

Hepatoprotective and antioxidant activity of *Enicostemma littorale* against CCl₄ induced liver damage in rats

Abstract:

Objective

To investigate the hepatoprotective and antioxidant effects of ethanol extract of *Enicostemma littorale* Blume (Ens) against CCl₄ induced hepatic injury in albino rats.

Methods

Male albino rats of six numbers in each group were undertaken for study. Hepatoprotective and antioxidant effect of *E. littorale* Blume (Ens) ethanol extract at a dosage of 100 & 200 mg/kg body weight was evaluated.

Results

The degree of hepatoprotection was assessed by measuring the activity levels of the marker enzymes such as serum aspartate transaminase (AST), alanine transaminase (ALT) alkaline phosphatase (ALP) , acid phosphatase (ACP) and total bilirubin. Free radicals generated lipid per oxidation was assessed by measuring the activity levels of the tissue antioxidant enzymes such as glutathione peroxidase,(GPX) catalase (CAT), superoxide dismutase (SOD). The CCl₄ administered rats recorded elevated activity levels of serum AST,ALT,ALP and ACP revealing CCl₄ induced hepatotoxicity. In the groups treated with 100mg/kg and 200mg/kg of the extract, the above bio-chemical markers of hepatotoxicity were found to be decreased when compared to CCl₄ treated control group. Both the doses of EEEL used in the study showed significant protective property than control. (*p<0.01, **p<0.001 vs. control)..In the groups treated with 100mg/kg and 200mg/kg of the extract, GPX, SOD and catalase were found to be increased when compared to CCl₄ treated control group.(p<0.01 vs control).

Conclusions

It can be concluded that the ethanol extract of *E. littorale* Blume is not only hepatoprotective but also possess significant antioxidant property.

Keywords: Hepatoprotective, Antioxidant enzymes, *Enicostemma littorale* Blume,

34 INTRODUCTION:

35 Hepatic system is very vital organ system involved in the body's metabolic activities. As
 36 a result the chemical reactions in the liver may generate several reactive species like free
 37 radicals. These reactive species form covalent bond with the lipids of the tissue. However
 38 inbuilt protective mechanisms combat the hazardous reactions associated with the free
 39 radicals. Due to excessive exposure to hazardous chemicals, the free radicals generated will
 40 be so high such that they overpower the natural defensive system leading to hepatic damage
 41 and cause jaundice, cirrhosis and fatty liver, which remain one of the serious health problems.
 42 Carbon tetrachloride (CCl₄) is one such hazardous chemical which induces hepatopathy
 43 through membrane lipid per oxidation by its free radical derivative, (CCl₃·, CCl₃O₂·).
 44 Excessive production of the reactive species manifests in tissue-thiol depletion, lipid per
 45 oxidation, plasma membrane damage etc., culminating into severe hepatic injury¹. Much of
 46 the cell damage that occurs during liver degeneration is believed to be caused by free
 47 radicals, highly reactive oxygen species liberated during alcohol metabolism. These radicals
 48 react with cell membrane and induce lipid peroxidation, which has been implicated as
 49 important pathological mediation² in many clinical disorders such as heart disease, diabetes,
 50 cancer and liver disease. The management of liver diseases is still a challenge to the modern
 51 medicine. In the background of the above, it is realized that antioxidant activity or inhibition
 52 of generation of free radicals plays a crucial role in providing protection against such hepatic
 53 damage. Several herbs and herbal products are known to possess antioxidant principles and
 54 may be useful as organ protective agents. Plant drugs are frequently considered to be less
 55 toxic and free from side effects.³ Numerous medicinal plants and their formulations are used
 56 for liver disorders in ethnomedical practices as well as in traditional systems of medicine in
 57 India. There are a number of evidences indicating that natural substances from edible and
 58 medicinal plants exhibit strong antioxidative activity, and could work against hepatic toxicity
 59 caused by various toxicants⁴⁻⁵ *Enicostemma littorale*(*E. littorale*) Blume (Family:
 60 Gentianaceae) is a perennial, tropical traditional medicinal herb with sessile lanceolate
 61 leaves, flowers arranged in clusters, fruit in a capsule. The plant is locally used for its
 62 medicinal properties in Tamilnadu, Kerala India, such as antiinflammatory, antiulcer
 63 activity⁶, hypoglycaemic⁷, and antimalarial activities⁸. The present study aims to investigate
 64 the hepatoprotective and antioxidant effects of *E. littorale* on CCl₄ induced hepatotoxicity in
 65 male albino rats.

