# **Original Research Article**

### BIOREMEDIATION OF HEAVY METALS IN THE SOIL BY PSEUDOMONAS AERUGINOSA AND TRICHODERMA HARZIANUM USING SOLANUM LYCOPERSICUM AS TEST PLANT

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### 7 ABSTRACT

8 This study determined the heavy metal concentrations of contaminated stream water and assessed the heavy metal 9 contents of pre- and post-cropped sterilized soil. It also determined the heavy metal uptake of the S. 10 lycopersicum plant. This was with a view to assessing the potential of Pseudomonas aeruginosa and Trichoderma 11 harzianum for degrading heavy metals in heavy metal contaminated stream water. Experimental pots containing 12 3000 g of sterilized soil was used for this experiment whereby 60 sample pots were used with various treatments in 13 this study. Solanum lycopersicum seeds were raised in the nursery for a period of 3 weeks and treatments applied 14 just before transplanting into the experimental pots. The plants were left for a week so as to be established properly 15 and overcome transplanting shock before watering with the contaminated stream water. Heavy metal analysis using 16 Atomic Absorption Spectroscopy (AAS) method was carried out on the contaminated stream water to determine the 17 amount of heavy metal in the stream water before the commencement of the experiment. The contaminated stream 18 water was applied to the pots in measured quantities; 0, 5 and 0%. Pre and post soil heavy metal analysis were 19 carried out on the soil samples. At harvest, plant tissues were analysed for heavy metals using AAS method. The 20 results showed that heavy metals were present in high concentration in the stream water sample. The values of the 21 heavy metals in the stream water sample used for watering were Iron - 138.15 mg/L, Zinc - 68.4 mg/L, Lead - 7.89 22 mg/L and Copper – 8.98 mg/L. Heavy metal analysis of the soil and all the treatments revealed that treatments with 23 P. aeruginosa inoculation had the lowest level of Iron, Copper, Zinc and Lead followed by treatments inoculated 24 with T. harzianum. The study concluded that the use of contaminated stream water for irrigation could be a potential 25 source of heavy metals in tomato. However, inoculation of microorganisms for the treatment of the heavy metal 26 contaminated sites was effective for increased health, growth and yield of tomato fruits.

27 KEWORDS: Bioremediation, Pseudomonas aeruginosa, Trichoderma harzianum, Solanum lycopersicum

### 28 INTRODUCTION

29 Heavy metals represent a great environmental concern, because of their widespread use and distribution, and 30 particularly their toxicity to human beings and the biosphere. However, they also include some elements that are 31 essential for living organisms at low concentrations (Alloway, 1990). These elements are usually transition metals. 32 They have high densities (>5 g cm-3) when compared with other materials (Baird and Cann, 2005). Human 33 activities such as industrial production, mining, agriculture and transportation lead to release of high amount of 34 heavy metals into the biosphere. The primary sources of metal pollution are the burning of fossil fuels, smelting of metal like ores, municipal wastes, fertilizers, pesticides and sewage (Nriagu, 1979, 1996; Pendias and Pendias, 1989; 35 36 Rai, 2009). Heavy metal contamination may occur due to factors which could include irrigation with contaminated 37 water, addition of fertilizers and metal based pesticides, industrial emissions, and transportation (Radwan and 38 Salama, 2006; Tuzen and Soylak, 2007; Duran et al., 2007). Heavy metal pollution does not only affect the 39 production and quality of crops, it also influences the quality of the atmosphere and water bodies. This threatens the 40 health and life of animals as well as human beings by the way of food chain and most phenomenal is that, this kind 41 of pollution is covert, long term and non-reversible (Zhang, 1999). Heavy metals are also one of the major 42 contaminating agents in our food supply (Zaidi et al., 2005; Khair, 2009). Bioremediation is a process that uses

43 naturally occurring micro-organisms to transform harmful substances to nontoxic compounds, these processes which 44 take advantage of microbial degradation of organic and inorganic substances can be defined as the use of micro-45 organisms to remove environmental pollutants of soils, water and sediments (Pala et al., 2006). Bioremediation 46 involves the use of organisms for the treatment of polluted soils. These organisms which could be micro-organisms 47 or green plants eliminate, attenuate or transform the harmful substances via biological processes to a less harmful 48 substance (Mrayyana and Battikhi, 2005). Micro-organism breaks down organic molecules to carbondioxide, 49 fattyacid and water in order to obtain energy and nutrients. Bioremediation occurs naturally (even though it could be 50 enhanced by a number of processes), thus, it is widely accepted by the general public as a safe way of treating 51 polluted soils. Trichoderma harzianum has potential in stimulating phytoremediation directly and indirectly and 52 therefore, inoculation of plants with this fungus could be a feasible approach to enhance the degradation of 53 hydrocarbons in polluted soil. T. harzianum also have the ability to solubilize metal ions and produce siderophores 54 to chelate iron, making metal ions required for plant growth more available to the plant (Harman et al., 2004). The 55 fungus is thought to colonize roots of annual plants for their entire lifetime by penetrating the outer layers of the 56 roots (Harman et al., 2004). This makes the plants release more root exudates to the surrounding soil, thus, 57 stimulating microbial degradation of pollutants. Trichoderma harzianum has been shown to induce the production of 58 larger and deeper root systems, and plants inoculated with Trichoderma harzianum also produce greater plant 59 biomass. Such plants are more resistant to abiotic stress and take up nutrients more effectively (Harman et al., 60 2004). Edwards et al., (2006) noted that various bacteria such as Pseudomonas aeruginosa produce surfactants that 61 aid in the biodegradation. A recent study has found a P. aeruginosa strain that actually supports plant growth. This 62 characteristic, along with the fact that P. aeruginosa can degrade polycyclic aromatic hydrocarbons, suggests the 63 future uses of P. aeruginosa for environmental detoxification of synthetic chemicals and pesticides and for industrial

