

# Antifungal Potentials of *Acacia nilotica*, *Ziziphus jujube* Linn and *Lawsonia Inermis*

## ABSTRACT

**Background:** The increasing emergence of resistance to conventional antimicrobial drugs and the complicity of their usage is a serious challenge in Nigeria. In our previous report, it was demonstrated that methanol leave extracts of *Acacia nilotica*, *Ziziphus jujube* Linn and *Lawsonia inermis* exhibited antibacterial activity against *Escherichia coli*, *Pseudomonas flourecense*, *Streptococcus* and *Staphylococcus aureus*.

**Methodology:** In this study, Agar Well Diffusion Method was employed to assess the antifungal potency of these plant extracts and were tested against *Aspergillus flavus*, *Trichophyton rubrum* and *Candida albicans*.

**Results:** Exclusive of *L. inermis* extract against *T. rubrum* at 100 mg/ml (zone of inhibition 34.33±1.89 mm). 100 mg/mL of all the extracts investigated have significantly lower (P<0.05) antifungal activity when compared to standard antifungal drug (Nystatin, 100 mg/ml). The activity of *L. inermis* against *A. flavus* was comparatively similar (P>0.05) to the control drug, but significantly higher (P<0.05) against both *T. rubrum* and *C. albicans* at 150 mg/ml. Conversely, the antifungal activity of *A. nilotica* extract against *T. rubrum* and *C. albicans* significantly surpass (P<0.05) that of the control drug, while *Z. jujube* Linn extract activity against *C. albicans* was comparatively similar (P>0.05) to it, but significantly higher (P<0.05) against *T. rubrum*. A dose dependent antifungal activity of the plants were observed, and *L. inermis* extract was the most potent antifungal agent with an MIC and MCF values of 5 mg/ml. **Conclusion:** This study reveals that *L. inermis* leaves extract could be used as a sources of potential antifungal agents.

**Key words:** Antifungal, *Acacia Nilotica*, *Ziziphus Jujube* Linn and *Lawsonia Inermis*.

## 1.0 INTRODUCTION

The leaves, roots and stem back of *Lawsonia inermis* (henna), *Ziziphus jujube* Linn and *Acacia Nilotica* are traditionally used for the management of bacterial and fungal infections [1]. Recent study on quantitative phytochemical analysis of crude methanol leave extracts of these plants revealed the presence of glycoside, tannins, phenols saponins and flavonoids

**Comment [P1]:** 100 mg/ml mic as a control is too high, what is recommended mic of Nystatin in vitro and stick to that.

**Comment [P2]:** I wonder why you are silent on your mic results, rather than loud on 100 and 150 mg/ml results?

31 [2]. The antibacterial potency of these plant extracts against *Escherichia coli*, *Pseudomonas*  
32 *fluorecense*, *Streptococcus* and *Staphylococcus aureus* was efficient [2]. In the present  
33 study, the antifungal property of the individual plant extracts was tested against *Aspergillus*  
34 *flavus*, *Trichophyton rubrum* and *Candida albicans*.

## 35 2.0 MATERIALS AND METHODS

### 36 2.1 Collection of Plant Material

37 Fresh leaves of *A. nilotica*, *Z. jujube* Linn and *L. inermis* were collected from Achida, Wurno  
38 Local Government Area of Sokoto State. The samples were thoroughly washed with distilled  
39 water, then air-dried under shade

### 40 2.2 Preparation of Plant Material

41 The plant leaf samples were pulverized to powder using pestle and mortar. The pulverized  
42 plant materials were mixed with 95 % methanol. The mixture was kept at room temperature  
43 for 72 hours and was then filtered with a Whatman filter paper No.1 of pore size 11 µm. The  
44 filtrates were evaporated at 45°C using rotary evaporator. Then the methanol extracted  
45 material was dissolved in distilled water and the solution was used for antimicrobial studies

