

Evaluation of Heavy metal Content of Soil Sample from Njere River in Umuakam Okaiuga Nkwoegwu, Umuahia North L.G.A of Abia State

ABSTRACT

Aim: To evaluate the heavy metal content of soil samples from Njere river bank.

Study design: Soil samples were sourced from three different locations, upstream, midstream and downstream of the river bank.

Place and Duration of Study: Department of Chemistry, Michael Okpara University of Agriculture, Umudike, between June 2009 and December 2009.

Methodology: In each location, soil sample was collected from various points and pooled together. The soil samples were air-dried and used for Atomic Absorption Spectroscopy.

Results: Total Ni and Zn concentrations were below W.H.O standard in the upstream and midstream samples. Fe and Pb were not detected in the midstream and downstream samples. Soluble concentrations of Ni, Fe and Zn and exchangeable concentrations of Ni, Cd, Fe, Zn and Pb were below the standard.

Conclusion: The results reveal that the heavy metal contamination of the soil around the Njere river bank is very low. The soil can support production of healthy crops for food security in the area.

Keywords: Soil, Heavy metals, metalloid, Environment, Locations, River

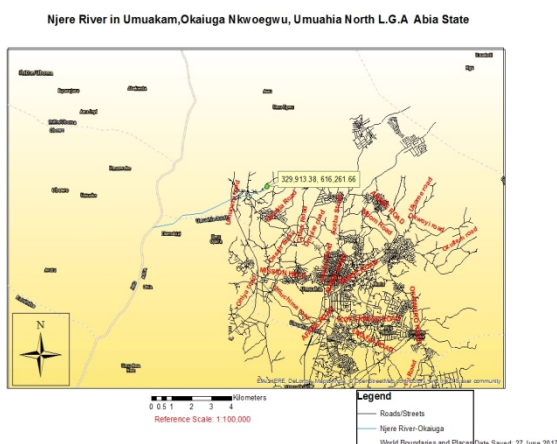
1. INTRODUCTION

The increase in pollution of soil and environment by heavy metals as well as its effect on food security is an issue of great concern all over the world [1]. Heavy metals are usually found naturally in soils at concentrations that are non-toxic [2] [3]. Their concentrations in the environment often become high primarily due to man-made activities which eventually becomes detrimental to plants and living organisms [4] [5] [6] [7]. In developing countries where industrial activities is heightening, indiscriminate disposal of wastes and effluents containing heavy metal compounds coupled with inefficient method of remediation, help to increase their concentrations in the environment. Heavy metals are usually found as single elements or in combination with soil components. The concentration of heavy metals is often higher in the top soil region [8] where nutrient uptake mainly occurs [9]

Heavy metals have been defined to include transition metals, some metalloids, lanthanides and actinides and metallic elements whose specific density is greater than that of water (5 g cm^{-3}) [10]. They are broadly divided into essential and non-essential heavy metals. Essential heavy metals are necessary for some biochemical and physiological functions in plants and animals. They include: cobalt (Co), copper (Cu), chromium (Cr), iron (Fe), magnesium (Mg), manganese (Mn), molybdenum (Mo), nickel (Ni), selenium (Se) and zinc (Zn) [11]. Heavy metals with no verified biological role such as aluminium (Al), antimony (Sb), arsenic (As), bismuth (Bi), cadmium (Cd), gallium (Ga), germanium (Ge), gold (Au), indium (In), lead (Pb), mercury (Hg), nickel (Ni), platinum (Pt), silver (Ag), tellurium (Te), thallium (Tl), tin (Sn), titanium (Ti), vanadium (V) and uranium (U) are classified as non-essential [12].

Reports from some researchers have shown that certain physicochemical properties of soils like dissolved organic carbon, cation exchange capacity, oxidation-reduction potential as well as pH influence the bioavailability of heavy metals in soil [13][14]. Heavy metals are more tightly bound to soils with high

clay and organic matter content [15]. When heavy metals combine with soil at even low concentrations, they hinder nutrient availability to plants [16]. Heavy metals resist chemical and microbial degradation and can enter the food chain. The frequency, toxicity and potency of As, Pb, Hg and Cd has been rated first, second, third and seventh respectively in humans [17]. Arao et al. [18], in a study conducted in Japan reported that soils and agricultural products became contaminated when heavy metal levels are beyond permissive levels. Khan et al. [19], reported that ingestion of food crops grown on soils irrigated with waste water stand the risk of accumulating significant amounts of metals. Ayodele et al. [20], assessed the heavy metal content of soil and water from an abandoned granite quarry. Another researcher Adama et al. [21] assessed the heavy metal content of soils around a hospital waste incinerator. The present study assessed the total, soluble and exchangeable metal ions in the soils from around the Njere river bank located in Umuakam Okaiuga, Nkwoegwu Umuahia South L.G.A, Abia state (Longitude N 05°34'24.3" and latitude E 007°27'52.0").



