (CAT) is a H₂O₂ splitting enzyme. The activity of CAT (EC 1.11.1.6) was assayed
58 as described by Bisht et al. (1989) in a reaction mixture (10 ml) containing 500 µM H₂O₂ and 1.0
59 mmol KH₂PO₄ buffer at pH 7.0 stabilized at 25°C. The reaction was allowed to proceed for 5 min and
60 stopped by adding 5.0 ml 2N H₂SO₄. The reaction mixture was titrating with 0.1N potassium
61 permanganate (KMnO₄). CAT activity is expressed as µmol H₂O₂ decomposed in the given conditions of
62 the assay.

63 Peroxidase (POX) (EC 1.11.1.7) activity was assayed by method of Luck (1963). The
64 reaction mixture (10 ml) contained 5.0 ml of 0.1M of KH₂PO₄ buffer (pH 6.5), 1.0 mL of 0.5% p-
65 phenylenediamine, 1.0 mL of 0.01% (v/v) H₂O₂, and 1.0 mL tissue extract and reaction proceed for
66 5.0 min and was stopped by adding 2.0 ml 5N H₂SO₄. The colour intensity of the supernatant after
67 centrifugation at 10,000xg for 5.0 mints was measured at 485nm.

68 Carbonic anhydrase (CA) is a zinc-containing carbonate hydro-lyase enzyme. CA (EC
69 4.2.1.1) used 2 ml of 0.02 M vermol buffer (pH 8.15), 0.10% bromothymol blue, 0.2 ml enzyme extract
70 and 2 ml distilled water saturated with CO₂. The change in colour blue to greenish yellow measured to
71 record the time in second and unit of CA activity expressed enzyme unit 100 mg mg⁻¹ protein (Wilbur
72 et al.1948).

73 Plants were harvested at 90d and studied number of pod plant⁻¹, number of seed pod⁻¹, seed
74 weight pod⁻¹ and weight of 100 seeds. Plants were sampled and separated into various parts for

determination of dry matter, Zn and Fe tissue concentration. Harvested plants were washed properly under running tap water, and finally rinsed with distilled water to avoid surface contamination. Plants of all treatments were separated into their parts, which were chopped and placed in paper bags to be dried in an electric oven at 70 °C for 48 h. Concentrations of iron and zinc in cowpea were measured in different plants parts by atomic absorption spectrophotometer (Thermo Jerrell Ash Video 12E: AA/AE Chicago, IL) in clear digests after a nitric acid–perchloric acid (10:1) digestion (Piper 1942). After harvesting the crop soil was sampled for analysis of soil Zn and Fe concentration (Lindsay et al. 1978).

The values presented in tables and figures are the mean of three observations ($\pm$ SE; n = 3). The data were analyzed by one way ANOVA using software program Sigma Stats 3.5. It was followed by comparison of mean value using Holm Sidak method at $P \leq 0.05$.

3. Results

Number of pod plant⁻¹, number of seed pod⁻¹, seed weight pod⁻¹ and weight of 100 seed were increased in all treatment as compared to control (table 1). Single treated Fe was more significant as compare to combined treatment, its obtained maximum number of pod plant⁻¹ (12.66), number of seed pod⁻¹(13.00), seed weight pod⁻¹ (1.95g) and weight of 100 seeds (12.98g) at 35d. Single and combined treated Zn was more significant increased at 45d and 35d respectively (table 1).

Biomass of plant increased significantly and variably in single and combined treatment of iron and zinc. The increased biomass was more pronounced in combined treatment (Zn+Fe) at 35d (table 1). Zn and Fe tissue concentration in seed were significantly increased in all treatment as compared to control. Combined Zn treated plants might be obtained maximum Zn translocation and accumulate in seed. The data in table 2 indicated that significantly effect on translocation and accumulation of Zn and Fe concentration in seed in both treatment as compared to without any treatment at different time periods.

Chlorophyll concentration in leaves was increased with foliar application of Fe single and combined Zn and Fe treatment as compared to single treated zinc. The effect of foliar application of Fe and Zn on total chlorophyll, chlorophyll a, chlorophyll b and carotenoid were shown in figure1 (a-d). With increase in period of foliar application, chlorophyll content was decreased and carotenoid was slightly increased.

In both single and combined treatment of Fe, the specific activity of catalase (CAT) and peroxidase (POX) showed a significant result at 45d. Foliar application after 45d marked decrease in activity of CAT and POX was found in single and combined treated Fe plants (figure 2a-b). The specific activity of carbonic anhydrase increased with supply of Zn single and combined treatment. Maximum enzyme activity of CA was obtained at 45d in single Zn treatment. The specific activity of that enzyme was marked significant effect in single and combined treatment as compare to control (figure 2c).