- 64 purposes (Botzenhardt and Doring, 1993).
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### 66 MATERIALS AND METHOD

### 67 Collection of Contaminated water, seeds and microorganisms

- Heavy metals contaminated stream water was obtained from a flowing stream. It is situated at 7°30' Northern
   latitude and 4°28' Eastern longitude. The sampling point was located at the back of the Ife Iron and Steel Nigeria
- 70 Limited along Ife-Ibadan expressway. Surface water samples was collected at downstream into clean plastic kegs.
- 71 The water samples were collected during the month of April, 2015. Seeds of *Solanum lycopersicum* cultivar (ROMA
- 72 VF) were obtained from Institute of Agricultural Research and Training, Moor Plantation, Ibadan.

### 73 Culturing of Organisms

- 74 A culture of Pseudomonas aeruginosa was obtained from the Department of Microbiology, Obafemi Awolowo
- 75 University (OAU), Ile-Ife. A culture of Trichoderma harzianum was also obtained from the Mycology unit of the
- 76 Department of Crop Production and Protection, OAU, Ile-Ife, A single colony of P. aeruginosa was subcultured by
- (177) (using nutrient agar in petri dishes and kept in the incubator for 48 hours at 37°C to a medium after which it was
- (harvested by flooding with sterile distilled water. The bacterium inoculum was prepared by streaking a single colony
- 79 of *P. aeruginosa* earlier isolated on plated nutrient agar plate and incubated at 37°C for 48 hours. Cells of *P.*
- 80 *aeruginosa* were harvested from agar plates by flooding with sterile distilled water and standardized using a
- 81 colorimeter to  $10^8$  CFU/ml. Spores of *Trichoderma harzianum* was subcultured by using potato dextrose agar in 82 petri dishes and kept in the incubator for 7 days at 37°C to a medium after which it was harvested by flooding with
- (plated potato dextrose agar plate and incubated at 37°C for 7 days. Spores of *T. harzianum* were harvested from agar
- 85 plates by flooding with sterile distilled water and standardized using a colorimeter to  $10^7$  spores/ml.
- 86 Planting of seeds and contamination of experimental pots
- 87 Seedlings of S. lycopersicum were raised on nursery beds for a period of three weeks. Sixty pots, each containing
- 88 three kilograms of soil from sterilized soil was used for this study. *Pseudomonas aeruginosa* inoculum solution (30
- 89 ml) was poured into a hole that was made in the middle of a set of 15 experimental pots containing sterized soil
- 90 before S. lycopersicum seedlings are transplanted to it. Trichoderma harzianum spore solution (30 ml) was also

91 poured into a hole that was made in the middle of another set of 15 experimental pots before S. lycopersicum 92 seedlings are transplanted to them. The third set of 15 pots received dual inoculation of Trichoderma harzianum

93 spore solution (15 ml) and P. aeruginosa innoculum before S. lycopersicum seedlings were transplanted into it; with

- 94 the final set of 15 pots acting as control at various levels. Thereafter, pot preparation was arranged in a completely
- 95 randomized design in the screenhouse.

96 Seedlings were left for a week to establish and overcome transplanting shock before wetting with the contaminated

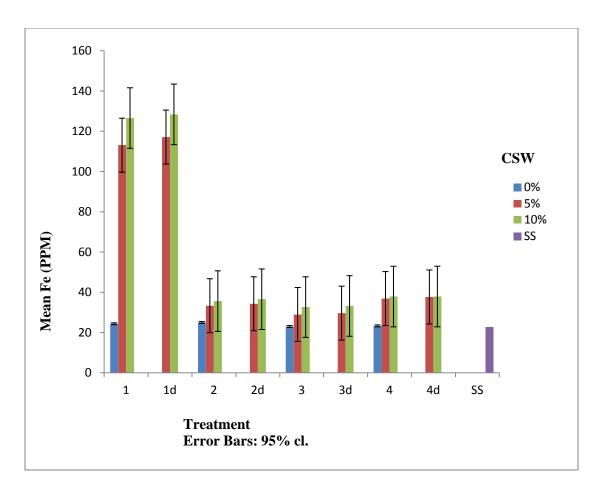
97 stream water at various concentrations of 0%, 5% and 10% v/v. Contaminated stream water was quantified using the 98 formula: percentage soil contamination = (Volume of polluted stream water applied / Volume of soil) x 100. Each

