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ABSTRACT

Aims: This study was conducted to determine the toxicity and tolerance to lead by seedling growth of an important country legume crop cowpea (*Vigna unguiculata* <u>L</u>).

Effects of Lead on Different Seedling Growth

Attributes of Cow Pea (Vigna unguiculata) L.

Study Design: The seedlings of cowpea were grown in sand culture at 0, 20, 40, 60, 80 and 100 ppm of metal ions of lead salt as lead acetate.

Place and Duration of Study: The experimental site is located in the Department of Botany at the Karachi University Campus, Pakistan in 2011.

Methodology: The healthy seeds of *Vigna unguiculata* L, were surface sterilized with 0.2% solution of sodium hypochlorite (NaOCL) for one minute to avoid any fungal contamination. The sand was collected from the construction site of the Karachi University washed 2-3 times with tap water, distilled water and also with 5% HCl to remove any types of impurities from the soil. Seedlings were grown in sand culture at 0, 20, 40, 60, 80 and 100 ppm of metal ions of lead salt as lead acetate. The Hoagland solution was used for the supply of nutrient elements. The experiment was conducted for six weeks. **5** ml of lead concentration were poured weekly and before given concentration of lead the materials of the tray were drained out to avoid any algal contaminations. The experiment was completely randomized and consists of six treatments replicated six times. After six weeks the seedlings were harvested and morphological parameters shoot, root, seedling length (cm), number of leaves and leaf area (sq. cm) was noted. The biomass production such as shoot, root, leaf and total seedling dry weight (g) was also observed along with root / shoot, leaf weight, leaf area ratio and specific leaf area. The seedlings of cow pea were dried in an oven at 80° C for 24 hours until the seedlings were completely oven dried. Leaf area, Root / shoot ratio, leaf weight ratio, specific leaf area, leaf area ratio was determined and a tolerance index was determined. The data obtained was statistically analyzed.

Results: The effects of different concentrations (0, 20, 40, 60, 80, 100 ppm) of lead on seedling growth performance of cow pea (Vigna unguiculata L) as compared to control were observed. Lead treatment in the form of lead acetate at 100 ppm highly affected seedling growth and biomass production of V. unguiculata L. as compared to control Lead treatment at 40 ppm produce significant (P = .05). % reduction in seed germination of V. unguiculata L. as compared to control. Lead treatment at 20 ppm concentration produced significant reduction in shoot length as compared to control. Root growth is an important growth variable and found negatively affected by different concentration of lead treatment. The results also showed that lead treatment in the substrate at same concentration (20 ppm) produced significant effect on root and seedling growth of V. unguiculata L... The treatment of lead at 20 ppm produced significant (p<0.05) on seedling dry weight of V. unguiculata L. as compared to control. 80 ppm concentration of lead treatment was found sufficient to cause significant % reductions in seedling dry weight of V. unguiculata L. as compared with control. The seedlings of V. unquiculata L. were also tested for percentage of tolerance to lead. The results showed that V. unguiculata L. has high tolerance to lead at 20 ppm and lowest at 80 ppm of lead. V. unguiculata seedlings showed highest percentage of tolerance (92.50 %) to lead at 20 ppm. The lowest V. unquiculata seedlings was 64.50 % at 80 ppm of lead, but better tolerance of V. unquiculata L. seedlings by 73.25 % at 60 ppm of lead.

Conclusion: In conclusion, lead was toxic causing significant reduction (P = .05). to the seedling and biomass production of cowpea. Increased in lead concentration deceased the tolerance indices of cow pea seedlings growth.

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1. INTRODUCTION

Key words: Crop, germination, growth, lead, tolerance, toxicity.

Plants are an integral part of life in many indigenous communities and modified the plant cover over wide areas [1-2]. Lead is considered highly toxic to the growth of plants. Lead (Pb) is an environmental pollutant extremely toxic to plants and other living organisms including humans [3]. A high concentrations of Pb eventually may cause cell death [4]. Soil pollution with lead is a problem of concern [5].