In soil analysis after mixing DTPA extractable Zn and Fe were varied from 7.31 to 6.23 mg kg⁻¹ and 12.07 to 11.56 mg kg⁻¹ respectively and after harvesting the crop it varied from 6.04 to 5.22 mg kg⁻¹ Zn and 10.74 to 9.67 mg kg⁻¹ Fe in single and combine treated soil. The results indicate that 16.22% to 17.38% applied Zn and 11.02% to 16.35% applied Fe was available to plants.

Table 1: Effect of single and combined Zn and Fe foliar application at different time period on seed yield and biomass of cowpea plants.

Days of foliar spray	Treatments	No. of Pod plant ⁻¹	No. of Seed pod ⁻¹	Seed weight pod ⁻¹ (g)	Weight of 100 seed (g)	Dry weight(g plant ⁻¹)**
35d	Control	7.33±0.666	9±0.577	1.32±0.032	11.75±0.017	16.87±0.054
	Zn	10.66±0.666 [*]	11.66±0.333 [*]	1.44±0.024	11.92±0.005	24.48±0.043 [*]
	Fe	12.66±1.33 [*]	13.00±0.577 [*]	1.95±0.017	12.98±0.004	34.81±0.012 [*]
	Zn+Fe	11.33±0.666 [*]	11.33±0.333 [*]	1.64±0.033	11.97±0.006	28.09±0.014 [*]
45d	Control	7.33±0.666	9±0.577	1.32±0.032	11.75±0.017	16.87±0.054
	Zn	11.66±0.666 [*]	12.00±0.577 [*]	1.53±0.015	12.52±0.014	28.67±0.034 [*]
	Fe	12.00±1.15 [*]	12.66±0.666 [*]	1.80±0.011	12.20±0.020	28.29±0.023 [*]
	Zn+Fe	10.66±1.33 [*]	10.00±0.577 [*]	1.58±0.005	11.96±0.006	25.04±0.021 [*]
55d	Control	7.33±0.666	9±0.577	1.32±0.032	11.75±0.017	16.87±0.054
	Zn	9.33±0.666 [*]	11.00±0.333 [*]	1.51±0.013	11.98±0.041	23.10±0.010 [*]
	Fe	10.00±1.15 [*]	11.33±0.333 [*]	1.41±0.021	11.82±0.003	26.86±0.018 [*]
	Zn+Fe	9.66±0.333 [*]	10.00±0.577 [*]	1.37±0.008	11.85±0.012	23.03±0.026 [*]

*Significant difference from the control at P≤0.05

** including pod

Table 2: Effect of single and combined Zn and Fe foliar application at different time period on tissue Zinc and iron concentration in different plant parts of cowpea.

Days of foliar spray	Treatments	Root	Stem	Leaf	Pod
Zinc concentration(µg g⁻¹ dry matter)					
35d	Control	8.91±0.620	25.15±0.606	18.44±0.692	29.86±0.092
	Zn	31.81±0.219 [*]	31.6±0.103 [*]	31.74±0.976 [*]	33.80±0.346 [*]
	Zn+Fe	32.07±0.875 [*]	32.20±0.499 [*]	29.92±0.798 [*]	37.63±0.941 [*]
45d	Zn	29.11±0.793 [*]	32.44±0.049 [*]	32.02±0.470 [*]	35.06±0.199 [*]
	Zn+Fe	30.61±0.155 [*]	31.17±0.586 [*]	30.41±0.280 [*]	35.41±0.536 [*]
55d	Zn	19.14±0.333 [*]	30.05±0.890 [*]	33.47±0.733 [*]	31.96±0.369 [*]
	Zn+Fe	28.50±0.800 [*]	30.99±0.505 [*]	31.39±0.176 [*]	34.32±0.730 [*]

Iron concentration ($\mu\text{g g}^{-1}$ dry matter)					
35d	Control	105.75 $\pm$ 0.016	52.52 $\pm$ 0.660	85.56 $\pm$ 1.11	58.08 $\pm$ 0.396
	Fe	116.95 $\pm$ 1.48 [*]	85.01 $\pm$ 0.626 [*]	106.20 $\pm$ 0.680 [*]	98.10 $\pm$ 0.736 [*]
	Zn+Fe	109.45 $\pm$ 0.396 [*]	69.69 $\pm$ 0.733 [*]	100.75 $\pm$ 0.666 [*]	60.20 $\pm$ 0.386 [*]
45d	Fe	117.85 $\pm$ 0.548 [*]	93.41 $\pm$ 0.303 [*]	109.50 $\pm$ 0.00 [*]	95.07 $\pm$ 0.726 [*]
	Zn+Fe	121.15 $\pm$ 0.375 [*]	78.95 $\pm$ 0.623 [*]	95.30 $\pm$ 1.40 [*]	61.45 $\pm$ 0.416 [*]
55d	Fe	108.75 $\pm$ 0.750 [*]	78.57 $\pm$ 0.763 [*]	104.00 $\pm$ 0.750 [*]	78.57 $\pm$ 0.946 [*]
	Zn+Fe	111.55 $\pm$ 0.779 [*]	83.67 $\pm$ 0.333 [*]	92.90 $\pm$ 0.660 [*]	63.21 $\pm$ 0.196 [*]

*Significant difference from the control at $P \leq 0.05$.