99 treatment of the experiment was replicated three times. Twenty four pots were watered with the contaminated stream

- 100 water once during the experiment and another 24 pots watered daily with the contaminated stream water. The
- 101 remaining 12 pots which served as the control experiment were watered daily with distilled water. Pots containing S.
- 102 lycopersicum was watered regularly to ensure adequate moisture. Heavy metal analysis on the contaminated stream
- 103 water was carried out using AAS (Atomic Absorption Spectrophotometer) for Iron, Copper, Lead, and Zinc pre
- 104 experiment. Plant samples were also subjected to heavy metal analysis using AAS (Atomic Absorption
- 105 Spectrophotometer) for Iron, Copper, Lead, and Zinc post experiment. Pre and post - soil tests were carried out to
- 106 determine soil nutrients. Soil samples were also subjected to heavy metal analysis using AAS (Atomic Absorption
- 107 Spectrophotometer) for Iron, Copper, Lead, and Zinc pre and post - soil tests. Data obtained was subjected to
- 108 statistical analysis using descriptive and inferential methods.

#### 109 RESULTS

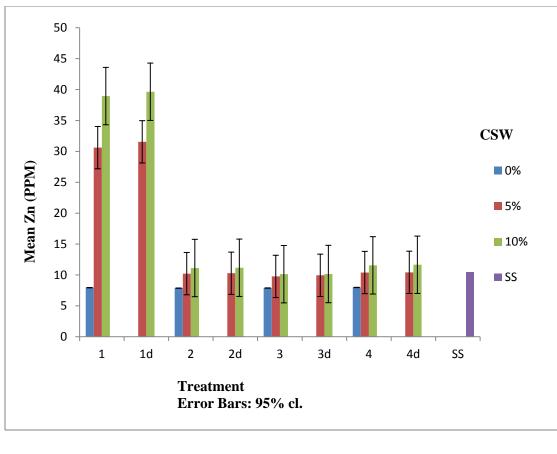
- 110 The heavy metals analysis of the stream water showed that heavy metals (Iron, Zinc, Copper and Lead) were present
- 111 in high concentration in the water. Iron (Fe) had the highest concentration of 138.15 mg/L followed by zinc (Zn) 112 which had a concentration of 68.4 mg/L. The order of concentration was Fe>Zn>Cu>Pb.
- 113 After the soils were subjected to heavy metal analysis, it was observed that iron concentration of the soil increased
- as the contaminated stream water concentration increased in all the treatments without any inoculation of 114 115 microorganism (Fig. 1). Treatments 3 and 3d inoculated with P. aeruginosa were lower in concentration of iron
- compared to treatments 2 and 2d which were inoculated with T. harzianum. Treatment 1d without any inoculation of 116
- 117 microorganisms had highest iron concentration followed by treatment 1 also without any inoculation of
- 118 microorganisms at 5% and 10% contaminated stream water concentration. The order of iron concentration across the
- 119 treatments with 5% and 10% contaminated stream water concentration was 1d>1>4d>4>2d>2>3d>3 and
- 120 1d>1>4d>4>2d>2>3d>3 respectively.
- 121 Treatment 1d had the highest zinc concentration at 10% contaminated stream concentration followed by treatment 1
- at the same 10% concentration. Treatment 3 at 0% concentration had the lowest iron concentration. Soil samples 122
- 123 treated with single or both micro-organisms had the lowest value in zinc compared to soil polluted with
- 124 contaminated stream water without any treatment with microorganisms (Fig 2). Treatment 3 had the lowest copper 125
- level of 2.46 part per million (ppm) at 5% contaminated stream water concentration while treatment 1d had the
- 126 highest level of copper with 3.86 ppm at the same concentration (Fig 3). The order of copper concentration in 0% 127 and 10% was treatment4>2>2>1>3and 1d>1>4d>4>2d>2>3d>3 respectively. Lead analyses in the soil indicated
- 128 that the order of the concentration in 5% and 10% was 1d>1>4d>4>>2d>2>3d>3 and 1d>1>4d>4>>2d>2>3d>3
- 129 respectively, treatment 1d had the highest level of lead concentration followed by treatment 1 both at 10%
- 130 contaminated stream water concentration while treatment 2 had the lowest at 0% (Fig. 4).