2. MATERIALS AND METHODS

2.1 SAMPLE COLLECTION AND PREPARATION

Soil samples were collected from three different points: in the upstream, midstream and downstream locations around the bank of the Njere river. In each location, soil was collected from different points with the aid of a clean trowel and pooled together to form a composite sample for each location. The samples were stored in clean polythene bags and carried to the laboratory. Thereafter, the samples were air-dried, milled and sieved using 0.01 μm sieve. The soil samples were collected in June, 2009.

2.2 DETERMINATION OF TOTAL METAL CONCENTRATION

One gram of soil sample was weighed into a conical flask. To it was added 15 ml of HNO_3 and 10 ml of HCl . The resulting mixture was digested by heating it in a fume cupboard at 100°C for 30 minutes. After this, 25 ml of deionized water was added and heated for another 90 minutes. Thereafter, it was allowed to cool, filtered into 50 ml volumetric flask and made up to mark with deionized water. Thereafter, 10 ml of the above solution was poured into clean sample bottles for determination of total metal concentration using atomic absorption spectrophotometer.

2.3 DETERMINATION OF WATER SOLUBLE METAL CONCENTRATION

Twenty mls of deionized water was poured into a flask containing 1g of soil sample and left to stand for 30 minutes. The content was filtered into 50mls volumetric flask using whatmann no 1 filter paper and made up to mark with deionized water. Ten of the filtrate was poured into clean sample bottle for determination of water soluble metal concentration with AAS.

2.4 DETERMINATION OF EXCHANGEABLE METAL FRACTION

Into a clean conical flask was added 1 g of soil sample and 20 ml of 1M solution of $MgCl_2$ and the pH was adjusted using potassium phosphate buffer. After that, it was boiled inside a fume cupboard for 1 hour, filtered with whatmann no 1 filter paper into a volumetric flask and deionized water was added to make up the content to the 50 ml mark. Ten ml of the filtrate was measured out into clean sample bottle for determination of exchangeable metal fraction using AAS.

STATISTICAL ANALYSIS

The results are means of duplicate determinations and analyzed using Microsoft excel package, 2010.

3. RESULTS AND DISCUSSION

Table 1 shows the total metal ion concentration of soils from around the Njere river. The concentration of Ni and Zn were below the W.H.O standard. Cd was above the W.H.O standard of < 0.1 in the upstream and midstream samples. Fe and Pb were not detected in the midstream and downstream samples. However, the concentration of Fe and Pb in the upstream sample did not exceed the W.H.O standard given.

Table 1. Total metal ion concentration in soil samples from around Njere river bank

Type of Metal Ion	Sample Location	Mean metal ion conc (ppm)	WHO (ppm)(2005)	Standard
Ni	Upstream	0.118	10-50	
Ni	Midstream	0.076	10-50	
Ni	Downstream	0.075	10-50	
Cd	Upstream	0.315	<0.1	
Cd	Midstream	0.196	<0.1	
Cd	Downstream	0.018	<0.1	
Fe	Upstream	1.688	10-1000	
Fe	Midstream	ND	10-1000	
Fe	Downstream	ND	10-1000	
Zn	Upstream	0.674	60-780	
Zn	Midstream	0.122	60-780	
Zn	Downstream	0.142	60-780	
Pb	Upstream	0.208	2-13.4	
Pb	Midstream	ND	2-13.4	
Pb	Downstream	ND	2-13.4	

Values are means of duplicate determinations.

Table 2 is the result of soil water soluble metal ions from Njere river. Ni, Fe and Zn levels were below W.H.O standard. Pb was not detected in the soil samples from the three locations. Cd was not detected in the upstream and midstream sample. Cd level was above the W.H.O standard in the downstream sample.

Table 2. Exchangeable metal ion concentration of soil samples from around Njere river bank

Type of metal ion	Sample Location	Mean metal ion conc (ppm)	WHO Standard (ppm)
Ni	Upstream	0.152	10-50
Ni	Midstream	0.181	10-50
Ni	Downstream	0.179	10-50
Cd	Upstream	0.02	<0.1
Cd	Midstream	0.033	<0.1
Cd	Downstream	0.088	<0.1
Fe	Upstream	0.079	10-100
Fe	Midstream	0.099	10-100
Fe	Downstream	0.08	10-100
Zn	Upstream	0.016	60-780
Zn	Midstream	0.123	60-780
Zn	Downstream	0.125	60-780
Pb	Upstream	0.023	2-13.4
Pb	Midstream	0.019	2-13.4
Pb	Downstream	0.026	2-13.4

Values are means of duplicate determinations.