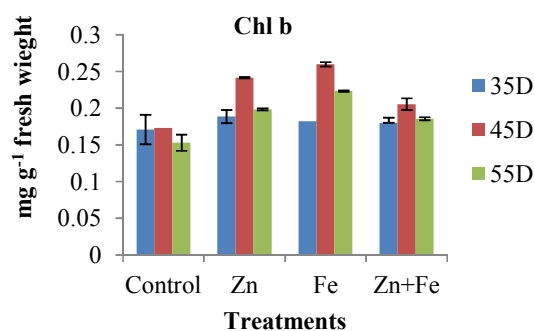
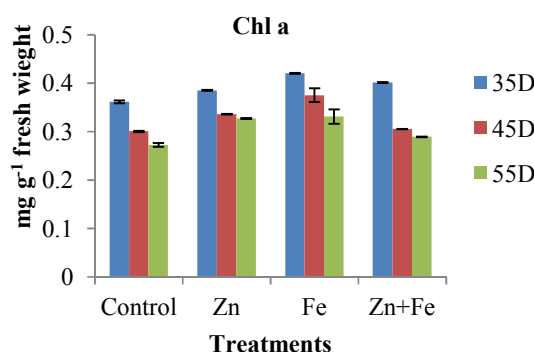


Fig: 1(a)

Fig: 1(b)

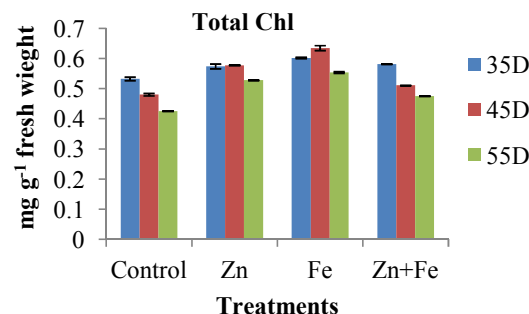
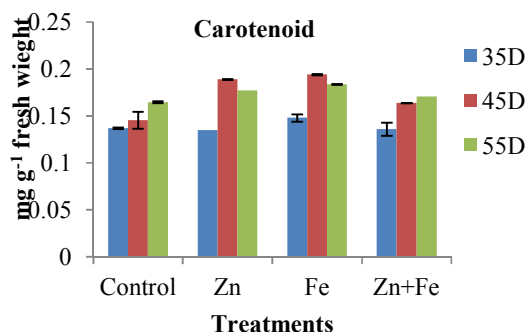


Fig: 1(c)

Fig: 1(d)

Figure 1(a-d): Effect of foliar application of Zn, Fe and Zn+Fe on chlorophyll a, chlorophyll b, carotenoid and total chlorophyll in cowpea leaf at different time period of plant growth. Vertical line in each represents $\pm$ SE.

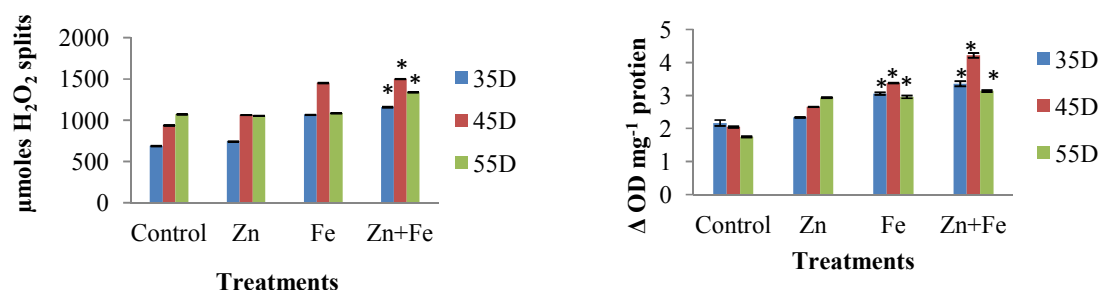


Fig: 2(a)

Fig: 2(b)