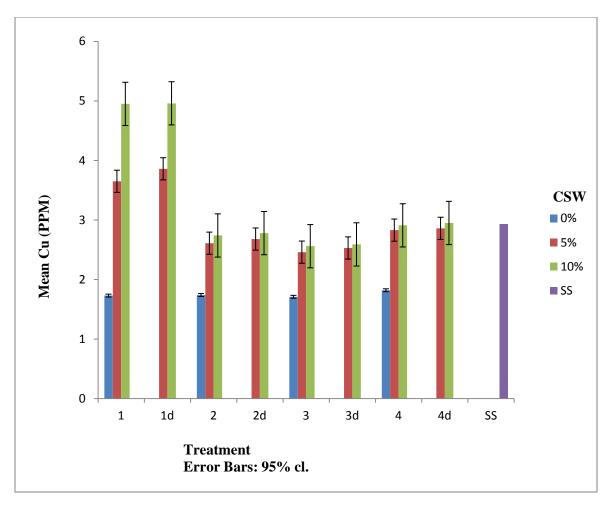
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### 133 Figure 1: Iron (PPM) content of Pre and Post Planting Soil Samples

- 134 Legend
- 135 1-SS + TP
- 136 1d-SS + TP
- 137 2-SS + TH+ TP
- 138 2d-SS + TH + TP
- **139** 3-SS + PA + TP
- 140 3d-SS + PA + TP
- $141 \qquad 4-SS + TH + PA + TP$
- $142 \qquad 4d\text{-}SS + TH + PA + TP$
- 143 SS Sterilized soil before planting
- 144 d Daily wetting of plants with contaminated stream water
- 145 Cl- Confidence level
- 146 TH T. harzianum
- 147 PA P. aeruginosa
- 148 TP Test Plant
- 149 CSW- Contaminated Stream Water

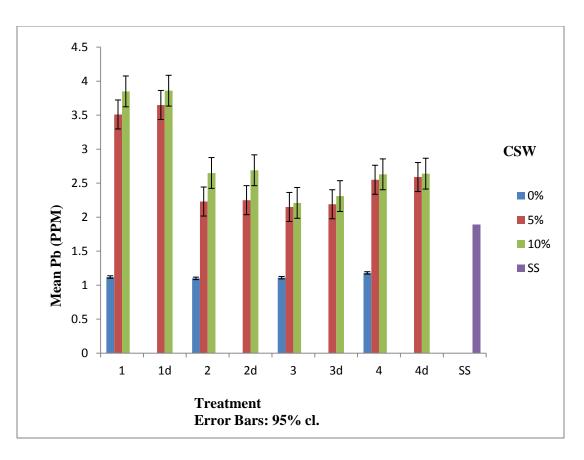


152 Figure 2: Zinc (PPM) content of Pre and Post Planting Soil Samples



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158 Figure 3: Copper (PPM) content of Pre and Post Planting Soil Samples



### 159 160

### 161 Figure 4: Lead (PPM) content of Pre and Post Planting Soil Samples

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Heavy metal analysis carried out on plant samples showed that plants from soil samples without inoculation of
micro-organisms had the highest heavy metal uptake as the concentration of contaminated stream water increased.
For 5% contaminated stream water concentration, treatment 1 had the highest level of iron at 77.78 ppm followed by
treatment1d with 77.71 ppm while treatment 3 had the lowest concentration of iron with 13.91 ppm (Fig 5).

167 The order of concentration in iron at 10% was sample 1>1d>4d>4>2d>2>3d>3. Zinc at 10% contaminated stream 168 water concentration had the highest concentration in treatment 1d and the lowest at treatment 3 at same 10%. The 169 order of zinc concentration at 5% was 1d>1>4d>4>2d>2>3d>3 while 10% was 1d>1>4d>2>2d>2>3d>3 (Fig. 6).

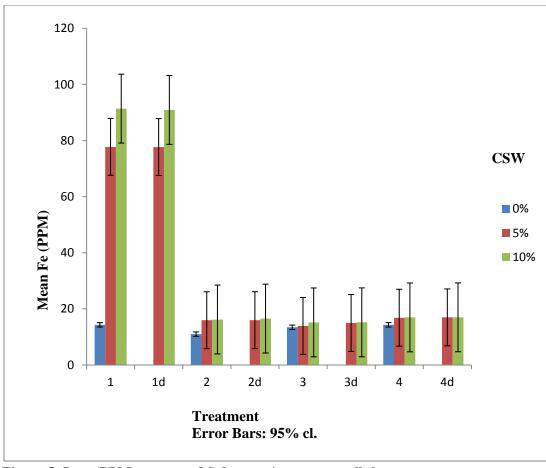
170 Copper in treatment 1d without any inoculation had the highest concentration at 5% and 10% followed by treatment

171 1 at same concentrations with treatment 3 inoculated with *P. aeruginosa* having the lowest value (Fig.7).