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67 **Materials and methods:**

68 **Plant material:**

69 The whole plant of *E. littorale* were collected during January (2018) in and around Palakkad
 70 District, Kerala, India. The plant were cleaned, shade dried, authenticated by Dr Jayaraman,
 71 Plant anatomy research centre, Chennai. A voucher specimen was deposited in the
 72 Department of Pharmacognosy, Sanjo college of Pharmaceutical studies, Palakkad, Kerala.
 73 (SCOPS/P.COG/07/2018)

74 **Preparation of plant extract:**

75 For preliminary phytochemical analysis, extract was prepared by weighing 500 grams of the
 76 dried powdered leaves were subjected to hot successive continuous extraction using Soxhlet
 77 apparatus with different solvents as per the polarity, petroleum ether, benzene, chloroform,
 78 ethanol and finally with aqueous. The extracts were filtered in each step using Whatman
 79 filters paper. The filtrate was concentrated using a rotary evaporator at low temperature (40-
 80 45°C) and pressure. The presence or absence of the primary and secondary phytoconstituents
 81 was detected by usual prescribed method⁹

82 **Chemicals and Drugs:**

83 Carbon tetra chloride and Ecoline kits for serum aspartate transaminase (AST), alanine
 84 transaminase (ALT) alkaline phosphatase (ALP) and acid phosphatase (ACP), total bilirubin,
 85 were purchased from Sigma Co. (Sigma St. Louis, MO). Standard Silymarin was obtained
 86 from Ranbaxy Ltd, New Delhi. Absolute ethanol was of analytical grade and was purchased
 87 from Merck (German). The other reagents were of analytical grade.

88 **Animals**

89 Western albino rats 180-230 gm maintained in the Animal house facility of the Department of
 90 Pharmacology, Sanjo College of pharmaceutical studies, were used in these experiments. The
 91 animals were maintained on standard small animal feeds (Excel feed, Ilorin) and water *ad*
 92 *libitum*. This research was carried out in accordance with the rules governing the use of
 93 laboratory animals as accepted internationally. The experiment was conducted between the
 94 hours of 900 h and 1600 h. The experimental groups consisted of six animals. They were
 95 maintained at constant room temperature (22° ± 1 °C) and submitted to 12 h light/dark cycle
 96 with free access to food and water.

97 **Experimental procedure**

98 **Acute oral toxicity study**

Acute oral toxicity was conducted as per OECD guidelines (Organization of Economic Cooperation and Development) 423 (Acute toxic class method). The acute toxic class method is a step wise procedure of three animal of a single sex per step. Depending on the mortality and / or moribund status of animals, on the average 2-4 steps may be necessary to allow judgment on the acute toxicity of the test substance. This procedure results in the use of a minimal number of animals while allowing for acceptable data based scientific conclusion. The method uses defined doses, (5, 50, 300, 2000 mg/kg body weight) and the results allow a substance to be ranked and classified according to the globally harmonized system (GHS) for the classification of chemicals which causes acute toxicity. The method previously described by Lorke ¹⁰ was adopted

Hepatoprotective activity:

