Antifungal Potentials of Acacia nilotica, Ziziphus jujube Linn and Lawsonia Inermis 3

ABSTRACT 4

Background: The increasing emergence of resistance to conventional antimicrobial drugs 5 and the complicity of their usage is a serious challenge in Nigeria. In our previous report, it 6 7 was demonstrated that methanol leave extracts of Acacia nilotica, Ziziphus jujube Linn and 8 Lawsonia inermis exhibited antibacterial activity against Escherichia coli, Pseudomonas 9 flourecense, Streptococcus and Staphylococcus aureus.

10 Methodology: In this study, Agar Well Diffusion Method was employed to assess the antifungal potency of these plant extracts and were tested against Aspergillus flavus, 11 12 Trichophyton rubrum and Candida albicans.

13 **Results**: Exclusive of *L. inermis* extract against *T. rubrum* at 100 mg/ml (zone of inhibition 34.33±1.89 mm). 100 mg/mL of all the extracts investigated have significantly lower 14 15 (P<0.05) antifungal activity when compared to standard antifungal drug (Nystatin, 100 16 mg/ml). The activity of L. inermis against A. flavus was comparatively similar (P>0.05) to the 17 control drug, but significantly higher (P<0.05) against both T. rubrum and C. albicans at 150 18 mg/ml. Conversely, the antifungal activity of A. nilotica extract against T. rubrum and C. albicans significantly surpass (P<0.05) that of the control drug, while Z. jujube Linn extract 19 20 activity against C. albicans was comparatively similar (P>0.05) to it, but significantly higher 21 (P < 0.05) against T. rubrum. A dose dependent antifungal activity of the plants were 22 observed, and L. inermis extract was the most potent antifungal agent with an MIC and MCF 23 values of 5 mg/ml. **Conclusion**: This study reveals that *L. inermis* leaves extract could be 24 used as a sources of potential antifungal agents.

25 Key words: Antifungal, Acacia Nilotica, Ziziphus Jujube Linn and Lawsonia Inermis.

26 **1.0 INTRODUCTION**

The leaves, roots and stem back of Lawsonia inermis (henna), Ziziphus jujube Linn and 27 28 Acacia Nilotica are traditionally used for the management of bacterial and fungal infections 29 [1]. Recent study on quantitative phytochemical analysis of crude methanol leave extracts 30 of these plants revealed the presence of glycoside, tannins, phenols saponins and flavonoids

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31 [2]. The antibacterial potency of these plant extracts against *Escherichia coli*, *Pseudomonas*

32 flourecense, Streptococcus and Staphylococcus aureus was efficient [2]. In the present

- 33 study, the antifungal property of the individual plant extracts was tested against Aspergillus
- 34 *flavus*, *Trichophyton rubrum* and *Candida albicans*.

35 2.0 MATERIALS AND METHODS

36 **2.1 Collection of Plant Material**

Fresh leaves of *A. nilotica*, *Z. jujube* Linn and *L. inermis* were collected from Achida, Wurno
Local Government Area of Sokoto State. The samples were thoroughly washed with distilled
water, then air-dried under shade

40 2.2 Preparation of Plant Material

The plant leaf samples were pulverized to powder using pestle and mortar. The pulverized plant materials were mixed with 95 % methanol. The mixture was kept at room temperature for 72 hours and was then filtered with a Whatman filter paper No.1 of pore size 11 μ m. The filtrates were evaporated at 45°C using rotary evaporator. Then the methanol extracted material was dissolved in distilled water and the solution was used for antimicrobial studies

46 2.3 Antifungal Screening Using Agar Well Diffusion Method

47 Clinical isolates of A. flavus, T. rubrum and C. albicans were collected from Microbiology 48 Unit of Specialist Hospital Sokoto, Nigeria. The isolates were subjected to antifungal studies 49 by agar well diffusion method. Sabouraud dextrose agar plates were inoculated with fungal culture (10 days old) by point inoculation. Standard solution (50, 100 and 150 mg/ml) of the 50 51 extracts were added onto test organism-seeded plates. The plate containing distilled water 52 (100 ml) was used as positive control while Nystatin (a standard fungicide) (100 mg/ml) was 53 used as Negative control. Antifungal activity was determined at 28 °C in 7 days incubation. 54 The zones of inhibitions (mm) were defined by area of complete visible growth inhibition [3].

