1 2	Gastrointestinal Tract Effects of Aqueous Stem Bark Extract of <i>Citrus aurantifolia</i>
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4 5 6 7 8	Abstract
9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29	 Objective: The anti-diarrhea activity of the aqueous extract of the stem bark of <i>C. aurantifolia</i> was investigated in this study. Materials and Methods: Acute toxicity evaluation of the aqueous extract of <i>C. aurantifolia</i> stem bark was carried out using rats according to Lorke's method. Experimental diarrhea was induced in rats with castor oil, and the effect of the extract on castor oil-induced gastrointestinal motility and enteropooling was consequently investigated. Results: In the acute toxicity study, the extract produced no signs of toxicity or mortality in rats up to a dose of 5000 mg/kg. The oral LD₅₀ of the aqueous stem bark extract of <i>C. aurantifolia</i> was therefore taken to be >5000mg /kg. The extract significantly (p<0.05) decreased the frequency of defecation as well as in the number of unformed feces produced by castor oil-induced diarrhea in a dose- dependent manner. The extract also decreased the distance travelled by activated charcoal in the gastrointestinal tract of treated rats when compared to control rats. Results of castor oil-induced enteropooling revealed slight reduction in the weight of intestinal contents of treated rats compared to control rats. Conclusion: This study therefore, clearly shows that <i>citrus aurantifolia</i> stem bark possess significant anti-antidiarrheal potential and could be useful in the treatment of diarrhea. Key words: Gastrointestinal tract, Aqueous, Stem Bark, <i>Citrus aurantifolia</i>
30	1.0 INTRODUCTION
31	Diarrhea can be defined as an alteration in the normal bowel movement, characterized by a
32	situation in which an adult daily stools exceeds 300 g and contains $60 - 95$ % water ^[1] . The
33	WHO estimation revealed that diarrhea causes 45 million deaths annually throughout the world.
34	80% of these deaths are reported in developing countries including Nigeria. In Nigeria, diarrheal
35	infection remains the number one killer disease among children under 5 years, while 7-12 month

old babies remain the most susceptible ^[2]. Acute diarrhea being the most common is usually

37 caused by an infectious agent, even though drugs, poisons or acute inflammatory reactions are

38 contributing factors ^[3]. Rotavirus is the major causative agent of infectious diarrhea, particularly

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in young children now a days, however, other viral (*Enterovirus, norovirus* and adenovirus),
bacterial (*Salmonella* sp., *Shigella* sp., *Escherichia coli, Camphylobacter* and *Vibrio cholerae*)
and parasitic (*Cryptosporidium* and *Giardia*) agents are important pathogens ^[4].

Despite the effective and simple cheap treatment of oral dehydration therapy, majority of the 42 local populace still rely on herbs to treat diarrhea ^[5]. The use of herbal drugs in the treatment of 43 diarrhea is a common practice in many developing countries; here we attempt to investigate the 44 folklore claim of Citrus aurantifolia stem bark extract for antidiarrheal activity. Citrus 45 *aurantifolia* belongs to the family Rutaceae (orange family)^[6]. *Citrus aurantifolia* is a Perennial 46 Tree, with evergreen leaves, thorny stem, whitish flowers, globase fruits with many seeds, green 47 when unripe and greenish yellow when ripe, with sour taste ^[7]. C. aurantifolia is used in the 48 folklore medicine as an antiseptic, anthelmintic, mosquito bite repellent, for stomach ailments, 49 tonic, antiscorbutic, astringent, diuretic, headache, arthritis, digestive and appetite stimulant, and 50 for colds, coughs and sore throats ^{[8], [9]}. C. aurantifolia alleviates anxiety and nervousness, 51 relieves stress related disorders such as insomnia or nervous originated digestive disorders, and 52 53 also possesses anti-inflammatory potential/ an anticoagulant property, which renders it very valuable for people with cardiovascular risks. It is also used against fever, headaches and cold 54 [10] 55

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60 **2.0 MATERIALS AND METHODS**

- 61 **2.1 Materials**
- 62 **2.1.1** Chemicals and drugs

All chemicals used in this study were of analytical grade and were purchased from Sigma
Chemical Co. Ltd (USA) through a local vendor while the drugs were purchased from a local
pharmacy shop.