The method of Ko et al ¹¹ was used for screening the hepatoprotectivity of the test extract. Adult Wistar rats of either sex were randomly assigned into 5 groups of 6 animals. The animals of Group I served as normal control and received only the vehicle normal saline (10 ml/kg i.p). Group II served as toxic control and administered CCl₄ (1ml/kg) by subcutaneous injection. The animals of Group III and IV received *Enicostemma littorale* extract (100 mg/kg BW and 200mg/kg BW p.o. respectively) for 15 days. Group V served as Standard and was treated with Silymarin (25 mg/kg BW i.p., for 15 days). Animals (except Group 1) were treated CCl₄ at a dose of 1 ml/kg BW by subcutaneous injection. Blood samples were collected after last dose CCl₄ administration by direct cardiac puncture under light ether anesthesia and animals were sacrificed by cervical decapitation and hepatic tissue was collected. Heparinized blood sample were taken and assessed for serum enzyme markers and hepatic tissue was taken and subjected to histopathological study and further tissue was analyzed for Glutathione and lipid per oxidation.

Group 1-Normal animals (10mg/kg i.p)

Group 2-CCl₄ (1ml/kg s.c) treated animals

Group 3-CCl₄ + EEEL (100mg/kg ,p.o.) treated animals.

Group 4-CCl₄ + EEEL (200mg/kg, p.o.) treated animals.

Group 5-CCl₄ + Silymarin (25mg/kg i.p.) treated animals

Estimation of biochemical parameters:

Separated serum was analyzed. serum aspartate transaminase (AST), alanine transaminase (ALT) alkaline phosphatase (ALP) and acid phosphatase (ACP), and total bilirubin are

estimated¹² the tissue levels of enzymatic antioxidants viz. superoxidase dismutase (SOD), catalase (CAT), glutathione peroxidase (GPX), were estimated.¹³⁻¹⁶

Statistical analysis

The results are analysed by one way ANOVA followed by Dunnet's test and p value <0.001 was considered significant

Results:

Acute toxicity:

The results showed no clinical signs and mortality of the animal therefore an LD₅₀ > 2000 mg/kg body weight may be assume.

Hepatoprotective and antioxidant activity:

The estimated values of serum AST, ALT, ALP, ACP and Total bilirubin values in control (saline) group of rats are tabulated in Table 1. A remarkable elevation was observed in Serum AST, ALT, ALP, ACP, total bilirubin and values in CCl₄ intoxicated rats (Toxic Control group). In the groups treated with 100mg/kg and 200mg/kg of the extract, the above biochemical markers of hepatotoxicity were found to be decreased when compared to CCl₄ treated control group. Evidently, the hepatoprotective effects of higher dose of Ethanolic extracts of *Enicostemma littorale* (200mg/kg) were near to that of standard i.e. Silymarin (25mg/kg). Both the doses of EEEL used in the study showed significant protective property than control. (*p<0.01, **p<0.001 vs. control) However the test extract was found to be less potent than that of standard,

TABLE: 1 SERUM ENZYME PROFILE

Treatment	AST(U/I)	ALT(U/I)	ALP (IU/L)	ACP(U/L)	Total Bilurubin(mg/100ml of blood)
Control (saline)	97.3±1.18	35.08±0.2	15.92±0.72	10.5±0.064	0.39±0.04
CCl ₄ (1ml/kg)	186.7±1.82	136.9±1.94	98.3±7.9	38.6±2.9	0.89±0.76
EEEL(100mg/kg)	138.64±5.92	72.4±6.8	63.9±5.8	25.4±0.95	0.74±0.06
EEEL(200mg/kg)	106.8±8.28	47.5±4.1	43.6±3.4	16.5±0.18	0.57±0.04
Silymarin25mg/kg	105.3±4.3	49.4±3.6	34.8±2.9	16.2±1.2	0.24±0.03

