Original Research Article

Antimicrobial susceptibility profiles of *Pseudomonas aeruginosa* isolates from patients attending health care facilities, Ebonyi Sate, Nigeria

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7 ABSTRACT

8 Aim: To assess age group related level of infection and antimicrobial susceptibility profiles of
9 *Pseudomonas aeruginosa* isolates from wound infection.

10 Study Design: This is a cross sectional study conducted among patients suspected of having wound

infection to determine age group related level of infection and antimicrobial susceptibility profile of the *P*.
 aeruginosa isolates.

Place and Duration of Study: The study was conducted between May, 2015 and June, 2016 at the
 Microbiology Laboratory of Akanu Ibiam Federal Polytechnic, Unwana, Ebonyi State, Nigeria.

15 Methodology: A total of 165 wound swabs were analysed for the presence of *P. aeruginosa*. Standard

16 microbiology laboratory tests were used to isolate and identify the isolates. Antibiotic susceptibility testing

17 of the isolates were carried out using the disc diffusion method.

18 Results: A total of 56 (33.94 %) P. aeruginosa isolates were identified. The age group 31 - 40 years 19 recorded the highest number 28 (50.00 %) of P. aeruginosa infection. Within this age group, females 15 20 (51.72 %) were slightly more infected than males 13 (48.15 %). In the tertiary hospital (MMH), the highest 21 sensitivity was seen to ofloxacin 32 (78.05 %) followed by ciprofloxacin, 29 (70.73 %) and ceftazidime, 26 22 (63.41 %). The number of the isolates resistant to amoxicillin/clavulanate, cefixime and cefuroxime were 34 23 (82.93 %), 26 (63.41%) and 23 (56.10 %) respectively. The number of sensitive P. aeruginosa from the 24 teaching hospital (FETHA) to ofloxacin and ciprofloxacin were 9 (60.00 %) and 8 (53.33 %) respectively. In 25 FETHA, the isolates showed the highest resistance to amoxicillin/clavulanate 14 (93.33 %) and cefixime 12 (80. 00 %). Exactly, 32 (57.14 %) of *P. aeruginosa* isolates were found to be multidrug resistant (MDR). 26 27 Regular monitoring of antimicrobial susceptibility profile is essential to guide the physicians in drug 28 prescription against P. aeruginosa strains.

29 Keywords: Susceptibility, Infection, P. aeruginosa, Multidrug resistance.

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31 1. INTRODUCTION

Wound infection is universal and the bacterial type varies with geographical location, resident flora of the skin, clothing at the site of wound, time between wound and examination [1]. The development of wound infection depends on the integrity and protective function of the skin [1]. Many bacteria pathogens are involved in wound infection [2].

P. aeruginosa can be found in most environments including soil, water and various types of vegetation. The organism is a clinically important pathogen, responsible for a variety of systemic infections such as urinary tract infections, respiratory system infections, gastrointestinal infections, dermatitis, bacteremia, soft tissue infections, bone and joint infections [3]. *P. aeruginosa* causes infection in immunocompromised patients such as those suffering from burn wounds or receiving cancer chemotherapy [4]. It is a major nosocomial pathogen, particularly dangerous to cystic fibrosis patients [5].

42 This organism is resistant to many antibacterial drugs [6]. Multidrug-resistant P. aeruginosa 43 phenotype is defined as resistant to one anti-microbial agent in three or more anti-pseudomonal anti-44 microbial classes (carbapenems, fluoroquinolones, penicillins/cephalosporins and aminoglycosides) [7, 8]. 45 MDR P. aeruginosa is an important public health issue because the organism is inherently resistant to 46 many drug classes and is able to acquire resistance to all effective antimicrobial drugs [9]. Multidrug-47 resistant P. aeruginosa develops resistance by various mechanisms like multidrug resistance efflux pumps, 48 biofilm formation, production of β-lactamases and aminoglycoside modifying enzymes [8]. Broad-spectrum 49 anti-pseudomonal drugs such as imipenem, ceftazidime, amikacin have been recommended for treatments 50 of infections caused by multiple drug resistant P. aeruginosa [10]. However, resistance to one or more of 51 these anti-pseudomonal drugs during therapy has been widely observed.

The alarming rate of resistance in bacteria pathogens raises concern for the effectiveness of antibiotic therapy [11]. The spread of multidrug resistant pathogens is a real threat to public health and a major concern for infection control practitioners globally [12]. This spread has paved way for the reemergence of previously controlled diseases and a high frequency of opportunistic and chronic infection cases in developing countries like Nigeria [12].