### 46 2.3 Antifungal Screening Using Agar Well Diffusion Method

47 Clinical isolates of *A. flavus*, *T. rubrum* and *C. albicans* were collected from Microbiology  
48 Unit of Specialist Hospital Sokoto, Nigeria. The isolates were subjected to antifungal studies  
49 by agar well diffusion method. Sabouraud dextrose agar plates were inoculated with fungal  
50 culture (10 days old) by point inoculation. Standard solution (50, 100 and 150 mg/ml) of the  
51 extracts were added onto test organism-seeded plates. The plate containing distilled water  
52 (100 ml) was used as positive control while Nystatin (a standard fungicide) (100 mg/ml) was  
53 used as Negative control. Antifungal activity was determined at 28 °C in 7 days incubation.  
54 The zones of inhibitions (mm) were defined by area of complete visible growth inhibition [3].

### 55 2.4 Determination of Minimum Inhibitory Concentration (MIC)

56 Minimum inhibitory concentration (MIC) was determined for *A. nilotica*, *Z. jujube* Linn, and  
57 *L. inermis* against fungal species using Broth Dilution Method. A stock suspension of each  
58 organism was adjusted to  $1.5 \times 10^5$  spores/ml in sabouraud dextrose broth. Test tubes  
59 containing only the media were used as negative control, while those containing only  
60 sabouraud dextrose broth and fungi inoculums served as positive control. Visible turbidity  
61 were determined after incubation at room temperature for 72 hours and 37°C for 24 hours for

Comment [P3]: How possible

62 moulds and yeast (*C. albicans*) respectively. The MIC values were extrapolated from the  
63 lowest concentration of extract that inhibited the visible growth of the tested organism [3].

## 64 2.5 Determination of Minimum Fungicidal Concentration (MFC)

65 In order to determine minimum fungicidal concentration (MFC), plates with no visible  
66 growth in the MIC assay were further subcultured in fresh sterile Sabouraud dextrose agar  
67 plates. The plates were incubated at room temperature until growth was detected in the  
68 growth control subculture. The MFC was then taken as the lowest concentration or highest  
69 dilution of the samples that did not show any visible growth [3].

## 70 2.6 DATA ANALYSES

71 Results were expressed as mean  $\pm$  standard deviation and presented in tabular form. Data was  
72 analysed using In Stat Software package 3.0 version; San Diego USA. Differences between the  
73 means were established by One way ANOVA followed by Duncan's, multiple comparison test.  
74 Statistical significance was set at  $p < 0.05$

## 75 3.0 RESULTS

76 The antifungal activity of the methanol leaf extracts of *A. nilotica*, *L. inermis* and *Z. jujube*  
77 Linn against *A. flavus*, *T. rubrum* and *C. albicans* are presented in Table 1. At 50 and 100  
78 mg/ml, the antifungal **patency** of the extracts were significantly lower ( $P < 0.05$ ) than that of  
79 standard antifungal agent, Nystatin 100 mg/ml), except for *L. inermis* against *T. rubrum* at  
80 100mg/ml with mean zone of inhibition value of  $34.33 \pm 1.89$  mm. However, concentration of  
81 150 mg/ml, the effect *L. inermis* extract against *A. flavus* was comparatively similar ( $P > 0.05$ )  
82 to that of the control drug. However, similar concentration of *L. inermis* extract exhibited  
83 significantly higher activity ( $P < 0.05$ ) against both *T. rubrum* and *C. albicans* than Nystatin.  
84 On the other hand, the antifungal activity of *A. nilotica* extract against *T. rubrum* and *C.*  
85 *albicans* significantly surpass ( $P < 0.05$ ) that of the control drug, while *Z. jujube* Linn extract  
86 activity against *C. albicans* was comparatively similar ( $P > 0.05$ ) to that of Nystatin, but  
87 significantly higher ( $P < 0.05$ ) against *T. rubrum*.