55 **2.4 Determination of Minimum Inhibitory Concentration (MIC)**

56 Minimum inhibitory concentration (MIC) was determined for *A. nilotica*, *Z. jujube* Linn, and 57 *L. inermis* against fungal species using Broth Dilution Method. A stock suspension of each 58 organism was adjusted to 1.5×10^5 spores/ml in sabouraud dextrose broth. Test tubes 59 containing only the media were used as negative control, while those containing only 50 sabouraud dextrose broth and fungi inoculums served as positive control. Visible turbidity 51 were determined after incubation at room temperature for 72 hours and 37^{0} C for 24 hours for 62 moulds and yeast (C. albicans) respectively. The MIC values were extrapolated from the

63 lowest concentration of extract that inhibited the visible growth of the tested organism [3].

64 **2.5 Determination of Minimum Fungicidal Concentration (MFC)**

In order to determine minimum fungicidal concentration (MFC), plates with no visible growth in the MIC assay were further subcultured in fresh sterile Sabouraud dextrose agar plates. The plates were incubated at room temperature until growth was detected in the growth control subculture. The MFC was then taken as the lowest concentration or highest dilution of the samples that did not show any visible growth [3].

70 2.6 DATA ANALYSES

71 Results were expressed as mean \pm standard deviation and presented in tabular form. Data was 72 analysed using In Stat Software package 3.0 version; San Diego USA. Differences between the 73 means were established by One way ANOVA followed by Duncan's, multiple comparison test. 74 Statistical significance was set at p< 0.05

3.0 RESULTS

The antifungal activity of the methanol leaf extracts of A. nilotica, L. inermis and Z. jujube 76 77 Linn against A. flavus, T. rubrum and C. albicans are presented in Table 1. At 50 and 100 78 mg/ml, the antifungal patency of the extracts were significantly lower (P < 0.05) than that of 79 standard antifungal agent, Nystatin 100 mg/ml), except for L. inermis against T. rubrum at 80 100mg/ml with mean zone of inhibition value of 34.33±1.89 mm. However, concentration of 150 mg/ml, the effect L. inermis extract against A. flavus was comparatively similar (P>0.05) 81 82 to that of the control drug. However, similar concentration of *L. inermis* extract exhibited significantly higher activity (P < 0.05) against both T. rubrum and C. albicans than Nystatin. 83 84 On the other hand, the antifungal activity of A. nilotica extract against T. rubrum and C. 85 albicans significantly surpass (P < 0.05) that of the control drug, while Z. jujube Linn extract 86 activity against C. albicans was comparatively similar (P>0.05) to that of Nystatin, but 87 significantly higher (P<0.05) against *T. rubrum*.

Table 1: Antifungal Activities of *A. nilotica, L. inermis* and *Z. jujube* Linn Methanol Leaf

Extracts (50, 100 and 150 mg/ml) and Nystatin (100 mg/ml)

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Extract Conc.	P 1	Zone of Inhibition (mm)										
	Fungal <i>spp</i> .	A. nilotica	L. inermis	Z. jujube Linn	Nystatin	Distilled Water						
	A. flavus	11.00±1.00 ^a	11.67 ± 2.89^{a}	11.67 ± 1.73^{a}	53.00±2.65 ^b	0.00						
50mg/ml	T. rubrum	$9.00{\pm}2.44^{c}$	17.33 ± 1.89^{d}	$7.67 \pm 0.45^{\circ}$	30.00±2.33 ^e	0.00						
	C. albicans	11.45±0.89 ^a	11.00 ± 0.45^{a}	14.33 ± 0.67^{f}	31.00±2.89 ^e	0.00						
	A. flavus	22.33+2.50 ^a	41.67±2.90 ^b	11.00±1.73°	53.00±2.65 ^d	0.00						
100mg/ml	T. rubrum	28.00 ± 0.44^{e}	$34.33{\pm}1.89^{f}$	27.67±0.33 ^e	30.00±2.33 ^g	0.00						
	C. albicans	$24.67{\pm}0.33^h$	17.67 ± 1.45^{i}	20.33±2.33 ^a	31.00±2.89 ^g	0.00						
	A. flavus	28.00 ± 2.00^{a}	50.67±1.15 ^b	26.67±2.65°	53.00±2.65 ^b	0.00						
150mg/ml	T. rubrum	36.00 ± 0.44^{d}	44.67±1.89 ^e	$37.33{\pm}0.33^{\rm f}$	30.00±2.33 ^g	0.00						
	C. albicans	35.30 ± 0.33^{d}	$38.33{\pm}1.45^{\rm f}$	$30.33{\pm}0.33^{g}$	30.33 ± 2.89^{g}	0.00						