57 This study aims at assessing age group related level of infection and antimicrobial susceptibility 58 profiles of *P. aeruginosa* isolates from wound infection.

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60 2. MATERIALS AND METHODS

61 2.1 Study Area

62 One tertiary hospital (MMH) and one teaching hospital (FETHA) in Ebonyi State, Nigeria was used 63 for this study. These hospitals were selected because they receive large number of patients seeking 64 medical attention and also serve as referral centre for the state and neighbouring states.

65 2.2 Sample Collection

A total of 165 wound swabs were collected from patients aged 11 years and above. No specimen was got from the age group 0 -10 years. All the wound swabs collected at the hospitals were inoculated into 10 ml cooked meat broth and incubated at 37 °C for 48 hours [13].

69 2.3 Isolation of Bacteria

A loopful of the organisms growing in the cooked meat broth above was inoculated onto Cetrimide agar plates using the streaking technique. The inoculated Petri dishes were incubated at 37 °C for 24 hours [14]. Suspected discrete colonies of *P. aeruginosa* that appeared green on cetrimide agar were inoculated into nutrient broth and nutrient agar slants and incubated at 37 °C for 24 hours. The organisms were confirmed by adopting standard microbiological procedure which includes: colony morphology, Gram stain reaction and biochemical reaction such as catalase test, indole test, motility test, citrate utilization test, oxidase test, triple sugar iron agar test and urease test.

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79 2.4 Antibiotic Susceptibility Testing

In vitro antibiotic susceptibility testing of the isolates were carried out using the disc diffusion method as recommended by the Clinical and Laboratory Standard Institute [15]. The antibiotic disc used include: Ceftazidime 30 µg, Cefuroxime 30 µg, Gentamycin 10 µg, Cefixime 5 µg, Ofloxacin 5 µg, Amoxycillin/clavulanate 30 µg, and Ciprofloxacin 5 µg (Abtek Biologicals Ltd, Liverpool, U.K). A sterile Pasteur pipette was used to drop 0.2 ml of the standardized inoculum equivalent of 0.5 McFarland turbidity standards (1.0 x10⁸ cfu/ml) on the surface of dry Mueller-Hinton agar. The inoculum was evenly spread 86 using Hockey stick shaped glass rod. The agar was left for about 10 minutes for the inoculum to dry and 87 thereafter, antibiotic discs were aseptically placed on the surface of the inoculated Mueller-Hinton agar plate using heat sterilized forceps. They were incubated at 37 °C for 18-24 hours. The diameter of zones of 88 89 inhibition for each antibiotic was measured in millilitre and compared with values provided by the Clinical 90 and Laboratory Standard Institute [15]. 91 92 2.5 Statistical Analysis 93 Statistical analysis was carried using the statistical package for social sciences (SPSS) version 15. P < 94 0.05 was considered significant. 95

96 2. RESULTS AND DISCUSSION

Out of 165 wound swabs analysed, 56 (33.94 %) *P. aeruginosa* isolates were obtained (Table 1).
The most frequent isolates of *P. aeruginosa* were from trauma 39 (44.32 %) followed by burn wound 3
(25.00 %).

100 **Table 1**: Distribution of *P. aeruginosa* from wound swabs

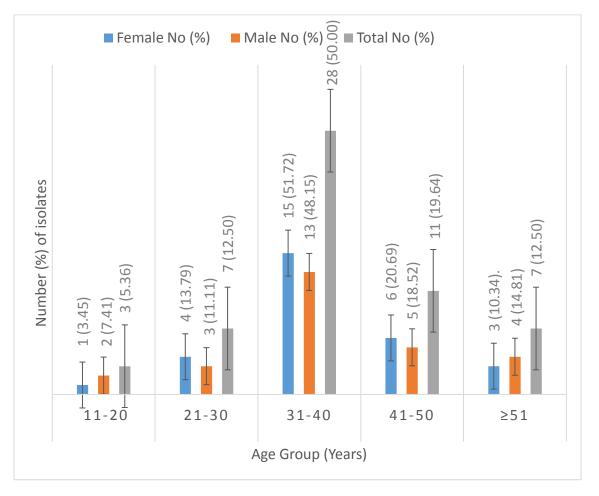
| Type of wound | Number | Number of bacterial isolates obtained | Percentage bacterial isolates |
|---------------|--------|---------------------------------------|-------------------------------|
| Burn wound | 12 | 3 | 25.00 |
| Abscess | 65 | 14 | 21.54 |
| Trauma | 88 | 39 | 44.32 |
| Total | 165 | 56 | 33.94 |

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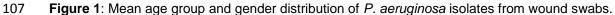
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103 The age group 31 – 40 years recorded the highest number 28 (50.00 %) of *P. aeruginosa* infection.