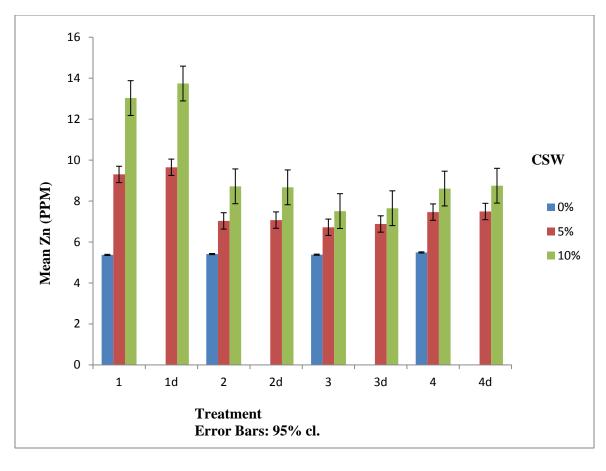
Lead content in the plant samples was highest in treatment 1d, followed by those from treatment 1 but lowest in
treatment 3. Order of increase of lead is treatment 1d>1>4d>4>2d>2>3d>3 (Fig 8). Treatments 2 and 2d inoculated

with *T. harzianum* had more of the heavy metal in plant tissue compared to treatments 3 and 3d treated with *P. aeruginosa*.

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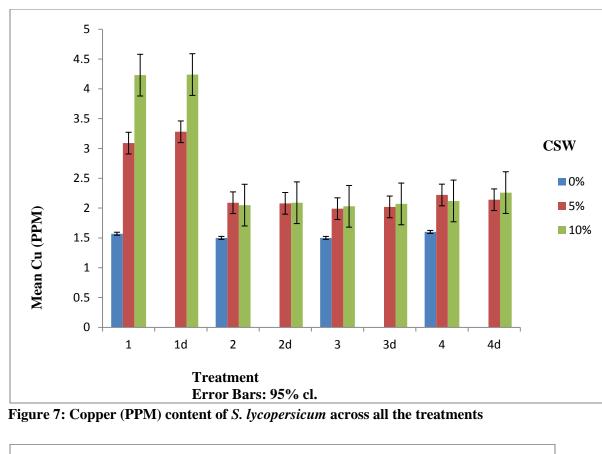
180 181 Figure 5: Iron (PPM) content of S. lycopersicum across all the treatments

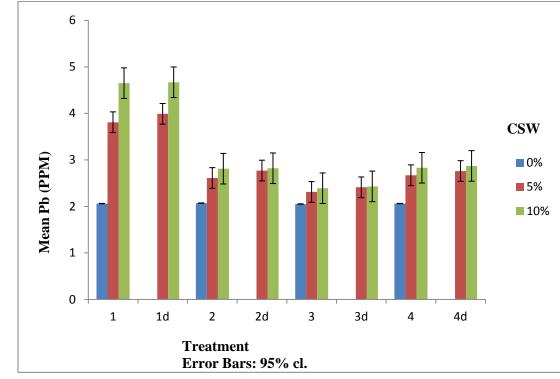


184 Figure 6: Zinc (PPM) content of *S. lycopersicum* across all the treatments

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191 Figure 8: Lead (PPM) content of *S. lycopersicum* across all the treatments

### 192

### 193 **DISCUSSION**

194 Heavy metals are elements that exhibit metallic properties such as ductility, malleability, conductivity, cation 195 stability, and ligand specificity (Opaoluwa, 2010). They are characterized by relatively high density and high 196 relative atomic weight with an atomic number greater than 20. Industrial effluents are usually considered as 197 undesirable for arable soil, plants, animals and human health. This is due to the contained heavy and trace metals 198 like Cr, Mn, Fe, Cu, Co, Zn, Ni, As, Cd and Pb that are discharged continuously into water source (streams/ nullahs, 199 canals and rivers). These are allowed to spread on agricultural lands. The unplanned disposal of these effluents has 200 increased the threat of environmental pollution (Gulfraz et al., 2003). Soils, whether in urban or agricultural areas 201 represent a major sink for metals released into the environment from a wide variety of anthropogenic sources 202 (Niragu, 1991).

203 Su et al. (2014) reported that low concentration of heavy metals could stimulate microbial growth and increase 204 microbial biomass, while high concentration could decrease soil microbial biomass significantly. The 205 microorganisms used in this study (T. harzianum and P. aeruginosa) were highly effective in biodegrading heavy 206 metals. The bio-sorption potential of the organisms used in this study showed that T. harzianum and P. aeruginosa 207 posses effective heavy metal absorption capacity. It was discovered in this study that at higher concentrations of 208 these metals, there were reductions in plant growth. This may be due to the decrease in growth parameters of S. 209 lycopersicum as the contaminated stream water concentration increased in this study. Heavy metals of soil in all the 210 soil samples showed an increase as the contaminated stream water increased in concentration. Treatments inoculated 211 with P. aeruginosa were found to have lower concentration of heavy metals (Fe, Zn, Cu and Pb) followed by 212 treatments inoculated with T. harzianum. Due to a change in their oxidation state, heavy metals can be transformed 213 to become either less toxic, easily volatilized, more water soluble (and thus can be removed through leaching), less 214 water soluble (which allows them to precipitate and become easily removed from the environment) or less 215 bioavailable (Marques et. al., 2009).