92 Values are mean inhibition zones (mm) \pm S.D of three replicate experiment. Mean value having 93 different superscript letters along the rows are significantly different (*P*<0.05)

Table 2 shows the minimum inhibitory concentration (MIC) of methanol leaf extracts of A.

95 nilotica, L. inermis and Z. jujube Linn against A. flavus, T. rubrum and C. albicans. The

96 extract of *L. inermis* was most effective against the three fungi species with an MIC value of

97 5 mg/ml. the least antifungal potency was observed in Z. jujube Linn with visible C. ablicans

- 98 growth at MIC value 30mg/ml.
- 99

100 Table 2: Minimum Inhibitory Concentration (MIC) of Methanol Leaf Extracts of A. nilotica,

101 *L. inermis* and *Z. jujube* Linn

			(Concer	ntratior	n of Ex	tract (r	ng/ml)				
Samples	Fungal spp.	50	45	40	35	30	25	20	15	10	5	MIC
	A. flavus	-	-	-	-	-	-	-	-	-	+	10
A. nilotica	T. rubrum	-	-	-	-	-	-	-	-	-	+	10
	C. ablicans	-	-	-	-	-	-	-	-	-	+	10
	A. flavus	-	-	-	-	-	-	-	-	-	-	5
L. inermis	T. rubrum	-	-	-	-	-	-	-	-	-	-	5
	C.ablicans	-	-	-	-	-	-	-	-	-	-	5
	A. flavus			-	-			-	-	+	+	15

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<i>Z. jujube</i> Linn	T. rubrum	-	-	-	-	-	-	-	+	+	+	20
	C. ablicans	-	-	-	-	+	+	+	+	+	+	35
	A. flavus	-	-	-	-			-	-	+	+	15
Nystatin	T. rubrum	-	-	-	-	-	-	-	-	+	+	15
	C. ablicans	-	-	-	-	-	-	-	-	+	+	15

102 Key: (-) indicate no visible growth of the organisms and (+) indicate visible growth of the organisms.
103

104	The extract of L. inermis exhibited least MFC at 5 mg/ml on the three fungi species. The
105	potency of A. nilotica was comparable to that of the standard antifungal agent, lower than that
106	of L. inermis but higher than Z. jujube Linn extract (Table 3). Also, the least antifungal
107	activity with MFC value of 35 mg/ml was observed in the activity of Z. jujube Linn against
108	C. ablicans.

Table 3: Minimum Fungicidal Concentration (MFC) of Methanol Leaf Extracts of *A*. *nilotica, L. inermis* and *Z. jujube* Linn

			Concentration of Extract (mg/ml)										
Samples	Fungal spp.	50	45	40	35	30	25	20	15	10	5	MFC	
	A. flavus	-	-	-	-	-	-	-	-	+	+	15	
A.nilotica	T. rubrum	-	-	-	-	-	-	-	+	+	+	20	
	C. ablicans	-	-	-	-	-	-	-	-	-	+	10	
	A. flavus	-	-	-	-	-	-	-	-		-	5	
L.inermis	T. rubrum	-	-	-	-	-	-	-	-	-	-	5	
	C. ablicans	-	-	-	-	-	-	-	-	-	-	5	
	A. flavus		-	-	-	-	-	+	+	+	+	25	
<i>Z.jujube</i> Linn	T. rubrum	-	-	-	-	-	-	+	+	+	+	25	
	C. ablicans	-	-	-	-	+	+	+	+	+	+	35	
<u> </u>	A. flavus		-	-				-	-	+	+	15	
Nystatin	T. rubrum	-	-	-	-	-	-	-	-	+	+	15	
	C. ablicans	-	-	-	-	-	-	-	-	+	+	15	