66 **2.1.2** Animals

Adult Wistar rats of either sex weighing 150–200g were used for this study. They were kept in stainless steel cages under standard laboratory conditions. They were maintained on clean water

69 and standard rodent feed.

70 **2.2 Methods**

71 2.2.1 Plant Collection and Identification

The stem bark of *Citrus aurantifolia* was collected from a natural habitat in Agbeji Area of Kogi
State, Nigeria. The plants were identified and authenticated at the herbarium unit of Biological
Sciences Department, Federal University, Lokoja.

75 2.2.2 Preparation of Extracts

The plant material was shade- dried for twenty- one (21) days and blended using a blender. One thousand and five hundred (1500) gram of the blended stem bark was soaked in distilled water for 72- hours. The resulting mixture was filtered using Whatmann filter paper (Size No1) and the extract was concentrated using a free- dryer. The extract was labelled as Aqueous stem extract of *Citrus aurantifolia* (SECA).

81 **2.2.3** Acute Toxicity Study

The oral median lethal dose (LD₅₀) of the extract was determined in rats according to the method of
Lorke ^[11].

84 2.2.4 Experimental Design

85 2.2.4.1 Castor oil-induced diarrhea

The method of Offiah and Chikwendu ^[12] was adopted. Twenty- five (25) rats of both sexes were fasted 86 overnight but allowed free access to water. They were randomized into five groups of five rats each. 87 Group I served as control and were administered 2 ml normal saline (0.9%). Groups II- IV were 88 administered 125, 250 and 500 mg/kg of SECA orally while Group V was administered diphenoxylate 89 hydrochloride (5 mg/kg) intraperitoneally. All rats were housed singly in a cage lined with white blotting 90 paper. 1 h after treatments, each of the rats was treated with 1 ml of castor oil orally. Rats were then 91 92 observed for 6-h and the number of water (wet) feces counted via fecal spots on the white bloated paper 93 lining the cage where individual rat was kept. Percentage protection was calculated as follows

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Mean number of defecation of control-Mean number of treated group

96 % Protection =
97 Mean number of defecation of control
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X100

99 2.2.4.2 Effect of castor oil-induced gastrointestinal motility

The method of Chitme et al., ^[13] was adopted. Rats were fasted overnight and then randomized into five 100 groups of five rats each and allowed free access to water. Group I served as control and was administered 101 2 ml normal saline (0.9%) orally while group V was administered 3 mg/kg of atropine intraperitoneally. 102 Groups II-IV was administered 125, 250 mg/kg, and 500 mg/kg of the SECA orally. After 10 min of 103 administering the extract and drug, 1 ml of 5% activated charcoal suspension in 10% aqueous solution of 104 105 Acacia powder was administered to treated rats. Rats were then sacrificed 30 min later and the abdomen was opened to measure the distance travelled by the activated charcoal. The results were expressed as 106 107 percentage of the total length of the intestine from the pylorus to the caecum.

108 2.2.4.3 Effect of castor oil-induced enteropooling

The method of Robert *et al.*, ^[14] was adopted. The intraluminal fluid accumulation due to the effect of 109 castor oil was determined. Rats were fasted overnight t but allowed access to fresh drinking water. The 110 rats were randomized into five groups of five rats each. Group I served as control and was administered 2 111 ml normal saline. Groups II- IV were administered 125, 250 and 500 mg/kg of SECA orally. Group V 112 was administered atropine (3 mg/kg) intraperitoneally. An hour later, 1 ml of castor oil was administered 113 to each of the treated rats. They were then sacrificed after 1h post castor oil administration. The small 114 intestines were removed, tied at both ends with thread and weighed. Intestinal contents were collected by 115 116 milking and the volume measured.