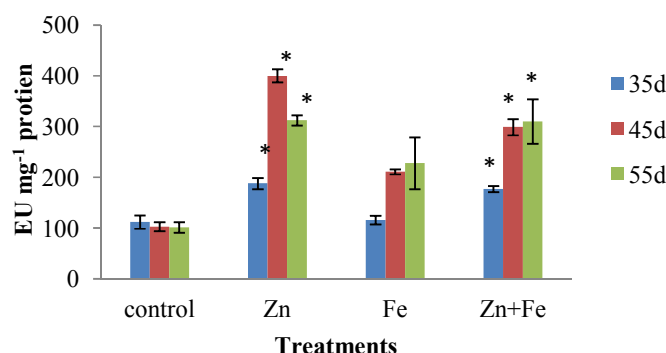


Fig: 2(c)

Figure 2(a-c): Effect of foliar application of Zn, Fe and Zn+Fe on catalase, peroxidase and carbonic anhydrase activity in cowpea leaf at different time period of plant growth. Vertical line in each represents $\pm$ SE. Asterisk (*) denotes significant difference from the control at $P \leq 0.05$

4. Discussion

After harvesting crop from experimental soil, zinc becomes slightly available to the plant might be due to high capacity of fixation (Takkar and Randhwa 1980 and Singh et al. 1994) and iron due to its oxidizing nature ($\text{Fe}^{2+} = \text{Fe}^{3+} + e^-$) (Guerinot and Yi 1994; Mori 1999; Zuo and Zhang, 2011; Samaranayake et al. 2012). In the present experiment the plants grown without any application of nutrients later showed reduction in the number of pod due to suppression of branching; reduction in number of seeds per plant and marked delay in flowering. Flower buds and open flowers were shed prematurely which resulted into reduced pod formation (table 1). Seed yield might be due to increase in shoot branching, early flowering and decrease in premature fall of flower buds and flowers. Metal accumulation in the edible plant part (seed) is correlated with the metal concentration in the substrate. In the foliar application leaves are the primary sites to which metal entered through stomata and showed that foliar application of Zn in leaves directly related to Zn concentration in both treatments single and combined (table 2). In combined treatment of Zn and Fe enhanced the mobility of Zn in their interaction. The leaves showed least accumulation Zn in combined treatment of Zn+Fe and Fe in single treatment which might reflect that most of metal is translocated to edible plant part (seeds). Gravel et al. (2000) reported least content of heavy metals accumulation in stem and most of metal

translocated to leaves in some medicinal plants. In combined foliar application of Fe and Zn concentration there was decrease in Fe content in the various parts of cowpea might be due to interaction between Zn and Fe for the same physiological binding site and interference the availability of Fe, leading to impairment of Fe metabolism at same concentration (Rosen et al. 1977). In combined treatment, Fe assess competitive effect, showed less translocation and accumulation from leaf to seed and stem as compare to single Fe treated plants. The data indicated that optimum soil Zn and Fe in experimental soil and foliar application of single treatment of Zn at day of 45 and combined treatment Zn+Fe at 35d markedly increased the Zn seed density in cowpea respectively. These results are in support with the results of Phattarakul et al. (2012).

The chlorophyll contents were increased in leaves of cowpea treated with Fe in single and combined with Zn. The chlorophyll contents in leaves are directly linked to Fe concentration applied to the plants (Bisht et al. 2002). The increase in chlorophyll concentration ultimately increased photosynthesis and production of photosynthetic product, might be a probable reason for the enhanced plant biomass. Catalase and peroxidase are free radical scavenging enzymes; their activity might depend upon availability of oxidizing substrate (H_2O_2). Foliar treatment of Fe at 45d was more significant and increased catalase activity, level of H_2O_2 decreased in leaves which resulted in enhancement of plant growth and yield. The increased specific activity of peroxidase might be attributed to the concentration of free radicals and re-synthesis of membrane bound enzyme (Crichton et al. 2002). Peroxidase decomposes H_2O_2 by oxidation of phenolic compounds which might be accumulated under Fe stress condition at 55 d and Fe supply is not useful for plant growth. Carbonic anhydrase is hydro-lyase enzyme that catalyzes the reversible hydration of carbon dioxide, increasing absorbtion of carbon dioxide per unit leaf area, thereby increasing photosynthesis and biomass production (Marschner 1986). At 45d Zinc foliar application significantly and increased yield and tissue Zn density in cowpea seed.

5. Conclusion

From the observed results in cowpea it finds that combined treatment of Zn with Fe significantly increased biomass, seed yield, enzyme activities (catalase and peroxidase) and tissue Zn concentration in cowpea seed at 35d. But carbonic anhydrase activity was more significant in single treatment of Zn. Fe in single treatment was more significant as compared to combined treatment of Zn+Fe. The observed data of foliar application of Zn and Fe at 35d might be more profitable for seed Zn and Fe density in cowpea.

7. References

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