The concentrations of heavy metals in the environment have increased at alarmingly level. Metals are toxic to plants and fungi [6]. There are few detail reports on the impacts of metals on seed metabolism, germination, particularly roots and shoots recorded [7]. Lead is a global environmental pollutant that is present in soil, water, air and biota. The increase in concentration of heavy metal decreased plant growth and responsible to death [8]. Lead is naturally occur in substantial quantities in the earth's surface food, water urban soil and air and lead stress causes multiple direct and indirect
 effects on plant growth, metabolism and also alters some physiological processes [9-10]. The metals
 absorbed by plants and prove toxic to plants that can be observed as growth retardation, alteration in
 biochemical processes [11-13].

25 In Pakistan substantial quantities of agricultural chemical are used annually to enhance yield 26 [14]. The ever increase of lead concentration over the wide areas of Karachi and rural areas raising a serious questions as to its effects plant growth. Although the data on the effects of lead on cowpea 27 seems scanty. The response of plant growth to toxic effects of heavy metals has become the subject 28 29 of great interest in the field of ecology. Attention has been given in developed countries, about the 30 effects of metal toxicities on crop growth. Therefore, this study was carried out with the aim to determine the effect of lead on seedling growth of an important country legume crop cowpea (Vigna 31 32 unguiculata L.).

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2. MATERIALS AND METHODS

35 The healthy seeds of Vigna unguiculata L. were obtained from the market and were surface 36 sterilized with 0.2% solution of sodium hypochlorite (NaOCI) for one minute to avoid any fungal contamination. The experimental site is located in the Department of Botany at the Karachi University 37 38 Campus. The sand was collected from the construction site of the Karachi University. The sand was 39 sieved through 2.0 mm sieve and after that it was washed 2-3 times with tap water and later with distilled water. The sand was also washed with 5% HCl to remove any types of impurities. The pots 40 41 with 7.3 cm in diameter and 9.6 cm in depth were filled with sand upto 2/3. All the pots were then placed in trays. Seedlings were grown in sand culture at 20, 40, 60, 80 and 100 ppm of metal ions of 42 43 lead salt as lead acetate. In control, no treatment was given except distilled water. As nutrients elements were absent in sand therefore, Hoagland solution was used for the supply of nutrients. The 44 45 Hoagland solution was applied for 3-4 days. Three uniform size seedlings of cowpea were 46 transplanted in each pot. Initially, 5 ml solution of lead acetate Pb(CH₃COO)₂ were applied. Every week the appearance of seedlings growth was recorded. The irrigation was carried out with the tap 47 water on daily basis. The experiment was conducted for six weeks. The nutrient solution (The 48 49 Hoagland solution) was given after three days time interval for nutrition and proper growth of plant. 5 ml solution of lead concentration were poured weekly and before given concentration of lead the 50 materials of the tray were drained out to avoid any algal contaminations. The experiment was 51 completely randomized and replicated six times. 52

After six weeks, the seedlings were harvested. The shoot, root, seedling length (cm), number of leaves and leaf area, specific leaf was recorded. The biomass production such as shoot, root, leaf and total seedling dry weight (g) was also determined alongwith root / shoot, leaf weight, leaf area ratio and specific leaf area. The seedlings of cowpea were dried in an oven at 80° C for 24 hours until the seedlings were completely oven dried.

Root / shoot ratio, leaf weight ratio, specific leaf area, leaf area ratio were determined by the following
 formulae, respectively.

- 60 Root/ shoot ratio = root dry weight / shoot dry weight
- 61 Leaf weight ratio = leaf dry weight / total plant dry weight
- 62 Specific leaf area = Leaf area / leaf dry weight
- 63 Leaf area ratio = Leaf area / Total plant dry weight

A tolerance index was determined by the following formulae as described by Iqbal and Rahmati [15]:

- 65 Mean root length in metal solution/Mean root length in distilled water X 100
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67 Analysis of variance and Duncan's Multiple Range Test using personal computer software 68 packages SPSS version 14.0 statistically analyzed the data obtained. 69

3. RESULTS

Lead treatment was found toxic to the all seedling growth parameter of *V. unguiculata* L. The seedling growth performance of *V. unguiculata* L. were tested in different concentrations (0, 20, 40, 60, 80 and 100 ppm) of lead as compared to control (Table 1-3, Fig.1). Lead treatment was found highly toxic to all seedling growth parameter of *V. unguiculata* L. at 100 ppm concentration. Lead treatment at 40 ppm produced significant (p<0.05) effects on shoot length of *V. unguiculata* L. as compared to control (Table 1). Lead treatment at 20 ppm affected root growth of *V. unguiculata* L. 77 Lead treatment at 20 ppm did not produced significant effects on seedling length, while increase in lead concentration at 40 ppm significantly decreased seedling length of V. unguiculata L. as 78 compared to control. Lead treatment at 40 ppm concentration produced toxic effects on number of 79 leaves of V. unguiculata L. and highly affected at 100 ppm lead. The results also showed that lead 80 treatment at similar concentration 40 ppm produced significant effect on leaf growth of V. unquiculata 81 82 L.