216 The biodegrading ability of *P. aeruginosa* which showed the most efficient heavy metal uptake from the soil is in 217 agreement with report of Lewis et al. (2002) and Odeyemi et al. (2011) which stated that Psedomonas spp have a 218 high biodegrading ability. Report from Jankiewicz et al. (2000) also support the findings from this study which 219 noted that *P. aeruginosa* cells grown in biofilms accumulate higher amounts of heavy metals. Also, many species of 220 soil fungi including *Trichoderma* are able to dissolve through the release of chelating compounds of organic acids. 221 The fungus releasing organic acids causes acidification of the environment, which helps increase the mobility of 222 heavy metals (Barea et al., 2005; Ledin, 2000; Wang and Chen, 2009). This study confirms this reports. Treatments 223 inoculated with dual inoculation of T. harzianum and P. aeruginosa were found to have slightly higher 224 concentration of heavy metals than treatments inoculated with P. aeruginosa or T. harzianum. However treatments 225 with no inoculation of one or two microorganisms showed very high concentration of heavy metals in the soil in comparison with treatments with dual microorganisms. This confirms that the microorganisms used in this study 226 227 biodegraded the heavy metals in the soil. This also revealed that there is positive and productive interaction between

228 *T. harzianum* and *P. aeruginosa* in bioremediation of heavy metals heavy metals polluted soil.

229 Many species of plants have been successful in absorbing contaminants such as lead, cadmium, chromium, arsenic,

and various radionuclides from soils. Some metals with unknown biological function (Cd, Fe, Zn, Cu, Cr, Pb, Co,

Ag, Se, Hg) can also be accumulated (Cho-Ruk *et al.*, 2006). Contaminant uptake by plants and its mechanisms have

been being explored by several researchers. It could be used to optimize the factors to improve the performance of
plant uptake. According to Sinha *et al.* (2008), the plants act both as "accumulators" and "excluders". Accumulators

survive despite concentrating contaminants in their aerial tissues. They biodegrade or biotransform the contaminants

into inert forms in their tissues. The excluders restrict contaminant uptake into their biomass. Plant has a lot of

consequences from heavy metal pollution in soil (Liao 1993, Su *et al.*, 2014, Wu *et al.*, 1998), plants were also seen

to be polluted by heavy metals (Yin *et al.*, 1999), which consequently threatens the health of animals and human

beings via the food chain (Wang *et al.*, 2001).

239 Heavy metals such as cadmium and lead are non-essential elements for plants. Microbial populations are generally 240 higher in the rhizosphere than in the root-free soil. This is due to a symbiotic relationship between soil 241 microorganisms and plants. This symbiotic relationship can enhance some bioremediation processes. Plant roots also 242 may provide surfaces for sorption or precipitation of metal contaminants (Sas-Nowosielska et al., 2008). This study 243 were found to show reduction in growth parameters as heavy metals increased which is brought by increase in 244 contaminated stream water concentration. Iron, Zinc, Copper and Lead level were higher in plant tissues from soil 245 samples containing no inoculation of microorganisms at 5% and 10% contaminated stream water concentration. This 246 was discovered to affect the growth of the plants. Su et al., (2014) reported that dicots, leafy vegetable crops are 247 sensitive to Zn toxicity, especially spinach and beet; because of their inherent high Zn uptake capacity. However soil 248 samples containing P. aeruginosa was generally the lowest in plant heavy metal uptake of iron, zinc, copper and 249 lead followed by samples containing T. harzianum. This may be an indication that the heavy metals in the soil had 250 been degraded by the microorganisms used which also showed there is low amount of heavy metals in soil left for 251 the plant to absorb. This result was found to be consistent with the work of Soumitra et al. (2014) which 252 demonstrated that P. aeruginosa reduced heavy metal uptake in Oryza sativa L. and increase its growth. Also 253 Trichoderma spp. produces organic acids such as gluconic acid, fumaric acid, and citric acid, which can decrease the 254 pH of the soil and allow for the dissolution of phosphate, as well as macro- and micronutrients such as iron, 255 manganese, and magnesium, which are necessary for plant metabolism (Ociepa, 2011; Cao et al., 2008) Treatments 256 inoculated with a combination of T. harzianum and P. aeruginosa in this study had lower concentration of heavy 257 metals in their plant tissue compared to treatments without inoculation of microorganisms. This may insinuate that 258 there is positive and effective interaction between T. harzianum and P. aeruginosa in the reduction of heavy metals

build up in plant cultivated on heavy metals polluted soil. concentrations of metals were attributed to the

- 260 contaminated stream water irrigation. The results from this study indicates that there is a serious potential health risk
- associated with heavy metals in tomato by using contaminated water for irrigation by farmers for tomato production.