104 (Fig. 1). Within this age group, females 15 (51.72 %) were slightly more infected than males 13 (48.15 %).



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108 **Key**: % = Percentage in bracket

Antibiotic susceptibility profile of *P. aeruginosa* from MMH is as shown in Table 2. The highest sensitivity was seen to ofloxacin 32 (78.05 %) followed by ciprofloxacin, 29 (70.73 %) and ceftazidime, 26 (63.41 %). The number of the isolates resistant to amoxicillin/clavulanate, cefixime and cefuroxime were 34 (82.93 %), 26 (63.41%) and 23 (56.10 %) respectively.

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| Antibiotic | Disc Content | Number of sensitive Isolates (%) | Number of Intermediate Isolates (%) | Number of Resistant Isolates (%) |
|-----------------------------|--------------|--|---|--|
| Ceftazidime | 30 µg | 26 (63.41) | 4 (9.76) | 11 (26.83) |
| Cefuroxime | 30 µg | 13 (31.71) | 5 (12.20) | 23(56.10) |
| Gentamycin | 10 µg | 15 (36.59) | 5 (12.20) | 21 (51.22) |
| Cefixime | 5 µg | 6 (14.63) | 9 (21.95) | 26 (63.41) |
| Ofloxacin | 5 µg | 32 (78.05) | 0 (0.00) | 9 (21.95) |
| Amoxicillin/ Clavulanate | 30 µg | 3 (7.32) | 4 (9.76) | 34 (82.93) |
| Ciprofloxacin | 5 µg | 29 (70.73) | 3 (7.32) | 9 (21.95) |

Table 2: Antibiotic Susceptibility Profile of *P. aeruginosa* isolates from Wound swabs, MMH.

Key: % = Percentage in bracket; MMH = Code of tertiary hospital used

120 The number of sensitive *P. aeruginosa* from FETHA to ofloxacin and ciprofloxacin were 9 (60.00 %) 121 and 8 (53.33 %) respectively (Table 3). The isolates showed the highest resistance to 122 amoxicillin/clavulanate 14 (93.33 %) and cefixime 12 (80. 00 %).

| Antibiotic | Disc Content | Number of sensitive Isolates (%) | Number of Intermediate Isolates (%) | Number of Resistant Isolates (%) |
|-----------------------------|--------------|--|---|--|
| Ceftazidime | 30 µg | 6 (40.00) | 1 (6.67) | 8 (53.33) |
| Cefuroxime | 30 µg | 5 (33.33) | 1 (6.67) | 9(60.00) |
| Gentamycin | 10 µg | 4 (26.67) | 2 (13.33) | 9(60.00) |
| Cefixime | 5 µg | 2 (13.33) | 1 (6.67) | 12 (80.00) |
| Ofloxacin | 5 µg | 9 (60.00) | 1 (6.67) | 5 (33.33) |
| Amoxicillin/ Clavulanate | 30 µg | 1 (6.67) | 0 (0.00) | 14 (93.33) |
| Ciprofloxacin | 5 µg | 8 (53.33) | 0 (0.00) | 7 (46.67) |

| 134 Table 3 : Antibiotic Susceptibility Profile of <i>P. aeruginosa</i> isolates from Wound swabs, FETH | 134 | Table 3: Antibiotic Susce | ptibility Profile of P. ae | eruginosa isolates from Wour | nd swabs, FETHA |
|--|-----|---------------------------|----------------------------|------------------------------|-----------------|
|--|-----|---------------------------|----------------------------|------------------------------|-----------------|

135 **Key**: % = Percentage in bracket; FETHA = Code of tertiary hospital used

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137 Exactly, 57.14 % *P. aeruginosa* isolates from the two hospitals were found to be multidrug resistant.

138 (Table 4). The prevalence was slightly higher in MMH (58. 54 %) than FETHA (53.33 %).