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	Table 1. Effects of lead on seedling growth of cow pea (V. unquiculata)				
Treatments (ppm)	Shoot length (cm)	Root length (cm)	Seedling length (cm)	No of leaves	Leaf area (sq. cm)
0	48.03±0.28d	22.39±0.28e	70.33 ±0.52d	12.50 ±0.76c	28.43±0.281d
20	47.05±0.26d	21.15±0.13d	68.20 ±0.37d	11.00 ±0.77c	27.40±0.35bd
40	42.73±0.74c	19.50±0.25c	62.23 ±0.99c	9.33 ±0.95ab	25.10±0.58c
60	39.38±1.44b	17.81±0.14b	57.20±1.82b	8.66 ±0.80ab	20.31 ±1.35b
80	32.96±1.38a	16.78±0.53ab	49.75±1.89a	7.83 ±0.87a	17.63 ±0.40a
100	30.71±1.39a	16.60±0.45a	47.31±1.75a	7.66 ±0.42a	16.66 ±0.18a

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86 Lead treatment at all concentration decreased high percentage of seedling dry weight of V. 87 unquiculata L. (Table 2). The treatment of lead at 40 ppm produced significant (p<0.05) on shoot dry weight of V. unguiculata L. as compared to control. However, increase in concentration of lead 88 treatment at 80 -100 ppm further decreased shoot dry weight of V. unguiculata L.as compared with 89 control. Lead treatment at 20 ppm concentration significantly affected root dry weight cowpea. Lead 90 treatment at 20 and 40 ppm concentration was less toxic for decrease in leaf dry weight of cowpea. 91 Lead treatment at 40 ppm concentration showed more decrease in leaf dry weight of cowpea. Lead 92 93 concentration of 20 ppm showed high percentage of decrease in total plant dry weight of V. 94 unquiculata L.

Treatments (ppm)	Shoot dry weight (g)	Root dry weight (g)	Leaf dry weight (g)	Total plant dry weight (g)
0	1.74±0.04b	0.86±0.02d	0.95±0.01c	3.55±0.06e
20	1.56±0.19b	0.84±0.03c	0.83±0.041c	3.24±0.21d
40	1.23±0.18a	0.74±0.02ab	0.77±0.04c	2.70±0.18c
60	1.03±0.07a	0.69±0.04ab	0.67±0.03ab	2.43±0.12bc
80	0.92±0.05a	0.64±0.03a	0.63±0.02a	2.20±0.08ab
100	0.86±0.03a	0.50±0.03a	0.48±0.04a	1.84±0.09a

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98 Lead concentration of 20 ppm showed high percentage of decrease in root / shoot ratio, leaf 99 weight, specific leaf area and leaf area ratio of V. unguiculata L. (Table 3).

Table 3. Effects of lead on root /shoot, leaf weight, specific leaf area and leaf area ratio cow pea (V. unguiculata)				
Treatments (ppm)	Root / shoot ratio	Leaf weight ratio	Specific leaf area	Leaf area ratio
0	0.49±0.01b	0.26±0.01b	27.16± 0.14a	7.27±0.09a
20	0.58±0.09a	0.26±0.02a	29.18±1.08b	7.64±0.64b
40	0.66±0.08a	0.29±0.03a	31.19±1.48bc	8.87±0.62d
60	0.67±0.01a	0.29±0.01a	31.14±1.38b	9.02±0.43c
80	0.70±0.02a	0.28±0.01a	30.62±1.43b	8.78±0.36bc
100	0.58±0.02a	0.26±0.01a	36.61±2.79c	9.30±0.37bc

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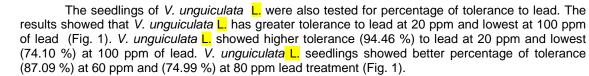
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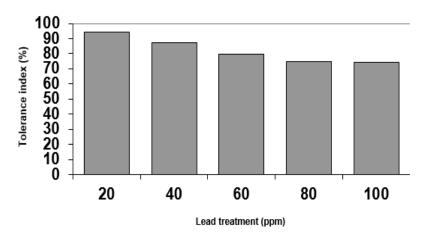


Fig. 1. Percentage tolerance index in seedlings of cow pea (Vigna unguiculata) to different concentration of lead.