### 263 **REFRENCES**

- Alloway, B.J. (1990). *Heavy metals in soils*. Glasgow & London: Blackie and Son Ltd, pp 12-15.
- Baird, C. and Cann, M. (2005). Environmental Chemistry. 3rd Ed. New York: W.H. Freeman and Company, pp. 89.
- Barea, J.M., Pozo, M.J., Azcón, R. and Azcón-Aguilar, C. (2005). Microbial co-operation in the rhizosphere. *Journal of Experimental Botany*, vol. 56, no. 417, pp. 1761–1778.
- Botzenhardt, K., and Doring, G. (1993). Ecology and epidemiology of *Pseudomonas aeruginosa*. *Pseudomonas aeruginosa as an Opportunistic Pathogen*. p. 1-7.
- 270 Cao, L., Jiang, M., Zeng, Z., Du, A., Tan, H. and Liu, Y. (2008). Trichoderma atroviride F6improves
- phytoextraction efficiency of mustard (Brassica juncea (L.) Coss. var. foliosa Bailey) in Cd, Ni contaminated
  soils, *Chemosphere*, vol. 71, no. 9, pp. 1769–1773,.
- 273 Cho-Ruk, K., Kurukote, J., Supprung, P., and Vetayasuporn, S. (2006). Perennial plants in the phytoremediation of
- 274 lead-contaminated soils, *Biotechnology*, vol. 5, no. 1, pp. 1–4.
- 275 Duran, A., Tuzen, M. and Soylak M. (2007). Trace element levels in some dried fruit samples from Turkey.
  276 *International Journal of Food Science and Nutrition*, 59:581–589.
- Edward, R.B.M., Brian J.T., Vitor, A.P.M., Dietmar, H.P., Juan-Luis, R. and Norberto, J.P.(2006). Nonmedical:
  Pseudomonas. *Prokaryotes*, 6:646-703.
- Gulfraz, M., Mussaddeq, Y., Khanum, R. and Ahmad T. (2003). Metal contamination in wheat crops (*Triticum estivum L.*) irrigated with industrial effluents. *Journal of Biological Science*, 3(3): 335-339.
- Harman, G.E, Lorito, M., Lynch, J.M. (2004). Uses of *Trichoderma* spp. to alleviate or remediate soil and water
   pollution. *Advanced Applied Microbiology*, 56:313–330.
- 283 Harman, G.E., Howell, C.R., Viterbo, A., Chet, I., Lorito, M. (2004). *Trichoderma* species—opportunistic avirulent
- 284 plant symbionts. *Nature Reviews Microbiology*, 2 (1): 43–56.
- 285 Jankiewicz, B., Ptaszyński, B. and Wieczorek, M. (2000). Spectrophotometric Determination Of Cadmium (Ii) In
- 286 Samples Of Soil From Selected Allotment Gardens In Lodz, *Polish Journal of Environmental Studies*, 9:83.