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120 2.2.5 Statistical Analysis

121 Statistical analysis was carried out using SPSS version 20.0. All the data were expressed as mean 122 \pm SEM and the statistical differences between the means were determined by one way analysis of 123 variance (ANOVA) which was followed by Fishers test and difference between means at P< 0.05 124 were considered significant.

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- 129 **3.0 RESULTS**
- 130 **3.1 Acute Toxicity**

The results of acute toxicity studies showed no mortality or signs of toxicity up to a dose of 5000 mg/kg of aqueous extract of *Citrus aurantifolia*. The oral LD_{50} of the extract was therefore taken

133 to be > 5000 mg/kg.

134 3.2 Effect of Aqueous Stem Bark Extract *Citrus aurantifolia* (SECA) on Castor Oil 135 induced Diarrhea in Albino Rats

Table-1 shows the frequency of defecation by the rats within 6-h of administration of SECA and 136 castor oil. There was a significant (p<0.05) difference in the frequency of defecation between the 137 control group and treated groups. The extract showed a dose- dependent effect as the group 138 139 treated with 500 mg/kg SECA had the lowest frequency of defecation and the highest percentage of inhibition (65.01%) followed by the 250 and 125 mg/kg SECA- treated groups with 59.06 and 140 53.25% of inhibition respectively. There was no significant (p>0.05) difference in percentage 141 inhibition between group V (standard drug- diphenoxylate hydrochloride -treated rats) and the 142 groups admitted SECA at all doses. 143

Table 1: Effect of Aqueous Stem Bark Extract *Citrus aurantifolia* (SECA) on Castor Oil induced Diarrhea in Albino Rats

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Group	Treatment (mg/kg)	Mean number of Defecation in 6 h	Percent Protection (%)
Ι	Control	7.23±1.14	-
II	SECA 125mg/ kg+ CO	3.38±1.01 ^a	53.25
III	SECA 250mg/ kg+ CO	2.96±0.96 ^a	59.06
IV	SECA 500mg/ kg+ CO	$2.53{\pm}0.55^{a}$	65.01
V	Diphynoxylate + CO	$2.22{\pm}0.47^{a}$	69.30

147 Data are presented as mean \pm SD. Data was analysed by one- way ANOVA followed by Fisher's 148 test, (n=5). ^a Statistically significant at p< 0.05. CO= Castor oil

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3.3 Effect of Aqueous Stem Bark Extract of *Citrus aurantifolia* (SECA) on Gastrointestinal Motility

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The effect of the extract on gastrointestinal transit of activated charcoal is shown in Table 2. There was a significant (p<0.05) decrease in the intestinal transit of activated charcoal in SECAtreated groups compared to the control group. The charcoal travelled very rapidly in the control group while the rate of movement was significantly (p<0.05) reduced in rats treated with SECA in a dose- dependent manner. The 500 mg/kg SECA –treated rats had a charcoal movement rate comparable to the 3 mg/kg atropine- group. The transit of charcoal in the groups treated with 125 and 250 mg/kg SECA were also statistically similar to the atropine- treated group.

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Table 2: Effect of Aqueous Stem Bark Extract of *Citrus aurantifolia* (SECA) on Charcoal
 Gastrointestinal Transit in Albino Rats

Group	Treatment (mg/kg)	Length of Intestine (cm)	Distance Travelled by Charcoal (cm)	Percent Intestinal Transit (%)
Ι	Control	35.2±1.14	30.7±3.21	87.22
II	SECA 125mg/ kg+ Ch	33.4±1.18	20.8±2.12	62.28 ^a
III	SECA 250mg/ kg+ Ch	34.6±2.33	18.3±2.33	52.89 ^a
IV	SECA 500mg/ kg+ Ch	34.1±1.43	16.1±1.98	47.21 ^a
V	Atropine 3mg/kg + Ch	36.4±2.17	16.6±1.56	45.60 ^a