4. DISCUSSION

109 The plants under stress condition are likely may be affected by high concentrations of pollutants. The growth in plants or any other living organisms affected directly or indirectly in the 110 presence of heavy metals [16]. Lead is a toxic heavy metal. In present study, the effect different 111 concentrations of lead on root, shoot, seedling length and seedling dry weight of an important bean 112 113 crop cowpea (Vigna unguiculata L.) were recorded. Kasim et al., [17] also showed that lead treatment 114 at 150 mM produced significant physiological, photosynthetics and utltra structural changes in seedlings of Vigna unguiculata L. The seedling growth parameters of cowpea responded differently to 115 lead treatment at higher concentration as compared to control. High percentage of decrease in shoot 116 growth of cowpea at 40-60 ppm lead treatment provided evidence that the treatment of lead in excess 117 was inhibitory to plant growth and development. In present study, a significant decrease in number of 118 119 leaves and leaf area of cowpea at 100 ppm lead treatment was also observed. Excessive amount of 120 toxic element usually caused reduction in plant growth [18]. The permeability of metals can decreased 121 the growth of plants. The reduction in the seedling growth of V. unguiculata L, with the increase in 122 concentration of lead in the immediate environment provides further evidence that the lead in excess 123 may be inhibitory to plant growth and development. Toxic metal ions enter cells by means of the same 124 uptake processes as essential micronutrient metal ions. The amounts of metal absorbed by a plant depend on the concentrations and speciation of the metal in the soil solution. Excessive 125 concentrations of metals result in phytotoxicity [19]. Similarly, drastic decrease in seedling growth of 126 127 Vigna radiata (L.) Wilczek cv. Pusa Baisakhi in the presence of 1.0 mM lead acetate [20]. The root 128 elongation tests have been used to evaluate the damage caused by toxic compounds [21] and many

129 species including cabbage, lettuce, carrot, cucumber, tomato and oats have been recommended for 130 the phytotoxicity test [22]. The roots are normally has the ability to supply water and nutrients to the 131 plants. They are also required to produce hormones, which may regulate the growth and performance 132 of both root and shoot [23]. In present study, the root growth of *V. unguiculata* **L**, was found highly 133 decreased at 80 ppm lead concentration. These findings are also agree with the work of [24] and 134 reported that a low concentration of 1 μ M lead (Pb) reduced root, shoot growth of cowpea (*Vigna* 135 *unguiculata* **L**). The primary site of Pb²⁺ toxicity was the root, caused severe reductions in root 136 growth, loss of apical dominance, the formation of localized swellings behind the root tips.

The significant decrease in seedling dry weight of *V. unguiculata* L. due to metal toxicity of lead was 137 also recorded. The treatment of lead in V. unguiculata L. provided evidence that the trace element in 138 nutrient medium if present in excess may be inhibitory to plant growth and development especially at 139 140 more than 60 ppm. Toxicants accumulate in the plant when soluble forms are present in high quantities. The exact amount of accumulation depends upon the solubility of the pollutants in the soil 141 142 [25]. The biomass production of V. unquiculata L. was initially non significant and decreased and with 143 increasing the lead concentrations upto 100 ppm significantly. Heavy metals at higher concentrations are toxic to plant growth [26] by modifying metabolic processes, oxidative stress in plants [27] and 144 145 excessive accumulation in plant tissue may disturb the physiological processes such as inhibition of root 146 growth recorded in Vigna mungo (L.) [28].

5. CONCLUSION

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According to tolerance test, it could be seen that tolerance to lead was higher at low concentration of lead in the seedlings of *Vigna unguiculata* L. These results showed that the reason of tolerance against heavy metals might be a physiological association of the tolerance mechanism to these metals. The seedling growth of *V. unguiculata* L. showed high percentage of tolerance to lead at 20 ppm concentration. The treatment of 100 ppm concentration of lead produced lowest percentage of tolerance in seedling of cowpea.