Table 3 is the result of the exchangeable metal ions of soil samples from around the Njere river. Ni, Cd, Fe, Zn and Pb levels were below the W. H.O standard.

Table 3. Exchangeable metal ion concentration of soil samples from around Njere river bank

Type of metal ion	Sample Location	Mean metal ion conc (ppm)	WHO Standard (ppm)
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Zn	Upstream	0.016	60-780
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Zn	Downstream	0.125	60-780
Pb	Upstream	0.023	2-13.4
Pb	Midstream	0.019	2-13.4
Pb	Downstream	0.026	2-13.4

Values are means of duplicate determinations.

Ni, Cd, Fe and Zn are transition metals. Ni, Fe and Zn are essential at low concentrations. Above the maximum permissible level, Ni can be very toxic. It is responsible for decreased chlorophyll content, nutrient uptake, inhibition of enzyme growth and activities in plants; alteration of cell membrane and oxidative stress in microorganisms; and health disorders such as cancer, cardiovascular, skin, respiratory and kidney diseases [22] [23]. The total Ni concentration in the soil samples is well below the W.H.O standard and a possible indication of an environment with poor industrial activities like painting that contributes to the natural level of Ni in the soil [24]. The higher concentration of soluble Ni ion in the downstream soil sample could be attributed to bioaccumulation due to the direction of the flow of the river. The Ni concentration in this study is within the safe limits.

Cadmium has no known biological role [25]. Cd causes chlorosis, decrease in plant nutrient content, growth inhibition, and reduced seed germination in plants. It damages nucleic acid, denatures proteins, and inhibits cell division and transcription, carbon and nitrogen mineralization in micro-organisms. It is responsible for bone disease, coughing, emphysema, headache, hypertension, itai-itai, kidney diseases, lung and prostate cancer, lymphocytosis, microcytic hypochromic anemia, testicular atrophy, vomiting in man [26] [27].

Zinc affects photosynthesis, inhibits growth rate, reduced chlorophyll content, germination rate and plant biomass. In humans, it is a leading cause of Ataxia, depression, gastrointestinal irritation, haematuria, icterus, impotence, kidney and liver failure, lethargy, macular degeneration, metal fume fever, prostate cancer, seizures, vomiting [28]. Lead is a group IV metal that affects photosynthesis and growth, chlorosis, inhibit enzyme activities and seed germination. It causes Anorexia, chronic nephropathy, damage to neurons, high blood pressure, hyperactivity, insomnia, learning deficits, reduced fertility, renal system damage, risk factor for Alzheimer's disease, shortened attention span [29] [30]. In the present study, the concentrations of the soluble and exchangeable metal ions were all higher in the downstream soil samples. This could be because the downstream location can be said to be a dumping site and hence the accumulation of various heavy metals. However, the observed concentrations were still within the safe limits.

4. CONCLUSION

Soils around the Njere river are still relatively safe for agricultural purposes. It will enhance production of quality agricultural food products and help attain food security.