- 287 Khair, M.H., (2009). Toxicity and accumulation of copper in Nannochloropsisoculata (Eustigmatophycea,
  288 Heterokonta). *World Applied Sciences Journal*, 6(3):378–384.
- 289 Khairiah, J., Zalifah, M.K., Yin, Y.H. and Aminah, A. (2004). The uptake of heavy metal by fruit vegetables grown
- in selected agricultural areas. *Pakistan Journal of Biological Science*, 7(8):1438 1442.
- Khan, S., Cao, Q., Zheng, Y.M., Huang, Y.Z. and Zhu Y. G. (2008). Health risks of heavy metals in contaminated
- soils and food crops irrigated with wastewater in Beijing, China. *Environmental Pollution*, 152(3):686–692.
- Khoudadoust, A.P, Reddy, K.R. and Maturi, K. (2004). Removal of nickel and phenanthrene from kaolin soil
  using different extraction. *Journal of Environmental Engineering Science*, 21(6): 691-704.
- 295 Kirpichtchikova, T.A., Manceau, A., Spadini, L., Panfili, F., Marcus, M.A. and Jacquet, T. (2006). Speciation and
- solubility of heavy metals in contaminated soil using X-ray microfluorescence, EXAFS spectroscopy, chemical
- extraction, and thermodynamic modeling, Geochimica et Cosmochimica Acta, vol. 70, no. 9, pp. 2163–2190.
- Ledin, M. (2000). Accumulation of metals by microorganisms—processes and importance for soil systems. *Earth Science Reviews*, vol. 51, no. 1–4, pp. 1–31.
- Lewis, T.A., Newcombe, D.A. and Crawford R.I. (2004). Bioremediation of oil contaminated with explosives.
   *Journal of Environmental Management*. 70: 291-307.
- 302 Marques, A.P.G.C, Rangel, A.O.S.S. and. Castro P.M.L. (2009). Remediation of heavy metal contaminated soils:
- 303 phytoremediation as a potentially promising clean-up technology. *Critical Reviews in Environmental Science and*
- **304** *Technology*, vol. 39, no. 8, pp. 622–654.
- 305 Mrayyan, B. and Battikhi, M.N. (2005). Biodegradation of total organic carbons (TOC) in Jordanian petroleum
   306 sludge. *Journal of Harzardous Materials* B 120: 127-134.
- 307 Nriagu, J.O. (1979). Global inventory of natural and anthropogenic emission of trace metals to the atmosphere.
   308 *Nature* 279:409–411.
- Nriagu, J.O. (1991). Human influence on the global cycling of the metals. In J.G. Farmer (ed.) heavy metals in the
   environment. *CEP consultants Limited.*, *Edinburgh, UK.* 1: 1-5.
- 311 Ociepa, E., (2011). The effect of fertilization on yielding and heavymetals uptake by maize and Virginia fanpetals
- 312 (*Sida hermaphrodita*). Archives of Environmental Protection, vol. 37, no. 2, pp. 123–129.
- 313 Odeyemi, A.T., Faweya, E.B., Agunbiade, O.R and Ayeni, S.K. (2011). Bacteriological, mineral and radioactive
- contents of leachate samples from dumpsite of Ekiti State
   *of Applied Science Research*, 3 (4): 92-108.
- 316 Opaoluwa O. D. and Umar, M.A. (2010). Bulletin of pure and applied sciences, 2010, 29:1, 39-55.
- Pala, M.B., DeCarvalho, D., Pinto, J.C. and Sant Anna Jr, G. (2006). A suitable model to describe bioremediation of
   a petroleum-contaminated soil. *Journal of International Biodeterioration and Biodegradation*, 58(6): 254-260
- 319 Pendias H. and Pendias K. (1989). Trace elements in soil and plants. Florida: CRC. Peplow, D. (1999).
- 320 Environmental Impacts of Mining in Eastern Washington, Center for Water and Watershed Studies Fact Sheet,
- University of Washington, Seattle.Persoon, C.H. (1974). Disposita methodica fungorum. *Römer's Neues Mag Bot*,
   1:81–128.
- Radwan, M. A. and Salama A. K., (2006). Market basket survey for some heavy metals in Egyptian fruits and vegetables. *Food and Chemical Toxicology*, 44:1273–1278.
- Rai, P. K. (2009). Heavy metal phytoremediation from aquatic ecosystems with special reference to macrophytes.
- 326 *Critical Reviews in Environmental Science and Technology*, 39(9): 697–753.
- 327 Sas-Nowosielska, A., Galimska-Stypa, R., Kucharski, R., Zielonka, U., Małkowski, E. and Gray, L. (2008).
- 328 Remediation aspect of microbial changes of plant rhizosphere in mercury contaminated soil. Environmental
- 329 Monitoring and Assessment, vol. 137, no. 1–3, pp. 101–109.
- 330 Sinha, R.K., Herat, S. and Tandon, P.K. (2004). Phytoremediation: role of plants in contaminated site management.
- Book of Environmental Bioremediation Technologies, pp. 315–330, Springer, Berlin, Germany.
- 332 Soumitra, N., Bibhas, D., Indu, S. and Piyush, P. (2014). Role of Cadmium and Lead Tolerant Pseudomonas
- aeruginosa in Seedling Germination of Rice (*Oryza sativa* L.). *Environmental and Analytical Toxicology*, 4:4.
- 334 Su, C., Jiang, L., and Zhang, W. (2014). A review on heavy metal contamination in the soil worldwide: Situation,
- impact and remediation techniques. *Environmental* Skeptics and Critics, 3(2): 24-38.

- 336 Tuzen, M. and Soylak, M., (2007). Evaluation of trace element contents in canned foods marketed from Turkey.
- **337** *Food Chemistry* 102:1089–1095.
- 338 Wang, J. and Chen, C. (2009). Biosorbents for heavy metals removal and their future. *Biotechnology Advances*, vol.
- **339** 27, no. 2, pp. 195–226.
- Wang, S., Li, J., Shi, S., et al. (2001): Geological disease caused by ecological environment: An example of cancer
- 341 village in Shanxi Province. *Environmental Protection* 5:42-46.
- 342 Wu, Y., Wang, X. and Liang, R.(1998): Dynamic migration of Cd, Pb, Cu, Zn and As in agricultural ecosystem.
- 343 Acta Scientiae Circumstantiae, 18:(4) 407-414 (In Chinese with English abstract).
- Yin, C., Peng, L., Wang, G, *et al.* (1999). The characteristics on contents of harmful elements in natural herbs in
  Kunming western suburb. *Pratacultural Science*, 16 (5) 24-26 (In Chinese with English abstract).
- Zaidi, M.I., Asrar, A., Mansoor, A. and Farooqui, M.A. (2005). The heavy metals concentration along roadside trees
   of Quetta and its effects on public health. *Journal of Applied Sciences*, 5(4):708–711.
- Zhang, N. (1999). Advance of the research on heavy metals in soil plant system. *Advance in environmental science*,
  7(4):30-33.
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- 352
- 353
- 354
- 355
- 356