88 **Table 1:** Antifungal Activities of *A. nilotica*, *L. inermis* and *Z. jujube* Linn Methanol Leaf  
89 Extracts (50, 100 and 150 mg/ml) and Nystatin (100 mg/ml)

Comment [P4]: What does this mean



|                       |                    |   |   |   |   |   |   |   |   |   |   |    |
|-----------------------|--------------------|---|---|---|---|---|---|---|---|---|---|----|
| <i>Z. jujube</i> Linn | <i>T. rubrum</i>   | - | - | - | - | - | - | - | + | + | + | 20 |
|                       | <i>C. ablicans</i> | - | - | - | - | + | + | + | + | + | + | 35 |
|                       | <i>A. flavus</i>   | - | - | - | - | - | - | - | - | + | + | 15 |
| Nystatin              | <i>T. rubrum</i>   | - | - | - | - | - | - | - | - | + | + | 15 |
|                       | <i>C. ablicans</i> | - | - | - | - | - | - | - | - | + | + | 15 |

102 Key: (-) indicate no visible growth of the organisms and (+) indicate visible growth of the organisms.

103

104 The extract of *L. inermis* exhibited least MFC at 5 mg/ml on the three fungi species. The  
105 potency of *A. nilotica* was comparable to that of the standard antifungal agent, lower than that  
106 of *L. inermis* but higher than *Z. jujube* Linn extract (Table 3). Also, the least antifungal  
107 activity with MFC value of 35 mg/ml was observed in the activity of *Z. jujube* Linn against  
108 *C. ablicans*.

109 **Table 3:** Minimum Fungicidal Concentration (MFC) of Methanol Leaf Extracts of *A.*  
110 *nilotica*, *L. inermis* and *Z. jujube* Linn

| Samples               | Fungal spp.        | Concentration of Extract (mg/ml) |    |    |    |    |    |    |    |    |   | MFC |
|-----------------------|--------------------|----------------------------------|----|----|----|----|----|----|----|----|---|-----|
|                       |                    | 50                               | 45 | 40 | 35 | 30 | 25 | 20 | 15 | 10 | 5 |     |
| <i>A. nilotica</i>    | <i>A. flavus</i>   | -                                | -  | -  | -  | -  | -  | -  | -  | +  | + | 15  |
|                       | <i>T. rubrum</i>   | -                                | -  | -  | -  | -  | -  | -  | +  | +  | + | 20  |
|                       | <i>C. ablicans</i> | -                                | -  | -  | -  | -  | -  | -  | -  | -  | + | 10  |
| <i>L. inermis</i>     | <i>A. flavus</i>   | -                                | -  | -  | -  | -  | -  | -  | -  | -  | - | 5   |
|                       | <i>T. rubrum</i>   | -                                | -  | -  | -  | -  | -  | -  | -  | -  | - | 5   |
|                       | <i>C. ablicans</i> | -                                | -  | -  | -  | -  | -  | -  | -  | -  | - | 5   |
| <i>Z. jujube</i> Linn | <i>A. flavus</i>   | -                                | -  | -  | -  | -  | -  | +  | +  | +  | + | 25  |
|                       | <i>T. rubrum</i>   | -                                | -  | -  | -  | -  | -  | +  | +  | +  | + | 25  |
|                       | <i>C. ablicans</i> | -                                | -  | -  | -  | +  | +  | +  | +  | +  | + | 35  |
| Nystatin              | <i>A. flavus</i>   | -                                | -  | -  | -  | -  | -  | -  | -  | +  | + | 15  |
|                       | <i>T. rubrum</i>   | -                                | -  | -  | -  | -  | -  | -  | -  | +  | + | 15  |
|                       | <i>C. ablicans</i> | -                                | -  | -  | -  | -  | -  | -  | -  | +  | + | 15  |