REFERENCES

1. Kong XB. China must protect high-quality arable land. *Nature*, 2014; 506:7-7.
2. Kabata-Pendias A, Pendias H. *Trace Metals in Soils and Plants*, CRC Press, Boca Raton, Fla, USA, 2nd edition, 2001.
3. Zhao Q, Kaluarachchi JJ. Risk assessment at hazardous waste-contaminated sites with variability of population characteristics," *Environment International*, 2002;28(1-2):41–53.
4. J. J. D'Amore SR, Al-Abed KG, Scheckel KG, Ryan JA. Methods for speciation of metals in soils: a review. *Journal of Environmental Quality*, 2005; 34(5):1707–1745.
5. Nouri J, Khorasani N, Lorestani B, Karami M, Hassani H, Yousefi N. Accumulation of heavy metals in soil and uptake by plants species with phytoremediation potential. *Environ Earth Sci*. 2009;59:315–323.
6. Olowoyo JO, Okedeyi OO, Mkolo NM, Lion GN, Mdakane STR. Uptake and translocation of heavy metals by medicinal plants growing around a waste dump site in Pretoria, South Africa. *S Afr J Bot*. 2012;78:116–121. doi: 10.1016/j.sajb.2011.05.010.
7. Wójcik M, Sugier P, Siebielec G. Metal accumulation strategies in plants spontaneously inhabiting Zn-Pb waste deposits. *Sci Total Environ*. 2014;487:313–322. doi: 10.1016/j.scitotenv.2014.04.024.
8. Q. Hou, Z. Yang, J. Ji, T. Yu, G. Chen, J. Li, X. Xia, M. Zhang, X. Yuan Annual net input fluxes of heavy metals of the agro-ecosystem in the Yangtze River delta, China *J. Geochem. Explor.*, 139 (2014), pp. 68–84
9. T. Kismányoky, Z. Tóth Effect of mineral and organic fertilization on soil fertility as well as on the biomass production and N utilization of winter wheat (*Triticum aestivum* L.) *Arch. Agron. Soil Sci.*, 56 (4) (2010), pp. 473–479.
10. Babula P, Adam V, Opatrilova R, Zehnalek J, Havel L, et al. (2008) Uncommon heavy metals, metalloids and their plant toxicity: a review. *Environmental Chemistry Letters* 6(4): 189-213.
11. WHO/FAO/IAEA. World Health Organization. Switzerland: Geneva; 1996. Trace Elements in Human Nutrition and Health.
12. Chang LW, Magos L, Suzuki T, editors. *Toxicology of Metals*. Boca Raton. FL, USA: CRC Press; 1996.
13. Violante A, Cozzolino V, Perelomov L, Caporale AG, Pigna M. Mobility and bioavailability of heavy metals and metalloids in soil environments. *J Soil Sci Plant Nutr*. 2010;10:268-92.
14. An J, Jho EH, Nam K. Effect of dissolved humic acid on the Pb bioavailability in soil solution and its consequence on ecological risk. *J Hazard Mater*. 2015;286:236-41. doi:10.1016/j.jhazmat.2014.12.016
15. Kulikowska D, Gusiati ZM, Bułkowska K, Klik B. Feasibility of using humic substances from compost to remove heavy metals (Cd, Cu, Ni, Pb, Zn) from contaminated soil aged for different periods of time. *J Hazard Mater*. 2015; 300:882-891.

16. Gebreyesus ST. Heavy metals in contaminated soil: Sources and washing through chemical Extractants. American Scientific Research Journal of Engineering, Technology and Sciences (ASRJETS),2014;10(1):54-60.
17. Agency for Toxic Substances and Disease Registry (ATSDR) (2015) CERCLA priority list of hazardous substances.
18. Arao T, Ishikawa S, Murakami M, Abe K, Maejima Y, Makino T. Heavy metal contamination of agricultural soil and counter measures in Japan. Paddy and water Environment, 2010;8(3):247-257.
19. Kahn S, Cao Q, Zheng YM, Huang YZ, Zhu YG. Health risks of heavy metals in contaminated soils and food crops irrigated with wastewater in Beijing, China. Environmental Pollution, 2008; 152 (3):686-692.
20. Ayodele OJ, Shittu OJ, Balogun T. Heavy metal pollution assessment of granite quarrying operations at Ikole Ekiti, Nigeria. Int. J. Env. Monit. Anal. 2014;2(6):333-337.
21. Adama M, Esena R, Fosu-Mensah BK, Yirenya-Tawiah, D. Heavy metal contamination of soils around a hospital waste incinerator bottom Ash dumps site. J. Environ Public Health; 2016;1-6.
22. Fashola M, Ngole-Jeme V, Babalola O. Heavy metal pollution from gold mines: Environmental effects and bacterial strategies for resistance. Int. J. Environ. Res. Public Health 2016; 13 (1047): 2-20.
23. Chibuike G, Obiora S. Heavy metal polluted soils: Effect on plants and bioremediation methods. Appl. Environ. Soil Sci. 2014;1-12.
24. Malik A. Metal bioremediation through growing cells. Environ. Int. 2004;30: 261-278.
25. Manahan SE. *Toxicological Chemistry and Biochemistry*, CRC Press, Limited Liability Company (LLC), 3rd edition, 2003.
26. Sebogodi KM, Babalola OO. Identification of soil bacteria from mining environments in Rustenburg, South Africa. Life Sci. J. 2011; 8: 25-32.
27. Sankarammal M, Thatheyus A, Ramya D. Bioremoval of cadmium using pseudomonas fluorescens. Open J. Water Pollut. Treat. 2014;1: 92-100.
28. Gumpu MB, Sethuraman S, Krishnan UM, Rayappan JBB. A review on detection of heavy metal ions in water—An electrochemical approach. Sens. Actuators B Chem. 2015; 213: 515-533.
29. Wuana RA, Okieimen FE. Heavy metals in contaminated soils: A review of sources, chemistry, risks and best available strategies for remediation. ISRN Ecol. 2011; 1-20.
30. Mupa M. Lead content of lichens in metropolitan Harare, Zimbabwe: Air quality and health risk implications. Greener J. Environ. Manag. Public Saf. 2013; 2:75-82.