165 Data are presented as mean \pm SD. Data was analysed by one- way ANOVA followed by Fisher's 166 test, (n=5). ^a Statistically significant at p< 0.05. Ch = charcoal

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3.4 Effect of Aqueous Stem Bark Extract of *Citrus aurantifolia* (SECA) on Castor oil Induced Enteropooling

The effect of the extract on castor oil-induced enteropooling is shown in Table 3. The result showed that there was a significant decrease (p<0.05) between the volume of intestinal contents in the control group and the treated groups. The volume of fluid in the group treated with 500 mg/kg of SECA had comparable result to that of atropine-treated group. The 125 mg/kg treated group had the highest percentage intestinal fluid inhibition of 49.01% followed by the group treated with 250 mg/kg of SECA while the group treated with 500 mg/kg had the least percentage intestinal fluid inhibition.

Table 3: Effect of Aqueous Stem Bark Extract of *Citrus aurantifolia* (SECA) on Castor oil induced Diarrhea in Albino Rats

Group	Treatment (mg/kg)	Wt. of Full Intestine (g)	Wt. of Empty Intestine (g)	Wt. of Intestinal Content (g)	Percentage Inhibition of Fluid (%)
Ι	Control	4.32±0.58	1.79 ± 0.45	2.53±0.11	-
II	SECA 125mg/ kg+ Ch	4.10±0.73	2.81±0.34	1.29±0.01 ^a	49.01
III	SECA 250mg/ kg+ Ch	4.12±0.69	2.61±0.39	1.51 ± 0.23^{a}	40.32
IV	SECA 500mg/ kg+ Ch	4.08±0.52	$1.94{\pm}0.45$	$2.14{\pm}0.23^{a}$	15.42
V	Atropine 3mg/kg + Ch	3.99±0.48	1.88±0.43	2.11±0.31 ^a	16.60

183 Data are presented as mean \pm SD. Data was analysed by one- way ANOVA followed by Fisher's 184 test, (n=5). ^a Statistically significant at p< 0.05. Ch = Charcoal

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187 **4.0 DISCUSSION**

In this study, acute toxicity of the extract revealed that oral administration of the extract produced no immediate and/or delayed signs of toxicity. The LD_{50} of the extract was therefore, above 5000 mg/kg according to Lorke's method ^[11]. This implies that the extract can be administered with some degree of safety, especially through oral route, where absorption might not be complete due to inherent factors limiting gastrointestinal tract absorption.

193 The aqueous stem bark extract of *Citrus aurantifolia* exhibited a dose-dependent protective 194 effect against diarrhea. Diarrhea induced by castor oil results from the action of ricinoleic acid

which causes the irritation and inflammation of the intestinal mucosa leading to prostaglandins 195 (PGE2 α) release. The released PGE2 stimulates gastrointestinal motility and secretion of water 196 and electrolytes ^[15], thus inducing an increase in the peristalsis and an intestinal hyper secretion 197 of fluid. The inhibition of prostaglandins biosynthesis prolongs the time of induction of diarrhea 198 by castor oil ^[16]. In addition to the increase in the latency time and a decrease in the frequency of 199 defecation, the administration of SECA to rats also caused a significant reduction of total fresh 200 201 weight of deposit, of water content and of the surface of impregnation of deposit. These results are similar to those obtained with diphenoxylate used as standard drug and suggest that SECA 202 might act as diphenoxylate. In fact, the antidiarrheal activity of diphenoxylate results from its 203 antispasmogenic and antisecretory properties on the intestine ^[17]. 204

In this study, it was also observed that C. aurantifolia significantly produced a significant 205 reduction in the progression of charcoal meal and in the intestinal transit time dose- dependently. 206 The 500 mg/ kg SECA produced a reduction comparable to that of atropine used here as 207 reference drug and which is known to reduce intestinal motility ^[18]. Since the extract has 208 demonstrated the ability to inhibit castor oil-induced diarrhea, its anti-diarrheic effect might in 209 part be due to decreased gastrointestinal secretion and/or inhibition of gastrointestinal motility. 210 The decreased intestinal motility and intestinal charcoal transit time might be due to increased re-211 absorption of water as earlier reported by Sahoo et al. [19]. 212