111 Key: (-) indicate no visible growth of the organisms and (+) indicate visible growth of the organisms.

112

113

#### 4.0 DISCUSSION

The increasing resistance of microorganisms to conventional antimicrobial drug, has lead to a increase/wider usage of plant, and other folk medicines as alternative ailment [4, 5, 6, 7]. As previously reported, the methanol leaves extract of *A. nilotica*, *Z. jujube* Linn and *L. inermis* are rich in phytochemicals and showed antibacterial potency [2]. In this study, the antifungal activities of the plant extracts were assessed. At 100 mg/mL the antifungal patency of the extracts was below that of standard antifungal agent (Nystatin), exclusive of *L. inermis* against *T. rubrum*. At a concentration of 150 mg/ml, the effect *L. inermis* extract was significantly increases against the fungi species beyond that of Nystatin. More so, the activity of *A. nilotica* extract against *T. rubrum* and *C. albicans* is higher than that of the control drug, while that *Z. jujube* Linn extract against *C. albicans* was comparable to that of Nystatin, but higher against *T. rubrum*.

Comment [P5]: led

The outcome of this study conforms to earlier report suggesting a dose dependent antifungal activity of *A. nilotica* against *A. flavus* [8, 9, 10] and *C. albicans* [11, 12]. The results of this study also conform to the findings of Yigit [13] which reported a strong antifungal activity of *L. inermis* against fungal isolates. This antifungal activity is attributed to the rich naphthoquinone content of it leaves extract [14, 15, 16]. The methanol leaves of *Z. jujube* Linn exhibited the lowest antifungal activity in comparison to the other samples investigated. These findings conform to the earlier reports of Manoj *et al.*, [17] which revealed that the plant leaves extract had no effect against both *A. niger* and *C. albicans*. Whereas a report of Elaloui *et al.* [18] indicated promising antifungal effect of the plants leaves extract against *F. culmorum*, *F. solani* and *B. cinerea*. Similarly Naz *et al.* [19] reported a moderate activity by methanol leave extracts of *Z. Jujuba mill* against *G.lucidum* but lower activity against *A. flavus*, *A. niger* and *A. alternate*. Abalaka *et al.* [20] reported the resistance of *A. niger* and *C. albicans* to ethanolic extracts of two *Ziziphus* species.

Comment [P6]: meaning

#### 5.0 CONCLUSION

The findings of this study suggest that the leave extracts of *L. inermis*, *Z. jujube* Linn and *A. nilotica* showed antifungal activities against *A. flavus*, *T. rubrum* and *C. ablicans*. Thus, these plants could served as potential sources of antifungal agents.

#### REFERENCE

1. Sameera NS, Mandakini BP. Investigation into the antimicrobial activity of *Ziziphus mauritina* Lam. And *Ziziphus xylopyra* (Retz) wild. Nagpur, Mahashtra, India, 2015; 5:188-297.