111 Key: (-) indicate no visible growth of the organisms and (+) indicate visible growth of the organisms.
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114 **4.0 DISCUSSION**

115 The increasing resistance of microorganisms to conventional antimicrobial drug, has lead to a 116 increase/wider usage of plant, and other folk medicines as alternative ailment [4, 5, 6, 7]. As previously reported, the methanol leaves extract of A. nilotica, Z. jujube Linn and L. inermis 117 118 are rich in phytochemicals and showed antibacterial potency [2]. In this study, the antifungal 119 activities of the plant extracts were assessed. At 100 mg/mL the antifungal patency of the 120 extracts was below that of standard antifungal agent (Nystatin), exclusive of L. inermis against T. rubrum. At a concentration of 150 mg/ml, the effect L. inermis extract was 121 122 significantly increases against the fungi species beyond that of Nystatin. More so, the activity 123 of A. nilotica extract against T. rubrum and C. albicans is higher than that of the control 124 drug, while that Z. jujube Linn extract against C. albicans was comparable to that of 125 Nystatin, but higher against T. rubrum.

126 The outcome of this study conforms to earlier report suggesting a dose dependent antifungal 127 activity of A. nilotica against A. flavus [8, 9, 10] and C. albicans [11, 12]. The results of this 128 study also conform to the findings of Yigit [13] which reported a strong antifungal activity 129 of L. inermis against fungal isolates. This antifugal activity is attributed to the rich naphtoquinone content of it leaves extract [14, 15, 16]. The methanol leaves of Z. jujube Linn 130 131 exhibited the lowest antifungal activity in comparison to the other samples investigated. 132 These findings conform to the earlier reports of Manoj *et al.*, [17] which revealed that the 133 plant leaves extract had no effect against both A. niger and C. albicans. Whereas a report of 134 Elaloui et al. [18] indicated promising antifungal effect of the plants leaves extract against F. 135 culmorum, F. solani and B. cinerea. Similarly Naz et al. [19] reported a moderate activity by 136 methanol leave extracts of Z. Jujuba mill against G. lucidum but lower activity against A. 137 flavus, A. niger and A. alternate. Abalaka et al. [20] reported the resistance of A. niger and C. 138 albicans to ethanolic extracts of two Ziziphus species.

139 **5.0 CONCLUSION**

The findings of this study suggest that the leave extracts of *L. inermis, Z. jujube* Linn and *A. nilotica* showed antifungal activities against *A. flavus, T. rubrum* and *C. ablicans*. Thus,
these plants could served as potential sources of antifungal agents.

143 **REFERENCE**

Sameera NS, Mandakini BP. Investigation into the antimicrobial activity of *Ziziphus mauritina* Lam. And *Ziziphus xylopyra* (Retz) wild. Nagpur, Mahashtra, India, 2015;
 5:188-297.