Data are expressed as mean ±S.E (n=6) *p<0.01 vs.control, **p<0.001 vs. control

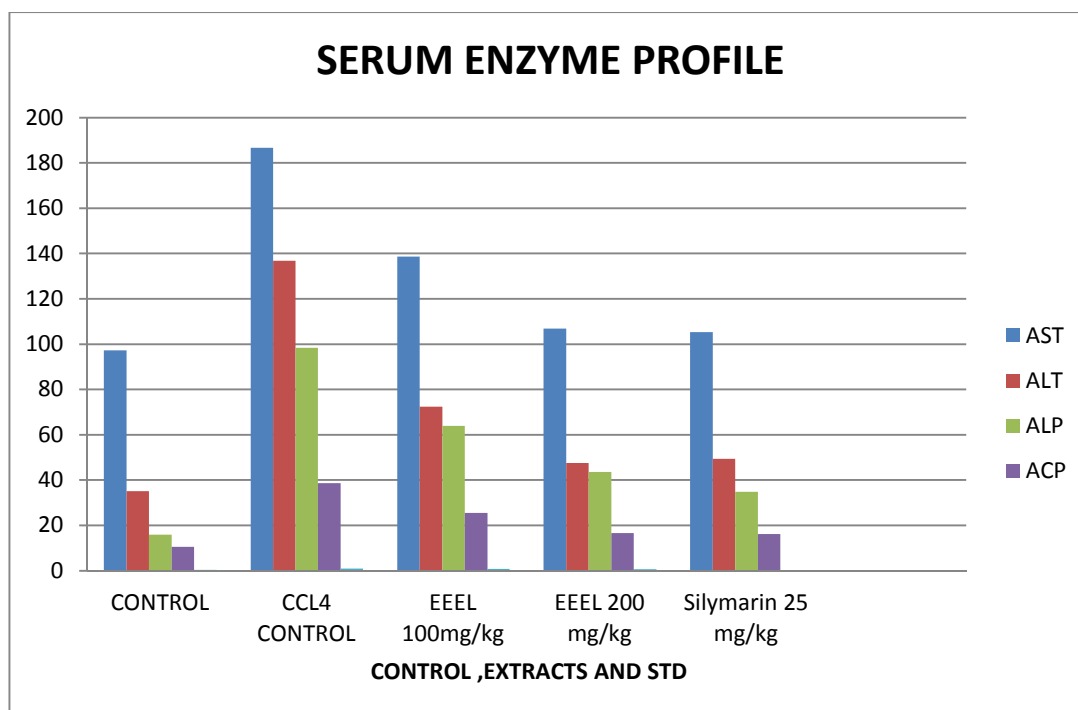
EEEL- Ethanolic extract of *Enicostemma littorale*

AST-Aspartate transminase ACP-Acid phosphatase

ALT- Alanine transaminase

ALP- Alkaline phosphatase

FIG : 1 SERUM ENZYME PROFILE



The tissue glutathione was found to be depleted upon CCl_4 intoxication, indicate that the tissue damage is due to over powering the inbuilt free radical scavenger mechanisms. This tissue GSH depletion was inhibited by the pretreatment with test extract in a dose dependant manner. Similarly lipid peroxidation induced by CCl_4 treatment was reversed by test extract in a dose dependant manner. The results are compiled in table 2. In the groups treated with 100mg/kg and 200mg/kg of the extract, GPX, SOD and catalase were found to be increased when compared to CCl_4 treated control group. Evidently, the hepatoprotective effects of higher dose of Ethanolic extracts of *Enicostemma littorale* (200mg/kg) were near to that of standard i.e. Silymarin (25mg/kg). Both the doses of EEEL used in the study showed significant protective property than control. (* $p < 0.01$, vs. control) However the test extract was found to be less potent than that of standard drug.

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TABLE-2: ANTIOXIDANT ACTIVITY

Treatment	Glutathione peroxidase (mg liver protein) ⁻¹	SOD (mg liver protein) ⁻¹	Catalase (mg liver protein) ⁻¹
Control (saline)	0.992±0.05	75.81±1.94	296.83±10.05
CCl ₄ (1ml/kg)	0.61±0.03	47.84±0.50	179.73±5.78
EEEL(100mg/kg)	0.85*±0.07	67.73*±0.54	266.27*±8.74
EEEL(200mg/kg)	0.92*±0.06	86.97*±0.75	281.38*±9.92
Silymarin25mg/kg	0.95*±0.03	88.34*±2.54	268.27*±6.46

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Data are expressed as Mean ±S.E (n=6) *p<0.01 Vs control