2. Abubakar AL, Dandare A, WASagu RSU, Yerima M, Abubakar HI. Antimicrobial Activities of *Acacia Nilotica*, *Ziziphus Jujube* Linn and *Lawsonia Inermis*. Accepted for publication, Nigerian Journal of Basic and Applied Sciences. 2018; 27(1).
3. Magaldia S, Mata-Essayaga S, Hartung de Capriles C, Perez C, Colella MT, Carolina O, Yudith O. Well diffusion for antifungal susceptibility testing International Journal of Infectious Diseases. 2004; 8 (1):39-45
4. Sharmin T, Chowdhury SR, Mian MY, Hoque M, Sumsujjaman M, Nahar F. Evaluation of antimicrobial activities of some Bangladeshi medicinal plants. World Journal of Pharmaceutical Sciences. 2014; 2 (2): 170-175.
5. Ngoci NS, Ramadhan M, Ngari MS, Leonard OP. Screening for antimicrobial activity of *Cissampelos pareira* L. methanol root extract. European Journal of Medicinal Plants, 2014; 4(1): 45-51.
6. Dhama KR Tiwari S Chakraborty *et al.*, "Global warming and emerging infectious diseases of animals and humans:current scenario, challenges, solutions and future perspectives—a review," International Journal of Current Research.2013;5(7):1942–1958
7. Malik SA, Kumar AK Verma *et al.*, "Incidence and drug resistance pattern of colibacillosis in cattle and buffalo calves in Northwest part of Utter Pradesh in India," Journal of Animal Health and Production, 2013; 1(2):15–19.
8. Gurjinder K, Sharma AK, Arun K. Antimicrobial activity of *Acacia Nilotica* against Various Clinical Isolates. Applied Botany. 2016; 97: 42260-42261
9. Rwarinda UA. Efficacy of *Acacia nilotica* Extracts towards Microbicidal Activity against Pathogens. International Journal of Current Microbiology and Applied Science. 2015; 4(10): 33-42
10. Sharma AK, Kumar A, Yadav SK, Rahal A. Studies on Antimicrobial and Immunomodulatory Effects of Hot Aqueous Extract of *Acacia nilotica* L. Leaves against Common Veterinary Pathogens. Veterinary Medicine International. 2014: 9
11. Abd-Ulgadir KS, El-Kamali HH. Antimicrobial Activity of *Acacia nilotica* ssp. *nilotica* against Some Causative Agents of Urogenital Infections. Annual Research & Review in Biology. 2017 19(5): 1-14
12. Ali Atif, Akhtar Naveed, Khan Barkat Ali, Khan Muhammad Shoaib, Rasul Akhtar, Shahiq-UZ-Zaman, Khalid Nayab, Waseem Khalid, Mahmood Tariq and Ali Liaqat *Acacia nilotica*: A plant of multipurpose medicinal uses. Journal of Medicinal Plants Research. 2012; 6(9): 1492-1496.
13. Yiğit D. Antifungal Activity of *Lawsonia inermis* L. (Henna) Against Clinical Candida Isolates. Journal of Science and Technology. 2017; 10(2): 196-202

- 183      **14.** Arun P, Purushotham KG, Jayarani J, Kumari V. In vitro antibacterial activity and  
184      flavonoid contents of *Lawsonia inermis* (henna). International Journal of Pharm Tech  
185      Research. 2010; 2, 1178-1181.
- 186      **15.** Babu PD Subhasree RS Anticandidal activity of *Lawsonia inermis*. Academic Journal  
187      of Plant Sciences. 2009; 2, 231-232.
- 188      **16.** Abdulmoneim MA Evaluation of *Lawsonia inermis* Linn. (Sudanese Henna) leaf  
189      extract as an antimicrobial agent. Research Journal of Biological Sciences. 2007; 2:  
190      417-423.
- 191      **17.** Manoj G, Badri PN, Dinakar S. Review on ethnomedicinal uses, pharmacological  
192      activity and phytochemical constituents of *Ziziphus mauritiana* (*Z. jujuba* Lam., non  
193      Mill). Spatula DD. 2012; 2(2):107-116
- 194      **18.** Elaloui M, Ennajah A, Ghazghazi H, Youssef IB, Othman NB, Hajlaoui M Rabeh,  
195      Khouja A, Laamouri A. Quantification of total phenols, flavonoides and tannins from  
196      *Ziziphus jujuba* (mill.) and *Ziziphus lotus* (L.) (Desf). Leaf extracts and their effects on  
197      antioxidant and antibacterial activities. International Journal of Secondary Metabolite,  
198      2017; 4(1): 18-26
- 199      **19.** Naz S, Sultana B, Shahid M, Khalil-ur-R.. Alteration in antioxidant and antimicrobial  
200      attributes of leaves of *Zizyphus* species in response to maturation. Journal of  
201      Medicinal Plants Research. 2013; 7(2): 61-70
- 202 **20.** Abalaka ME, Daniyan SY. Mann A. Evaluation of the antimicrobial activities of two  
203      *Ziziphus* species (*Ziziphus mauritiana* L. and *Ziziphus spinachristi* L.) on some microbial  
204      pathogens. African Journal of Pharmacy and Pharmacology, 2010; 4(4): 135-139