It is concluded, that concentration of heavy metals like lead and other metals in the 155 156 environment has been increased due to automobiles, industries, agro chemicals and anthropogenic 157 activities and responsible for limiting the crop yield. Lead is found highly toxic pollutant for plants. The 158 results showed that treatment of different concentrations of lead to the seedlings of V. unguiculata L. responded differently with the increasing contamination exposure as 20, 40, 60, 80 and 100 ppm. The 159 160 response of V. unguiculata L. seedlings in the form of tolerance indices to lead treatment was found suitable pollutant indicator to study the deleterious effects of the lead. The present findings proved the 161 deleterious effects of lead at higher concentration to the seedlings of V. unguiculata L. The 162 information from the present studies would be helpful in understanding the level of lead tolerance in 163 seedlings of V. unguiculata L. while growing in lead polluted areas. Heavy reliance on metals 164 containing agrochemicals such as fungicides, nematicide, and pesticides (Lead arsenate) can be 165 166 discouraged. The continuous release of lead into the immediate environment may endanger the 167 growth performance of other crops. Current research shows that lead treatment at different concentration has produced an important effect on seed germination and seedling growth of V. 168 169 unguiculata.

171 **REFERENCES**

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- Bhatia H, Sharma YP, Manhas RK, Kumar K.. Ethnomedicinal plants used by the villagers of district Udhampur, J&K, India. Journal of Ethnopharmacology, 2014, 151 (2): 1005-1018.
 Eyre SR. The outlook for wild nature. In "Vegetation and Soils". Edward Arnold, The Pitman Press,
 - Eyre SR. The outlook for wild nature. In "Vegetation and Soils". Edward Arnold, The Pitman Press, 2 Ed. U.K. p. 273-277. 1984.
 - Lamhamdi M, Bakrim A, Aarab A, Lafont R, Sayah F. Lead phytotoxicity on V. ungiculata (*Triticum aestivum L.*) seed germination and seedlings growth. Comptes Rendus Biologies, 2011, 334(2): 118-126.
 - WHO. Effects of heavy metals in plants at the cellular levels. In: Schuurmann G (ed). Ecotoxicology, ecological Fundamentals, Chemical Exposures and Biological effects. Pp 587-620. Heidelberg, Wiley. 2001.
- Pirzda H, Ahmad SS, Rashid A, Shah T. multivariate analysis of selected roadside plants
 (Dilbergia sisso and Cannabis sativa) for lead pollution monitoring. Pak. J. Bot. 2009, 41(4): 1729-1736.]
- Crane, S., Barkay, T. and Dighton, J. 2012. The effect of mercury on the establishment of *Pinus rigida* seedlings and the development of their ectomycorrhizal communities. Fungal Ecology, 5(2): 245-251.
- 188 7. Kranner, I. and Colville, L. 2011. Metals and seeds: Biochemical and molecular implications and 189 their significance for seed germination. Experimental and Experimental Botany, 72(1): 93-105.