2. Abubakar AL, Dandare A, WAsagu RSU, Yerima M, Abubakar HI. Antimicrobial 147 148 Activities of Acacia Nilotica, Ziziphus Jujube Linn and Lawsonia Inermis. Accepted for publication, Nigerian Journal of Basic and Applied Sciences. 2018; 27(1). 149 3. Magaldia S, Mata-Essayaga S, Hartung de Capriles C, Perez C, Colella MT, Carolina 150 O, Yudith O. Well diffusion for antifungal susceptibility testing International Journal 151 of Infectious Diseases. 2004; 8 (1):39-45 152 153 4. Sharmin T, Chowdhury SR, Mian MY, Hoque M, Sumsujjaman M, Nahar F. Evaluation of antimicrobial activities of some Bangladeshi medicinal plants. World 154 Journal of Pharmaceutical Sciences. 2014; 2 (2): 170-175. 155 156 5. Ngoci NS, Ramadhan M, Ngari MS, Leonard OP. Screening for antimicrobial activity of Cissampelos pareira L. methanol root extract. European Journal of 157 Medicinal Plants. 2014: 4(1): 45-51. 158 6. Dhama KR Tiwari S Chakraborty et al., "Global warming and emerging infectious 159 diseases of animals and humans:current scenario, challenges, solutions and future 160 161 perspectives—a review," International Journal of Current Research.2013;5(7):1942-1958 162 163 7. Malik SA, Kumar AK Verma et al., "Incidence and drug resistance pattern of collibacillosis in cattle and buffalo calves in Northwest part of Utter Pradesh in 164 India," Journal of Animal Health and Production. 2013; 1(2):15–19. 165 166 8. Gurjinder K, Sharma AK, Arun K. Antimicrobial activity of Acacia Nilotica against Various Clinical Isolates. Applied Botany. 2016; 97: 42260-42261 167 9. Rwarinda UA. Efficacy of *Acacia nilotica* Extracts towards Microbicidal Activity 168 against Pathogens. International Journal of Current Microbiology and Applied 169 170 Science. 2015; 4(10): 33-42 10. Sharma AK, Kumar A, Yadav SK, Rahal A. Studies on Antimicrobial and 171 172 Immunomodulatory Effects of Hot Aqueous Extract of Acacia nilotica L. Leaves against Common Veterinary Pathogens. Veterinary Medicine International. 2014: 9 173 174 11. Abd-Ulgadir KS, El-Kamali HH. Antimicrobial Activity of Acacia nilotica ssp. nilotica against Some Causative Agents of Urogenital Infections. Annual Research & 175 176 Review in Biology. 2017 19(5): 1-14 12. Ali Atif, Akhtar Naveed, Khan Barkat Ali, Khan Muhammad Shoaib, Rasul Akhtar, 177 Shahiq-UZ-Zaman, Khalid Nayab, Waseem Khalid, Mahmood Tariq and Ali Liaqat 178 Acacia nilotica: A plant of multipurpose medicinal uses. Journal of Medicinal Plants 179 Research. 2012; 6(9): 1492-1496. 180 13. Yiğit D. Antifungal Activity of Lawsonia inermis L. (Henna) Against Clinical 181 Candida Isolates. Journal of Science and Technology. 1017; 10(2): 196-202 182

183 184 185	14. Arun P, Purushotham KG, Jayarani J, Kumari V. In vitro antibacterial activity and flavonoid contents of <i>Lawsonia inermis</i> (henna). International Journal of Pharm Tech Research. 2010; 2, 1178-1181.
186 187	15. Babu PD Subhasree RS Anticandidal activity of <i>Lawsonia inermis</i> . Academic Journal of Plant Sciences. 2009; 2, 231-232.
188 189 190	16. Abdulmoneim MA Evaluation of <i>Lawsonia inermis</i> Linn. (Sudanese Henna) leaf extract as an antimicrobial agent. Research Journal of Biological Sciences. 2007; 2: 417-423.
191 192 193	17. Manoj G, Badri PN, Dinakar S. Review on ethnomedicinal uses, pharmacological activity and phytochemical constituents of <i>Ziziphus mauritiana</i> (<i>Z. jujuba</i> Lam., non Mill). Spatula DD. 2012; 2(2):107-116
194 195 196 197 198	18. Elaloui M, Ennajah A, Ghazghazi H, Youssef IB, Othman NB, Hajlaoui M Rabeh, Khouja A, Laamouri A. Quantification of total phenols, flavonoides and tannins from Ziziphus jujuba (mill.) and Ziziphus lotus (l.) (Desf). Leaf extracts and their effects on antioxidant and antibacterial activities. International Journal of Secondary Metabolite, 2017; 4(1): 18-26
199 200 201	19. Naz S, Sultana B, Shahid M, Khalil-ur-R Alteration in antioxidant and antimicrobial attributes of leaves of <i>Zizyphus</i> species in response to maturation. Journal of Medicinal Plants Research. 2013; 7(2): 61-70
	abalaka ME, Daniyan SY. Mann A. Evaluation of the antimicrobial activities of two Viziphus species (Ziziphus mauritiana L. and Ziziphus spinachristi L.) on some microbial

204 pathogens. African Journal of Pharmacy and Pharmacology, 2010; 4(4): 135-139