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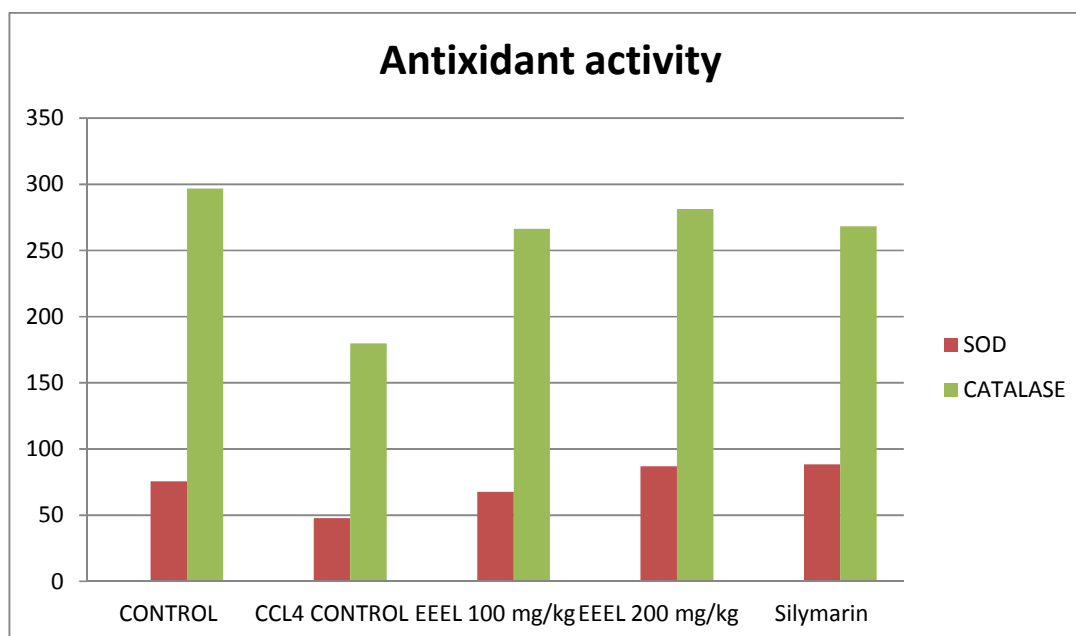
EEEL- Ethanolic extract of *Enicostemma littorale*

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SOD- Superoxidase dismutase

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FIG 2: ANTIOXIDANT ACTIVITY



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189 Discussion:

190 Since the extract has demonstrated dose dependant anti-oxidant activity in all the models of
 191 the study, the ethanol extract was taken for assessing the *in vivo* hepatoprotective properties.

192 Pretreatment with the test extract has reduced the elevated levels of biochemical markers of
 193 hepatotoxicity. Further it was also observed that the tissue GSH depletion due to CCl₄