190	8.	Kumar, P., Sudha, S., Ranjitha, T. and Kumari, B.D. 2011. Effect of chelators and mercury on
191		growth and development of Catharanthus roseus (L). G Don. Journal of Agricultural Technology,
192		7(2): 281-288.
193	9.	Diaz-Aguilar, I., Larque-Saavedra, M.U., Alcantar_Gonzalex, G. and Carrillo-Ganzalez, R. 2001.
194		Alternation of some physiological processes in V. ungiculata by lead additions. Revista
195		International-de-Contamination Ambiental, 17(2): 79-80.
196	10.	Haq, R., Khan, F.F.U. and Haq, E. 2013. Adverse effect of lead acetate on light with protein of
197		Bactrocera cucuitae. Journal of Basic and Applied Sciences, 9: 29-296.
198	11.	Arun KS, Cervantes C., Loza-Tavera H, Avudainayagam S. chromium toxicity in plants. Environ.
199		Int. 2005, 31:739-753.
200	12.	Shafiq M, Iqbal MZ, Athar M. Effect of lead and cadmium on germination and seedling growth of
201		Leucaena leucocephala. Journal of Applied Science and Environmental Management, 12 (3),
202		2008, 61-66.
203	13.	Shafiq M, Iqbal MZ. "Impact of Automobile Pollutants on Plants". ISBN 978-3-8443-8504-5. LAP
204		LAMBERT Academic Publishing GmbH & Co. KG Heinrich-Böcking-Str. 6-8, 66121, Saarbrücken,
205		Germany. 2012, 132 pp.
206	14.	Nuzhat, A., Jameela, A., Munawar, R. and Attaurrehman. 2005. Hydrolysis of a fungicides,
207		Buprimate by indigenous Achromobacter sp. International Journal of Biotechnology, 2(2): 357-363.
208	15.	Iqbal, M.Z. and Rahmati, K. 1992: Tolerance of Albizia lebbeck to Cu and Fe application. Ekologia
209		(CSFR) 11 (4): 427-430.
210	16.	Szollosi, R. 2014. Chapter 3 – Superoxide Dismutase (SOD) and Abiotic Stress Tolerance in
211	-	Plants: An Overview. Oxidative damage to plants. Antioxidant Networks and signaling, pp- 89-129.
212		Academic Press.
213	17.	Kasim, W.A., Abokassem, E.M., Rajab, G.A. and Sewelam, R.N. 2014. Allevation of lead stress
214		toxicity in Vigna unguiculata by salicylic acid. Egyptian Journal Exp. Biology (Bot.) 10(1): 37-49.
215	18.	Kubota, J. and Allaway, W.H. 1972. In "Micro nutrients in Agriculture". Mortvedt. J.J., Giordano,
216		P.M., Lindsay, W.L. (ed.). Soil Science Society of America, Madison. Wis. p. 525-554.
217	19.	Patra, M., Bhowmik, N., Bandopadhyay, B. and Sharma, A. 2004. Comparison of mercury, lead
218		and arsenic with respect to genotoxic effects on plant systems and the development of genetic
219		tolerance. Experimental and Experimental Botany, 52(3): 199-223.
220	20	Singh, R.P., Tripathi, R.D., Dabas, S., Rizvi, S.M.H., Ali, M.B., Sinha, S.K, Gupta, D.K., Mishra, S.
221	_0.	and Rai, U.N. 2003. Effect of lead on growth and nitrate assimilation of <i>Vigna radiata</i> (L.) Wilczek
222		seedlings in a salt affected environment. Chemosphere, 52(7): 1245-1250.
223	21.	U.S.E.P.A. (U.S. Environmental Protection Agency), 1982. Seed Germination/Root Elongation
224		Toxicity Test EG-12.
225	22	F.D.A. (Food and Drug Administration), 1982. Environmental Assessment of Technical Assistance
226		Document 4.06. The Center of Veterinary Medicine, U.S. Department of Health and Human
227		Services, Washington, D.C.
228	23	Blackman, P.G. and Davies, W.J. 1985. Root to shoot connection in maize plants and the effects
229	20.	of soil drying. J. Exp. Bot. 36: 39-48.
230	24	Kopittke, P.M. Asher, C.J., Kopittke, R.A. and Menzies, N.W. 2007. Toxic effects of Pb ²⁺ on growth
230	21.	of cowpea (Vigna unguiculata). Environmental Pollution, 150(2):280-287.
231	25	Treshow, M. 2010. Terrestrial Plants and Plant Communities. Chapter 10. Department of "Biology,
232	20.	University of Utah, Salt Lake City, Utah 84112, U.S.A. pp 225-236.
233	26	Konate, A., He, X., Rui, Y. and Zhang, Z. 2017. Magnetite (Fe_3O_4) nanoparticle alleviate growth
235	20.	inhibition and oxidative stress caused by heavy metals in young seedlings of cucumber (<i>Cucumis</i>
236		sativus L.). <i>ITM Web Conferences</i> , 12, 03034. 1-10.
230	27	Bharwana, S,A., Ali, S., Farooq, M.A., Iqbal, N., Abbas, F. and Ahmed, MSA. 2013.
	27.	
238		Alleviation of lead toxicity by Silicon is related to elevated photosynthesis, antioxidant
239		enzymes suppressed lead uptake and oxidative stress in cotton. J Bioremed Biodeg
240		4:187. doi: 10.4172/2155-6199. 100018.
241	28.	Hussain, K., Sahadevan, K.K. and Salim, N. 2010. Bioaccumulation and release of mercury in
242		Vigna mungo (L.) Hepper seedlings. Journal of Stress Physiology & Biochemistry, 6(3): 56-63.
243		