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140 **Table 4**: Prevalence of MDR Isolates of *P. aeruginosa* from Wound swabs.

| Hospital | Total No. of <i>P.</i> aeruginosa Isolates | Number of MDR <i>P.</i> aeruginosa Isolates | Percentage of MDR <i>P.</i> aeruginosa Isolates |
|----------|---|--|--|
| MMH | 41 | 24 | 58.54 |
| FETHA | 15 | 8 | 53.33 |
| Total | 56 | 32 | 57.14 |

141 **Key**: MDR = Multidrug Resistant

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P. aeruginosa is known to be associated with wound infections. The result of this study is in contrast with the reports of Garba et al. [2] who isolated 11 % of *P. aeruginosa* from wounds of patients attending Ahmadu Bello University Teaching Hospital, Zaria, Nigeria. Also, Hyder et al. [16] and Mama et al. [17] reported a lower prevalence of 6 (3.95 %) and 11 (8 %) respectively. Similarly, Ezebialu et al. [18]
isolated 27.3 % *P. aeruginosa* from wounds in Enugu, which is slightly lower than the 33.94 % recorded in
this study. The findings of this study is similar to the reports of Nwachukwu et al. [19] and Anupurba et al.
[1] that isolated 32.90 % and 32 % *P. aeruginosa* respectively from wound swabs.

In a similar study carried out by Kirecci and Kareem [20], the age group mostly infected with *P*. *aeruginosa* was 41 – 60 years representing 45.33 %. Their finding differ from the present study where the age group 31 – 40 years (50.00 %) recorded the highest number of *P. aeruginosa* infection. Rajat et al. [21] reported isolation rate of 29 % in the age group of 31–45 years which is similar to our study and that done by Chander and Raza [22] that had 20% in age group of 21–40 years.

155 P. aeruginosa has been reported to have innate resistance to several antibiotics due to the 156 presence of lipopolysaccharide in the outer membrane [16]. In Nigeria, the trend in resistance phenotype of 157 P. aeruginosa to commonly prescribed antibiotics in various hospitals is increasing since the last decade 158 [10]. In line with the present study, Jombo et al. [23] and Garba et al. [2] reported a high resistance to 159 amoxicillin/clavulanate 100 % and 81.8 % respectively among P. aeruginosa isolates from urine and wound 160 swabs. Rajeevan et al. [24] found 40 % and 20 % resistance to ceftazidime and gentamycin respectively. 161 This is low compared the finding of this study in FETHA with ceftazidime and gentamycin resistance of 162 53.33 % and 60.00 % respectively. In MMH, the resistance to gentamycin 51.22 % was high compared to 163 20 % resistance reported by Rajeevan et al. [24]. Ahmed et al. [8] reported a high (91 %) level resistance to ceftazidime. In comparison to the present study from the two hospitals. Hyder et al. [16] recorded a higher 164 165 level of resistance to gentamycin 70.83 % and a lower level of resistance to ciprofloxacin 31.25 % in Hlla 166 Teaching hospital, Babylon. The resistance observed against Ceftazidime is worrisome, being among the 167 broad-spectrum anti-pseudomonal drugs recommended for treatments of infections caused by MDR P. 168 aeruginosa [10]. High resistance of the isolates to antibiotics may be due to the practice of self-medication, 169 lack of diagnostic laboratory services or unavailability of guideline regarding the selection of drugs, thereby 170 leading to inappropriate use of antibiotics [17].

Various researchers have reported lower rates of MDR *P. aeruginosa* in their study. Ahmed et al. [8] detected a slightly low prevalence rate 52 % compared to this study in patients with nosocomial infections at a University hospital, Egypt. In Iran, Zahra and Moniri [25] reported 30 % prevalence of MDR

P. aeruginosa. Siveraj et al. [26] reported 12 % MDR among *P. aeruginosa* isolates from India. In Abeokuta, Nigeria, Okonko *et al.* [27] reported multidrug resistant to 5 antibiotics (ampicillin, chloramphenicol, cotrimoxazole, nitrofuratoin and tetracycline) by *P. aeruginosa*. The level of MDR recorded in this study shows that *P. aeruginosa* can develop resistance to many antibacterial. *P. aeruginosa* can develop resistance to many antibacterials both through the resistance genes on extrachromosomal genetic elements and through mutational processes [28].

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181 3. CONCLUSION

In this study, 56 (33.94 %) isolates of *P. aeruginosa* were obtained from the wound swabs analyzed. The age group mostly infected with *P. aeruginosa* was 31 – 40 years. The *P. aeruginosa* strains were more sensitive to ofloxacin while they showed the highest resistance to amoxicillin/clavulanate. Exactly, 57.14 % *P. aeruginosa* isolates were found to be multidrug resistant. A regular monitoring of antimicrobial susceptibility profile is essential to guide the physicians in prescribing right drugs and prevent the emergence of multidrug resistance strains of *P. aeruginosa*.

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189 COMPETING INTEREST

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Authors have declared that competing interest do not exist.

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193 CONSENT

All authors declare that written informed consent was obtained from the patients for publication of this case report. A copy of the written consent is available for review by the Editorial office/Chief Editor/Editorial Board members of this journal.

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