The extract also produced a reduction of castor oil-induced enteropooling in a dose- dependent 213 manner. This observation might be due to the ability of the extract to mediate a reduction in 214 weight gain of intestinal contents by preventing fluid and electrolyte secretion into the intestine 215 through the reduction of gastrointestinal motility. This is because reduction of the 216 gastrointestinal motility normally allows intestinal content ample time to be exposed to the 217 absorptive surface of the intestinal tract ^[20]. Diphenoxylate hydrochloride, an opioid, is known to 218 inhibit gastrointestinal secretions and motility, as exhibited by the study extract. Therefore, it 219 could be inferred from the study that the decrease in frequency of defecation and distance 220 travelled by the charcoal meal might be due to the inhibition of the gastrointestinal motility by 221 the extract. It can also be suggested that effects of the extract might be mediated through α -2 222 223 adrenergic receptor stimulation.

224 5.0 Conclusions

225 The aqueous extract of *Citrus aurantifolia* stem bark exhibited a dose-dependent effect against diarrhea; it significantly inhibited castor oil-induced intestinal fluid accumulation and the volume 226 227 of intestinal content and also significantly reduced the castor oil induced intestinal transit. This study therefore, clearly shows that citrus aurantifolia stem bark possess significant anti-228 antidiarrheal potential and could be useful in the treatment of diarrhea. 229 230 231 232 233 REFERENCES 234 [1] Guerrant RL, Van Gilder T, Steiner TS, Theilman MN, Slutsker L. 2001. Pratice guidelines 235 for the management of infectious diarrhea. 2001. J Infect Dis; 32: 331-351. 236 [2] Audu, R., Umilag, SA, Renner, JK, Awodiji A. 2000. Diarrhea Management. J. Nigeria 237 Infection Control Association: 3-15 238 [3] Thapar N, Sanderson IR. 2004. Diarrhea in children: An interface between developing and 239 developed countries. Lancet; 363: 641-53. 240 [4] Allen SJ, Okoko B, Martinez E, Gregorio G. Dans LF. 2004. Probiotics for treating infectious 241 diarrhea. Cochrane Database Syst Rev; 2: CD003048. 242 [5] Ahmadu, AA, Zezi, AK, and Yaro AH. 2007. Anti-diarrheal activity of the leaf extracts of 243 Hutch and Dalz (Fabaceae) and Mio (moraceae). 4(4): 524 528. 244 245 [6] Bakare, AA., Bassey, RB., Okoko, IE., Sanyaolu, AO., Ashamu, AE. and Ademola AO. 2012: Effect of Lime Juice (Citrus aurantifolia) on Histomorphological Alterations of the 246 Ovaries and Uterus of Cyclic Sprague-Dawley Rats. European Journal of Scientific 247 Research, 67(4): 607-616. 248 249 [7] Cheesbrough, M. 2006: Biochemical tests to identify bacteria. Laboratory practice in Tropical Countries. Cheesbrough M (ed.), Cambridge, Part II: Pp 63-90. 250 [8] Morton, J. 1987: Mexican Lime. In: Fruits of Warm Climates, 1st ed.; J.F. Morton: Miami, 251 FL, USA, Pp. 168-172. 252 [9] Aliyu, B. S. (2006): Some ethno-medicinal plants of the Savannah Regions of West Africa 253 Description and phytochemicals. Triumph publishing company. Pp 135-152. 254 [10] Chellaiah, M., Muniappan, A., Nagappan, R. and Savarimuthu, I. 2006: Medicinal plants 255 used by traditional healers in kancheepuram district of Tamil Nadu, India. J. Ethnobiol. 256 Ethnomedicine, 2 (43): 10. 257

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