challenge was reversed by the test extract and also reduced the extent of lipid peroxidation. Most of the mammals have an effective mechanism to prevent and neutralize the free radical induced damage, which is accomplished by a set of endogenous substances such as superoxide dismutase, catalase, and glutathione peroxidase and glutathione reductase. In biochemical system, superoxide radical and H_2O_2 react together to form the hydroxyl radical, this can attack and destroy almost all known biochemicals¹⁷. The hydroxyl radicals thus produced may attack the sugar of DNA base causing sugar fragmentation, base loss and DNA stand leakage. Ethanolic extract of *E.littorale* reduced the super oxide anions and also scavenge off the hydroxyl radicals and hence, inhibit the cellular damage. It is apparent from the present study that the test extract does not interfere with the generation of the free radicals but it scavenges off the free radicals. CCl_4 undergo hepatic metabolism to give rise to trichloromethyl radicals, which upon reacting with reactive oxygen species yields trichloromethyl peroxide radicals, which forms covalent bond with membrane lipids and destroy the membrane integrity. The observation of increased MDA formation in hepatic cells after CCl_4 challenge is in accordance with the earlier report which suggests involvement of trichloromethyl and tri-chloromethylperoxy radicals in the propagation of per-oxidation process¹⁸. The pretreatment with extract has prevented oxygen free radicals and thereby prevented the formation of peroxy radicals. This aspect of test extract also contributes to the hepatoprotectivity. The unpublished data on the hepatoprotective activity of this plant on other models like paracetamol and thiocetamide induced hepatotoxicity indicated that the hepatoprotectivity of the test extract is not model specific. SOD is metalloproteins catalyzing the dismutation of superoxide anion to hydrogen and oxygen. Numerous studies have shown the importance of SOD in protecting cells against oxidative stress¹⁹. The SOD activity could be decreased in tissue during alcohol ingestion. This decrease could be due to the feedback inhibition or oxidative inactivation of enzyme protein due to excess ROS generation²⁰. CAT, hemeprotein, catalyzes the reduction of hydrogen peroxides²¹ acts as preventive antioxidant and plays an important role in protection against the deleterious effects of lipid peroxidation²². The activity levels of catalase in tissue decreased in ethanol fed animals might be due to the inhibition of CAT activity, which is suggestive of enhanced synthesis of O_2^- during the ingestion of alcohol since O_2^- is a powerful inhibitor of catalase²³. GPX is an enzyme with selenium in the form of selenocysteine and can catalyze the reduction of hydrogen peroxide and hydroperoxides to non toxic products. GPX has a well-established role in protecting cells against oxidative injury. GPX is non-specific for H_2O_2 and lipid peroxide generated during alcohol ingestion which are efficiently scavenged by GPX activity. The depression of this

enzyme activity reflects perturbations in normal oxidative mechanism during alcohol ingestion. The cellular antioxidant defense enzymes *viz.* SOD, CAT, and GPX were significantly reduced in the CCl₄ administered rat. This might lead to decreased antioxidant defense and increased oxidative stress and thereby the tissue injury occur. Similar studies also indicate the failure of cellular antioxidant defense system during hepatotoxicity were recorded²⁴⁻²⁵

Conclusion:

The results of the present investigation, it may be concluded that the ethanolic extract of the whole parts of *Enicostemma littorale* possess significant hepatoprotective activity against carbon tetrachloride induced hepatotoxicity and antioxidant activity. The antioxidant potential may be attributed to the presence of polyphenolic compounds. Further studies like isolation and characterization of the active principle(s) responsible for such activity are needed to confirm.

References:

- 1) Vir Ji Chrungoo, Kuldeep Singh, Jaswant Singh. Differential biochemical response of freshly isolated rat hepatocytes to paracetamol, carbontetrochloride and D-galactosamine toxicity. I J Exp Bio, 1997; 35:603-610.
- 2) Muriel P. Role of free radicals in liver diseases. Hepatol Int. 2009;3:526–536
- 3) Pari L, Saravanan R. Antidiabetic effect of diasulin, an herbal drug, on blood glucose, plasma insulin and hepatic enzymes of glucose metabolism in hyperglycaemic rats. Diabetes Obes Metab. 2004;6:286–292.
- 4) Upur H, Amat N, Blazekovic B, Talip A. Protective effect of *Cichorium glandulosum* root extract on carbon tetrachloride-induced and galactosamineinduced hepatotoxicity in mice. Food Chem Toxicol. 2009;47:2022–2030
- 5) Zeashan H, Amresh G, Singh S, Rao CV. Hepatoprotective activity of *Amaranthus spinosus* in experimental animals. Food Chem Toxicol. 2008;46:3417–3421
- 6) Roy S, Niranjana C, Jyothi T, Shankrayya M, Vishawanath K, Prabhu K, et al. et al. Antiulcer and anti-inflammatory activity of aerial parts *Enicostemma littorale* Blume. J Young Pharm. 2010;2(4):369–373
- 7) Vishwakarma SL, Sonawane RD, Rajani M, Goyal RK. Evaluation of effect of aqueous extract of *Enicostemma littorale* Blume in streptozotocin induced type 1 diabetic rats. Indian J Exp Biol. 2010; 48:26–30.

- 262 8) Yghemonos S. The earth institute, University of Colombia; 2007. Malaria and
263 poverty: European alliance against malaria; pp. 1–4.
- 264 9) Kokate CK., Practical Pharmacognosy, M/S Vallabh Prakashan, Pune, pp 111-115,
265 1985.
- 266 10) Lorke D. A new approach to acute toxicity testing. Archives of toxicology 1983; 54:
267 275-287
- 268 11) Ko KM, Yick PK, Chiu TW, Hui TY, Cheng CHK, Kong YC. Impaired antioxidant
269 status in CCl₄ intoxicated rats: an in-vivo study. Fitotherapia, 1993; LXIV: 539-544.
- 270 12) Malloy HT and Evelyn KA. The determination of bilirubin with the photoelectric
271 colorimeter. J of Biol Chem, 1937; 19: 481-490.
- 272 13) Kakkar P, Das B, Viswanathan PN. A modified spectrophotometric assay of
273 superoxide dismutase. Indian J Biophys. 1984;21:130–132.
- 274 14) . Sinha AK. Colorimetric assay of catalase. Anal Biochem. 1972;47:389–394
- 275 15) Rotruck JT, Pope AL, Ganther HE, Swason AB, Hafeman DG, Hoekstra WG.
276 Selenium: biochemical role as a component of glutathione peroxidase. Science. 1973;
277 179:588–590.
- 278 16) Habig WH, Pabst MJ, Jakpoly WB. Glutathione-s-transferase: The first enzymatic
279 step in mercapturic acid formation. J Biol Chem. 1974; 249:7130–7139.
- 280 17) Sasanka Chakraborty, Asha Naik S, Gali Reddy R. Phenylhydra-zine mediated
281 degradation of Bovine serum albumin and mem-brane proteins of human
282 erythrocytes. Biochem et Biophys Acta 1990; 1028:89-94.
- 283 18) Indu Bala Koul, Aruna Kapil. Evaluation of the liver protective potential of Piperine,
284 an active principle of black and long pep-pers. Planta Med 1993; 59:413-417
- 285 19) Yang ES, Lee JH, Park JW. Ethanol induces peroxynitrite-mediated toxicity through
286 inactivation of NADP⁺-dependent isocitrate dehydrogenase and superoxide
287 dismutase. Biochimie. 2008;90(9):1316–1324. [[PubMed](#)]
- 288 20) Shanmugam KR, Ramakrishna CH, Mallikarjuna K, Reddy KS. Protective effect of
289 ginger against alcohol induced renal damage and antioxidant enzymes in male albino
290 rats. Indian J Exp Biol. 2010;48:143–149.
- 291 21) Dash DK, Yeligar VC, Nayak SS, Ghosh T, Rajalingam D, Sengupta P, et al. et al.
292 Evaluation of hepatoprotective and antioxidant activity of *Ichnocarpus*
293 *frutescens* (Linn.) R.Br. on paracetamol-induced hepatotoxicity in rats. Trop J Pharm
294 Res. 2007;6(3):755–765.

- 295 22) Dinkova-Kostova AT. Protection against cancer by plant phenyl propenoids:
296 induction of mammalian anticarcinogenic enzymes. Mini Rev Med
297 Chem. 2002;2:595–610
- 298 23) Popovic M, Janicijevic-Hudomal S, Kaurinovic B, Rasic J, Trivic S. Effects of
299 various drugs on alcohol-induced oxidative stress in the
300 liver. Molecules. 2008;13:2249–2259
- 301 24) Sakr SA, Mahran HA, Lamfon HA. Protective effect of ginger (*Zingiber officinale*)
302 on adriamycin-induced hepatotoxicity in albino rats. J Med Plant
303 Res. 2011;5(1):133–140.
- 304 25) Garouiel M, Fetoui H, Makni FA, Boudawara T, Zeghal N. Cobalt chloride induces
305 hepatotoxicity in adult rats and their suckling pups. Exp Toxicol Pathol. 2011;63(1–
306 2):9–15.